EFFECT OF *GYMNEMA SYLVESTRE* ON THE PHARMACOKINETICS OF SITAGLIPTIN PHOSPHATE IN TYPE II DIABETES MELLITUS

S. R. Dhande *, D. V. Lokegaonkar and S. P. Bhutkar

Bharati Vidyapeeth’s College of Pharmacy, Sector - 8, C.B.D. Belapur, Navi Mumbai - 400614, Maharashtra, India.

**ABSTRACT:** Practising complementary and alternative medicinal therapy has become a boom worldwide. There is a dearth in communication between doctors and the patients regarding concomitant use of complementary medicines like ayurvedic churnas and other drugs of natural origin with allopathic drugs. This usage might magnify or oppose the effect of synthetic drug leading to interactions. Use of herbals along with allopathic treatment is inevitable in treatment of chronic diabetes which might lead to drug interaction. The present study was undertaken to evaluate pharmacokinetic interaction between herb *Gymnema sylvestre* and allopathic drug Sitagliptin Phosphate in streptozotocin and high fat diet-induced type II diabetes mellitus in rat model. Female Sprague Dawley rats were grouped into following groups. Group 1: Vehicle Control (VC), Group 2: Disease Control (DC), Group 3: Sitagliptin Phosphate (STG) (20 mg/kg), Group 4: *Gymnema sylvestre* (GYM) (400 mg/kg), Group 5: Sitagliptin Phosphate (20 mg/kg) and *Gymnema sylvestre* (400 mg/kg) (STG + GYM). After giving proper treatment to the rats, blood samples were withdrawn from retro-orbital plexus at 0th, 2nd, 4th, 8th and 12th hours on first day of treatment with Sitagliptin Phosphate and combination of Sitagliptin Phosphate with *Gymnema sylvestre* in order to check alteration in absorption parameter by determining the change in area under curve (AUC) of STG, using a newly developed and validated HPTLC method. It was found that, AUC of STG gradually decreased in presence of *Gymnema sylvestre* at the given dose. Further studies need to be carried out at various other doses to evaluate possible interactions.

**INTRODUCTION:** Due to the increasing prevalence of diabetes at an alarming rate, the search for a holistic approach in treatment, and the attitude that some patients have about synthetic medicines are leading individuals to embrace alternative therapies. The accelerating trend and inclining mind set of the population towards usage of as CAM (Complementary Alternative Medicine) is well documented, especially among diabetic patients.

Although herbal medicines are widely used across the globe for prevention, treatment and management of disease, their standardisation, quality control and regulations still possess a great demur. Herbal products which lack quality control and standardisation checks, physicochemical testing of raw materials as well processed formulations may prove to be of great threat when consumed by an individual. In addition to this, the insufficient *in vivo* pharmacokinetic and pharmacodynamic data of the bioactives might rise complexities.

A major factor impeding the therapeutic utilisation of medicinal plants is the lack of information on various herb-drug interactions.
Except for the use and pharmacological activity of
the plant, not much information has been available
on their safety, mechanism of interactions with
other medicines. Gathering all the necessary
data has become utmost important in this area as
millions and millions of people consume herbal
drugs concomitantly along with prescription drugs.
Herbal medicines also follow the modern
pharmacological principles unlike synthetic drugs.
Hence, the herbal drug interactions are based on the
same pharmacokinetic and pharmacodynamic
mechanisms as drug–drug interaction. Drug-drug
or herb-drug interactions can occur in several
different ways. Pharmacodynamic interactions
occur when the object drug’s effect is altered by the
interfering drug or herb.

These interactions are not due to an alteration in the
plasma concentration of either drug but rather
because of the net effect that can be additive,
synergistic or antagonistic. These adjectives can
refer to alteration in the object drug's intended
therapeutic effect, or can refer to the change in the
toxicity levels and adverse side-effects as well.
On the other hand, pharmacokinetic interactions
witness changes in parameters like absorption,
distribution, metabolism or elimination of the
object drug due to the presence of the interfering
drug. Unlike pharmacodynamic interactions, these
interactions do result in changes in the plasma
concentration of the object drug, and as a
consequence, the toxic or sub-therapeutic levels
occur more frequently.

**Reported Herb-drug Interaction with medicinal
plants used in diabetes:** Herbs used in treating
diabetes have been reported to interact with some
of the prescription drugs. The interactions
mentioned either prove to be useful or deleterious
depending upon the mechanism of action of both
the herb as well as the drug. Few interactions
proved to be a boon to the patients consuming
herbs along with prescribed drugs, while some led
to deleterious outcomes. Bitter melon is said to
have a positive outcome in controlling diabetes.
Fenugreek is reported to have synergistic effect in
controlling hyperglycaemia and hyperlipidaemia
when given along with Sitagliptin Phosphate.
Garlic preparations have been found to increase the
bioavailability of Propranolol.

