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ASSESSMENT OF ANTIOXIDANT ACTIVITY OF DIFFERENT EXTRACTS OF *ANNONA RETICULATA* (L.)

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Keywords:

Annona reticulata,
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ABSTRACT: Objective: The present study has been carried out to evaluate the antioxidant property of different extracts obtained from successive extraction of *Annona reticulata* L. leave with solvents of increasing polarity. Effort also has been made to estimate the flavonoid and phenolics content of the extracts. **Method:** The antioxidant activity has been studied *in-vitro* by using DPPH, hydrogen peroxide and nitric oxide-scavenging assay. **Results:** DPPH, H₂O₂ and Nitric-oxide scavenging effect of *Annona reticulata* L. leaf extracts were found to be dose dependant with maximum inhibition at highest concentration. Methanolic extract of the leaves of *Annona reticulata* Linn. is found to have most potent anti-oxidant activity in DPPH, H₂O₂ and nitric-oxide Radical scavenging methods. IC₅₀ value DPPH, H₂O₂ and Nitric-oxide inhibition of methanolic extract at highest concentration (400µg/ml) are 62.58 ± 1.15, 68.27 ± 1.05 and 64.01 ± 1.02. **Conclusion:** The results suggest that all the tested extracts are having antioxidant property, but the methanol is having significantly higher flavonoid and phenol content. Due to presence of higher flavonoid and phenol content in methanol, it may be considered as the fraction with better pharmacological property in comparison to other tested extracts.

INTRODUCTION: For thousands of years mankind is using plant sources to alleviate or cure illness. Novel chemical compounds synthesis from the plant active constituents are of potential use in medicine and other usefull application. Herbal remedies are popular remedies for diseases used by a vast majority of the world's population. Herbal plants having many pharmacologically active compounds like flavonoids, alkaloids, tannin, steroids, glycosides, phenols, fixed oils, which is stored in their specific parts of leaves, bark, flowers, seed, fruits, root *etc.*

Annona reticulata Linn. (family- annonaceae) having different pharmacological activities such as abortifacient, anticancer, anti-inflammatory, chemo-protective, antidiabetic and wound healing activities of their different parts¹.

Reactive oxygen species (ROS) are oxygen free radicals and play dual role of being both deleterious and beneficial to biological systems. Apart from their role in the diseased conditions in the body, ROS are also known to have a role in the spoilage of food by the autoxidation of lipids, the enzymatic oxidation, during storage and processing in fats, oils and fat-containing foods. Antioxidants are defined as substances, present at low concentration relative to the oxidizable substrate, which significantly delay or prevent oxidation of substrate². Human body does not synthesize overwhelming amount of antioxidants to compensate with the damaging effects of ROS.

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Although synthetic antioxidants such as butylated hydroxy toluene, butylated hydroxy anisole, gallic acid esters and tertiary butylated hydroquinone have potential to neutralize free radicals, they have been criticized due to possible toxic effects, low solubility along with moderate antioxidant activity³. Hence there arises a need to discover new potential natural sources of antioxidants.

North East India has a vast and diverse genetic pool of various plant species and so, lots of plants are yet to be analyzed for their medicinal properties. *Annona*, a genus belonging to the Annonaceae family, is a very important medicinal plant. This is native of West Indies but *Annona reticulata* and *Annona squamosa* are widely grown throughout the tropics in India. *Annona reticulata* Linn. (Family – Annonaceae) commonly known as bullock's heart or raamphal plant, is widely distributed all over India and tropical parts of the globe. In traditional medicinal system different parts of the plant are used as insecticides, anthelmintic, suppurant, astringent, antidyseric and vermifuge. *Annona reticulata* L. is a highly apparent plant in Ayurvedic system of medicine for the treatment of various ailments. Ethanolic extracts of leaves and stem are reported to have an anticancerous activity. The aqueous leaf extract has also been reported to ameliorate hyperthyroidism, which is often considered as a causative factor of DM^{3,4}.

