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BISPHENOL- A INDUCED OXIDATIVE STRESS AND ITS FERTILITY ASPECTS

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ABSTRACT: Bisphenol-A (BPA), 2,2-bis(4-hydroxyphenyl) propane is an emerging environmental toxicant with endocrine disrupting properties and toxic effects on living organisms. BPA is ubiquitously present in consumer products current in our daily lives. As it is released from consumer products and deposited in the environment, thus creating the potential for human exposure through oral, inhaled, and dermal routes. BPA exposure might be able to cause oxidative damage by disturbing the balance between reactive oxygen species (ROS) and the antioxidant defense system of eukaryotic cells, resulting in the development of oxidative stress-related diseases. From the available information, it can be inferred that a wide variety of BPA intake through any mode results into a generation of reactive oxygen species (ROS), altering the antioxidant balance of eukaryotic cells, induces mitochondrial dysfunction, and affects cell signaling pathways related to oxidative stress. BPA induced oxidative stress might be able to cause sperm damage, mitochondrial dysfunction, and impairment of the structure and function of spermatozoa resulting in male infertility. Here, a review of the current literature examining literature related to BPA exposure, induction of ROS or oxidative stress and concludes that it alters reproductive system functions through induced oxidative stress pathways and negatively affects the fertility of male and females.

INTRODUCTION: Endocrine disrupting chemicals (EDCs) are both synthetic and natural compounds known to mimic natural hormones. Global industrialization has increased population exposure to EDC's. EDC's has found to have detrimental health effects on the living organisms by negatively disrupting their endocrine system functions. They are attracting high attention during the past two decades.

Endocrine disruptors because of their structural similarity, interfere with several processes of natural hormones as a result, they may found to be associated with different kinds of diseases¹.

Among potential EDCs, Bisphenol-A (BPA) [4, 4'-isopropylidenedi-phenol, 2, 2-bis (4- hydroxy phenyl)-propane); CAS # 80 b5-7] is synthetic organic compound firstly synthesized by A.P. Dianin in 1891. BPA is an anthropogenic compound mainly used as a precursor in the production of consumer products, including polymer synthesis (polycarbonate plastics and epoxy resins) thermal paper and non-polymer additives². Now it is one of the most used and produced synthetic compound all over the world. Every year, more than 8 billion lbs (1 lb= about

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0.45 kg) of BPA is produced worldwide but strongest growth in production of BPA is seen in Asia³. Predominant BPA product, polycarbonate used in the manufacture of food containers and epoxy resins are using for coating in, food and beverage cans⁴.

BPA is a solid matter, insoluble in water but well soluble in alcohols, ethers, and fats. The molecular weight of BPA is 228.28 g cm⁻³. Melting temperature of BPA is 156 °C; boiling temperature is 220 °C and the combustion temperature is 79.4 °C. BPA used in the production of safety helmets, sunglasses, road signs, baby bottles and dishes, food containers, lenses, infant incubators, fridges, hair dryers, CD and DVDs, cell phones, and computers; due to its properties of thermo-resistance and transparency. Epoxy resin containing BPA also used as protective films for automotive and marine applications⁵.

In recent years BPA is a subject of concern because of its harmfulness. It is found to be present in different animate matters such as urine, human milk, and blood. The United State Environmental Protection Agency (USEPA) provided much information that about 400,000 kilograms of BPA are leached into our ecosystem per year⁶. A study designed by Center for Disease Control and Prevention (CDC) confirmed a detectable concentration of 92.6% of BPA in ranging from 0.4 to 149 ng/mL and an average of 2.6 ng/ml in urine of 2517 Americans (≥6 years of age) from the year 2003-2004 National Health and Nutrition Examination Survey (NHANES)⁷. Recently, a study on BPA correlates relationships between urinary BPA concentrations and semen quality parameters and reproductive hormone levels were examined and found that BPA was detectable in 95.3% of the urinary samples⁸. Studies also confirmed that during the development of embryo there is a high risk of vulnerability to BPA exposure and it has been found in eight weeks human embryos after fertilization⁹.

BPA is commonly present in different food-related products that are commercially used, such as baby bottles, metal food cans, and food contact paper. BPA was banned in the production of baby bottles in the European Union in 2011, due to report, adverse effects on health. Primarily food is thought

to be a predominant source of exposure to BPA, because it migrates from food containers into water or food, at high temperatures and by repeated use¹⁰. Its maximum detectable concentration in canned based food is 842 ng/g. It is estimated that BPA exposure from food is 0.01-13 µg kg⁻¹ BW day⁻¹ and less than 4.2 µg kg⁻¹ b.w day⁻¹ for children and adults respectively^{4, 7, 11, 12, 13, 14, 15}. BPA can also be released from epoxy resin lining leading to an increase of BPA levels in drinking water and has been found in drinking water at concentrations ranging from 0.014 to 0.317 µg/L and in water bottles at concentrations from 0.07 to 4.21 µg/L^{16, 17}. BPA also reported being found in detectable level in amniotic fluid, umbilical cord, placenta and fetal serum¹⁸.

Human exposure to BPA is low since BPA does not bioaccumulate in the body; chronic exposure of BPA is dependent on routine exposure from different sources. Many studies predict that BPA acts as an endocrine disrupting compound (EDC) even in low doses^{19, 20, 21}. Low doses experiments on BPA predicts that it quickly metabolized into bisphenol A glucuronide (BPAG) or bisphenol A sulfate, (BPAS) and eliminated through urine in the relatively short period - virtually all administered BPA leaves the body within 48 h without any significant amount of retention in bodily tissues²². But studies demonstrated that free BPA also to be found circulating throughout the body^{23, 24, 25, 26}.

The presence of free BPA in maternal and fetal serum and breast milk may result in long-term exposure to BPA during the fetal and neonatal period that results long-term harmful effects on the fetus and neonate.

BPA is regarded as estrogenic EDC or xenoestrogen compound after its observed effects on different hormonal regulated processes. Xenoestrogens are compounds that may disturb several processes of endogenous estrogens, such as synthesis, transport, and activity of natural estrogen. BPA has structural similarity to the synthetic estrogen diethylstilbestrol (DES) and consequently has estrogenic activity but substantially weaker than DES and natural estrogen^{27, 28}. BPA is capable of mimicking like endogenous estrogens and interacting with their receptors in a variety of fashions, and because of this, adverse

neuro-developmental, reproductive, cancerous, and metabolic outcomes have been reported in various studies^{29, 30, 31}. Recently, studies reported that BPA believed to disturb bone metabolism. BPA exerts estrogen antagonistic effect on bone cells by binding to the non-classical ER γ receptor which reduces the differentiation and increases the apoptosis in the osteoblasts and osteoclasts³².

Experimental and epidemiological studies confirmed that the estimated daily intake of BPA by humans ranges from < 1 to 5 $\mu\text{g}/\text{kg}$ body weight (BW)/day^{4, 9, 12, 33}. It is very difficult to precisely estimate whether these levels of BPA can cause endocrine disruptive or toxic effects in humans. However, several studies have reported adverse endocrine disruptive or toxic effects of BPA in animal models in the range of < 1 $\mu\text{g}/\text{kg}$ b.w/day. The free form of BPA in biological samples is of concern because animal and human studies have identified adverse health effects, many of these reported effects on neurodevelopment, male and female reproductive systems alterations, metabolic diseases and oxidative stress^{34, 35, 36}.

The United States Environmental Protection Agency (EPA) has established, after toxicological testing studies, a reference dose of 50 $\mu\text{g}/\text{kg}/\text{day}$. The Canadian equivalent of this reference dose is known as, the tolerable dose intake (TDI), and was established at 25 $\mu\text{g}/\text{kg}/\text{day}$. The European Food Safety Authority (EFSA) has established a TDI of 4 $\mu\text{g}/\text{kg}/\text{day}$ ³⁷. After an analysis of different exposure sources to BPA, EFSA stated that at current levels of exposure below the tolerable daily intake (TDI), BPA does not constitute any threat to consumers³⁸. These doses mentioned above are approximately 1000-fold lower than the lowest used dose at which effects were noted in preliminary animal testing, and are well above to most estimates of the daily day today routine human exposure³⁷.