Onion consumption reduces oxidative stress which
proves to be great for the human body. Concomitant use of Ashwagandha along with
anxiolytics and anti-stressors lead to synergistic effects. Several reports have highlighted garlic’s
potential for increasing the risk of postoperative
bleeding if consumed along with NSAID’s. A
clinical trial suggested that garlic changed some
pharmacokinetic of Paracetamol. Animal and
clinical studies imply hypoglycaemic effects, which
could explain the fall in glucose levels in patients
taking chlorpropamide by producing active
nitrogen compounds. Garlic tend to reduce the
efficacy of anti-AIDS drugs like Saquinavir.

**Rationale behind the study:** Type II diabetes
mellitus is generally associated with alteration in
the lipid levels. There are various treatments
available for the same; be it the traditional
complementary medicines as well as allopathic
drugs. There might be interaction taking place
when both therapies are practiced concomitantly.
On the basis of folklore use and literature, the
following herb and drug of synthetic origin were
selected for the interaction study.

The antidiabetic activity of Gymnema sylvestre has
been documented in ayurvedic books and also
published in the literature, but very few literatures
are available to confirm the safety of consuming the herb along with oral antidiabetic drugs. Literature survey revealed few interactions of Gymnema sylvestre along with oral antidiabetic drugs. For example:

- Consumption of Gymnema sylvestre along with antidiabetic drug Metformin hydrochloride showed pharmacokinetic interaction at absorption and elimination level. The observations showed that Gymnema sylvestre contributed to a significant decrease in the bioavailability of Metformin, thus decreasing its hypoglycemic effect\(^{13,14}\).

- The interaction appeared to be a pharmacokinetic interaction at absorption, elimination yet again when Gymnema sylvestre was concomitantly administered to rats along with Gliclazide. The herb inhibited the absorption of Gliclazide which resulted in a significant decrease in the bioavailability of the latter\(^{15,16}\).

- *In-silico* docking studies have sighted probable interaction if Gymnema sylvestre is concomitantly consumed with Glimepiride\(^{17}\).

Since no studies were performed on evaluating the interaction of consumption of Gymnema sylvestre along with drug from new class of DPP-IV inhibitors i.e. Sitagliptin Phosphate, these two were selected as herb and drug respectively for studying herb-drug interaction at pharmacokinetic level.

**MATERIAL AND METHODS:**

**Drugs and Chemicals:** Sitagliptin Phosphate (STG) was obtained as a gift sample from Glenmark Generics, India. Herbal extracts of leaves of Gymnema sylvestre (GYM) (Family: Asclepiadaceae) were procured from Konark Herbal and Health Care, Daman, India. The extract contained 75.62% of mixture of gymnemic acids (by Gravimetric analysis). Streptozotocin (STZ) was procured from Huohua Industry Co. Ltd., China. High fat diet containing whole wheat (50.0g), yellow corn (50.0g), barley (25.0g), bone meal (2.5g), calcium chloride (2.5g), salt (2.5g), oil (25.0g), fat (25.0g), 1 tablet of Vitamin B\(_{12}\) and cholesterol (200mg/kg/day) was procured from D.S. Trading, Mumbai, India.

**Animals:** Female Sprague Dawley rats were procured from the Bombay Veterinary College and Glenmark Pharmaceuticals, Mumbai, India. Rats were housed in standard polypropylene cages (three animals per group) and kept under good hygienic conditions in the animal house of the institute. Animals were maintained under standard experimental conditions i.e. in a 12-hour light-dark cycle, temperature (24 ± 2°C) and relative humidity (60%) controlled facility of the institute. Standard laboratory diet and water ad-libitum was provided to rats throughout the study. The experiment was carried out in accordance with the ethical guidelines of committee for the purpose of control and supervision of experiments on animals (CPCSEA), Government of India. The study was conducted after the approval of protocol by Institutional Animal Ethics Committee (IAEC) affiliated to CPCSEA with protocol number IAEC/PR/2014-2015/03.

**Experimental Design:**

**Bioanalytical Method Development and Validation:** High Performance Thin Layer Chromatography (HPTLC) was selected as a chromatographic method in order to find the area under curve (AUC) of Sitagliptin Phosphate when given alone and in the presence of herbal extract-Gymnema sylvestre in rat plasma. A new bioanalytical HPTLC method was developed to determine Sitagliptin Phosphate in rat plasma and validated using EMA and US-FDA guidelines.