Ripe fruit is sweet, cooling, good tonic and sedative. It enriches the blood, increases muscular strength, lessens burning sensation, tendency to biliousness and vomiting. Leaf can be used for destroying lice⁴. Previously reported phytochemical constituents from the plant are anonaine, roemerine, norcorydiene, corydine, norisocorydine, Carvone, linalool, samoquasine A, squamocin-I, squamocin-B, squamocenin, motrilin, Kaurenic acid, phenolic and nonphenolic alkaloids, two crystalline alkaloids -muricine, muricinine, (2,4-cis and trans)-squamolinone, (2, 4-cis and trans)-9-oxoasimicinone, bullacin B 3,4 etc⁵.

Previous Experiments conducted revealed that, the methanolic extract of leave of *Annona reticulata* Linn. is having potential anti-hyperglycemic property in diabetes induced animal models⁶. Oxidative stress is known to play major role in development of diabetes and its related

complications as well as several other several metabolic disorders, and use of antioxidants may be considered as one of the approaches for management of those disorders^{7,8}. The present experiments were carried to evaluate the antioxidant potential of different extracts obtained from successive extraction of *Annona reticulata* Linn. leaves *in-vitro*.

Effort also has been made to estimate the flavonoid and phenolic content of the extracts.

MATERIALS AND METHODS:

Collection and Authentication of the Plant

Material: The leaves of *Annona reticulata* Linn. were collected in the month of October 2014 from Dibrugarh, Assam. The plant was authenticated by Dr. B.K. Sinha (HOO), Botanical Survey of India, Shillong. The plant parts specimen were submitted as herbarium with voucher specimen no. DU/PSc/HRB-01/2011. The leaves were dried initially under shade. It was preserved in air tight containers. The completely dried leaves of *Annona reticulata* Linn was coarsely powdered.

Preparation of Extracts: 2kg of powdered material was extracted in soxhlet apparatus with solvents of increasing order of polarity *i.e.* petroleum ether, ethyl acetate, acetone. Methanol and water. Extracts were dried in rotary evaporator and weighed. All the extracts obtained from successive extraction were subjected for estimation of anti-oxidant activity.

Estimation of Total Phenolic Compounds: The total phenolic content of the methanolic extract of *C. colebrookianum*, *S. edule* was determined by using the Folin-Ciocalteu reagent. The extract are dissolved in methanol and diluted. 0.5ml of extract dilution were mixed with 2.5ml of 0.2N Folin-Ciocalteu reagent for 5 min and 2.0 ml of 75 g/l sodium carbonate were then added. After incubation for 120 minutes at room temperature, the absorbance against prepared reagent blank was determined at 760nm with an UV-Vis Spectrophotometer. Standard curve of Gallic acid solution with different concentration was prepared using the similar procedure. Results were expressed as gallic acid equivalents⁹ and the calculations were done by using the following formula:

$$\text{TPC} = C \times V/m$$

Where, TPC= total phenol content, C= concentration of Gallic acid (mg/ml), V= volume of plant extract (ml) and m= weight of pure plant extract (g).

Estimation of Total Flavonoids Content:

Aluminum chloride colorimetric method was used for determination of total flavonoids. The methanolic extract of *Annona reticulata* Linn. are dissolved in methanol in the concentration of 1mg/ml and diluted. 1ml of the diluted extract solution was taken in a test tube and 1 ml of 2% AlCl₃ solution dissolved in methanol. The samples were incubated for an hour at room temperature. The absorbance was determined using spectrophotometer at $\lambda_{\text{max}} = 415$ nm. The calibration curve was prepared by using quercetin as a (standard compounds) solutions at different concentrations 10 to 90 $\mu\text{g/ml}$ in methanol. The content of flavonoids in extracts was expressed in terms of quercetin equivalent and the calculations were done by using the following formula¹⁰:

$$\text{TFC} = C \times V/m$$

Where, TFC= total flavonoid content, C= concentration of Quercetin (mg/ml), V= volume of plant extract (ml), m= weight of pure plant extract (g).

