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PRODUCTION OF XYLANASE BY *PENICILLIUM SP.* AND ITS BIOBLEACHING EFFICIENCY IN PAPER AND PULP INDUSTRY

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ABSTRACT: The main objective of this study is to evaluate the ability of production of xylanase by a fungal strain *Penicillium sp.* isolated from decaying wood. Xylanases are hydrolytic enzymes produced by many microorganisms which catalyze the hydrolysis of xylan polymer to xylose monomer. The xylanase production was carried out under submerged fermentation xylose as the main substrate, and the enzyme yield was regularly monitored. Maximum xylanase production was observed on the 4th day of incubation with 123.1 U/ml. The protein secretion was high on the 6th day with 787 µg/ml; total sugar was on the 7th day of incubation (982 µg/ml). Among natural substrates used in this study, corn cobs amended medium had yielded more xylanase production by *Penicillium sp.* after 6 days of incubation (167.2 U/ml) followed by sawdust and rice bran (128.1 and 120.8). The crude enzyme was used in bleaching of paper pulp and in biobleaching sample kappa number was reduced from 23.1 to 20.8 and brightness was increased to 40.1 from 39.2. It was also observed that chromophores and hydrophobic compounds were removed in xylanase treated pulp samples. Thus by this study, it is highlighted that xylanase by present fungus *Penicillium sp.* would be useful as a promising biobleaching agent in paper and pulp industry.

INTRODUCTION: The effluents generated from paper and pulping industries are showing the deleterious effect on mankind as well as on the environment. To avoid these environmental hazardous effects novel methods with biotechnology and enzyme technology are in use and are becoming more popular in various industrial sectors ¹. Xylanases (*O*-glycoside hydrolases, EC 3.2.1.) are one of such enzymes getting attention in recent years.

These are the hydrolytic enzymes that randomly split the β -1,4 strength of the complex plant cell wall polysaccharide xylan. The xylan, almost as ubiquitous as cellulose, is the most abundant polysaccharide present in wood, agricultural and several agro-industrial wastes.

The production of xylanases is reported by many different fungi and bacteria. But from an industrial point of view, filamentous fungi are interesting producers of these enzymes due to xylanases releasing and their easy cultivation ²⁻⁵. These enzymes have potential industrial, economic and commercial applications and are used in the pulp and paper, food, beverage, textile and animal feed industries ^{6, 7}. Mostly, xylanases are produced by *Trichoderma*, *Bacillus*, *Aspergillus*, *Penicillium*, *Aureobasidium* and *Talaromyces sp.* ^{8, 4, 5}.

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The production of xylanases and xylanolytic enzymes by *Penicillia* has been explored in some species^{9, 10, 11, 1}. According to Chávez *et al.*, *Penicillia* are saprophytic and have a rich source of enzymes for the biodegradation of xylan¹². Also, Cellulase-free xylanases are important in pulp biobleaching as alternatives to the use of toxic chlorinated compounds because of the environmental hazards and diseases caused by the release of the adsorbable organic halogens¹³. There are several reports of xylanase production from *Penicillia* from soil, litter and wood chips. In the present study, a laboratory isolate grown on a decayed wood was tested for its production of xylanases and an attempt was also made in testing its efficiency in paper pulp biobleaching.

MATERIALS AND METHODS:

Isolation of Microorganism: The fungal culture was isolated from decayed wood and was cultured on potato dextrose agar (PDA) medium. The colonies were observed for mycelium, spores arrangement and characterized morphologically. Stock cultures were cultured at 30 °C but stored at 4 °C on slants of solid potato dextrose agar (PDA) medium.

Screening of Fungal Culture for Xylanase Production: The fungus was screened for xylanase production efficacy by growing it on xylan-agar medium, containing 1% w/v of birchwood xylan and 2% agar incubated at 37 °C. Actively growing 4th-day old fungal strain was inoculated in solid medium and the plates were incubated at 37°C for 5 days. After the incubation period, the plates were screened for xylanase activity by staining with Congo red solution (0.5% w/v Congo red and 5% v/v ethanol in distilled water) for 15 min followed by destaining with 1M NaCl. The plates were then observed for the appearance of the yellow coloured zone around the colony on the red background

The same fungal strain was also tested for its ability of cellulase (CMCase) production on a screening medium on a minimal medium with 1% CMC, and 2% agar. After incubation for 6 days at 37 °C, the Petri plates were flooded with 0.1% Congo Red for 15 min and subsequently destained with 1M NaCl for 30 min. Formation of the zone around the fungal colony was observed for CMCase production.

