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ANTIBIOTIC RESISTANCE ISSUES IN CLINICAL PATHOGENS: A REVIEW

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ABSTRACT: Antibiotic resistance in pathogens is on the increase but in the past few decades this problem raised a deepening concern among scientific community in terms of the clinical management of infectious diseases. In this review, the common gram negative clinically significant pathogenic bacteria who have acquired resistance with passage of time like *Shigella dysenteriae*, *Sh. Boydii*, *Sh. Flexneri*, *Listeria monocytogenes*, *Acinetobacter baumannii*, *acinetobacter*, *A. baumannii*, *Morganella morganii*, *Enterobacteriaceae*, *Citrobacter freundii*, *Citrobacter braakii*, or *Citrobacter amalonaticus*, *P. stuartii*, *Providencia rettgeri*, *P. alcalifaciens*, *P. rustigianii*, *E. coli*, *Salmonella paratyphi*, *S. typhi*, *Salmonella enterica*, have been compiled with their plausible ways of causing resistance. We conclude from our literature review that resistance in gram negative bacteria is on the rise and new and advanced antibiotics need to be designed and introduced.

INTRODUCTION: Antibiotics literally means “against life”; in this case, against microbes. There are many types of antibiotics such as antibacterials, antivirals, antifungals, and antiparasitics. Some drugs are effective against many organisms; these are called broad-spectrum antibiotics.

Others are effective against just a few organisms and are called narrow spectrum antibiotics. The most commonly used antibiotics are antibacterial in nature¹. Ehrlich made the first antibiotic called Salvarsan, now known as Arsphenamine^{2,3,4}.

Later, the science of antibiotics entered into an age of progress and at the time of penicillin discovery it was thought that soon we would have an upper hand over pathogenic microbes. However in recent past several studies have demonstrated authentically that now these disease causing agents are developed resistance to most of these antibiotics at an alarmingly faster rate and the race once we thought would soon be over after penicillin discovery in 1928 seems had just started⁵.

Resistance Mechanisms: There are several mechanisms by which a microbe can get resistant to a particular antibiotic however among them the most studied and frequent one is the horizontal gene transfer between microbes⁶. The antibiotic action against the pathogen can be seen as an environmental pressure; those bacteria which have a mutation allowing them to survive will live on to reproduce. They will then pass this trait to their offspring, which will result in the evolution of a fully resistant colony.



Other four major type of resistance mechanism are; drug inactivation or modification, alteration of target site, alteration of metabolic pathway and reduced drug accumulation⁷⁻⁸.

Some gram negative bacteria involved in diahorreal infections: As in case of *Shigella* infection yet claims for a significant proportion of bacillary dysentery in many tropical and subtropical countries and review of research papers shows the global rise in the growing pattern of resistance of *Shigella* species particularly type-I strains that poses great problem not in antibacterial therapy but also in clinical management all over the world⁷. In Somalia, a studied conducted in which the resistance to conventional antibiotics of 240 *Shigella* strains (like ampicillin, chloramphenicol, streptomycin, tetra cycline, and sulfonamides) and particularly *Shigella dysenteriae* type-I were found to be resistant to more than one drug.

The study also suggested that polymyxin B or M sulfate could be used as they had found effective in in-vivo trails for which have proved to be effective for bacillary dysentery¹⁰. A similar but a bit at large scale study conducted in London and Wales in which two thousand three hundred and seventy strains of *Sh. dysenteriae*, *Sh. flexneri*, and *Sh. boydii* were tested for resistance to a panel of a dozen of antimicrobial drugs and eighty per cent of strains were resistant to one or more drugs, with significantly sulphonamide resistance was observed, except in *Sh. Soneii*⁹.

Another study in Central Isreal over a period of approximately 10 years difference indicated the significantly increased resistance to tetracycline from 23% to 87%, and high resistance to trimethoprim-sulfamethoxazole (94%) and ampicillin (85%) were also noted⁹. In Nepal, a study was conducted with the same aim a couple of years and results indicated that resistance to cotrimoxazole was highest (80.7%), followed by tetracycline (74.7%), gentamicin (55.4%), ampicillin (53%), chloramphenicol (39.7%) and nalidixic acid (31.3%)¹². In a vast study conducted in Pakistan, where, 79.3% *S. typhi* and 59.9% *S. paratyphi* A were isolated from patients under 15 years of age, with the MDR rate increased in *S. typhi* (34.2 to 48.5% p<0.001), but decreased in *S. paratyphi* A (44.5 to 8.6%) in the first line of drugs.