Evaluation of Anti-oxidant Activity:

DPPH Radical Scavenging Activity: The DPPH (1, 1-Diphenyl -2-picrylhydrazyl) assay method is based on the reduction of DPPH, a stable free radical. The free radical DPPH with an odd electron gives a maximum absorption at 517 nm (purple color). When Antioxidants react with DPPH, which is a stable free radical becomes paired off in the presence of a hydrogen donor (e.g., a free radical-scavenging antioxidant) and is reduced to the DPPH and as consequence the absorbance's decreased from the DPPH.

The stock solution of extracts and ascorbic acid (standard compound) were prepared in methanol to achieve the concentration of 1 mg/ml, then it was diluted to different concentration. 1 ml each of the diluted solutions were in a test tube and mixed with 1 ml of methanolic solution of DPPH in concentration of 1 mg/ml. After 30 min incubation

in darkness at room temperature, the absorbance was recorded at 517 nm¹¹.

% inhibition =

$$\frac{\text{Absorbance of control} - \text{Absorbance of sample}}{\text{Absorbance of control}} \times 100$$

Nitric Oxide Free Radical Scavenging Activity:

The principle behind this procedure is that sodium nitro-prusside in aqueous solution at physiological pH spontaneously generates nitric oxide which interacts with oxygen to produce nitrite ions that can be estimated using Griess reagent. Scavengers of nitric oxide compete with oxygen, leading to reduced production of nitrite ions. Large amounts of NO may lead to tissue damage.

The stock solution of extracts and ascorbic acid (standard compound) were prepared in methanol to achieve the concentration of 1 mg/ml, then it was diluted to different concentration. 0.5 ml each of the diluted solutions were taken in a separate test tube. To each tube 2.0 ml of sodium nitroprusside (10 mM) in phosphate buffer saline was added. The solutions were incubated at room temperature for 150 minutes.

The similar procedure was repeated with methanol as blank which served as control. After the incubation, 5 ml of griess reagent (1% sulphanilamide, 2% H₃PO₄ and 0.1% naphthyl ethylene diamine dihydrochloride) was added to each tube including control. The absorbance was measured at 546 nm on UV-visible spectrometer Shimadzu, UV-1800, Japan¹².

% inhibition =

$$\frac{\text{Absorbance of control} - \text{Absorbance of sample}}{\text{Absorbance of control}} \times 100$$

Hydrogen Peroxide Scavenging Activity:

The ability of the extracts to scavenge hydrogen peroxide was determined according to the method of Ruch *et al.*, (1989). A solution of hydrogen peroxide (40 mM) was prepared in phosphate buffer (pH 7.4). Extracts (100 $\mu\text{g/mL}$) in distilled water were added to a hydrogen peroxide solution (0.6 mL, 40mM). Absorbance of hydrogen peroxide at 230 nm was determined 10 minutes later against a blank solution containing the phosphate buffer without hydrogen peroxide¹².

The percentage of hydrogen peroxide scavenging of both *C. monogyna* extracts and standard compounds were calculated:

% inhibition =

$$\frac{\text{Absorbance of control} - \text{Absorbance of sample}}{\text{Absorbance of control}} \times 100$$

Statistical Analysis: The data were subjected to statistical analysis. All the values are expressed as mean \pm SD and data was analyzed by One-way ANOVA, using Graphpad INSTANT. The post-hoc analysis was carried out by Dunnett's multiple comparison tests to estimate the significance of difference between individual groups (**P<0.01). Confidence interval has been considered as 99% and $p < 0.01$ were considered significant. IC₅₀ value was calculated by plotting a graph with percent inhibition on y-axis and concentration on x-axis.

RESULTS AND DISCUSSION

Determination of Total Phenolic Content: The Petroleum ether, ethyl acetate, acetone, Methanolic and water extract of leaves of *Annona reticulata* Linn. were found to contain 0.08, 0.12, 0.266, 0.300 and 0.281 mg/ml equivalent to Gallic acid **Table 2**. The phenolic compound may contribute directly to the antioxidant activity.

