(Research Article)

1

## IJPSR (2015), Vol. 6, Issue 1





Received on 12 May, 2014; received in revised form, 07 August, 2014; accepted, 16 September, 2014; published 01 January, 2015

# *IN-VITRO* CYTOTOXIC ACTIVITY OF LEAF EXTRACTS OF *IPOMOEA* JACQ. SPECIES AGAINST A 549AND HEP-G2CELL LINES

Vinaya A. Rane\* and Behnaz B. Patel

Department of Botany, R. Ruia College, Matunga, Mumbai - 400 019, M.H., India.

**Keywords:** 

Cytotoxicity Activity, SRB Assay, Ipomoea species, HEP G2, A549

**Correspondence to Author:** 

#### Vinaya A Rane

Department of Biotechnology, Ramnarain Ruia College, L.N. Road, Matunga (East), Mumbai - 400 019, MH, India.

E-mail:\_mhatre\_vinaya@yahoo.com

**ABSTRACT:** Members of the genus *Ipomoea* of family Convolvulaceae besides being some of the most beautiful ornamental plants cultivated for their bell shaped flowers, are also of medicinal value, with some of them reported as cytotoxic agents against some cancer cell lines. The present study evaluates the anticancer activity of the leaf extracts of ten *Ipomoea* Jacq. Species growing in and around Mumbai, India, on the *A* 549 cellline and *Hep-G2* cell line evaluated by the SRB assay. The crude aqueous extracts of leaves of *Ipomoea* species studied had significant cytotoxic effect on *HEP G2* Cell Line and showed significant inhibitory effect on A549 cell line., *I. Cairica* was the best growth inhibitor (GI<sub>50</sub>at 30.6µg/ml) for HEP G2 cell line followed by *I. Hederifolia* (GI<sub>50</sub> at 38.1 µg/ml), *I. Carnea* (GI<sub>50</sub> at 40.0 µg/ml). Considering the A549 cell line, none of the crude extracts showed GI<sub>50</sub> at 75.6 µg/ml.

**INTRODUCTION:** Folk medicine since times immemorial has incorporated the use of plants for their medicinal properties. Many of the plants species used for this purpose have been found to contain therapeutic substances which can be extracted and used in preparation of drugs, but the plant itself can also be used either directly or as an extract for medication, a practice that is particularly popular in developing countries<sup>1, 2</sup>.

Plewa and Wagner<sup>3</sup> have stated that many plants contain cytotoxic and genotoxic substances. Many drugs that are used for cancer treatment are obtained from plant sources the most common examples being Taxol from *Taxus*, Campothecin from *Camptothecaacuminate*, vinca alkaloids, vincristine and vinblastin, isolated from *Catharanthusroseus*;



etoposide and teniposide derivatives of epipodophyllotoxin, isolated from *Podophyllum*. <sup>4</sup>, <sup>5,6</sup>

They are effective against proliferating cellsand exhibit cytotoxic effects either by damaging DNA or by blocking the formation of mitotic spindle during stages of cell division<sup>7</sup>. There is an emphasis thus on the importance of the search for improved cytotoxic agents in the discovery of modern anti-cancer drugs<sup>8</sup>. Herbal medicines, as an alternative cancer therapy have attracted a great deal of attention due to their low cost and low toxicity<sup>9</sup>.

Madhuri and Pandey<sup>10 in</sup> their review article on medicinal plants with anti-cancer properties have cited examples of various medicinal plants effective in various types of malignant (cancer) and benign tumours of humans and experimental animals like, Agrimoniapilosa in sarcoma, Ailanthusaltissimafor intestinal cancer, sarcoma-180. sarcoma-37 and leukaemia-16: Akebiaquinatafor sarcoma-180and sarcoma-37; Chelidoniumjajusvar. Asiaticum for stomach

### Rane and Patel, IJPSR, 2015; Vol. 6(1): 294-299.

cancer; *Chimaphilaumbellate* for breast tumour; Coix lachrymal-jobi for ascites cancer and sarcoma; Fritillariathunbergii Yoshida's for tumours of the throat, chest, neck and breast; *Larrea tridentate* for various cancers, especially leukaemia; Sophoraflavescens and S. Subprostrata for sarcoma-180, leukaemia and cervical cancer-14 cells; Taraxacum mongolicum for ascites cancer, sarcoma-180 and lung cancer cells, and *Vitexrotundifolia* for lung tumour. Jain *et al.*<sup>11</sup> have proved the cytotoxic activity of several Indian medicinal plants against human breast cancer (MCF-7) and human leukemia (HL-60) tumor cell lines.

