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IN-VITRO EVALUATION OF THE ANTIDIABETIC & CYTOTOXIC ACTIVITIES OF EARTHWORM (LUMBRICUS RUBELLUS) EXTRACTS

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ABSTRACT: This in-vitro antidiabetic and cytotoxic activity of earthworm Lumbricus rubellus extracts were investigated. In the present study, three types of solvent (petroleum ether, 95% ethanol and phosphate buffer) extracts of earthworm paste were prepare and subjected to preliminary screening. The antidiabetic activity was determined in terms of its ability to inhibit the α-amylase activity. Different concentrations ranging from 100-1000µg/mL of earthworm extracts were incubated with enzyme substrate solution and subjected in the determination of absorbance to assess the antidiabetic activity. Also Acarbose was used as the standard inhibitor. The 95% ethanol extracts showed the highest percentage inhibition of α -amylase activity, followed by petroleum ether and phosphate buffer. The IC50 value of 95% ethanol, petroleum ether and phosphate buffer extracts were 242.46µg/mL, 304.15µg/mL and 689.68µg/mL respectively, when compared with Acarbose (IC50 value 193.68µg/mL). The cytotoxic activity was determined towards brine shrimp lethality assay. Different concentrations ranging from 10-100µg/mL of earthworm extracts were subjected to determine the mortality of brine shrimps. Potassium dichromate was used as positive control. Results showed that the petroleum ether extract was potent against the brine shrimp with LC50 values of 7.13µg/mL, followed by 95% ethanol extract (7.28µg/mL) and phosphate buffer extract (73.50µg/mL). While the positive control exhibited LC50 value with 11.96µg/mL. The antidiabetic and cytotoxic activity of the earthworm paste produced by Lumbricus rubellus seems to be a promising new direction in drug delivery research.

INTRODUCTION: Diabetes and cancer are prevalent disease with tremendous impact on health worldwide whose incidences are increasing year-on-year. However, there is a demand for new compounds for the treatment of diabetes and cancer due to unwanted side effects of the efficacies of antidiabetic and chemotherapy agents are debatable. Earthworms are not vectors of disease nor are they annoying pests.



There are about 7000 species of earthworms on the Earth which are originated majority in the Middle East, Europe and Asia ¹. They are vital component of traditional Chinese medicine and have been widely used for thousands of years. However, the research on pharmacological effects of earthworms has been initiated in the past few decades along with the development of biochemical technologies. Science has already proven that earthworm and its extracts can be used as medicinal resources but only a few species of earthworm can be used as medicine.

Several research studies were conducted to prove that earthworm protein and its coelomic fluid exhibited a variety of pharmacological activities include anti-inflammatory, anti-oxidative, antitumor and antibacterial effect 2, 3. Some active proteins and compounds in earthworm are responsible in pharmacological effect such as fibrinolytic enzymes, lumbrokinase, collagenase, superoxide dismutase, cholinesterase, catalases, glycosidases, metallothionein, calmodulin-binding protein, protein with proliferation improving activity, lysenin, antitumor protein and so on ⁴. In fact, earthworms are inexpensive and require less ethical concerns. They have been used as medicine for centuries to cure several diseases. They are worthy of critical attention for analyzing the immune and endocrine Therefore, the following study was conducted to identify the potential antidiabetic and anticancer activity of earthworm (Lumbricus rubellus) extract with different methods.

MATERIAL AND METHODS:

cultured **Collection of Earthworms:** 1kg Lumbricus rubellus was collected from the vermin composting unit near Klang, Approximately 500 earthworms were washed with running water and drained to remove dirt from the body surface. The earthworms then were soaked in distilled water for 6 to 8 hours to allow the soil in its tract to be extracted. Later the earthworms were washed thoroughly with distilled water and collected in a petridish, which then kept in an incubator at 55°C. The drying process took around 3 days. The earthworm pastes were stored in refrigerator at normal temperature ⁵.