However, Quinolone resistance (MIC>1µg/ml) increased in both *S. typhi* (1.6 to 64.1%) and *S. paratyphi* A (0 to 47%) in older patients. Apart from endemic and sporadic cases, and MDR strains have been responsible for numerous outbreaks on the Asian continent. The Asian isolates, eighteen of 25 isolates of *Salmonella enterica* serovar Typhi were multidrug resistant and contained class 1 integrons with a single cassette, *dfrVII* or *aadA1* on an IncHI1 plasmid. *Salmonella* serovar Typhi could become resistant to broad-spectrum cephalosporins by integrating cassettes, such as *veb-1*, a commntegron-Associated Antibiotic Resistance in *Salmonella enterica* Serovar Typhi from Asia¹³.

In an extensive molecular study, seventy-eight *Salmonella typhi* strains isolated in 1994 and 1995 from patients living in Dhaka, Bangladesh, were subjected to phage typing, ribotyping, IS200 fingerprinting, and PCR fingerprinting. The data indicated a significant number of the *S. typhi* strains (67%) were demonstrated to be multiple drug resistant (MDR) strains were resistant to chloramphenicol, ampicillin, trimethoprim, streptomycin, sulfamethoxazole, and tetracycline (R type CATmSSuT), that could be conjugated to *Escherichia coli* and resulted in the complete transfer of the MDR phenotype¹⁴.

High resistance, most probably plasmid mediated a serious public health concern and could be responsible for this increased mortality¹⁵. In a study conducted, *L. monocytogenes* was isolated in 4.5% of fish samples and 8.3% of seawater samples. Multi-resistant environmental strains as 6% of *L. monocytogenes* strains isolated showed multi-resistance to ampicillin, erythromycin, tetracycline, dicloxacillin, and trimethoprim-sulfamethoxazole, that a serious threat to human health¹⁶.

Resistant Nosocomial Infection, A Potential Risk for Health Care Professionals: *Acinetobacter baumannii*, is a worldwide nosocomial pathogen associated with opportunistic infections, community and healthcare-associated infections (HAIs) like pneumonia, urinary tract, bloodstream, skin and soft tissue infections¹⁷. Thus, it constitutes a major public health problem due to its propensity to develop resistance to numerous drugs¹⁸. A study was conducted to determine the possible resistant genes in case of MDR acinetobacter starins and found about eight resistance gene determinants the genes

*bla*OXA-23 and *ampC*, resistance to carbapenems and cephalosporins, respectively. However, resistance to quinolones and fluoroquinolones was conferred by an S83L mutation in GyrA and *bla*TEM-1, which was found, associated to β -lactam resistance, and *strB*, which contributed to aminoglycoside resistance that give rise to the MDR phenotype both in sporadic and epidemic cases¹⁹.

A similar research on a tertiary care 1000 bed hospital in Iran was carried out the results indicated that the *A. baumannii* strains showed high rate of resistance to ceftriaxone (90.9%), piperacillin (90.9%), ceftazidime (84.1%), amikacin (85.2%) and ciprofloxacin (90.9%) however, Imipenem was the most effective antibiotic against *A. baumannii* strains and the rate of resistance for imipenem was 4.5%²⁰ among various uro-pathogens responsible for urinary infections.

The natural antibiotic susceptibility of 38 *Providencia rettgeri*, 35 *P. stuartii*, 23 *P. alcalifaciens* and 20 *P. rustigianii* strains was examined. MIC values were determined by a microdilution procedure and evaluated by a table calculation programme. *P. stuartii* was the least susceptible *Providencia* sp. and was naturally resistant to tetracyclines, some penicillins, older cephalosporins, sulphamethoxazole and fosfomycin and to antibiotics to which other species of Enterobacteriaceae are also resistant. It was naturally sensitive to modern penicillins and cephalosporins, carbapenems and aztreonam²¹.