To confirm with surety that a particular compound or plant had anti-cancer properties, it is essential to carry out the test on human cancer cell lines, of various types as an extract can be cytotoxic on a particular cell line but not on another, that is the plant extracts exhibit selective cytotoxicity. Several workers <sup>12, 13, 14, 15</sup>, have tested the anti- cancer activity of plant extracts of herbal medicines and have observed the selective cytotoxicity.

Considering *Ipomoea* species, the whole plant of *I. nil* has been reported to exhibit anticancer activity against human epidermoid carcinoma of the nasopharynx in tissue culture and  $P_{388}$  lymphocytic leukemia in mice<sup>16</sup>. The leaf extracts of *I. aquatic* have exhibited cytotoxic activity against Hep -2 and A549 cell lines<sup>17</sup>. The methanolic extract of the whole plant of *I. pes-caprae* exhibited proliferative activity on murine spleen cells<sup>18</sup> *I. cairica* has been reported as having cytotoxic activity against Lung adenocarcinoma epithelial cell line A549<sup>19</sup>. Cytotoxic activity of *I. obscura*<sup>20</sup> has also been reported.

Considering the medicinal values of *Ipomoea* species and the reported cytotoxic effect of the extracts of *I. aquatica, I. pes-caprae, I. Hederifolia* and *I. carnea* on human cell lines cytotoxic activity involving ten species growing in and around Mumbai, India was studied on the *A 549* cell line and Hep-G2 cell line to assess the anti -cancer activity of their leaf extracts.

## **MATERIALS AND METHODS:**

Ten species of *Ipomoea* were collected from wild habitat, in and around Mumbai, identified and authenticated in consultation with Blatter Herbarium, St. Xavier's College, Mumbai, India. One copy of the herbarium was deposited in the Blatter Herbarium and the other copy is preserved in The Late Yashwant rao Adbal Herbarium, of the Department as voucher specimen (**Table 1**).

TABLE 1: THE TEN SPECIES OF IPOMOEA COLLECTEDAND DEPOSITED IN THE LATE YASHWANT RAO ADBALHERBARIUM, OF THE DEPARTMENT

Species	Locality	Voucher Specimen	
Ipomoea nil (Linn.) Roth.	Yeoor, Thane Dist.	1	
Ipomoea aquaticaForsk.	Vikhroli, Mumbai	2	
Ipomoea pes-caprae	Arnala, Virar, Thane	2	
(Linn.) R.Br.	Dist.	5	
Ipomoea carneaJacq.	Alibaug, Raigad Dist.	4	
Ipomoea hederifolia Linn.	Yeoor, Thane Dist.	5	
Ipomoea triloba Linn.	Yeoor, Thane Dist.	6	
Ipomoea cairica ( Linn.) sweet	Goregaon, Mumbai	7	
Ipomoea turbinata Lag.	Yeoor, Thane Dist.	8	
Ipomoea quamoclit Linn.	Yeoor, Thane Dist.	9	
Ipomoea violacea Linn.	Airoli, Navi Mumbai	10	

**Plant Powder Preparation:** The leaves of the ten *Ipomoea* species under study were collected during flowering season washed in running water, rinsed in deionized water and air dried for 10 days. They were powdered and sieved through BSS mesh No. 85. Loss on drying was determined. The powders were stored in a cool dry place.