Xylanase Production from Fungal Isolate: Secondary screening of xylanase production was quantitatively determined by growing *Penicillium* sps in Malt yeast extract glucose (MYG) medium (0.2% malt extract, 0.2% yeast extract, 2% glucose) containing 1% birchwood xylan and incubated at 28 °C for 7 days at an agitation of 180 rpm. During incubation, the extracellularly secreted enzyme (crude) was recovered by centrifuging regularly for 7 successive days and was estimated.

Enzyme Assay: Xylanase activity was determined by measuring the amount of reduced sugar released in the reaction mixture by the method of Bailey *et al.*¹⁴ One unit of enzyme activity was expressed as 1 µmol of xylose released per minute per milliliter of enzyme solution.

Biomass Determination: Mycelial mat from fermentation broth was collected and then dried at 70 °C in an oven for overnight. The weight difference between the initial weight of the filter paper and filter paper having mycelial mat given biomass of the fungal mat and was expressed in terms of mg/flask.

Estimation of Total Protein: The amount of Protein content in culture broth was determined according to Lowry *et al.*,¹⁵ using bovine serum albumin (BSA), as the standard.

Determination of Reducing Sugars: Released reducing sugars in the culture filtrate was determined by using the 3, 5-dinitrosalicylic acid (DNS) method using glucose as standard¹⁶.

Production of Xylanase with Agro-industrial Residues as Carbon Sources: Enzyme production on different carbon sources was tested with the addition of different agricultural residues as Carbon sources like corncobs, groundnut shell, rice bran and sawdust at the concentration of 1% (w/v). After inoculation, flasks were incubated for 7 days at 28°C at 150 rpm. Xylanase activity was determined in each case as described previously.

Effect of Xylanase on Waste Paper Pulp: Waste office paper was finely ground and treated with 1ml of crude xylanase enzyme and incubated for 1 h at 30 °C. A similar type of pulp was prepared and treated with distilled water and kept as a control.

After incubation, the efficiency of the bioleaching process was measured in terms of reduction in kappa number and brightness of the pulp^{17, 18}. The pulp free filtrate as used for measurement of reducing the sugar by the DNS method and measurement of chromophores, hydrophobic compounds infiltrate was done at 237, 465 and 950 nm.¹⁹

Characterization of Pulp: The difference between control and enzyme-treated pulp were analyzed by SEM (JEOL JSM- 6701F scanning electron microscope, JAPAN). The results were explained below.

RESULTS AND DISCUSSION: In this study, the fungal strain, *Penicillium sp.* was screened for enzymes, cellulase and xylanase activities. Qualitative screening on MYG agar medium resulted in the appearance of a clear yellow zone on a red background and revealed that strain was xylanase positive whereas on CMC agar medium there was not any zone formation and found to be cellulase negative. To check the production of xylanase activity, quantitative submerged fermentation of the strain was cultured in liquid PDA broth. Enzyme production was determined for 1 week continuously, and activity exhibited was shown in **Fig. 1**.

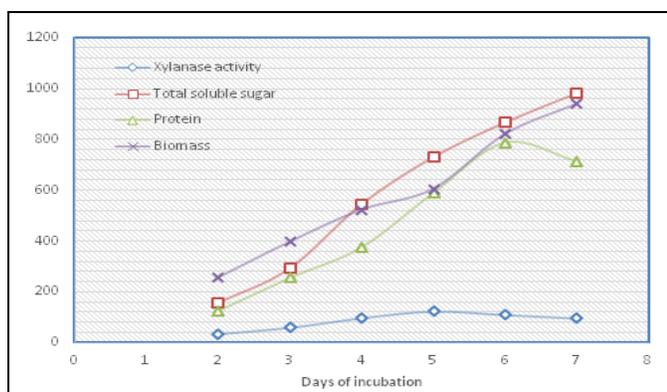


FIG. 1: PRODUCTION OF XYLANASE BY *PENICILLIUM* IN DIFFERENT DAYS OF INCUBATION

Maximum xylanase production was observed on the 4th day of incubation with 123.1 U/ml, and a little decline was observed after that. In addition to enzyme production total protein and reducing sugars were estimated in the same incubation time. The protein secretion was high on the 6th day with 787 µg/ml whereas total sugar was on the 7th day of