Scientists provided much attention to find out the molecular mechanism by which BPA shows its susceptibility to a wide range of disorders, and there is numerous evidence suggests that BPA exposure causes induction of Reactive oxygen species (ROS) and contributes to the predisposition to a variety of toxicity^{28, 38}. A plethora of evidence suggested that increased BPA exposure is

associated with increased oxidative stress, which could be one of the possible mechanisms causing reproductive, hepatotoxicity and genetic toxicity^{6, 38}. In a eukaryotic cell, reactive oxygen species (ROS) form as a result of normal physiological conditions in which molecular oxygen is reduced partially. ROS that is, superoxide anions ($\text{O}_2^{\cdot-}$), hydrogen peroxide (H_2O_2), peroxy (ROO^{\cdot}), and hydroxyl (OH^{\cdot}) radicals, arise in many ways, as a product of the respiratory chain in mitochondria during aerobic respiration by enzymatic reactions. These highly reactive molecules including reactive oxygen species (ROS) and reactive nitrogen species (RNS) are most widely studied species and play essential roles in cell death or to acceleration in aging and age-related diseases³⁹.

A cell's ability to keep balance in the pro-oxidant and antioxidant levels is essential for the normal cellular metabolism, cell survival, and cell proliferation^{40, 41}. But as a result of the exposure to UV light, ionizing radiation, EDC (e.g., BPA) or heavy metal ions, this balance is disrupted and leading to a predisposition to a wide range of diseases including accelerated aging, cardiovascular problems, neuronal disorders, and the onset of cancer⁶. The health issues resulting from BPA exposure have been widely studied; however, the safety of BPA in consumer products is evaluated, and studies indicate that BPA is not only widely spread in the environment due to its wide uses, but also toxic even in low doses level. Several lines of evidence suggest that BPA-induced cytotoxicity caused by oxidative stress occurs in both cell culture studies and an animal model. Various doses and durations of BPA exposure through an imbalance of pro-oxidants/antioxidants in cultured cells and rodents observed increased oxidative stress in multiple tissues^{42, 45}.

Furthermore, Bindhumol *et al.*, demonstrated that BPA induced oxidative stress in the liver of rats by decreasing antioxidant enzymes and increasing hydrogen peroxide and lipid peroxidation and co-administration of antioxidant vitamin C reversed this BPA-induced oxidative stress⁴⁶. In the body, BPA also processed enzymatically by cytochrome P450, and as similar to natural estrogen metabolism BPA converted into quinone form that is reactive to DNA that is lethal to the viability of eukaryotic cells⁴⁷. BPA is structurally similar to estradiol and

thus interferes with steroid signaling with different possible outcomes on reproductive health depending on doses, life stage, mode, and timing of exposure. BPA exerts its epigenetic effects in both male and female reproductive system. In males, BPA affects spermatogenesis and sperm quality and possible trans-generational effects on the reproductive ability of the offspring. In females, BPA affects ovary, embryo and gamete development. It is now investigated that BPA induced oxidative stress as a result of an imbalance between oxidants and antioxidants in the semen can lead to sperm damage, mitochondrial dysfunction⁴⁸, impairments of the structure and function of spermatozoa eventually lead to male infertility^{49, 50, 51}. In this review, we will address BPA-induced oxidative stress and its possible role in fertility problems in humans by animal studies.

Dose Level of BPA: Affecting the dose level of BPA is highly controversial. Both *in-vitro* and *in-vivo* data also has contradictory health effects of BPA and create problems for regulatory agencies in evaluating the adverse health effect of BPA⁵². A plethora of evidence indicates that adverse effects of BPA on health are varying with duration, doses, exposure route and sex difference⁵³. A cut-off for lowest observable adverse effect level (LOAEL) is 50 µg/kg b.w/day (*in-vivo*) and 50 ng/ml or 2.19×10^{-7} M (*in-vitro*) BPA^{25, 29, 38, 54, 55}.

The European Food Safety Authority (EFSA) defined the maximum tolerable daily intake (TDI) for BPA at 4 µg/kg of body weight per day, and FDA established the “no observed adverse effect level” (NOAEL) of 5 mg/kg of body weight per day^{56, 57}. Based on these data Gassman, 2017 conclude in their review of 2017 that doses that are mostly used in experiments for animal studies range from 0.2 to 50 µg/kg/bw and in cell culture studies it ranges from 10^{-15} to 10^{-4} M³⁸. Based on these doses used in experiments on BPA it is predicted that a variety of BPA doses induced oxidative stress studies says that these doses affect differently in inducing oxidative stress.

BPA Induced Oxidative Stress and Generation of ROS: From the last decade, oxidative stress has become the most interesting research topic of many scientists working in the field of biology because of its role in the initiation of various diseases.

Oxidative stress describes the condition in which levels of oxygen and oxygen-derived free radicals overwhelm the natural antioxidant defenses naturally present in the cell and results in the damage of cellular components, inactivate essential metabolic enzymes and disrupt signal transduction pathways. Chemically, oxidative stress is associated with increased production of reactive oxidizing species (ROS) or a significant decrease in the effectiveness of antioxidant defenses^{41, 58, 59}.

The eukaryotic cell has a variety of natural antioxidant defenses, such as reduced glutathione, ascorbic acid, thioredoxins, and α -tocopherol, and enzymes, like superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx), play critical roles in redox reactions in the cell. Oxidative stress induces modifications of cellular proteins, thereby altering their functions and causes a state of susceptibility to a wide range of disorders. It causes damage to biomolecules including DNA, lipids and proteins and contributes to the pathology of many diseases including neuronal degeneration, autoimmune diseases, cardiovascular dysfunction, accelerated aging, the progression of cancer and conditions of the reproductive system including both male and female infertility^{41, 60}.

Reactive oxygen species (ROS) are cytotoxic agents causing oxidative damage by disrupting cell membrane and cause harm to DNA^{43, 60}. Reactive oxygen species (ROS) are highly reactive oxidizing free radical agents represent a broad category of molecules including organic (hydroxyl ion, superoxide, NO, peroxy, *etc.*) and non-radical (ozone, singlet oxygen, lipid peroxide, hydrogen peroxide) and oxygen derivatives. ROS play important roles in cell signaling, a process termed redox signaling. Thus, to maintain proper cellular homeostasis, a balance must be struck between reactive oxygen production and consumption. Free radical scavengers are involved in the defense mechanism of the organisms against the pathologies associated with the attack of free radicals. Free radicals in the body have a physiological as well as the pathological role, antioxidant molecule due to its free radical scavenging activity sometimes may act as disease promoter, by neutralizing the physiologically desired ROS molecules, and as disease mediator by removing the excessive levels of ROS species⁴¹.

ROS also have been shown to play an important role in the defense mechanisms against pathological conditions, but the excessive generation of free oxygen radicals may damage tissues and also damage proteins, leading to the structural alteration and functional inactivation of many enzymes and receptor proteins involved in cell signaling. From some recent studies, it is now clear that this generation of ROS by BPA exposure depends on cell types and which hormone receptor is being found in that particular cell type. Koong and Watson work on androgen-dependent and -independent (LAPC-4 and PC3) prostate cell lines made it clear that the same dose shows a difference in generation of ROS in both cell lines^{38, 61}. Some tests are available to measure the extent of oxidative stress, and these generally involve quantification of ROS levels, total antioxidant capacity (TAC) and levels of biomarkers associated with oxidative damage to biomolecules⁴⁵.

Environmentally persist low level of BPA exposure might be able to cause oxidative damage by disturbing the balance between reactive oxygen species and antioxidant defense system, resulting in the development of oxidative stress-related diseases **Table 1**. Experimental data have shown BPA can induce the generation of ROS through the enzymatic and non-enzymatic formation of radicals^{61, 62}. Doses of BPA below the NOAEL induce mitochondrial dysfunction in the liver, and this is thought to be caused by an increase in oxidative stress and inflammation⁶³. Cell death, DNA mutation, replication errors, and genomic instability can occur if the oxidative DNA damage is not repaired before DNA replication^{64, 65}. Recently, it has been reported that BPA exposure significantly induced DNA damage in zebrafish, with a significant increase in ROS production⁶⁶.

BPA exposure to adult male and female rats below the NOAEL dose (5.0 mg) and even at 10 mg, which is very close to environmental exposure, led to a significant increase in the chromosome breaks and fragments in bone marrow cells as well as DNA fragmentation in blood lymphocyte thereby showing genotoxic effects⁶⁷. ROS thought to create stable base lesions and basic sites in genomic DNA. The base excision repair (BER) is the main repair system responsible for removal of modified bases [such as 8-oxo-guanine (8-oxoGua) and 2, 6-

diamino -4 -hydroxy -5 -formamide-pyrimidine (FapyGua)] that formed upon oxidative stress^{68, 69}.