**Plant Extract preparation:** 10g of the dry leaf powders of the ten species were weighed and extracted in 100ml Distilled water for 3h in Soxhlet Extractor at 100°C. The extract was filtered using Whatman filter paper no. 41. Solvent extract was concentrated on Rotavapor (under vacuum) and was considered as 10% crude aqueous extract.

# Testing the extracts for anti-cancer activity on Human cell lines:

The anticancer activities of the crude aqueous extracts were studied at Advanced Center for Treatment, Research and Education in Cancer (ACTREC), Mumbai, India. Two cancer cell lines namely, Lung adenocarcinoma epithelial cell line A549 and Human Hepatoma cell line, Hep-G2 were used for the assay. The SRB Assay procedure of Skehan *et al.*<sup>19</sup> as given by Gaidhani*et al.*<sup>20</sup> was followed. The two cell lines were maintained in

ideal laboratory conditions. The cell lines were grown in RPMI 1640 medium containing 10% fetal bovine serum and 2mM L-glutamine. The cells were inoculated into 96 well microtiter plates of 90 $\mu$ l/ well capacity, with appropriate plating densities, depending on the doubling time of individual cell lines, followed by incubation at 37°C, in 5% CO<sub>2</sub>, 95% air and 100% relative humidity for 24 h prior to addition of plant extracts.

After 24 h, the cells from one well, of each cell line were fixed *in situ* with TCA, to represent a measurement of the cell population for each cell line at the time of drug addition (Tz). Plant extracts were solubilized in DMSO as 10mg/ml concentration and stored frozen prior to use. At the time of drug addition, an aliquot of frozen concentrate was thawed and diluted to 10 times, to make it 1000  $\mu$ g/ml with complete RPMI medium.

This complete RPMI medium containing test extract was further diluted with complete RPMI medium to adjust the concentration at 100, 200, 400 and 800  $\mu$ g/ml. Aliquots of 10  $\mu$ l of these different dilutions were added to the appropriate microtiter wells already containing 90  $\mu$ l of cell suspension, resulting in the required final drug concentrations of 10, 20, 40 and 80  $\mu$ g/ml. For each of the microtitre plate the well known anticancer drug 'ADR' (Adriamycin) was used as a positive control.

**Endpoint Measurement:** After compound addition, the plates were incubated at standard conditions for 48 hours and assay was terminated by the addition of cold TCA. Cells were fixed *in situ* by the gentle addition of 50  $\mu$ l of cold 30 % (w/v) TCA (final concentration, 10 % TCA) and incubated for 60 minutes at 4°C. The supernatant was discarded; the plates were washed five times with tap water and air dried.

The SRB Assay: Sulforhodamine B (SRB) solution (50  $\mu$ l) at 0.4 % (w/v) in 1 % acetic acid was added to each of the wells, and plates were incubated for 20 minutes at room temperature. After staining, unbound dye was recovered and the residual dye was removed by washing five times with 1% acetic acid. The plates were air dried. Bound stain was subsequently eluted w ith 10 mM

Trizma base, and the absorbance was read on an ELISA Plate Reader at a wavelength of 540 nm with 690 nm reference wavelength.

Calculations: Percent growth was calculated on a plate-by-plate basis for test wells relative to control wells.

 $\% Growth = \frac{Averageabsorbanceofthetestwell*100}{Averageabsorbanceofcontrolwells}$ 

Using the six absorbance measurements [time zero (Tz), control growth (C), and test growth in the presence of drug at the four concentration levels (Ti)]; the percentage growth was calculated at each of the drug concentration levels.

Percentage growth inhibition was calculated as:  $%GI = \frac{(T_i - T_z) * 100}{(C - T_z)}$ 

**Growth inhibition of 50 % (GI**<sub>50</sub>) was calculated from  $[(Ti-Tz)/(C-Tz)] \times 100 = 50$ , which is the drug concentration resulting in a 50% reduction in the net protein increase (as measured by SRB staining) in control cells during the drug incubation.