Preparation of Earthworm Extracts: About 10-20g of earthworm pastes were homogenized separately in different solvents used according to decreasing polarity such as that is phosphate buffer (0.2M, pH 7.0), 95% ethanol and petroleum ether. The homogenized mixtures prepared separately in different solvents were filtered and the filtrates obtained were condensed in water-bath at 35°C 6. The crude extracts obtained were diluted in 10% dimethyl sulphoxide (DMSO). The earthworm extracts were first diluted to make a stock concentration of 1000µg/mL for evaluation of antidiabetic activities. While the remaining extracts were diluted in artificial seawater (38g NaCl in 1L distilled water). The earthworm extracts were then diluted to make a stock solution of 100µg/mL.

Preliminary Phytochemical Screening:

Test for Steroids:

Salkowski Test: In 2mL of extract, 2mL of chloroform was added. Equal volume of concentrated sulphuric acid (H₂SO₄) was added by sides of the test tube. Chloroform layer appeared red and acid layer greenish yellow fluorescent. This confirms the presence of sterols ⁷.

Test for Terpenoids: 2mL of extract was added to 2mL of acetic anhydride. 3mL concentrated H₂SO₄ was carefully added to form a layer. A reddish brown colouration of the interface is formed to show positive result of the presence of terpenoids ⁷.

Test for Glycosides:

Keller-Kiliani Test: In 2mL extract, glacial acetic acid, one drop of 5% FeCl3 and concentrated H₂SO₄ were added. Reddish brown colour appears at junction of the two layers and upper layer appears bluish green, confirming the presence of glycosides ⁷.

Test for Carbohydrates:

Fehling's Test: Fehling A and Fehling B reagents were mixed and few drops of extract are added and boiled. A brick red coloured precipitate of cuprous oxide forms, hence confirming the presence of carbohydrates ⁷.

Test for Saponin:

Foam Test: The extract was diluted with 20mL distilled water and it was shaken in a test tube for 15 minutes. A layer of foam was formed which indicated the presence of saponin ⁷.

Test for Phenols:

Ferric Chloride Test: Extracts were treated with 3-4 drops of ferric chloride solution. Formation of bluish black colour indicates the presence of phenols ⁸.

Test for Flavonoids: A small quantity of extract is heated with 10mL of ethyl acetate in boiling water for 3 minutes. The mixture is filtered and the filtrated are used for the following test.

Aluminium Chloride Test: The filtrates were shaken with 1mL of 1% aluminium chloride solution and observed for light yellow colour, which appeared indicating the presence of

flavonoids. When dilute NaOH and HCl is added, the yellow solutions turns colourless ⁷.

Test for Tannins:

Ferric Chloride Test: 2mL of aqueous extract was added to 2mL of water. 1 to 2 drops of diluted ferric chloride solution was added. A dark green or blue green colouration which appear indicates the presence of tannins ⁹.

Test for Alkaloids:

The extract was warmed with 2% H₂SO₄ for 2 minutes. It is filter and few drops of reagents were added separately 7 .

Mayer's reagent: a creamy white coloured precipitation appeared giving a positive result. Wagner's reagent: a reddish brown precipitate appeared which also confirms the presence of alkaloids in the extract.

Antidiabetic Activity-Inhibition of a-Amylase Assay: 1mL of each earthworm extracts (100, 200, 400, 800 and 1000µg/mL) and 1mL of the enzyme solution were mixed in a test tube and incubated at 25°C for 30min¹⁰. To 1mL of this mixture was added 1mL of the 0.5% w/v starch solution and the tube was further incubated at 25°C for 3min. Then 1mL of the color reagent was added and the tube was placed into an 85°C water bath. After 15min, the reaction mixture was removed from the water bath and cooled thereafter, diluted with 9mL distilled water. The absorbance was measured at 540nm using spectrophotometer. Individual blanks were prepared for correcting the background absorbance. The color reagent solution was added before the addition of starch solution. Then the tube was placed into the water bath and the method was followed as described above. Controls were conducted in identical manner, replacing the extracts with 1mL DMSO. Acarbose solution was used as positive control. The experiment was carried out 6 replicates and the percentages inhibition was calculated as:

Inhibition of
$$\alpha$$
-amylase (%) = $\frac{Abs540 (control) - Abs540 (sample)}{Abs540 (Control)}$ X 100%

$$Abs_{540}$$
 (control) = Abs_{test} – Abs_{blank}
 Abs_{540} (sample) = Abs_{test} – Abs_{blank}

The inhibition of α -amylase (%) was plotted against sample concentration and a logarithmic regression curve was obtained in order to calculate the IC₅₀ value which is concentration of sample (μ g/mL) necessary to decrease the absorbance of α -amylase solution by 50%.