incubation (982 µg/ml). The organism had reached maximum growth of 939 mg/flask on the 7th day of incubation. The production of xylanase by *Penicillium sp.* in this study was compared to other species of *Penicillium*. Oat spelt xylan was found to be the best inducer in submerged fermentation for others species of genus *Penicillium* in studies of Medeiros & others^{9, 10, 11, 1, 20}. Several other reports related to xylanase production by submerged fermentation using bacteria and fungi stating that submerged fermentation is advantageous and scale up is easy¹².

Influence of Other Carbon Sources on Xylanase Production: The production of xylanases has been extensively studied in submerged and SSF culture processes. But the industrial application of xylanases is hindering due to their high cost. To overcome this disadvantage production of the enzyme on agricultural and agro-industrial wastes as carbon sources may be advantageous. In this attempt, the study was performed experimenting by using Sawdust, corncobs, rice bran, and groundnut shell at 1% to medium instead of xylan.

Of these corn cobs amended medium had yielded more xylanase by *Penicillium sp.* after 6 days of incubation followed by sawdust and Rice bran. Ground shell amendment had not increased the enzyme production than control. Similar studies also indicated that wheat bran showed to be the best inducer for xylanase production by *P. sclerotiorum* as for units of activity per volume and specific activity as well. *Penicillium expansum*, *Penicillium sp.* ZH-30 and *Penicillium chrysogenum* xylanases also were induced by this carbon source^{21, 22}.

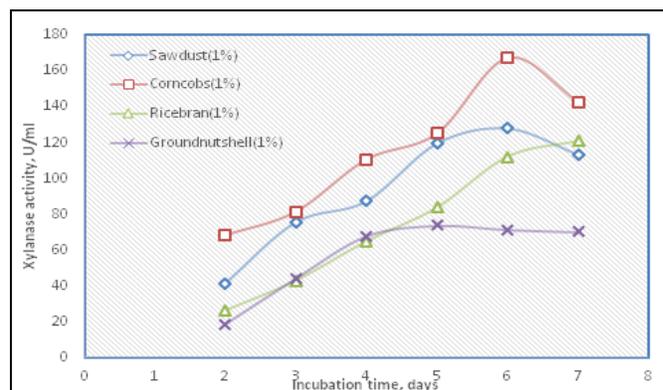


FIG. 2: EFFECT OF CARBON SOURCES ON XYLANASE PRODUCTION BY *PENICILLIUM* SPS

Effect of Xylanase on Waste Paper Pulp:

Enzyme treated and control pulp samples were analyzed for Kappa number and brightness. It was observed that in bioleached sample kappa number was reduced from 23.1 to 20.8 and brightness was increased from 39.2 to 40.1 **Table 1**. It is due to the release of lignin from the pulp as by biological treatment²³. It was also observed that removal of chromophores and hydrophobic compounds were in xylanase treated pulp samples **Table 1**.