**Induction of Type II Diabetes Mellitus:** After one week of acclimatisation of rats, study was initiated. Rats weighing above 250 grams were randomly divided into groups containing equal number of animals (n=6) and were fed with high fat diet throughout the study except for the vehicle control group. Intraperitoneal shot of 40 mg/kg Streptozotocin (STZ) was administered to overnight fasted rats except for the vehicle control group. After 24 hours of STZ challenge, the rats were subjected to blood withdrawal for the estimation of serum glucose levels. The rats found to be hyperglycaemic were selected for the study.
Treatment Design:
Group 1: Vehicle Control (VC)
Group 2: Disease Control (DC)
Group 3: Sitagliptin Phosphate (STG) (20 mg/kg; p.o.)
Group 4: Gymnema sylvestre (GYM) (400 mg/kg; p.o.)
Group 5: Sitagliptin Phosphate (20 mg/kg; p.o.) and Gymnema sylvestre (400 mg/kg; p.o.) (STG+GYM)

Pharmacokinetic Analysis:
1. Blood samples were withdrawn from retroorbital plexus at 0th, 2nd, 4th, 8th and 12th hours from rats on first day of treatment with Sitagliptin Phosphate (Group 3) and combination of Sitagliptin Phosphate with Gymnema sylvestre (Group 5).
2. The blood was collected directly into the centrifuge tubes containing anticoagulant and centrifuged for 15 minutes at 4500 rpm.
3. The supernatant clear plasma obtained was transferred carefully with help of micropipette into small test tubes for estimation of desired parameters.
4. Parameter evaluated for pharmacokinetic interaction was Absorption (AUC-Area under Curve)
5. Mean peak area and percentage increase or decrease in the area at gradual increase in the time points are tabulated and interpreted.

RESULT:
Bioanalytical Method Development and Validation: Bioanalytical procedure was optimized using spiked sample of STG (100 μg/ml). In eppendorf tube of 2 ml capacity, 250 μl of plasma was transferred and spiked with 50 μl of 100 μg/ml methanolic solution of Sitagliptin Phosphate using a micropipette. To this 50 μl of 30 μg/ml standard solution of IS Zolpidem Tartrate was added. To this 1 ml of extracting solvent i.e. methanol was added and the tube was vortex mixed for 2 minutes followed by centrifugation at 6000 rpm for 10 minutes at 5°C. Supernatant (1ml) was transferred to a glass tube and evaporated to dryness using solvent evaporator under stream of nitrogen gas. The temperature of water bath in evaporator was set at 60°C. The residue was reconstituted using 100 μl of methanol and 30 μl was applied on chromatographic plate for analysis. Extracts of blank plasma were prepared using above procedure in which drug solution was replaced with 50 μl of methanol. The optimized procedure used is summarized below.

Optimized Bioanalytical Procedure:
Extracting method: Protein Precipitation
Extracting solvent: Methanol (1 ml)
Internal Standard: Zolpidem Tartrate
Vortex Time: 2 minutes
Centrifugation: 6000 rpm
Centrifugation time: 10 minutes
Centrifugation temperature: -5°C
Extracting solvent for reconstitution: Methanol (100 μl)
Injection Volume: 50 μl
Stationary phase: Silica Gel 60 F_{254} precoated TLC plates
Mobile phase: Ethyl acetate: toluene: methanol: triethylamine (4.2:3.8:1.5:0.5 v/v/v/v)
Chamber saturation time: 20 minutes
UV maxima for detection: 267 nm

The final optimized method was statistically validated and was found to be precise, accurate, stable, robust and rugged. Few of the validated parameters are mentioned below:

Selectivity: No interferences from plasma components were seen at the R_{t} of Sitagliptin Phosphate and IS- Zolpidem Tartrate. The method was found to be selective at LLOQ (10 μg /ml) for Sitagliptin Phosphate. The blank responses were found to be less than 20 % of area response of LLOQ and 5 % for IS Zolpidem Tartrate. Fig. 1 and 2 denote densitograms of blank plasma and drug respectively.
**Carry-over Effect:** Carry over in the blank sample following the high concentration standard was not greater than 20% of LLOQ and 5% of IS. The calculated value of Sitagliptin Phosphate and IS-Zolpidem Tartrate were found to be 2.3% of Sitagliptin Phosphate and 1.82% of IS-Zolpidem Tartrate.