Cytotoxic Activity-Brine Shrimp Lethality Assay: About 100mg dried cysts were allowed to hatch in 100mL seawater at 28-30°C, under conditions of continuous illumination and strong aeration ¹¹. 10 *nauplii* were drawn through a glass capillary and placed in each 12-well culture plate containing 4.5mL of brine solution. The earthworm extract 0.5mL (10, 50, 100, 500 and 1000μg/mL) of the earthworm extracts were added to 4.5mL brine solution ¹². After the incubation for 24 hours at 25 °C, the number of survivors was counted under microscope and the percentage of mortality was calculated. The percentage of mortality (% M) was calculated as:

M (%) = % survival in the control - % survival in the treatment.

 LC_{50} (50% lethal concentration, $\mu g/ml$) was determined from 6 replicates experiments. The logarithm of concentration versus percentage of mortality was plotted and the values of LC_{50} were calculated by using Microsoft Excel program. Potassium dichromate was used as positive control and solvent was used as negative control.

RESULTS & DISCUSSION:

Percentage Yield: The petroleum ether *Lumbricus rubellus* extract yielded a brown paste at a concentration of 42.1mg/g dry weight while the 95% ethanol *Lumbricus rubellus* extract yielded a brown paste at a concentration of 83.7mg/g dry weight. The phosphate buffer *Lumbricus rubellus* extract yielded a dark brown paste at a concentration of 206.3mg/g dry weight. The percentage yield of the petroleum ether, 95% ethanol and phosphate buffer extracts were calculated and noted in **Table 1**.

Preliminary Phytochemical Screening: In this study, the results of preliminary phytochemical screening were summarized in **Table 2**.

TABLE 1: PERCENTAGE YIELD OF LUMBRICUS RUBELLUS EXTRACTS IN DIFFERENT SOLVENTS

Earthworm Extracts	Percentage Yield (%)
Petroleum Ether	4.21
95% Ethanol	8.37
Phosphate Buffer	20.63

TABLE 2: QUALITATIVE ANALYSIS OF THE PHYTOCHEMICAL COMPONENTS OF *LUMBRICUS RUBELLUS* EXTRACTS IN DIFFERENT SOLVENTS

Serial No.	Phytochemical Components	Petroleum Ether Extract	95% Ethanol Extract	Phosphate Buffer Extract
1.	Steroids	++	++	+
2.	Terpenoids	++	++	+
3.	Glycosides	++	++	+
4.	Carbohydrates	+	+	++
5.	Saponin	-	-	+
6.	Phenol	-	-	-
7.	Flavonoids	+	+	+
8.	Tannins	-	-	-
9.	Alkaloids	-	-	-

(++) Strongly present, (+) Present, (-) absent

Evaluation of Antidiabetic Activity of Lumbricus rubellus extracts: Different extracts of Lumbricus rubellus exhibited different percentage of inhibition of α -amylase activity when compared to the standard drug. The results were presented in the **Table 3** and graphically presented in **Fig. 1**. The percentage inhibition of α -amylase activity at 100-1000µg/mL concentrations of Lumbricus rubellus extracts showed a concentration-dependent increase in percentage inhibition. While the phosphate buffer extract of Lumbricus rubellus was increased in a less significant manner.