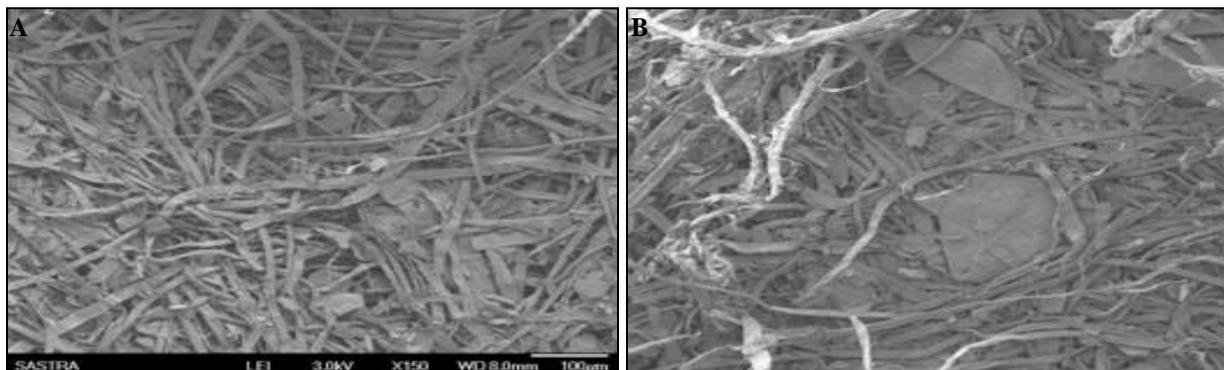
Crude xylanase biobleaching of paper pulp was also studied by *T. asperellum* and *A. niger*^{4, 5} but with better efficiency than in present study. Similarly, xylanases is utilized as a part of pulp and paper biobleaching to substitute dangerous chlorinated compounds^{24, 25}. Thus, biobleaching efficiency was found to be effective to ensure minimal damage to the pulp fibers and to generate superior quality of paper pulps.

TABLE 1: EFFECT OF XYLANASE ON PAPER PULP SHOWING DIFFERENCES IN BLEACHING EFFICIENCY

Parameter	Kappa number	Brightness (ISO Units)	Chromophoric compounds (λ_{237})	Hydrophobic compounds (λ_{465})	Reducing Sugar (mg/g)
Untreated	23.1	39.2	0.189	0.091	1.23
Xylanase (20 U/g)	20.8	40.1	0.828	0.394	1.94

Pulp Characterization by SEM Analysis: The Untreated and crude xylanase treated pulp samples were analyzed by scanning electron microscope. The scanning electron microscopy revealed that xylanase biobleaching on pretreated pulp had created pores which are not visible in the control sample **Fig. 3a & b**.

Change in the surface of the pulp indicates the efficiency of the enzyme. Sridevi et al., (2016 & 2017) had also reported with crude xylanase treatment to a paper pulp with *A. niger* and *T. Asperellum* there was a change in morphology of pulp fibers with breaking and appearance of pores and loosening of pulp fibers.

**FIG. 3 A & B: SEM IMAGES SHOWING MORPHOLOGICAL CHANGES AND PORES ON PAPER PULP**

CONCLUSION: The xylanase production by the strain *Penicillium* sp. was investigated in this study. The yield of the enzyme was enhanced in the medium when amended with natural agro wastes as substrates than xylan. Crude xylanase application as biobleaching agent on paper pulp was observed as effective after confirming the change in kappa number, brightness, and hydrophobic compounds. Future work will focus on enhancing the enzyme yield and biobleaching efficiency through optimized conditions.

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CONFLICT OF INTEREST: Nil

REFERENCES:

1. Walia A, Guleria S, Mehta P, Chauhan A and Parkash J: Microbial xylanases and their industrial application in pulp and paper biobleaching: a review. 3 Biotech 2017; 7: 11.
2. Wong KKY, Tan LUL and Saddler JN: Multiplicity of β -1,4 xylanase in microorganisms: functions and applications. Microbiology Reviews 1988; 52: 305-317.
3. Dekker M: Handbook of Fungal Biotechnology, Dilip K. Arora ed., New York 2003.
4. Sridevi A, Ramanjaneyulu G and Devi PS: Biobleaching of paper pulp with xylanase produced by *Trichoderma asperellum*. 3 Biotech 2017; 7(4): 266.
5. Sridevi A, Sandhya A, Ramanjaneyulu G, Narasimha G and Devi PS: Biocatalytic activity of *Aspergillus niger* xylanase in paper pulp biobleaching. 3 Biotech 2016; 6: 165.