Several studies also suggest that BPA exposure causes an increase in oxidative stress biomarker, such as 8-OHdG, white blood cell count, and C-reactive protein, as well as malondialdehyde (MDA)⁷⁰. Eid *et al.*, found that early life exposure to BPA significantly increased oxidative/nitrosative stress, decreased antioxidant enzyme activities, inducing DNA damage and severe chronic inflammation in the hepatic tissue of female rat offspring in a time-dependent manner⁷¹. Tiwari and Vanage, predicts in their study that both low and high doses (0.01 and 5.0 mg/kg/b.w) of BPA exposure generate excess of ROS by decreasing the levels of SOD, CAT, reduced GSH and increasing LPO in bone marrow cells, blood lymphocyte, and reproductive organs, thereby causing oxidative stress⁷². *In-vivo* and *in-vitro* studies of BPA observed the formation of BPA-DNA adducts after high dose exposure of BPA^{73, 74}.

About the effects of BPA on metabolism studies demonstrate exposure to low or high doses of BPA induces abnormal glucose metabolism and insulin resistance. Insulin resistance may be thought to be associated with induced by decreased ADP production and increased oxidative damage⁷⁵. A study of Moghaddam *et al.*, indicated that BPA dose-dependently increased the levels of blood glucose, lipid profile and MDA in the tested groups compared with the control group ($p \leq 0.001$)⁷⁶.

BPA injection increased the levels of MDA and decreased the levels of GSH and TAS, and also the activities of SOD and CAT in the pancreas of exposed mice compared with the control group ($P \leq 0.05$). Results suggest that BPA exposure might induce hyperglycemia and its complications in adult male mice by induction of oxidative stress.

Epidemiological, animal and *in-vitro* studies following BPA exposure data demonstrate that BPA promotes adipogenesis, lipid and glucose dysregulation and adipose tissue inflammation, thus contributing to the pathophysiology of obesity. Induction of oxidative stress correlated with 8-hydroxydeoxyguanosine (8-OHdG) or malondialdehyde (MDA) levels has been confirmed further by population studies^{70, 77}.

Recently an study of Lv Y *et al.*, observed that due to the high levels of BPA on thermal receipts and their wide applications in our daily life, some amount of BPA may be transferred to our skin that has serious health issues confirmed by urinary BPA, TCS and 8-hydroxy-2'-deoxyguanosine (8-OHdG) concentrations determined by High-Performance Liquid Chromatography / Tandem Spectrometer (LC/MS/MS) ⁷⁸. Moreover, they investigate the potential oxidative DNA damage from exposure to BPA and TCS, ninety-six urine samples of children (aged 3–6) and 57 dust samples were collected and concentrations of urinary BPA, TCS, and 8-hydroxy-2'-deoxyguanosine (8-OHdG, a biomarker of oxidative DNA damage) in urine were determined using a liquid chromatography tandem mass spectrometer. Results showed that both BPA exposures were associated with oxidative damage.

Additionally, Ferguson *et al.*, investigated an association between BPA exposure and oxidative stress and inflammation in 482 pregnant women by analyzing urine and plasma ⁷⁹. Results suggest that BPA and two biomarkers of oxidative stress (8-hydroxydeoxyguanosine and 8-isoprostane) were

found in the urine of women. Besides this Inflammation markers, including C-reactive protein and cytokines were also measured in their plasma. Han and Hong in 2016 suggest effects of BPA exposure such as endocrinal disturbance, induction of oxidative stress and inflammation, epigenetic change, and links with other chronic diseases may highlight a possible association between BPA exposure, hypertension, and cardiovascular diseases ⁸⁰. It is reported that BPA has an adverse effect on the heart of rats which is mediated principally by the generation of ROS and reduction of antioxidant defenses of the heart aggravating a state of oxidative stress.

Study revealed that BPA administration induced a state of oxidative stress in the heart of rats as evident from the increase in MDA levels and decrease in catalase activity at the two tested doses (10 and 25 mg/kg) after 6 weeks and the decrease in GSH levels after the administration of the two doses of BPA at all tested time segments. Increased lipid peroxidation may indicate an increased oxygen free radical generation, and BPA induced ROS production significantly compromises mitochondrial function ⁸¹.

TABLE 1: INDUCTION OF OXIDATIVE STRESS BY BPA

S. no.	Model system	Dose (per-day)	Duration	Effects	Reference
1	Male Holtzman Rats	0.01, 5.0 mg/kg	6 days	Increased lipid peroxidation, decreased antioxidant enzyme activity	72
2	<i>Drosophila melanogaster</i>	0.1, 1.0, 2.5 and 5.0 µg/mL	4 h	Increased ROS, lipid peroxidation and depletion of superoxide dismutase (SOD), catalase (CAT), glutathione (GSH) and glutathione-s-transferase (GST)	82
3	Bone mesenchymal stem cells	200 IM to 500	18 h	Increased lipid peroxidation, decreased antioxidant enzyme activity	83
4	Rats	150, 250, 500 mg/kg	14 days	Hepatotoxicity by increased lipid peroxidation	84
5	Mice	0.5, 2 mg/kg	28 days	Increased lipid peroxidation, decreased antioxidant enzyme activity	85
6	Rats	0, 2, 10, 50 mg/kg	30 days	Hepatotoxicity by increased lipid peroxidation	85
7	Sprague-dawley rats	200 mg/kg	10 days	Increased lipid peroxidation, decreased antioxidant enzyme activity and induce DNA damage	86
8	Rats (Wistar)	0.1, 1, 10, 50 mg/kg	28 days	Decreased antioxidant enzyme activity	45
9	Female mice (CD-1)	0.1, 1, 5, and 10 lg/ml	8 days	Increased ROS observed	87
10	Male mice (C57BL/6)	0.05 and 1.2 mg/kg	5 days	MDA increased at both doses, GPx decreased at both doses	63

BPA Effects on Reproduction Process and Fertility: In human populations, most of the studies point towards an association between

exposure to EDCs and male and female reproductive system disorders, such as infertility, breast cancer, testicular cancer, poor sperm quality,

and function. In recent years the detrimental effects of BPA on reproductive function, following in experimental exposure, have been widely studied in laboratory animals such as rodents⁸⁸. Humans are mainly exposed to BPA through food ingestion⁸⁹, and increasing evidence supports its association with impairment of reproductive function, as well as other health problems and diseases; such diseases include diabetes, obesity, cardiovascular diseases, and cancer^{36, 90-95}. Epidemiological studies also report that BPA has a toxic effect on male and female reproductive organs at the environmentally relevant level of exposure³⁸. Even at a low dose of BPA, it is supposed to disturb the semen production in men and oocyte production in women by the interrupting the synthesis of sex steroids⁹⁶. But the effects of BPA on reproductive hormones and semen quality in different epidemiologic studies are inconsistent⁹⁷.

Association between BPA exposure and spermatogenesis is quite clear yet. BPA has been considered as a possible risk factor for fertility because it might induce testicular toxicity⁹⁸. It is reported that a low dose of BPA impairs spermatogenesis by suppressing reproductive hormone production and promoting germ cell apoptosis in adult rats²⁷. Additionally in this sense reported earlier that there is a reduction in the testosterone level after the exposure of BPA^{99, 100}. BPA also reported to interrupts the process of formation and maturation of sperm¹⁰¹. The decreased sperm count at the lowest tested BPA group (25 µg/kg) is to some extent supported by other studies reporting decreased sperm count in rodents after low-dose developmental exposure to BPA^{102, 103}. Salian *et al.*, performed a three-generation study to assess the effects of very low doses of BPA (1.2 or 2.4 µg/kg bw/day administered by gavage) in Holtzman rats (n = 8 litter per group/generation)¹⁰³. Sperm count and motility were significantly reduced in the F1, F2 and F3 male offspring, with a dose-related reduction in sperm count.