The maximum percentage inhibition of α -amylase activity was shown in 95% ethanol extract with 93.60±3.46% which is comparatively higher than the percentage inhibition of Acarbose as a standard drug with 87.90±4.85%, followed by petroleum ether extract (84.18±4.72%) and phosphate buffer extract (53.85 \pm 3.80). The IC₅₀ value for ethanol, petroleum ether and phosphate buffer extracts of Lumbricus rubellus were $242.46 \mu g/mL$, 304.15µg/mL and 689.68µg/mL respectively which comparatively higher than IC₅₀ of Acarbose as a standard with the value of 193.68µg/mL. Thus, it becomes evident that Acarbose produced more inhibition of α -amylase activity in lower concentration.

Narkhede *et al.* (2011) conducted a study on invitro antidiabetic activity of *Caesalpina digyna* methanol root extract by using a similar method ¹³. The results showed that the maximum percentage inhibition was 61.31±0.3729% with the IC₅₀ value of 686.94±3.98µg/mL. The IC₅₀ of Acarbose as a standard drug was found to be 325.50±4.7µg/mL. This revealed that *Lumbricus rubellus* extracts were able to perform a better antidiabetic activity compared to other plants.

Many bioactive compounds, especially phenolic, triterpenoids and flavonoids from different plants which have been reported to have hypoglycemic effect have a positive correlation as antidiabetic agents ^{14, 15}. In another study done by Hridi *et al*. (2013) showed that the ethanol extracts of *Hiptage* benghalensis which exhibited the presence of flavonoids and terpenoids have been reported to possess antidiabetic properties ¹⁶. Pradeep (2013) also reported that the maximum possibly of antidiabetic activity of the ethanolic extract of Strobilanthes asperrimus leaves was the presence of tannins, flavonoids and alkaloids ¹⁷. These revealed that the inhibitory α-amylase activity of Lumbricus rubellus extracts may be as a result of the existence of various phytochemicals like flavonoids, steroids, terpenoids and glycosides.

TABLE 3: IN-VITRO ANTIDIABETIC ACTIVITY OF LUMBRICUS RUBELLUS EXTRACTS AND ACARBOSE

Sample	Concentration (µg/mL)	% Inhibition Mean ± SD (n=6)	$IC_{50} (\mu g/mL)$
Petroleum Ether	100	20.91±4.81	304.15
	200	31.63±3.64	
	400	59.66±3.62	

	800	79.94±3.90	
	1000	84.18±4.72	
95% Ethanol	100	28.63±3.42	242.46
	200	38.97±4.44	
	400	59.65±4.59	
	800	90.59±3.49	
	1000	93.60±3.46	
Phosphate Buffer	100	13.02±3.59	689.68
•	200	27.71±4.23	
	400	47.08 ± 4.92	
	800	51.25±4.03	
	1000	53.85±3.80	
Acarbose	100	26.49 ± 4.02	193.68
	200	58.33±3.98	
	400	73.81±4.33	
	800	80.95±4.64	
	1000	87.90±4.85	

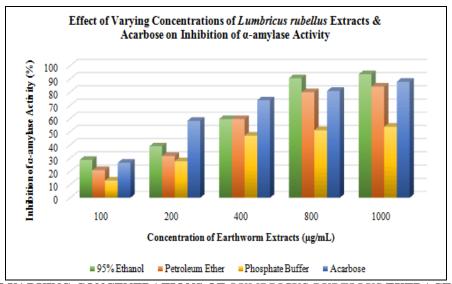


FIG. 1: EFFECT OF VARYING CONCENTRATIONS OF LUMBRICUS RUBELLUS EXTRACTS AND ACARBOSE ON INHIBITION OF α -AMYLASE ACTIVITY