Determination of Total Flavonoid Content: The flavonoid content as analyzed by the aluminum chloride colorimetric method was found to fall in the range of 5.72 to 77 mg quercetin equivalent/g of dry extract. The Petroleum ether, ethyl acetate, acetone, Methanolic and water extract of leaves of *Annona reticulata* Linn. were found to contain 0.235, 0.238, 0.518, 0.418 and 0.214 mg/ml equivalent to quercetin **Table 4**. Acetone extract of *Annona reticulata* Linn. showed maximum flavonoid content, followed by Methanol extract. Water extracted least amount of flavonoids, as opposed to notable amount of phenols extracted in it. Acetone appeared best for flavonoid extraction. Suitability of acetone for extraction of flavonols has already been reported¹³.

Evaluation of Anti-oxidant Activity: DPPH, H₂O₂ and Nitric-oxide scavenging effect of *Annona reticulata* Linn. leaf extracts were found to be dose dependant with maximum inhibition at highest concentration **Table 5, 6, 7**.

Methanolic extract of the leaves of *Annona reticulata* Linn. is found to have most potent anti-oxidant activity in DPPH, H₂O₂ and Nitric-oxide radical scavenging methods. IC₅₀ value DPPH, H₂O₂ and Nitric-oxide inhibition of methanolic extract at highest concentration are 62.58 \pm 1.15, 68.27 \pm 1.05 and 64.01 \pm 1.02. Water and Acetone extracts also showing remarkable anti-oxidant activity. Hence *Annona reticulata* Linn. leaf extracts have been found to show promising effect against DPPH, hydrogen peroxide and nitric-oxide.

DISCUSSION: The results of this study, clearly indicated that Methanolic extract of *Annona reticulata* Linn. have high antioxidant activity and radical scavenging activity against various antioxidant systems *in vitro* models. It has notable amount of flavonoid and phenolic compounds. These assays have important applications for the food and pharmaceutical industry. Moreover, Methanolic extract of *Annona reticulata* Linn. can be used as an easily accessible source of natural antioxidants and as a possible food supplement.

Total Phenolic Contents:

TABLE 1: OBSERVATION OF GALLIC ACID

S. no.	Conc.(mg/ml)	Absorbance
1	2	0.019
2	4	0.039
3	6	0.060
4	8	0.085
5	10	0.110

TABLE 2: TOTAL PHENOLIC CONTENTS OF EXTRACTS

S. no.	Extracts	Absorbance	Phenolic content in terms of Gallic acid equivalent(mg/ml)
1	Pet. ether	0.012	0.080
2	Ethyl acetate	0.018	0.120
3	Acetone	0.034	0.266
4	Methanol	0.045	0.300
5	Water	0.039	0.281

Total Flavonoid Contents:

TABLE 3: OBSERVATION OF QUERCETINE

S. no.	Conc.(mg/ml)	Absorbance
1	2	0.040
2	4	0.067
3	6	0.118
4	8	0.156
5	10	0.190

TABLE 4: TOTAL FLAVONOID CONTENTS OF EXTRACTS

S. no.	Plant Extracts	Absorbance	Total flavonoid content (in terms of quercetin equivalent) mg/ml
1	Pet Ether	0.045	0.235
2	Ethyl acetate	0.048	0.251
3	Acetone	0.099	0.518
4	Methanol	0.080	0.418
5	Water	0.040	0.214

TABLE 5: EFFECT OF LEAF EXTRACTS OF ANNONA RETICULATA LINN. IN DPPH ANTIOXIDANT MODEL

S. no.	Conc. (in µg/ml)	% Inhibition					Standard (Ascorbic acid)
		Pet. ether extract	Ethyl acetate extract	Acetone extract	Methanol extract	Water extract	
1	25	12.24±1.03**	20.35±1.11**	23.82±1.09**	41.57±1.02**	33.63±1.01**	55.19±1.01
2	50	20.54±1.12**	23.63±1.01**	35.08±0.97**	47.54±1.04**	43.10±1.04**	57.27±1.04
3	100	23.71±1.07**	36.76±1.01**	40.50±0.94**	54.97±1.05**	47.48±1.03**	65.68±1.05
4	200	30.05±1.18**	43.98±1.02**	48.62±0.98**	58.42±1.12**	52.84±1.02**	69.18±1.11
5	400	33.55±1.04**	50.98±1.01**	51.69±1.01**	62.58±1.15**	57.72±1.01**	72.46±1.02