Additionally, a recent study has shown that pre- and postnatal exposure of Sprague Dawley rats (n = 3) to 5 µg/kg b.w/day of BPA also decreased epididymal sperm count and motility at 70 days of age¹⁰⁴. Other studies have not found significant effects of low-dose BPA on sperm count¹⁰⁵⁻¹⁰⁸. In

2015 Johnson *et al.*, in their experiment selected three doses of BPA (2.5, 25 and 2500 µg/kg bw/day) or a 0.5 µg/kg/day ethinyl estradiol and doses were given to Sprague Dawley dams by oral gavage on gestational days 6-21, whereas offspring were dosed directly from birth to weaning. These findings indicate that developmental exposure to BPA can disrupt aspects of spatial navigational learning and memory in a sex-dependent manner¹⁰⁹. Study of Rahman *et al.*, investigated the effects of varying concentrations of BPA (0.0001, 0.01, 1, and 100 µM for 6 h) on sperm function, fertilization, embryonic development, and on selected fertility-related proteins in spermatozoa¹¹⁰.

Results suggest that BPA at concentrations of 0.0001, 0.01, and 1 µM did not produce significant or partial toxic effects on spermatozoa; however, 100 µM BPA affected motility parameters, the acrosome reaction, fertilization, and early embryonic development, which are closely associated with down-regulation and phosphorylation of fertility-related proteins in spermatozoa. Other study demonstrated that BPA induces subacute toxicity in wistar rats and there was significant (P≤0.05) reduction in the epididymal sperm count in 200 mg/kg and 600 mg/kg dose group. Sperm motility percentage, dead count percentage, head, and tail abnormality percentage were found to be significantly (P≤0.01) increased in rats of BPA-treated groups as compared to rats of control groups.

About the effect of BPA on accessory reproductive organs, BPA also has been reported to affect the prostate gland. It is reported that developmental exposure to BPA has been associated with the increased susceptibility to the prostate cancer¹¹¹. Additionally, Wu *et al.*, reported that at low dose BPA has an effect on the prostate in Sprague-Dawley rats by changing the estrogen to androgen ratio and also affect testosterone metabolism¹¹². Experimental studies suggest that BPA's mechanism of action is related to life stage and that its effect on the female reproductive system may involve agonism with estrogen nuclear receptors as well as steroid biosynthesis inhibition¹¹³. It is already mentioned that BPA exposure is able to disturb the hormonal balance due to its endocrine disrupting effects at low doses, so it may also affect the reproductive process in women. Experimental

results of *in-vivo* and *in-vitro* studies confirms BPA shows a significant effect on female reproduction. Earlier reported that BPA interrupts the process of meiosis and affect the development of oocyte¹¹⁴. Even at low doses, BPA is able to disturb the process of oocyte maturation¹¹⁵. Additionally, BPA exposure increased the formation of multi-oocyte follicles and caused an increase in some apoptotic oocyte^{116, 117}. Another study of Zhang *et al.*, found that low dose BPA exposure causes an increase in ovarian weight, atretic follicles, and peri-nuclear oocytes¹¹⁸. Cultural oocyte experiments also suggest that BPA might disturb the process of the prophase stage of meiosis and survival of the oocytes¹¹⁹. Ganesan and Keating observed that after one-day exposure to BPA, DNA damage is also observed in rat ovaries. *In-vitro* studies also indicated the impairment of uterine cell proliferation after BPA exposure¹²⁰.

The epidemiological study confirms that BPA level was found higher in infertile females as compared to fertile, but results are often contradictory. Its effects on female reproductive organs depend upon exposure level, dose and duration, and model used in the study. Some results suggest that it has a toxic effect on female reproduction and affect female fertility by affecting reproductive organs. Studies using animal model provide evidence that BPA has effects on litter size of mice¹²¹. Other studies indicate at a dose of 50 mg/kg/day during gestation period BPA has no significant effect on litter size of mice¹²². BPA is in higher concentration in the blood plasma of infertile women and thought to be having hormone effect on fertility¹²³. According to case studies concentration of BPA in the serum of women may be related to the onset of endometriosis, PCOS and pre-eclampsia and shows an association between BPA exposure and fertility outcomes¹²⁴⁻¹²⁸. Cohort studies also found as an association between urine BPA concentration and increased pre-term birth compared to full-term birth¹²⁹.

About the cohort study on male, Li *et al.* observed BPA concentration in urine of men and found significantly lower sperm count, sperm concentration, sperm vitality and sperm motility¹³⁰. Another study of Li *et al.*, observed BPA in the urine of 427 men and evaluated the significant decrease of libido, erection problem and lower

ejaculation intensity⁹⁰. It was also observed that increased BPA concentration in urine is also found to be related to greater sperm DNA damage⁵⁰. Viktu *et al.*, observed blood plasma and also found a correlation with BPA level in plasma and alteration in sperm parameters¹³¹. Recently reported that there is an interaction between BPA exposure and obesity on sperm count and sperm concentration as observed in mice confirming the toxic effect of BPA on male reproduction^{132, 133}.

BPA exposure induced oxidative stress, demonstrated by an increase in lipid peroxidation and a decrease in activity of various enzymatic and non-enzymatic antioxidants in bone marrow cells, blood lymphocytes, and testicular and epididymal tissues of male Holtzman rats illustrates one of the possible mechanisms causing reproductive and genetic toxicity⁷². The reproductive health effects resulting from BPA exposure due to its endocrine disrupting properties it is thought to have a pronounced effect on human reproduction even at low doses that are below the LOAEL. For example, BPA at its lower doses has been reported to induce complete degeneration of epididymal epithelium with reduction in the number of spermatozoa due to either a decrease in serum testosterone or dihydrotestosterone (DHT) or even as a result of reduction in 5 α - reductase, an enzyme required to convert testosterone to DHT of male Wistar rats⁸⁸.

Studies to date have explored associations between maternal exposure to BPA in pregnancy and indicators of inflammation and oxidative stress with an increase in maternal biomarkers of oxidative stress, including indices of oxidative DNA and lipid damage⁷⁹. About oxidative stress effects of BPA on reproductive organs, the literature shows an association between BPA exposure and its effects on reproduction. BPA at doses of 50 or 70 μ m induces apoptosis by an excessive ROS generation and mitochondrial dysfunction in sertoli cells which are crucial for the development of sperm⁶⁶. The oxidative stress caused by BPA exposure has been found to affect the fertility of rats in a study where different doses of BPA (0, 2, 10, 50 mg/kg/b.w) that induces oxidative stress in liver and epididymis of treated rats and found to affect the semen quality by decreasing sperm count and quality⁵¹.

The study confirms that BPA exposure inhibits the reproductive function in the male rat and cause genotoxicity¹³⁴. Histopathological findings in the testes showed necrosis of the germinal layer and spermatogonial cells in the seminiferous tubules. Thus, it appears that BPA affects the germ cells leading to impairment in the spermatogenesis and thus having its property as reproductive toxicant¹³⁵. Hass *et al.*, reported that developmental exposure to 25 µg/kg b.w/day bisphenol A that is above the safe level described by the EFSA¹³⁶. This exposure can cause adverse effects on fertility (decreased sperm count), neurodevelopment (masculinization of spatial learning in females) and lead to increased female body weight late in life.

Results suggest that highly exposed humans may not be sufficiently protected about endocrine disrupting effects of BPA. Spermatozoa with damaged DNA have the minimum potential of fertilizing an ovum, and the presence of high levels of DNA damage in human spermatozoa has been correlated with adverse clinical outcomes including infertility, recurrent pregnancy loss, genetic disorders, and childhood cancers. ROS generation causes DNA damage in spermatozoa that are generated by spermatozoa themselves and by leukocytes present in the seminal plasma. Study now confirmed that human exposure to BPA might be associated with reduced semen quality and increased sperm DNA damage⁵⁰.

CONCLUSION: From the available information it can be inferred that a wide variety of BPA intake through any mode results into a generation of ROS, altering the anti-oxidant balance, induces mitochondrial dysfunction, and affects a number of cell signaling pathways related to oxidative stress. Moreover, it also alters the reproductive system functions through induced oxidative stress pathways and negatively affects the fertility of mammals. BPA induced oxidative stress negatively affects fertility and probably adding to increasing infertility worldwide. Still, a more precise study of BPA intake, dose, duration, and model system to reveals that wide variety of BPA doses promote the generation of ROS and in form affect the normal reproductive system functions.