In one of the study on multidrug-resistant Enterobacteriaceae (MDRE) in solid-organ transplant patients, as often in compromised hosts, resistant to cephalosporins due to overexpression of their chromosomal β -lactamase found that almost half of patients colonized with MDRE carried one or more cefpodoxime-resistant. *Citrobacter* species are infrequent nosocomial pathogens, and can cause a number of infections due to impairment of immune system²² and include urinary tract infections, neonatal sepsis, brain abscess, meningitis, bloodstream infections²³. *Citrobacter freundii*, *Citrobacter braakii*, or *Citrobacter amalonaticus* strains²⁴.

Our study shows that *qnr* gene has occurred in *Citrobacter freundii* isolates from Anhui Province, China. *qnr* gene was therefore present in both

quinolone-resistant and -susceptible isolates and some of them could be transferred by conjugation experiments. *qnr* positive isolates strains showed multi-resistance and no clone was spread found in these SHV- and TEM-derived extended-spectrum β -lactamases (ESBLs) have also been described for *Citrobacter* species in the context of outbreaks of clonal strains and plasmids²⁵.

Resistance Cases with some recent clinical importance: *Morganella morganii* is also involved in a number of clinical cases. The different strains of this species are usually resistant to ampicillin, to the amoxicillin-clavulanic acid combination, and to cephalothin, and usually they are susceptible to other antibiotics active against gram-negative bacilli²⁶. In some studies, the potential reason behind the resistance his contribution *M. morganii* is the production of extended-spectrum β -lactamases (ESBL) to drug resistance was examined in *Morganella morganii*²⁷ a high-molecular-weight (49,000) class I cephalosporinase, which contributes in the resistance with ampicillin, carbenicillin, and and broad-spectrum cephalosporins²⁸.

In the same way, in the hospital of Portugal, a strain of *M. morganii* (FFLM15), isolated from the urine of a neonate, showed resistance to ceftazidime and aztreonam and reduced susceptibility to cefotaxime by the disk diffusion method, however the synergistic results between expanded-spectrum cephalosporins and clavulanic acid was performed, and the positive result indicated the presence of an ESBL producer²⁹. Another pathogen, *Citrobacter* species also are infrequent nosocomial pathogens, and can cause a number of infections due to impairment of immune system and cause urinary tract infections, neonatal sepsis, brain abscess, meningitis, bloodstream infections.

In one of the study on multidrug-resistant Enterobacteriaceae (MDRE) in solid-organ transplant patients, resistant to cephalosporins due to overexpression of their chromosomal β -lactamase have been noticed and almost half of patients were also resistant with one or more cefpodoxime-resistant *Citrobacter freundii*, *Citrobacter braakii*, or *Citrobacter amalonaticus* strains^{22, 23}. Through research, *qnr* gene found in *Citrobacter freundii* isolates from Anhui Province, China associated with quinolone-resistance and some of them could be transferred by conjugation trials.

Upon screening, plasmid association with resistance, SHV- and TEM-derived extended-spectrum β -lactamases (ESBLs) have also been noted for *Citrobacter* species in case of the outbreaks of clonal strains and plasmids. The susceptibility profile of 38 clinical isolates of *Providencia rettgeri*, 35 *P. stuartii*, 23 *P. alcalifaciens* and 20 *P. rustigianii* strains was examined by a microdilution procedure. *P. stuartii* was the least susceptible among *Providencia* sp. and was naturally resistant to tetracyclines²³. A total of 238 isolates of *Providencia stuartii* obtained from infected patients in six Dublin hospitals. Both chromosome-encoded and plasmid-coded resistance mechanisms were important and most of the clinical isolates were resistant to several of these antibacterial agents including tetracycline, resistance to penicillin, moreover resistance to polymyxin was also noticed²³.

A total of 238 isolates of *Providencia stuartii* obtained from infected patients in six Dublin hospitals were grouped by using serological and bacteriocin typing methods and tested for sensitivity to a number of antimicrobial agents. Most isolates were resistant to several of these agents. Resistance to tetracycline, resistance to penicillin, resistance to polymyxin, and probably resistance to nitrofurantoin was intrinsic. Plasmid screening coupled with resistance transfer studies showed that both chromosome-encoded and plasmid-coded resistance mechanisms were clinically important. Ampicillin resistance was both chromosomally and plasmid encoded, whereas resistance to kanamycin and resistance to carbenicillin were exclusively plasmid encoded³⁰.