Values are expressed as Mean ± SEM.; (n = 6); One Way ANOVA followed by Turkey – Kramer Multiple Comparison test; **p<0.01 vs. standard drug

TABLE 6: EFFECT OF LEAF EXTRACTS OF ANNONA RETICULATA LINN. IN H₂O₂ RADICAL SCAVENGING ASSAY

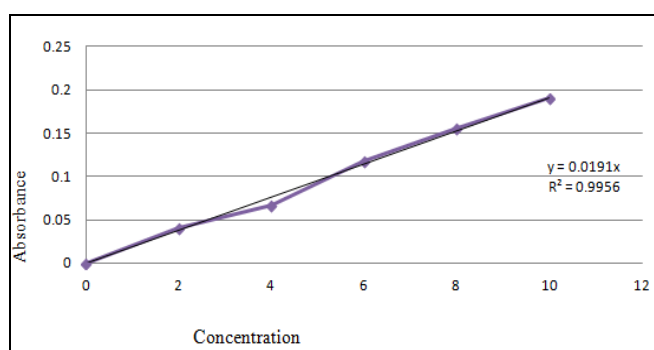
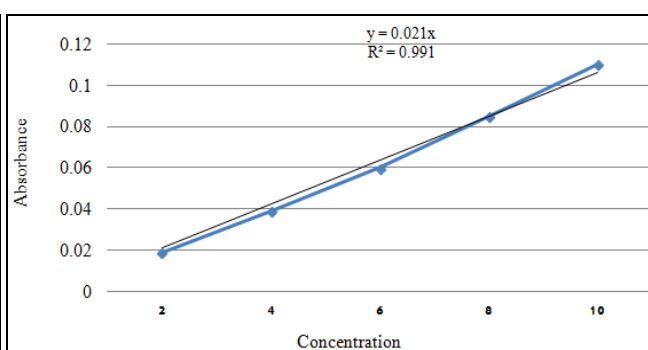
S. no.	Conc. (in µg/ml)	% Inhibition					Standard (Ascorbic acid)
		Pet. ether extract	Ethyl acetate extract	Acetone extract	Methanol extract	Water extract	
1	25	24.21±1.01**	40.35±1.01**	47.19±1.03**	51.01±1.01**	47.30±0.95**	67.82±1.05
2	50	28.13±0.98**	43.72±1.05**	51.56±1.04**	54.92±1.05**	51.68±1.05**	71.74±1.01
3	100	32.51±0.94**	46.07±1.06**	55.04±1.03**	58.00±1.09	54.93±1.03**	77.33±1.01
4	200	36.57±1.02**	51.45±1.09	58.07±1.05**	62.89±1.07**	57.06±1.09**	79.82±1.04
5	400	43.16±1.06**	55.49±1.04**	60.14±1.01**	68.27±1.05**	60.65±1.04**	81.95±1.05

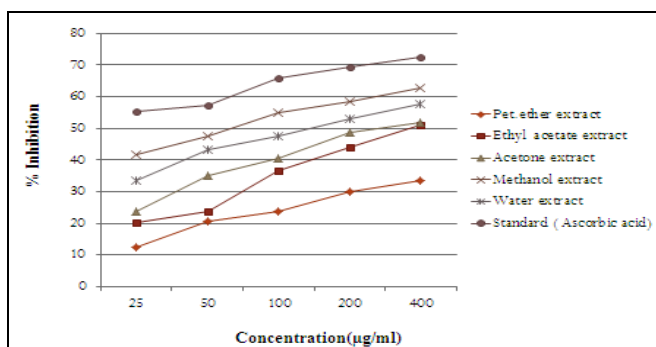
Values are expressed as Mean ± SEM.; (n = 6); One Way ANOVA followed by Turkey – Kramer Multiple Comparison test; **p<0.01 vs. standard drug