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REFERENCES:

1. Prins GS, Patisaul HB, Belcher SM and Vandenberg LN: CLARITY-BPA academic laboratory studies identify consistent low-dose Bisphenol A effects on multiple organ systems. *Basic & Clinical Pharmacology & Toxicology* 2018.
2. Michałowicz J: Bisphenol A-sources, toxicity and biotransformation. *Environ Toxicol Pharmacol* 2014; 37(2): 738-58.
3. Giulivo M, de Alda ML, Capri E and Barceló D: Human exposure to endocrine disrupting compounds: Their role in reproductive systems, metabolic syndrome and breast cancer. A review. *Environmental Research* 2016; 151: 251-64.
4. Kang JH, Kondo F and Katayama Y: Human exposure to bisphenol A. *Toxicology* 2006; 226(2-3): 79-89.
5. Kaur K, Simon AF, Chauhan V and Chauhan A: Effect of bisphenol-A on *Drosophila melanogaster* behavior-A new model for the studies on neurodevelopmental disorders. *Behavioral Brain Research* 2015; 284: 77-84.
6. Seachrist DD, Bonk KW, Ho SM, Prins GS, Soto AM and Keri RA: A review of the carcinogenic potential of bisphenol A. *Reproductive Toxicology* 2016; 59: 167-82.
7. Calafat AM, Ye X, Wong LY, Reidy JA and Needham LL: Exposure of the US population to bisphenol A and 4-tertiary-octylphenol: 2003-2004. *Environmental Health Perspectives* 2007; 116(1): 39-44.
8. Adoamnei E, Mendiola J, Vela-Soria F, Fernández MF, Olea N, Jørgensen N and Torres-Cantero AM: Urinary bisphenol-A concentrations are associated with reproductive parameters in young men. *Environmental Research* 2018; 161: 122-28.
9. Chen M, Fan Z, Zhao F, Gao F, Mu D, Zhou Y and Hu J: Occurrence and maternal transfer of chlorinated bisphenol A and nonylphenol in pregnant women and their matching embryos. *Environmental Science and Technology* 2015; 50(2): 970-77.
10. Konieczna A, Rutkowska A, Szczepańska N, Namieśnik J and Rachoń D: Canned food as a source of bisphenol a (BPA) exposure-estimation of consumption among young women from Gdańsk, Poland. *Medycyna Środowiskowa-Environmental Medicine* 2018; 1(21): 31-34.
11. Sajiki J, Miyamoto F, Fukata H, Mori C, Yonekubo J and Hayakawa K: Bisphenol A (BPA) and its source in foods in Japanese markets. *Food Additives and Contaminants* 2007; 24(1): 103-12.
12. Vandenberg LN, Hauser R, Marcus M, Olea N and Welshons WV: Human exposure to bisphenol A (BPA) *Reprod Toxicol* 2007; 24: 139-77.
13. Cao XL, Corriveau J, Popovic S, Coughlan MC, Chepelev N, Willmore W and Jin X: Background bisphenol A in experimental materials and its implication to low-dose *in-vitro* study. *Chemosphere* 2010; 81(6): 817-20.
14. Noonan GO, Ackerman LK and Begley TH: Concentration of bisphenol A in highly consumed canned foods on the US market. *Journal of Agricultural and Food Chemistry* 2011; 59(13): 7178-85.

15. Geens T, Neels H and Covaci A: Distribution of bisphenol-A, triclosan and n-nonylphenol in human adipose tissue, liver and brain. *Chemosphere* 2012; 87(7): 796-02.
16. Arnold SM, Clark KE, Staples CA, Klecka GM, Dimond SS, Caspers N and Hentges SG: Relevance of drinking water as a source of human exposure to bisphenol A. *Journal of Exposure Science and Environmental Epidemiology* 2013; 23(2): 137.
17. Colin A, Bach C, Rosin C, Munoz JF and Dauchy X: Is drinking water a major route of human exposure to alkylphenol and bisphenol contaminants in France? *Archives of Environmental Contamination and Toxicology* 2014; 66(1): 86-99.
18. Ikezuki Y, Tsutsumi O, Takai Y, Kamei Y and Taketani Y: Determination of bisphenol-A concentrations in human biological fluids reveals significant early prenatal exposure. *Human Reproduction* 2002; 17(11): 2839-41.
19. Völkel W, Colnot T, Csanády GA, Filser JG and Dekant W: Metabolism and kinetics of bisphenol A in humans at low doses following oral administration. *Chemical Research in Toxicology* 2002; 15(10): 1281-87.
20. Schug TT, Janesick A, Blumberg B and Heindel JJ: Endocrine disrupting chemicals and disease susceptibility. *The Journal of Steroid Biochemistry and Molecular Biology* 2011; 127(3-5): 204-15.
21. Fenichel P, Chevalier N and Brucker-Davis F: Bisphenol A: an endocrine and metabolic disruptor. In *Annales d'endocrinologie* 2013; 74(3): 211-20. Elsevier Masson.
22. Upmeier A, Degen GH, Diel P, Michna H and Bolt HM: Toxicokinetics of bisphenol-A in female DA/Han rats after a single IV and oral administration. *Archives of Toxicology* 2000; 74(8): 431-36.
23. Vandenberg LN, Chahoud I, Heindel JJ, Padmanabhan V, Paumgarten FJ and Schoenfelder G: Urinary, circulating, and tissue biomonitoring studies indicate widespread exposure to bisphenol A. *Ciencia and Saude Coletiva* 2012; 17: 407-34.
24. Vandenberg LN, Colborn T, Hayes TB, Heindel JJ, Jacobs Jr DR, Lee DH and Zoeller RT: Hormones and endocrine-disrupting chemicals: Low-dose effects and nonmonotonic dose responses. *Endocrine Reviews*, 2012; 33(3): 378-55.
25. Vandenberg LN, Hunt PA, Myers JP and Vom Saal FS: Human exposures to bisphenol A: mismatches between data and assumptions. *Reviews on Environmental Health* 2013; 28(1): 37-58.
26. Vandenberg LN and Prins GS: Clarity in the face of confusion: new studies tip the scales on bisphenol A (BPA). *Andrology* 2016; 4(4): 561-64.
27. Jin P, Wang X, Chang F, Bai Y, Li Y, Zhou R and Chen L: Low dose bisphenol A impairs spermatogenesis by suppressing reproductive hormone production and promoting germ cell apoptosis in adult rats. *Journal of biomedical research* 2013; 27(2): 135.
28. Rochester JR: Bisphenol A and human health: a review of the literature. *Reproductive Toxicology* 2013; 42: 132-155.
29. Wetherill YB, Akingbemi BT, Kanno J, McLachlan JA, Nadal A, Sonnenschein C and Belcher SM: *In-vitro* molecular mechanisms of bisphenol A action. *Reproductive Toxicology* 2007; 24(2): 178-98.
30. Marino M, Pellegrini M, La Rosa P and Acconcia F: Susceptibility of estrogen receptor rapid responses to xenoestrogens: Physiological outcomes. *Steroids* 2012; 77(10): 910-17.
31. Acconcia F, Fiocchetti M and Marino M: Xenoestrogen regulation of ER α /ER β balance in hormone-associated cancers. *Mole and Cell Endocrinology* 2017; 457: 3-12.
32. Thent ZC, Froemming GRA and Muid S: Bisphenol A exposure disturbs the bone metabolism: An evolving interest towards an old culprit. *Life Sci* 2018; 198: 1-7.
33. Vandenberg LN, Chahoud I, Heindel JJ, Padmanabhan V, Paumgarten FJ and Schoenfelder G: Urinary, circulating, and tissue biomonitoring studies indicate widespread exposure to bisphenol A. *Environmental Health Perspectives* 2010; 118(8): 1055-70.
34. Rubin BS: Bisphenol A: an endocrine disruptor with widespread exposure and multiple effects. *The Journal of Steroid Biochemistry and Mole Biol* 2011; 127(1-2): 27-34.
35. Gore AC, Chappell VA, Fenton SE, Flaws JA, Nadal A, Prins GS and Zoeller RT: EDC-2: the endocrine society's second scientific statement on endocrine-disrupting chemicals. *Endocrine Reviews* 2015; 36(6): E1-E150.
36. Srivastava S, Gupta P, Chandolia A and Alam I: Bisphenol A: a threat to human health? *J Env Health* 2015; 77: 20-26.
37. MacKay H and Abizaid A: A plurality of molecular targets: The receptor ecosystem for bisphenol-A (BPA). *Hormones and Behavior* 2018; 101: 59-67.
38. Gassman NR: Induction of oxidative stress by bisphenol A and its pleiotropic effects. *Environmental and Molecular Mutagenesis* 2017; 58(2): 60-71.
39. Rahman T, Hosen I, Towhidul Islam MM and Uddin Shekhar H: Oxidative Stress and Human Health. *Advances in Bioscience and Biotechnology* 2012; 3: 997-1019.
40. Huc L, Lemarié A, Guéraud F and Héliès-Toussaint C: Low concentrations of bisphenol A induce lipid accumulation mediated by the production of reactive oxygen species in the mitochondria of HepG2 cells. *Toxicology in-vitro* 2012; 26(5): 709-717.
41. Rahal A, Kumar A, Singh V, Yadav B, Tiwari R, Chakraborty S and Dhama K: Oxidative stress, pro-oxidants, and antioxidants: the interplay. *Bio Med Research International* 2014; 1-19.
42. Tyl RW, Myers CB, Marr MC, Thomas BF, Keimowitz AR, Brine DR and Joiner RL: Three-generation reproductive toxicity study of dietary bisphenol A in CD Sprague-Dawley rats. *Toxicological Sciences* 2002; 68(1): 121-46.
43. Kabuto H, Hasuike S, Minagawa N and Shishibori T: Effects of bisphenol A on the metabolisms of active oxygen species in mouse tissues. *Environmental Research* 2003; 93(1): 31-35.
44. Oh PS and Lim KT: Blocking of intracellular ROS production by phytylglycoprotein (30 kDa) causes anti-proliferation in bisphenol A-stimulated Chang liver cells. *Journal of Applied Toxicology: An International Journal* 2008; 28(6): 749-58.
45. Hassan ZK, Elobeid MA, Virk P, Omer SA, ElAmin M, Daghestani MH and AlOlayan EM: Bisphenol A induces hepatotoxicity through oxidative stress in rat model. *Oxidative Medicine and Cellular Longevity* 2012; 194829.
46. Bindhumol V, Chitra KC and Mathur PP: Bisphenol A induces reactive oxygen species generation in the liver of male rats. *Toxicology* 2003; 188(2-3): 117-24.
47. Cavalieri EL and Rogan EG: Is bisphenol A a weak carcinogen like the natural estrogens and diethylstilbestrol? *IUBMB Life* 2010; 62(10): 746-51.
48. Barbonetti A, Castellini C, Di Giammarco N, Santilli G, Francavilla S and Francavilla F: *In-vitro* exposure of human spermatozoa to bisphenol A induces pro-oxidative/apoptotic mitochondrial dysfunction. *Reproductive Toxicology* 2016; 66: 61-67.
49. Meeker JD, Calafat AM and Hauser R: Urinary bisphenol A concentrations in relation to serum thyroid and reproductive hormone levels in men from an infertility