**The LC**<sub>50</sub> (concentration of drug resulting in a 50% reduction in the measured protein at the end of the drug treatment as compared to that at the beginning) indicating a net loss of cells following treatment was calculated from:

$$LC_{50} = \frac{(Ti - Tz) + 100}{Tz} = -50$$

Values were calculated for each of these three parameters if the level of activity was reached; however, if the effect was not reached or was exceeded, the values for that parameter were expressed as greater or less than the maximum or minimum concentration tested.

# **RESULTS:**

In view of these observations and considering the ethno medicinal uses of several *Ipomoea* spp. and an earlier report that *I. aquatic* Forsk. and *I. bahiensis* possess anticancer activity against Vero, cell lines<sup>23</sup>, and against Hep-2, A-549<sup>17</sup>, the crude aqueous extracts of the ten species were tested for their cytotoxic properties on mammalian cell lines

Lung adenocarcinoma epithelial cell line A 549 and Human Hepatoma cell line, HEP G2. The crude extracts were more effective on the cell line HEP G2 than on A 549.

TABLE 2: EFFECT OF IPOMOEA	SPECIES AGAINST HEP G2 AND A549 CELL LINES
----------------------------	--

	Values derived from graph (µg/ml)						
Ipomoea spp.	Against HEP G2 cell lines			Against A549 cell lines			
	LC50	TGI	GI50	LC50	TGI	GI50	
I. nil	>80	>80	62.4	>80	>80	>80	
I. aquatica	>80	>80	75.6	>80	>80	>80	
I. pes-caprae	>80	>80	48.0	>80	>80	>81	
I. carnea	>80	>80	40.0	>80	>81	>82	
I hederifolia	>80	73.5	38.1	>80	>82	>83	
I. triloba	>80	>80	49.7	>80	>83	>84	
I. cairica	>80	65.5	30.6	>80	>84	>85	
I. turbinata	>80	>80	>80	>80	>85	>86	
I. quamoclit	>80	>80	>80	>80	>86	>87	
I. violacea	>80	>80	57.8	>80	>87	>88	
ADR	73.7	38.0	<10	34.2	12.3	<10	



FIG. 1 GROWTH CURVE: HUMAN HEPATOMA CELL LINE HEP-G2



FIG. 2 GROWTH CURVE: HUMAN LUNG CANCER CELL LINE A549

**DISCUSSION:** Considering the effect on HEP G2cell line (**Table 2, Fig 1**), *I. cairica* was the best Growth inhibitor (at 30.6µg/ml) followed by *I. hederifolia* (38.1 µg/ml), *I. carnea* (40.0 µg/ml). However *I. aquatic* with respect to HEP G2 it showed GI50 at 75.6 µg/ml and the cytotoxicity activity of *I. aquatica* was reported on the same cell line at 156 mg/ml by Prasad *et al*<sup>17.</sup> Such high concentrations were not tried out as according to Gaidhaniet al.<sup>22</sup> the range of the concentration of the compound being tested on human cell lines, should be from 10µg/ml to 80µg/ml.

Considering the A549 cell line (**Table 2, Fig.2**) none of the crude extracts showed GI50 inhibition upto  $80\mu$ g/ml. However by extending the graph, it was seen that higher concentrations would prove to be effective. Prasad *et al.*<sup>17</sup> have reported a concentration of 394mg/ml of *I. aquatic* leaves on the same cell lines and Lin *et al.*<sup>19</sup>have reported the cytotoxicity of *I. cairica* on the same cell line.

Arokiyaraj *et al.*<sup>23</sup> have reported that the triterpenes and flavonoids act as inhibitors of human lymphocyte proliferation. According to Pettit<sup>25</sup>*I.pes-caprae* contains betulinic acid which is a promising anti-cancer against melanomy by inhibiting the growth of cancer cells and inducing apoptosis. In the present study *I. pes-caprae* exhibited GI<sub>50</sub> activity at 48µg/ml for HEP G2anddid not exhibit any activity upto 80 µg/ml on A549 cell line. Values beyond 80µg/ml could be estimated from the graphs. Ancuceanu<sup>26</sup>in his review article on Lung Cancer, has pointed out anti cancer activity of a compound can be due to that microtubule interfering substances like *Vinca* alkaloids and colchicine, topoisomerase inhibitors like *Scutellaria baicalensis*, inhibitor of DNA synthesis like certain *Polygonum* species or an inhibitor of DNA polymerase I activity as Indirubin from *Indigofera suffrutticosa*.