**Calibration Curve (CC):** Using the selected model, the method developed was linear over the whole range of concentration from 10 to 200μg/ml. All the three calibration curves passed the criteria for linearity. The results of back calculated concentration of calibration standards are presented below in Table 1.

<table>
<thead>
<tr>
<th>Concentration (μg/ml)</th>
<th>Back Calculated Concentration (μg/ml)</th>
<th>Standard Deviation</th>
<th>% C.V.</th>
<th>% Nominal</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>CC 1</td>
<td>CC 2</td>
<td>CC 3</td>
<td>Mean</td>
</tr>
<tr>
<td>10 (LLOQ)</td>
<td>10.51</td>
<td>10.45</td>
<td>10.41</td>
<td>10.46</td>
</tr>
<tr>
<td>20 (LQC)</td>
<td>20.13</td>
<td>20.13</td>
<td>20.08</td>
<td>20.07</td>
</tr>
<tr>
<td>40</td>
<td>38.56</td>
<td>38.77</td>
<td>38.38</td>
<td>38.57</td>
</tr>
<tr>
<td>80</td>
<td>77.72</td>
<td>77.68</td>
<td>77.79</td>
<td>77.73</td>
</tr>
<tr>
<td>100 (MQC)</td>
<td>103.11</td>
<td>103.23</td>
<td>103.16</td>
<td>103.17</td>
</tr>
<tr>
<td>120</td>
<td>117.73</td>
<td>117.87</td>
<td>118.01</td>
<td>117.87</td>
</tr>
<tr>
<td>140</td>
<td>139.97</td>
<td>140.06</td>
<td>140.43</td>
<td>140.16</td>
</tr>
<tr>
<td>160 (HQC)</td>
<td>160.74</td>
<td>160.83</td>
<td>160.69</td>
<td>160.75</td>
</tr>
<tr>
<td>200 (ULOQ)</td>
<td>199.68</td>
<td>199.58</td>
<td>199.49</td>
<td>199.58</td>
</tr>
</tbody>
</table>

(% C.V. = Coefficient of Variation, LQC- Low Quality Control, MQC- Medium Quality Control, HQC- High Quality Control, ULOQ- Upper Limit of Quantitation)
Accuracy and Precision: The accuracy and precision values were within the acceptable limits at all concentration levels. The within run and between run accuracy and precision values are given below Table 2.

**TABLE 2: RESULTS OF WITHIN-RUN AND BETWEEN-RUN ACCURACY AND PRECISION**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>LLOQ (10 μg/ml)</th>
<th>LQC (20 μg/ml)</th>
<th>MQC (100 μg/ml)</th>
<th>HQC (160 μg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Within Run (n=5)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>9.93</td>
<td>19.28</td>
<td>101.84</td>
<td>158.98</td>
</tr>
<tr>
<td>% CV</td>
<td>1.036</td>
<td>0.516</td>
<td>0.173</td>
<td>0.048</td>
</tr>
<tr>
<td>% Nominal</td>
<td>99.39</td>
<td>96.44</td>
<td>101.84</td>
<td>99.36</td>
</tr>
<tr>
<td></td>
<td>Between Run (n=5)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>10.04</td>
<td>19.12</td>
<td>101.95</td>
<td>158.91</td>
</tr>
<tr>
<td>% CV</td>
<td>2.812</td>
<td>1.924</td>
<td>0.174</td>
<td>0.048</td>
</tr>
<tr>
<td>% Nominal</td>
<td>100.4</td>
<td>95.61</td>
<td>101.95</td>
<td>99.31</td>
</tr>
</tbody>
</table>

Where, n=5 indicate number of replicates

Recovery: The mean recoveries of Sitagliptin Phosphate at LQC, MQC and HQC levels were found to be 74.85 %, 73.36 % and 76.61 % respectively. The recovery of IS was found to be 76.41 %. Recoveries of both Sitagliptin Phosphate and IS were consistent, precise and reproducible. The results of recovery studies of Sitagliptin Phosphate and IS are summarized in Table 3.