- clinic. Environmental Science and Technology 2009; 44(4): 1458-63.
50. Meeker JD, Ehrlich S, Toth TL, Wright DL, Calafat AM, Trisini AT and Hauser R: Semen quality and sperm DNA damage in relation to urinary bisphenol A among men from an infertility clinic. Reproductive Toxicology 2010; 30(4): 532-39.
 51. Kourouma A, Peng D, Chao Q, Changjiang L, Chengmin W, Wenjuan F and Kedi Y: Bisphenol A-induced reactive oxygen species (ROS) in the liver and affect epididymal semen quality in adults Sprague-Dawley rats. Journal of Toxicology and Environmental Health Sciences 2014; 6(4): 103-12.
 52. Chapin RE, Adams J, Boekelheide K, Gray Jr LE, Hayward SW, Lees PS and Vandenberg JG: NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. Birth Defects Research Part B: Developmental and Reproductive Toxicology 2008; 83(3): 157-95.
 53. Song H, Zhang T, Yang P, Li M, Yang Y, Wang Y and Zhang K: Low doses of bisphenol A stimulate the proliferation of breast cancer cells *via* ERK1/2/ERR γ signals. Toxicology *in-vitro* 2015; 30(1): 521-28.
 54. US Environmental Protection Agency: Integrated Risk Information System (IRIS). Bisphenol A (CASRN 80-05-7). Available from www.epa.gov/iris/subst/0356.htm. Accessed 14 December 2010.
 55. Welshons WV, Nagel SC and Vom Saal FS: Large effects from small exposures. III. Endocrine mechanisms mediating effects of bisphenol A at levels of human exposure. Endocrinology 2006; 147(6): s56-s69.
 56. Aguilar F, Autrup H, Barlow S, Castle L, Crebelli R, Dekant W and Gürtler R: Toxicokinetics of bisphenol A scientific opinion of the panel on food additives, flavorings, processing aids and materials in contact with food (AFC). EFSA J 2008; 759: 1-10.
 57. EFSA Panel on Food Contact Materials, Enzymes, Flavorings and Processing Aids (CEF): Scientific Opinion on the risks to public health related to the presence of bisphenol A (BPA) in foodstuffs. EFSA Journal 2015; 13(1): 3978.
 58. Aitken RJ and Baker MA: Oxidative stress, sperm survival and fertility control. Molecular & Cellular Endocrinology 2006; 250(1-2): 66-69.
 59. Qiu W, Chen J, Li Y, Chen Z, Jiang L, Yang M and Wu M: Oxidative stress and immune disturbance after long-term exposure to bisphenol A in juvenile common carp (*Cyprinus carpio*). Ecotoxicology and Environmental Safety 2016; 130: 93-102.
 60. Bisht S and Dada R: Oxidative stress: Major executioner in disease pathology, role in sperm DNA damage and preventive strategies. Front Biosci (Schol Ed) 2017; 9: 420-47.
 61. Koong LY and Watson CS: Rapid, nongenomic signaling effects of several xenoestrogens involved in early- vs. late-stage prostate cancer cell proliferation. Endocrine Disruptors 2015; 3(1): e995003.
 62. Babu S, Uppu S, Claville MO and Uppu RM: Prooxidant actions of bisphenol A (BPA) phenoxyl radicals: implications to BPA-related oxidative stress and toxicity. Toxicology Mechanisms and Methods 2013; 23(4): 273-80.
 63. Moon MK, Kim MJ, Jung IK, Koo YD, Ann HY, Lee KJ and Jang HC: Bisphenol A impairs mitochondrial function in the liver at doses below the no observed adverse effect level. Journal of Korean Medical Science 2012; 227(6): 644-52.
 64. Cooke MS, Evans MD, Dizdaroglu M and Lunec J: Oxidative DNA damage: mechanisms, mutation, and disease. The FASEB Journal 2003; 17(10): 1195-14.
 65. Valko M, Rhodes C, Moncol J, Izakovic MM and Mazur M: Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chemico-biological interactions 2006; 160(1): 1-40.
 66. Wang H, Liu L, Wang J, Tong Z, Yan J, Zhang T and Shen H: Urinary sexual steroids associated with bisphenol A (BPA) exposure in the early infant stage: Preliminary results from a Daishan birth cohort. Science of the Total Environment 2017; 601: 1733-42.
 67. Tiwari D, Kamble J, Chilgunde S, Patil P, Maru G, Kawle, D and Vanage G: Clastogenic and mutagenic effects of bisphenol A: an endocrine disruptor. Mutation Research/Genetic Toxicology and Environmental Mutagenesis 2012; 743(1): 83-90.
 68. Atkinson A and Roy D: *In-vivo* DNA adduct formation by bisphenol A. Environmental and Molecular Mutagenesis 1995; 26(1): 60-66.
 69. Gassman NR, Coskun E, Stefanick DF, Horton JK, Jaruga P, Dizdaroglu M and Wilson SH: Bisphenol A promotes cell survival following oxidative DNA damage in mouse fibroblasts. PloS One 2015; 10(2): e0118819.
 70. Yang YJ, Hong YC, Oh SY, Park MS, Kim H, Leem JH, and Ha EH: Bisphenol A exposure is associated with oxidative stress and inflammation in postmenopausal women. Environmental Research 2009; 109(6): 797-101.
 71. Eid JI, Eissa SM and El-Ghor AA: Bisphenol A induces oxidative stress and DNA damage in hepatic tissue of female rat offspring. The Journal of Basic and Applied Zoology 2015; 71: 10-19.
 72. Tiwari D and Vanage G: Bisphenol A induces oxidative stress in bone marrow cells, lymphocytes, and reproductive organs of Holtzman rats. International Journal of Toxicology 2017; 36(2): 142-52.
 73. Atkinson A and Roy D: *In-vitro* conversion of environmental estrogenic chemical bisphenol A to DNA binding metabolite (s). Biochemical and Biophysical Research Communications 1995; 210(2): 424-33.
 74. Izzotti A, Kanitz S, D'Agostini F, Camoirano A and De Flora S: Formation of adducts by bisphenol A, an endocrine disruptor, in DNA *in-vitro* and in the liver and mammary tissue of mice. Mutation Research/Genetic Toxicology and Environmental Mutagenesis 2009; 679(1): 28-32.
 75. Song S, Song M, Zeng L, Wang T, Liu R, Ruan T and Jiang G: Occurrence and profiles of bisphenol analogs in municipal sewage sludge in China. Environmental Pollution 2014; 186: 14-19.
 76. Moghaddam HS, Samarghandian S and Farkhondeh T: Effect of bisphenol A on blood glucose, lipid profile and oxidative stress indices in adult male mice. Toxicology Mechanisms and Methods 2015; 25(7): 507-13.
 77. Li S, Jin Y, Zhao H, Jiang Y and Cai Z: Evaluation of bisphenol A exposure induced oxidative RNA damage by liquid chromatography-mass spectrometry. Chemosphere 2019.
 78. Lv Y, Lu S, Dai Y, Rui C, Wang Y, Zhou Y and Fan R: Higher dermal exposure of cashiers to BPA and its association with DNA oxidative damage. Environment International 2017; 98: 69-74.
 79. Ferguson KK, Cantonwine DE, McElrath TF, Mukherjee B and Meeker JD: Repeated measures analysis of associations between urinary bisphenol-A concentrations and biomarkers of inflammation and oxidative stress in pregnancy. Reproductive Toxicology 2016; 66: 93-98.