TABLE 7: EFFECT OF LEAF EXTRACTS OF ANNONA RETICULATA LINN. IN NITRIC-OXIDE-SCAVENGING ACTIVITY

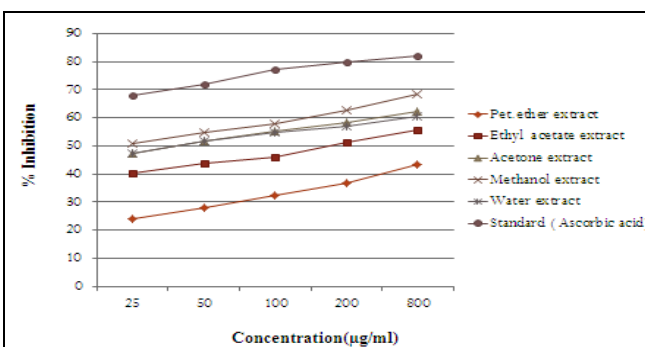
S. no.	Conc. (in µg/ml)	% Inhibition					Standard (Ascorbic acid)
		Pet. ether extract	Ethyl acetate extract	Acetone extract	Methanol extract	Water extract	
1	25	11.05±0.89**	20.30±1.10**	21.78±1.01**	43.58±1.12**	35.73±1.07**	56.65±1.14
2	50	23.87±0.97**	26.53±1.05**	31.91±1.02**	46.42±1.03**	41.92±1.02**	59.05±1.09
3	100	27.01±1.01**	33.90±1.09**	37.59±1.02**	55.81±0.97**	46.34±1.06**	66.78±1.12
4	200	34.05±1.02**	36.78±1.02**	43.42±1.01**	61.23±1.01**	52.56±1.03**	70.34±1.09
5	400	36.55±0.95**	39.95±1.13**	54.68±1.03**	64.01±1.02**	58.45±1.02**	73.45±1.10

Values are expressed as Mean ± SEM.; (n = 6); One Way ANOVA followed by Turkey – Kramer Multiple Comparison test; **p<0.01 vs. standard drug

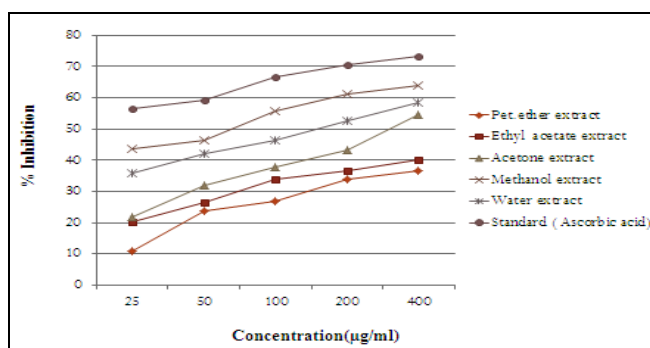
**GRAPH 1: TOTAL FLAVONOID CONTENTS: STANDARD CURVE OF QUERCETIN****GRAPH 2: TOTAL PHENOLIC CONTENTS: STANDARD CURVE OF GALLIC ACID**



GRAPH 3: COMPARATIVE EFFECT OF LEAF EXTRACTS OF *A. RETICULATA* LINN. (SAMPLE) AND ASCORBIC ACID ON DPPH ASSAY



GRAPH 4: COMPARATIVE EFFECT OF LEAF EXTRACTS OF *A. RETICULATA* LINN. (SAMPLE) AND ASCORBIC ACID ON H₂O₂ ASSAY



GRAPH 5: COMPARATIVE EFFECT OF LEAF EXTRACTS OF *A. RETICULATA* LINN. (SAMPLE) AND ASCORBIC ACID ON NITRIC OXIDE ASSAY

CONCLUSION: The plant *Annonasq reticulata* Linn. has a wide array of pharmacological activities. It is widely used in various traditional system of medicine as a medicine. It has been used since centuries a tonic, in treatment of diarrhoea, liver disorders, inflammation, leucorrhoea, urinary tract infections, malarial fever and diabetes, as antilice agent, wound healing activity.