6. Subramaniam S and Prema P: Biotechnology of microbial xylanases: Enzymology, molecular biology and application. *Critical Reviews in Biotechnology* 2002; 22: 33-64.
7. Polizeli MLTM, Rizzatti ACS, Monti R, Terenzi HF, Jorge JA and Amorim DS: Xylanases from fungi: properties and industrial applications. *Applied Microbiology and Biotechnology* 2005; 67: 577-591.
8. Medeiros RG, Hanada R and Ferreira-Filho EX: Production of xylan-degrading enzymes from Amazon forest fungal species. *International Biodeterioration and Biodegradation* 2003; 52: 97-100.
9. Krogh KBR, Morkeberg A, Jorgensen J, Crisvad JH and Olsson L: Screening genus *Penicillium* for producers of cellulolytic and xylanolytic enzymes. *Applied Biochemistry and Biotechnology* 2004; 114: 389-401.
10. Taibi Z, Saoudi B, Boudelaa M, Trigui H, Belghith H, Gargouri A and Ladjama A: Purification and biochemical characterization of a highly thermostable xylanase from *Actinomyces* sp. Strain Cpt20 isolated from poultry compost. *Appl Biochem Biotechnol* 2012; 166(3): 663-679.
11. Guleria S, Walia A, Chauhan A and Shirkot CK: Optimization of cultural conditions for cellulase-free xylanase production by mutant strain of alkalophilic *Cellulosimicrobium* sp. CKMX1 in submerged fermentation. *Appl Biol Res* 2013; 15(2): 137-144.
12. Chavez R, Bull P and Eyzaguirre J: The xylanolytic enzyme system from the genus *Penicillium*. *J Biotechnol*. 2006; 123: 413-433. doi: 10.1016/j.jbiotec.2005.12.036.
13. Viikari L, Ranua M, Kantelinen A, Sundquist J and Linko M: Bleaching with enzymes. *Proceedings of 3rd International Conference Biotechnology Pulp Paper Industry*, Stockholm, Sweden 1986: 66-69.
14. Bailey MJ, Biely P and Poutanen K: Interlaboratory testing of methods for assay of xylanase activity. *J Biotechnol* 1992; 23(3): 257-270.
15. Lowry OH, Rosebrough NJ, Farr AL and Randal RJ: Protein measurement with the folin phenol reagent. *Journal of Biological Chemistry* 1951; 193: 265-275.
16. Miller GL: Use dinitrosalicylic acid reagent for the determination of reducing sugars. *Analytical Chemistry* 1959; 31: 426-429.
17. TAPPI: Technical Association of the pulp and paper industry, standard T230. TAPPI Press, Atlanta, GA 1990.
18. Jordan BD and Popson SJ: Measuring the concentration of residual ink in recycled newsprint. *Journal of Pulp and Paper Science* 1994; 20: 161-167.
19. Manimaran A and Vatsala TM: Biobleaching of banana pulp using *Bacillus subtilis* C O1 xylanase produced from wheat bran under solid-state cultivation. *Journal of Industrial Microbiology and Biotech* 2007; 34: 745-749.
20. Querido ALS, Coelho JLC, Araújo EF and Chaves-Alves VM: Partial purification and characterization of xylanase produced by *Penicillium expansum*. *Brazilian Archives of Biology and Technology* 2006; 49: 475-480.
21. Li Y, Liu Z, Cui F, Xu Y and Zhao JH: Production of xylanase from a newly isolated *Penicillium* sp ZH-30. *World Journal of Microbiology and Biotechnology* 2007; 23: 837-843.
22. Okafor UA, Emezue TN, Okochi VI, Onyegeme-Okerenta BM and Nwodo-Chinedu S: Xylanase production by *Penicillium chrysogenum* (PCL501) fermented on cellulosic wastes. *African Journal of Biochemistry Research* 2007; 1: 48-53.
23. Fantahun W, Antar PV, Naveen G and Sharma P: Biobleaching of mixed wood kraft pulp with alkalophilic bacterial xylanase, mannanase and laccase-mediator system. *J Microbiol Biotech Res* 2013; 3(4): 32-41
24. Goluguri BR, Thulluri C, Cherupally M, Nidadavolu N, Achutannanda D, Mangamury LN and Addepally U: Potential of thermo and alkali stable xylanases from *Thielaviopsis basicola* (MTCC-1467) in biobleaching of wood kraft pulp. *Appl Biochem Biotechnology* 2012; 167: 2369-2380.
25. Azeri C: Thermoactive cellulase free xylanase production from alkalophilic *Bacillus* strains using various agro-residues and their potential in biobleaching of kraft pulp. *Journal of wood chemistry and technology* 2010; 309(1)-86-104.

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