**CONCLUSION:** The crude aqueous extracts of leaves of *Ipomoea* species studied had significant cytotoxic effect on *HEP G2* Cell Line and showed significant inhibitory effect on A549 cell line. , *I. cairica* was the best growth inhibitor (GI<sub>50</sub>at 30.6µg/ml) for HEP G2 cell line followed by *I. hederifolia* (GI<sub>50</sub> at 38.1 µg/ml), *I. carnea* (GI<sub>50</sub> at 40.0 µg/ml). Considering the A549 cell line, none of the crude extracts showed GI<sub>50</sub> inhibition upto80 µg/ml with the exception of *I. aquatic* which showed GI<sub>50</sub>at 75.6 µg/ml.

**ACKOWLEDGMENTS:** We are grateful to the Principal, R. Ruia College for providing the facilities for the conductance of the work. We thank Blatter Herbarium, St. Xavier's College for authentication of herbaria of *Ipomoea* species and also ACTREC, Mumbai for helping us in studying the anticancer activities.

## **REFERENCES:**

- Ishii, R., Yoshikawa, H., Minakata, N.T., Komura, K. &Kada, T. (1984). Specificity of biomutagensin the plant kingdom. *Agricultural and Biological Chemistry*, 48; 2587-2591.
- Hoyos, L.S., Au, W.W., Heo, M.Y., Morris, D.L. &Legator, M.S. (1992). Evaluation of the genotoxiceffects of a folk medicine, Petiveriaalliacea (anamu). *Mutation Research*, 280:29-34.
- 3. Plewa M. J., and Wagner, E. D. (1993). Activation of promutagens by green plants," *Annual Review of Genetics*.27; 93–113.
- Cragg, G.M., Schepartz, S.A., Stuffness, M., Grever, M.R. (1993). The taxol supply crisis. New NCI policies for handling the large-scale production of novel natural product anticancer and anti-HIV agents. *J. Nat. Prod.* 56: 1657-1668.
- Cragg, G.M., Boyd, M.R., Cardellina, J.H., Newman, D.J., Snader, K.M., McCloud, T.G. (1994). Ethnobotany and drug discovery experience of the US National Cancer Institute: In: Chadwick D.J, Marsh J (Eds.), Ciba foundation "*Ethnobotany and the search for new drugs*" symposium, 185. Wiley and Sons, Chichester, UK, pp. 178-196.
- 24. Arokiyaraj, S.; Perinbam, K.; Agastian, P. and Balaraju, K. (2007). Immunosuppressive effect of medicinal plants of