On the basis *in vitro* antioxidant activity this is found that leaf of *Annona reticulata* Linn. contains a wide range of phytoconstituent like alkaloids, tannins, phenolics, proteins, saponins *etc.* posses good antioxidant and free radical scavenging activity which is believe to be one of the most important component for many pharmacological activity. Further investigation for isolation and identification of the phytoconstituents responsible for antioxidant activity is desirable.

Such findings can contribute to the increasing database of the medicinal plants and may be of importance in varietal improvement, food preservatives, nutraceuticals, cosmetics and biopharmaceuticals in a race with the degenerative diseases like cancer, cardiovascular diseases and

neurodegenerative diseases. Hence proper isolation of more active principles might help in the findings of new lead compounds which will be effective against free radical mediated diseases.

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CONFLICT OF INTEREST: The authors declare no conflict of interests.

REFERENCES:

1. Liu J, Jia, Kan and Jin C: *In vitro* and *in vivo* antioxidant activity of ethanolic extract of white button mushroom (*Agaricus bisporus*). Food Chem. Toxicol 2013; 51: 310-316.
2. Islam MR, Rahman SM, Ahmed M, Das PR *et al*: Antinociceptive activity studies with methanol extract of *Annona reticulata* L. (Annonaceae) and *Carissa carandas* L. (Apocynaceae) leaves in Swiss albino mice. Adv Nat Appl Sci 2012; 6:1313-8.
3. Gupta S, Shukla R and Sharma KK: Antidiabetic, Antihypercholesterolemic and antioxidant effect of *Ocimum sanctum* Linn. Seed oil. Ind. Jr. Exp. Biol. 2006; 44(4):300-303.

4. Ramalakshmi S, Edayadulla N, Ramesh P and Muthuchelian K: Investigation on cytotoxic, antioxidant, antimicrobial and volatile profile of *Wrightia tinctoria* (Roxb.) R. Br. Flower used in Indian medicine. APJTD. 2012; D68-S75.
5. Swain: Anti-hyperglycemic effect *Annona reticulata* L. Leaves on experimental diabetic rat model. Asian J Pharm Clin Res 2013; 6(1): 56-60.
6. Praveen S, Sandhya M, Peeyush A *et al*: Oxidative Stress in Metabolic Syndrome. Indian J Clin Biochem 2005; 20(1):145-9.
7. Si J, Yongzhong W, Shiwei D, Jinxiang Z, Xiao QC: Oxidants and Antioxidants in Metabolic Syndrome and Cancer. Oxid Med Cell Longev 2014; 178962.
8. Bondet V, Brand-Williams W, Berset C: Kinetics and Mechanisms of antioxidant activity using the DPPH* free radical method. LWT-Food Sci Tech. 1997; 30(6): 609-15.
9. Brand-Williams W, Cuvelier ME and Berset C: Use of a free radical method to evaluate antioxidant activity. LWT-Food Sci Technol 1995; 28(1):25-30.
10. Dharmendra S, Manish M, Monika G *et al*: Nitric Oxide radical scavenging assay of bioactive compounds present in methanol Extract of *Centella asiatica*. Int J Pharm Pharm Sci Res 2012; 2(3): 42-4.
11. Mcdonald S, Prenzler PD, Autolovich M and Robards K: Phenolic content and antioxidant activity of olive oil extracts. Food Chem 2001; 73: 73-84.
12. Khatiwora Elija, Vaishali B, Adsul, Manik *et al*: Spectroscopic determination of total phenol and flavonoid contents of *Ipomoea carnea*. Int J Chem Tech Res 2010; 2(3):1698-701.
13. Rathore KS, Panchawat S and Sisodia SS: A review on herbal Antioxidants. International journal of Pharm Tech Research 2010; 2(1): 232-239.

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