80. Han C and Hong YC: Bisphenol A, hypertension, and cardiovascular diseases: epidemiological, laboratory, and clinical trial evidence. *Curr Hyper Rep* 2016; 18(2): 11.
81. Ezz HSA, Khadrawy YA and Mourad IM: The effect of bisphenol A on some oxidative stress parameters and acetylcholinesterase activity in the heart of male Albino rats. *Cytotechnology* 2015; 67(1): 145-55.
82. Anet A, Olakkaran S, Purayil AK and Puttaswamygowda GH: Bisphenol A induced oxidative stress mediated genotoxicity in *Drosophila melanogaster*. *Journal of hazardous materials* 2018.
83. Leem YH, Oh S, Kang HJ, Kim JH, Yoon J and Chang JS: BPA-toxicity *via* superoxide anion overload and a deficit in β -catenin signaling in human bone mesenchymal stem cells. *Environmental Toxicology* 2017; 32(1): 344-52.
84. Khan S, Beigh S, Chaudhari BP, Sharma S, Aliul Hasan Abdi S, Ahmad S and Raisuddin S: Mitochondrial dysfunction induced by Bisphenol A is a factor of its hepatotoxicity in rats. *Environmental Toxicology* 2016; 31(12): 1922-34.
85. Kourouma A, Quan C, Duan P, Qi S, Yu T, Wang Y and Yang K: Bisphenol A induces apoptosis in liver cells through induction of ROS. *Advances in Toxicology* 2015.
86. Wu HJ, Liu C, Duan WX, Xu SC, He, MD, Chen CH and Chen Y: Melatonin ameliorates bisphenol A-induced DNA damage in the germ cells of adult male rats. *Mutation Research / Genetic Toxicology and Environmental Mutagenesis* 2013; 752(1): 57-67.
87. Zhou C, Wang W, Peretz J and Flaws JA: Bisphenol A exposure inhibits germ cell nest breakdown by reducing apoptosis in cultured neonatal mouse ovaries. *Reproductive Toxicology* 2015; 57: 87-99.
88. Chitra KC, Latchoumycandane C and Mathur PP: Induction of oxidative stress by bisphenol A in the epididymal sperm of rats. *Toxico* 2003; 185(1-2): 119-27.
89. Aschberger K, Castello P, Hoekstra E, Karakitsios S, Munn S, Pakalin S and Sarigiannis D: Bisphenol A and baby bottles: challenges and perspectives. Luxembourg: Publications Office of the European Union 2010; 5-50.
90. Li X, Ying GG, Su HC, Yang XB and Wang L: Simultaneous determination and assessment of 4-nonylphenol, bisphenol A and triclosan in tap water, bottled water and baby bottles. *Environment International* 2010; 36(6): 557-62.
91. Salian S, Doshi T and Vanage G: Perinatal exposure of rats to bisphenol A affects fertility of male offspring-an overview. *Reproductive Toxicology* 2011; 31(3): 359-62.
92. Batista TM, Alonso-Magdalena P, Vieira E, Amaral MEC, Cederroth CR, Nef S and Nadal A: Short-term treatment with bisphenol-A leads to metabolic abnormalities in adult male mice. *PloS One* 2012; 7(3): e33814.
93. Shankar A and Teppala S: Urinary bisphenol A and hypertension in a multiethnic sample of US adults. *Journal of Environmental and Public Health* 2012.
94. Lee HR, Hyun SH, Jeung EB and Choi KC: 193 Bisphenol A and phthalate enhanced the growth of prostate cancer cells and altered TGF- β signaling molecules via an estrogen receptor or androgen receptor-dependent pathway in *in-vitro* and *in-vivo* models. *Reproduction, Fertility and Development* 2012; 25(1): 245-46.
95. Wang HX, Zhou Y, Tang CX, Wu JG, Chen Y and Jiang QW: Association between bisphenol A exposure and body mass index in Chinese school children: a cross-sectional study. *Environmental Health* 2012; 11(1): 79.
96. Zaid SSM, Othman S and Kassim NM: Protective role of *Ficus deltoidea* against BPA-induced impairments of the follicular development, estrous cycle, gonadotropin and sex steroid hormones level of prepubertal rats. *Journal of Ovarian Research* 2018; 11(1): 99.
97. Mínguez-Alarcón L, Gaskins AJ, Chiu YH, Souter I, Williams PL, Calafat AM and EARTH Study Team: Dietary folate intake and modification of the association of urinary bisphenol A concentrations with *in-vitro* fertilization outcomes among women from a fertility clinic. *Reproductive Toxicology* 2016; 65: 104-12.
98. Behmanesh MA, Najafzadehvarzi H and Poormoosavi SM: Protective effect of *Aloe vera* extract against bisphenol a induced testicular toxicity in Wistar rats. *Cell Journal (Yakhteh)* 2018; 20(2): 278.
99. Akingbemi BT, Sottas CM, Koulova AI, Klinefelter GR, and Hardy MP: Inhibition of testicular steroidogenesis by the xenoestrogen bisphenol A is associated with reduced pituitary luteinizing hormone secretion and decreased steroidogenic enzyme gene expression in rat Leydig cells. *Endocrinology* 2004; 145(2): 592-03.
100. Srivastava S, Gupta P, Chandolia A and Alam I: SPECIAL REPORT: Bisphenol A: A threat to human health? *Journal of Environmental Health* 2015; 77(6): 20-27.
101. Wisniewski P, Romano RM, Kizys MM, Oliveira KC, Kasamatsu T, Giannocco G and Romano MA: Adult exposure to bisphenol A (BPA) in Wistar rats reduces sperm quality with disruption of the hypothalamic-pituitary-testicular axis. *Toxicology* 2015; 329: 1-9.
102. Vom Saal FS, Cooke PS, Buchanan DL, Palanza P, Thayer KA, Nagel SC and Welshons WV: A physiologically based approach to the study of bisphenol A and other estrogenic chemicals on the size of reproductive organs, daily sperm production, and behavior. *Toxicology and Industrial Health* 1998; 14(1-2): 239-60.
103. Salian S, Doshi T and Vanage G: Perinatal exposure of rats to Bisphenol A affects the fertility of male offspring. *Life Sciences* 2009; 85(21-22): 742-52.
104. Yang YJ, Hong YP and Chae SA: Reduction in semen quality after mixed exposure to bisphenol A and isobutylparaben in utero and during lactation periods. *Human & Experimental Toxicology* 2016; 35(8): 902-11.
105. Tinwell H, Haseman J, Lefevre PA, Wallis N and Ashby J: Normal sexual development of two strains of rat exposed in utero to low doses of bisphenol A. *Toxicological Sciences* 2002; 68(2): 339-348.
106. Kato H, Furuhashi T, Tanaka M, Katsu Y, Watanabe H, Ohta Y and Iguchi T: Effects of bisphenol A given neonatally on reproductive functions of male rats. *Reproductive Toxicology* 2006; 22(1): 20-29.
107. Howdeshell KL, Furr J, Lambright CR, Wilson VS, Ryan, BC and Gray JLE: Gestational and lactational exposure to ethinyl estradiol, but not bisphenol A, decreases androgen-dependent reproductive organ weights and epididymal sperm abundance in the male long evans hooded rat. *Toxicological Sciences* 2007; 102(2): 371-82.
108. Delclos KB, Camacho L, Lewis SM, Vanlandingham M, Latendresse JR, Olson GR and Bryant MS: Toxicity evaluation of bisphenol A administered by gavage to Sprague Dawley rats from gestation day 6 through postnatal day 90. *Toxicological Sci* 2014; 139(1): 174-97.
109. Johnson SA, Javurek AB, Painter MS, Ellersieck MR, Welsh JTH, Camacho L and Rosenfeld CS: Effects of developmental exposure to bisphenol A on spatial navigational learning and memory in rats: A CLARITY-BPA study. *Hormones and Behavior* 2016; 80: 139-48.
110. Rahman MS, Kwon WS, Lee JS, Yoon SJ, Ryu BY and Pang MG: Bisphenol-A affects male fertility via fertility-related proteins in spermatozoa. *Sci Rep* 2015; 5: 9169.