**TABLE 3: RESULTS OF RECOVERY STUDIES OF SITAGLIPTIN PHOSPHATE AND IS- ZOLPIDEM TARTRATE:**

<table>
<thead>
<tr>
<th>QC Samples</th>
<th>Mean peak area of extracted samples</th>
<th>Mean peak area of un-extracted samples</th>
<th>% Mean recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td>IS</td>
<td>2028.4</td>
<td>2654.32</td>
<td>76.41</td>
</tr>
<tr>
<td>LQC</td>
<td>400.63</td>
<td>535.21</td>
<td>74.85</td>
</tr>
<tr>
<td>MQC</td>
<td>2124.12</td>
<td>2895.31</td>
<td>73.36</td>
</tr>
<tr>
<td>HQC</td>
<td>3310.67</td>
<td>4315.47</td>
<td>76.71</td>
</tr>
</tbody>
</table>

Pharmacokinetic Analysis: In 24-hour pharmacokinetic study model, blood samples were withdrawn at intervals- 0th, 2nd, 4th, 8th and 12th hours and plasma samples were analysed using developed and standardized bioanalytical method. The results are tabulated in Table 4.

**TABLE 4: RESULT OF PHARMACOKINETIC ANALYSIS- PERCENTAGE DECREASE IN AREA OF SITAGLIPTIN PHOSPHATE IN THE PRESENCE OF GYMNEMA SYLVESTRE IN RAT PLASMA**

<table>
<thead>
<tr>
<th>Time (in hours)</th>
<th>Mean Peak Area along with Spiked Area (n=3)</th>
<th>Mean Spiked Area (n=3)</th>
<th>Actual Area</th>
<th>Percentage decrease in Area (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>2124.5</td>
<td>2124.48</td>
<td>2124</td>
<td>0.5</td>
</tr>
<tr>
<td>2</td>
<td>2126.8</td>
<td>2125.9</td>
<td>2124</td>
<td>2.8</td>
</tr>
<tr>
<td>4</td>
<td>2132.87</td>
<td>2130.17</td>
<td>2124</td>
<td>8.87</td>
</tr>
<tr>
<td>8</td>
<td>2138.9</td>
<td>2133.1</td>
<td>2124</td>
<td>14.9</td>
</tr>
<tr>
<td>12</td>
<td>2124.28</td>
<td>2124.18</td>
<td>2124</td>
<td>0.28</td>
</tr>
</tbody>
</table>

Where, n=3 indicate number of replicates, STG: Sitagliptin Phosphate (20 mg/kg; p.o.), GYM: Gymnema sylvestre (400 mg/kg; p.o.)

RESULT: It was found that there was decrease in the area of Sitagliptin Phosphate when given in combination with Gymnema sylvestre as compared to the single administration of Sitagliptin Phosphate.

DISCUSSION: It can be inferred that the presence of Gymnema sylvestre decreased the bioavailability of Sitagliptin Phosphate when administered concomitantly in rats. Thus stating that there exists a pharmacokinetic interaction at absorption level.
This may be attributed to the fact that being an herb, Gymnema sylvestre might have altered the bioavailability of Sitagliptin Phosphate by any of the following mechanism of absorption interaction:

**Alteration in the gastro-intestinal pH:** The rate of drug absorption by passive diffusion is basically dependent on pH of gastric fluid. Basic drugs are absorbed in basic environment (intestine) whereas acidic drugs are absorbed in acidic environment (stomach). Therefore, compounds that create an acidic environment with a lower pH may decrease the solubility of compounds which are basic in nature. Since Gymnema contains gymnemic acids there might have been alteration in the gastrointestinal pH leading to increase in acidity, due to which the absorption of sitagliptin having pKa = 7.7 \(^{18}\) might have hampered due to limited absorption in an acidic environment.

**Effects of P-Glycoprotein:** P-Glycoprotein is a multidrug-resistance gene product found in a variety of human tissues, including the gastrointestinal epithelium. This efflux pump is expressed at the luminal surface of the intestinal epithelium and opposes the absorption of unchanged drug by transporting compounds out of enterocytes back into the gastrointestinal lumen. Sitagliptin being a substrate for P-glycoprotein, this might be the reason for decreased bioavailability of Sitagliptin.

**CONCLUSION:** The above result indicates somewhat significant drug-herb interaction between Sitagliptin Phosphate and Gymnema sylvestre for the proposed work. In presence of Gymnema sylvestre, the mean peak area of Sitagliptin Phosphate was found to decrease in the rats. This interaction needs to be evaluated through mechanistic studies at different doses. Further studies need to be carried out in human subjects to evaluate this interaction in order to adjust the appropriate dose of both- Sitagliptin Phosphate and Gymnema sylvestre. Such herb drug interaction studies would not only bring scientific knowledge but also enlighten the healthcare system about the practise of complementary therapies. This would educate the patients as well as keep them alert about the interactions between the medicines they are consuming.

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**CONFLICT OF INTEREST:** The authors report no conflict of interest.

**REFERENCES:**