Evaluation of Cytotoxic Activity of Lumbricus rubellus extracts: Different extracts of Lumbricus rubellus exhibited different percentage of mortality of brine shrimps when compared to the positive control. The results were presented in the Table 4 and graphically presented in **Fig. 2**. The percentage mortality of brine shrimps at 10-100µg/mL concentrations of Lumbricus rubellus extracts showed a concentration-dependent increase in percentage mortality. While the phosphate buffer extract of Lumbricus rubellus was increased in a less significant manner. The maximum percentage mortality of brine shrimps was shown in petroleum ether extract with 83.33±5.16% which comparatively lower than the percentage inhibition of potassium dichromate as a positive control with 88.33±7.53%, followed by 95% ethanol extract (73.33±5.16%) and phosphate buffer extract (53.33 ± 8.16) . The IC₅₀ value for petroleum ether and 95% ethanol extracts of *Lumbricus rubellus* were 7.13µg/mL and 7.28µg/mL which comparatively lower than IC₅₀ of potassium dichromate as a positive control with the value of 11.96µg/mL. Thus, it becomes evident that petroleum ether and 95% ethanol extracts of *Lumbricus rubellus* produced more cytotoxic effect in lower concentration.

A study done by Musa (2012) showed that the LC_{50} value of 80% acetone extract of *Cochlospermum tinctorium* was 240±3µg/mL, while the LC_{50} value of n-butanol extract of *Cochlospermum tinctorium* was 437±8µg/mL ¹⁸. Ahmad *et al.* (2013) reported that the rhizome extract has shown significant cytotoxic and antitumor activities with LC_{50} value of 13.57µg/mL towards brine shrimp assay ¹⁹.

This revealed that *Lumbricus rubellus* extracts were able to perform a better cytotoxic activity compared to other plants. The cytotoxic effect of plant is principally contributed by the presence of secondary metabolites such as alkaloids, glycosides, steroids, tannin, phlobatannin, terpenoids and flavonoids in their extracts ²⁰. Lay *et al.* (2014) reported that the high amount of

flavonoids, glycosides, saponin glycosides, phenolic compounds, tannins and terpenoids present in *P. macrocarpa* fruit may contribute to the antioxidant and cytotoxic properties ²¹. These are consistent with the present study as the preliminary phytochemical screening showed the presence of steroids, terpenoids, glycosides and flavonoids in *Lumbricus rubellus* extracts.

TABLE 4: IN-VITRO CYTOTOXIC ACTIVITY OF LUMBRICUS RUBELLUS EXTRACTS AND POTASSIUM DICHROMATE

Sample	Concentration (µg/mL)	% Mortality Mean ± SD (n=6)	LC ₅₀ (µg/mL)
Petroleum Ether	10	55.00±5.48	7.13
	20	61.67±4.08	
	40	71.67±7.53	
	80	80.00±6.32	
	100	83.33±5.16	
95% Ethanol	10	53.33±5.16	7.28
	20	58.33±7.53	
	40	65.00±5.48	
	80	71.67±7.53	
	100	73.33±5.16	
Phosphate Buffer	10	35.00±5.48	73.50
	20	36.67±5.16	
	40	43.33±5.16	
	80	51.67±4.08	
	100	53.33±5.16	
Potassium Dichromate	10	50.00±6.32	11.96
	20	55.00±5.48	
	40	70.00±6.32	
	80	83.33±5.16	
	100	88.33±7.53	

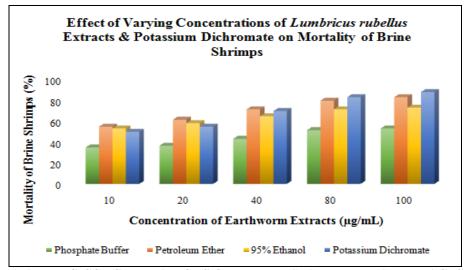


FIG. 2: EFFECT OF VARYING CONCENTRATIONS OF *LUMBRICUS RUBELLUS* EXTRACTS AND POTASSIUM DICHROMATE ON MORTALITY OF BRINE SHRIMPS

CONCLUSION: The earthworm *Lumbricus rubellus* showed significant antidiabetic and cytotoxic effect. Earthworms seem to be a promising new direction in drug delivery research. Hence, this finding indicates that the *Lumbricus*

rubellus extracts could be developed as an anticancer agent with further detailed study on compound isolation, purification and characterization.

Further toxicity studies could be conducted on individual cell lines to confirm the toxic effect of phytochemical groups.

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