111. Cheong A, Zhang X, Cheung YY, Tang WY, Chen J, Ye SH and Ho SM: DNA methylome changes by estradiol benzoate and bisphenol A links early-life environmental exposures to prostate cancer risk. *Epigenetics* 2016; 11(9): 674-689.
112. Wu J, Huang D, Su X, Yan H and Sun Z: Oral administration of low-dose bisphenol A promotes proliferation of ventral prostate and upregulates prostaglandin D2 synthase expression in adult rats. *Toxicology and industrial health* 2016; 32(11): 1848-58.
113. Caserta D, Di Segni N, Mallozzi M, Giovanale V, Mantovani A, Marci R and Moscarini M: Bisphenol A and the female reproductive tract: an overview of recent laboratory evidence and epidemiological studies. *Reproductive Biology and Endocrinology* 2014; 12(1): 37.
114. Pacchierotti F, Ranaldi R, Eichenlaub-Ritter U, Attia S and Adler ID: Evaluation of arogenic effects of bisphenol A in somatic and germ cells of the mouse. *Mut Res/Gen Toxico and Environ Mutagenesis* 2008; 651(1): 64-70.
115. Hunt PA, Koehler KE, Susiarjo M, Hodges CA, Ilagan A, Voigt RC and Hassold TJ: Bisphenol A exposure causes meiotic aneuploidy in the female mouse. *Current Biology* 2003; 13(7): 546-53.
116. Hunt PA, Lawson C, Gieske M, Murdoch B, Smith H, Marre A and Voort CAV: Bisphenol A alters early oogenesis and follicle formation in the fetal ovary of the rhesus monkey. *Proceedings of the National Academy of Sciences* 2012; 109(43): 17525-30.
117. Zhang T, Li L, Qin XS, Zhou Y, Zhang XF, Wang LQ and Shen W: Di-(2-ethylhexyl) phthalate and bisphenol A exposure impairs mouse primordial follicle assembly *in-vitro*. *Environ and Mole Mutagen* 2014; 55(4), 343-53.
118. Zhang Y, Gao J, Xu P, Yuan C, Qin F, Liu S and Wang Z: Low-dose bisphenol A disrupts gonad development and steroidogenic genes expression in adult female rare minnow *G. rarus*. *Chemosphere* 2014 112: 435-42.
119. Brieno-Enriquez MA, Robles P, Camats-Tarruella N, Garcia-Cruz R, Roig I, Cabero L and Caldés MG: Human meiotic progression and recombination are affected by Bisphenol A exposure during *in-vitro* human oocyte development. *Human Repro* 2011; 26(10): 2807-18.
120. Ganesan S and Keating AF: Bisphenol A-induced ovotoxicity involves DNA damage induction to which the ovary mounts a protective response indicated by increased expression of proteins involved in DNA repair and xenobiotic biotransformation. *Toxicological Sciences* 2016; 152(1): 169-80.
121. Berger RG and Shaw J: Impact of acute bisphenol-A exposure upon intrauterine implantation of fertilized ova and urinary levels of progesterone and 17 β -estradiol. *Reproductive Toxicology* 2008; 26(2): 94-99.
122. Xi W, Lee CKF, Yeung WSB, Giesy JP, Wong MH, Zhang X and Wong CK: Effect of perinatal and postnatal bisphenol A exposure to the regulatory circuits at the hypothalamus-pituitary-gonadal axis of CD-1 mice. *Reproductive Toxicology* 2011; 31(4): 409-17.
123. La Rocca C, Tait S, Guerranti C, Busani L, Ciardo F, Bergamasco B and Bordi G: Exposure to endocrine disrupters and nuclear receptor gene expression in infertile and fertile women from different Italian areas. *International Journal of Environmental Research and Public Health* 2014; 11(10): 10146-64.
124. Takeuchi T, Tsutsumi O, Ikezuki Y, Takai Y and Taketani Y: Positive relationship between androgen and the endocrine disruptor, bisphenol A, in normal women and women with ovarian dysfunction. *Endocrine journal* 2004; 51(2): 165-69.
125. Cobellis L, Colacurci N, Trabucco E, Carpentiero C and Grumetto L: Measurement of bisphenol A and bisphenol B levels in human blood sera from healthy and endometriotic women. *Biomedical Chromat* 2009; 23(11): 1186-90.
126. Kandaraki E, Chatzigeorgiou A, Livadas S, Palioura E, Economou F, Koutsilieris M and Diamanti-Kandarakis E: Endocrine disruptors and polycystic ovary syndrome (PCOS): elevated serum levels of bisphenol A in women with PCOS. *The Journal of Clinical Endocrinology and Metabolism* 2011; 96(3): E480-E484.
127. Cantonwine DE, Meeker JD, Ferguson KK, Mukherjee B, Hauser R and McElrath TF: Urinary concentrations of bisphenol A and phthalate metabolites measured during pregnancy and risk of preeclampsia. *Environmental health perspectives* 2016; 124(10): 1651.
128. Wang Y, Zhu Q, Dang X, He Y, Li X and Sun Y: Local effect of bisphenol A on the estradiol synthesis of ovarian granulosa cells from PCOS. *Gynecological Endocrinology* 2017; 33(1): 21-25.
129. Patel CJ, Yang T, Hu Z, Wen Q, Sung J, El-Sayed YY and Ling XB: Investigation of maternal environmental exposures in association with self-reported preterm birth. *Reproductive Toxicology* 2014; 45: 1-7.
130. Li DK, Zhou Z, Miao M, He Y, Wang J, Ferber J and Yuan W: Urine bisphenol-A (BPA) level in relation to semen quality. *Fertility and Sterility* 2011; 95(2): 625-30.
131. Vitku J, Heracek J, Sosvorova L, Hampl R, Chlupacova T, Hill M and Starka L: Associations of bisphenol A and polychlorinated biphenyls with spermatogenesis and steroidogenesis in two biological fluids from men attending an infertility clinic. *Environment International* 2016; 89: 166-73.
132. Vom Saal FS, Nagel SC, Coe BL, Angle BM and Taylor JA: The estrogenic endocrine-disrupting chemical bisphenol A (BPA) and obesity. *Molecular and Cellular Endocrinology* 2012; 354(1-2): 74-84.
133. Hu W, Dong T, Wang L, Guan Q, Song L, Chen D and Wang X: Obesity aggravates toxic effect of BPA on spermatogenesis. *Environment Inter* 2017; 105: 56-65.
134. Srivastava S and Gupta P: Genotoxic and infertility effects of bisphenol A on Wistar albino rats. *Int J Pharm Sci Rev Res* 2016; 41(1): 126-31.
135. Karnam SS, Ghosh RC, Mondal S and Mondal M: Evaluation of subacute bisphenol A toxicity on male reproductive system. *Veterinary World* 2015; 8(6): 738.
136. Hass U, Christiansen S, Boberg J, Rasmussen MG, Mandrup K and Axelstad M: Low-dose effect of developmental bisphenol A exposure on sperm count and behavior in rats. *Andrology* 2016; 4(4): 594-07.

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