- 6. Wang, H.K. (1998). Plant-derived anticancer agents currently in clinical use or clinical trials. *The Investigation Drugs Journal*.1: 92-102.
- 7. Gali- Muhtasib, H. and Bakkar, N. (2002).Modulating cell cycle: current applications and future prospects for future drug development. *Curr. Cancer Drug Targets* 2: 1-17.
- Nobili, S., D. Lippi, E. Witort, M. Donnini, L. Bausi, E. Mini and S. Capaccioli (2009).Natural compounds for cancer treatment and prevention. *Pharmacol. Res.*, 59: 365-378.
- Kang S. A.; Park , H.J. Kang, S.A.; Park, H.J.; Kim M. J. Lee, S. Y., Han, S. W. and K.H. Leem Leem, (2005).Citrireticulataeviridepercarpium extract induced apoptosis in SNCU-C4, human colon cancer cells. J. *Ethanopharmacol.*, 971: 231-235.
- MadhuriS.andPandey G. (2009). Some anticancer medicinal plants of foreign Origin.*Current Science*96(6): 779-783.
- 11. Jain R and Jain S. K (2011). Screening of *in vitro* cytotoxic activity of some medicinal plants used traditionally to treat cancer in Chhattisgarh state, India. *Asian Pacific Jour. of Tropical Biomedicine*. S147-S150.
- 12. Sakpak deejaroen I. and Itharat A. (2009) Cytotoxic Compounds against breast adenocarcinoma cells (MCF-7) from pikutbenjakul.*J Health Res*23 (2): 71-76.
- 13. Verma, S. K., Singh, S.K., Mathur, A. and Singh, S. (2010) In vitro cytotoxicity of Argemonemexicana Against Different Human Cancer Cell Lines. *International Journal of Chemical, Environmental and Pharmaceutical Research*, 1(1): 37-39,
- 14. Ampasavate, C., Okonogi, S. and Anuchapreeda S.(2010). Cytotoxicity of extracts from fruit plants against leukemic cell lines. *African Journal of Pharmacy and Pharmacology*.4 (1): 13-21.
- Mazumder, P.M., Das, S., Das, S. and Da, M.(2010) Cytotoxic Activity of Methanolic Extracts of Berberisaristata DC and Hemidesmusindicus R.Br. in MCF7 Cell Line. Journal of Current Pharmaceutical Research01: 12-15.'
- 16. Rastogi, R.P., Dhawan, B.N. (1990) Anticancer and antiviral activities in Indian medicinal plants: A review. *Drug Dev Res*.19:1–12.
- Prasad, K.N., Raghu, A.G., Shivamurthy G.R., Vijayan, P. and Aradhya, S.M. (2005). In vitro cytotoxic properties of Ipomoea aquatica. *Ind. J. Pharmacol.* 37: 397-398.
- Zandonai, R.H. (2010). Evaluation of the proliferative activity of methanol extracts from six medicinal plants in murine spleen cells. *Braz. J. Pharm. Sci.* 46(2): 323-333.
- 19. Lin, R.J., Chen, C.Y., Lo, W.L. (2008) Cytotoxic Activity of Ipomoea cairica. *Nat. Prod. Res.* 22(9): 747-53.
- Srinivasan R., Senthil RD., Ashok G. and Kumarappan C T (2014) Cytotoxic and antimicrobial activity of *Ipomoea* obscura. Aust Jour of Basic and Appl Sciences 8529.
- Skehan P., Storeng R., Scudiero D., Monks A., McMahon J., Vistica D., Warren J. T., Bokesch H., Kenney S. and Boyd M. R. (1990). New Colorimetric Cytotoxicity Assay for Anticancer-Drug Screening J. Natl. Cancer Inst, 82: 1107.
- 22. Gaidhani SN, Lavekar GS, Juvekar AS, Sen S, Singh A, Kumari S. (2009) In-vitro anticancer activity of standard extracts used in ayurveda. *Pharmacog Mag.* 5:425–429
- 23. Bieber, L. W., Filho AADS, Rosalia, M.O., Lima, C., Chiappeta, A.D, Nascimento, S.C.,.(1986).Anticancer and Antimicrobial glycosides from *Ipomoea bahiensis.Phytochemistry*25: 1077-81

Kolli hills on mitogen-stimulated proliferation of the human peripheral blood mononuclear cells *in vitro*. *Indian J. Pharmacol*.39:180-183.

- 25. Pettit, G. R. (1996). Progress in the discovery biosynthetic anticancer drugs. *J. Nat. Prod*.59: 812-821.
- 26. Ancuceanu, R and Istudor, V. (2004) Pharmacologically active natural compounds for lung cancer. *Altern. Med. Rev*.9(4): 402-19.s

#### How to cite this article:

Rane VA and Patel BB: *In-Vitro* Cytotoxic Activity of leaf Extracts of *Ipomoea* Jacq. Species against a 549 and HEP-G2 cell Lines. Int J Pharm Sci Res 2015; 6(1): 294-99.doi: 10.13040/IJPSR.0975-8232.6 (1).294-99.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)