CONCLUSION: Therefore to encounter these defense mechanisms of resistance, from the pathogens, we need to keep discovering new antibiotics against them so as to cure the diseases. However at the moment perhaps, the biggest problem with antibiotic resistance at the moment is the fact that it's inevitable i.e. no matter how exquisite a new drug is, microbes are going to develop resistance to it in one way or another and yet this matter does not get the kind of attention that other health treats like dengue and bird flu did. Perhaps the biggest obstacle in tackling resistance is the fact that too many people accept it as inevitable.

As Weber explained, "even if drugs were used exquisitely perfectly, the emergence of drug resistance is likely." Also, this problem does not receive much attention compared with other health issues-recent scares, such as severe acute respiratory syndrome (SARS), avian flu and bioterrorism, have clearly overshadowed the distant possibility of bacterial resistance with the short-term fear of imminent danger. However, Levy, who highlighted the dangers of antibiotic resistance more than a decade ago in his book *The Antibiotic Paradox*, feels that public awareness, is slowly increasing. "The message is out there," he said. The next step is overcoming resistance to change.

REFERENCES:

1. Immunizations & Infectious Diseases: An Informed Parent's Guide (Copyright © 2006 American Academy of Pediatrics)
2. Limbird LE: The receptor concept: a continuing evolution. *Molecular Intervention* 2004; 4 (6): 326–36.
3. Bosch F, Rosich L: The contributions of Paul Ehrlich to pharmacology: a tribute on the occasion of the centenary of his Nobel Prize. *Pharmacology* 2008; 82 (3): 17-19.
4. Calderon CB, Sabundayo BP: Antimicrobial Classifications: Drugs for Bugs. In Schwalbe R, Steele-Moore L, Goodwin AC. *Antimicrobial Susceptibility Testing Protocols*. CRC Press. Taylor & Frances group 2007; ISBN 978-0-8247-4100-6.
5. Leimane V. Clinical outcome of individualized treatment of multidrug-resistant tuberculosis in Latvia: a retrospective cohort study. *Lancet* 2005; 365: 318-326.
6. Ochiai K, Yamanaka T, Kimura K, Sawada O: Inheritance of drug resistance (and its transfer) between *Shigella* strains and between *Shigella* and *E.coli* strains (in Japanese). *Hihon Iji Shimpor* 1959; 34: 1861.
7. Li X, Nikadio H: Efflux-Mediated Drug Resistance in Bacteria: an Update. *Drug* 2009; 69 (12): 1555–623.
8. Morita Y, Kodama K, Shiota S, Mine T, Kataoka A, Mizushima T, Tsuchiya T: NorM, a Putative Multidrug Efflux Protein, of *Vibrio parahaemolyticus* and Its Homolog in *Escherichia coli*. *Antimicrobial Agents Chemotherapy* 1998; 42 (7): 1778–82.
9. Shai A, Itzhak L, Vered K and Zmira S: Growing antimicrobial resistance of *Shigella* isolates. *Journal of Antimicrobial Chemotherapy* 2003; 51(2): 427-429.
10. E. Mero: Resistance to antibiotics of *Shigella* strains isolated in Somalia. *Bulletin World Health Organization* 1976; 54(4): 473–474.
11. Gross RJ, Rowe B, Cheasty T, Thomas LV: Increase in drug resistance among *Shigella dysenteriae*, *Sh flexneri*, and *Sh.boydii*. *British Medical Journal (Clinical Research Ed)* 1981; 283(6291):575-6.
12. Godwin W, Joshy ME, Chiranjoy M and Shivananda PG: Isolation & antimicrobial susceptibility of *Shigella* from patients with acute gastroenteritis in western Nepal. *Indian Journal of Medical Research* 2006; 123:145-150.
13. Hasan R, Zafar A, Abbas Z, Mahraj V, Malik F, Zaidi A: Antibiotic resistance among *Salmonella enterica* serovars Typhi and Paratyphi A in Pakistan (2001-2006). *Journal of Infection in Developing Countries* 2008; 2(4): 289-294.

14. Threlfall EJ: Antimicrobial drug resistance in *Salmonella*: problems and perspectives in food- and water-borne infections. *FEMS Microbiology Review* 2002; 26:141-148.
15. Charpentier E and Courvalin P: *Antibiotic resistance in Listeria spp.* Antimicrobial Agents and Chemotherapy 1999; 43:2103-2108.
16. Rodas-Suárez OR, Flores-Pedroche JF, Betancourt-Rule JM, Quiñones-Ramírez EI and Vázquez-Salinas C: Occurrence and Antibiotic Sensitivity of *Listeria monocytogenes* Strains Isolated from Oysters, Fish, and Estuarine Water. *Applied and Environmental Microbiology* 2006; 72(11): 7410-7412.
17. Perez F: Global challenge of multidrug-resistant *Acinetobacter baumannii*. *Antimicrobial Agents Chemotherapy* 2007; 51:3471-3484
18. Fournier PE, Richet H: The epidemiology and control of *Acinetobacter baumannii* in health care facilities. *Clinical Infectious Diseases* 2006; 42:692-9.
19. Jennifer KM, Kim M, Pham J, Tapsall J, White PA: Antibiotic resistance determinants in nosocomial strains of multidrug-resistant *Acinetobacter baumannii*. *Journal of Antimicrobial and Chemotherapy* 2009; 63(1): 47-54.
20. Rahbar M, Mehrgan H, Aliakbari NH: Prevalence of antibiotic-resistant *Acinetobacter baumannii* in a 1000-bed tertiary care hospital in Tehran, Iran. *Indian Journal of Pathology and Microbiology* 2010; 53(2)290-293.
21. Stock I, Wiedemann B: Natural antibiotic susceptibility of *Providencia stuartii*, *P. rettgeri*, *P. alcalifaciens* and *P. rustigianii* strains. *Journal of Medical Microbiology* 1998; 47(7):629-42.
22. Lu CH, Chang WN, Chuang YC and Chang HW: Gram-negative bacillary meningitis in adult post-neurosurgical patients. *Surgical Neurology* 1999; 52:438-444.
23. Tellez I, Chrysant GS, Omer I and Dismukes WE: *Citrobacter diversus* endocarditis. *American Journal of Medical Sciences* 2000; 320:408-410.
24. Champs DC, Sirot D, Chanal C, Poupart MC, Dumas MP, and Sirot J: Concomitant dissemination of three extended-spectrum beta-lactamases among different Enterobacteriaceae isolated in a French hospital. *Journal of Antimicrobial Chemotherapy* 1991; 27:441-457.
25. Neuwirth C, Siebor E, Lopez J, Pechinot A and Kazmierczak A: Outbreak of TEM-24-producing *Enterobacter aerogenes* in an intensive care unit and dissemination of the extended-spectrum beta-lactamase to other members of the family Enterobacteriaceae. *Journal of Clinical Microbiology* 1996; 34:76-79.
26. Monnet D and Richard C: Autres Enterobacteriaceae. in Manuel de bacteriologie clinique, eds Frenay J, Renaud F, Hansen W, Bollet C. (E. S. Elsevier, Paris, France), Edition 2, 1994, 1053-1128.
27. Yang YJ and Livermore DM: Chromosomal beta-lactamase expression and resistance to beta-lactam antibiotics in *Proteus vulgaris* and *Morganella morganii*. *Antimicrobial Agents and Chemotherapy* 1988; 32(9):1385-1391.
28. Coudron PE, Moland ES, Sanders CC: Occurrence and detection of extended-spectrum beta-lactamases in members of the family Enterobacteriaceae at a Veterans medical center: seek and you may find. *Journal of Clinical Microbiology* 1997; 35:2593-2597.
29. Barroso H, Freitas-Vieira A, and Duarte A: Molecular Characterization of a Ceftazidime-Resistant *Morganella morganii* Isolate Producing a TEM-10 β -Lactamase. *Antimicrobial Agents and Chemotherapy* 1999; 43(2): 434-435.
30. McHale PJ, Keane CT, and Dougan G: Antibiotic resistance in *Providencia stuartii* isolated in hospitals. *Journal of Clinical Microbiology* 1981; 13(6):1099-104.

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