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AN EVALUATION ON GRAIN QUALITY UNDER POST HARVEST STORAGE IN SELECTED INDIAN WHEAT VARIETIES

Shiju Mathew

Ministry of Higher Education, Department of Natural Sciences, Aksum University, Aksum, Ethiopia

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Correspondence to Author:

Dr. Shiju Mathew

Ministry of Higher Education, Department of Natural Sciences, Aksum University, Aksum, Ethiopia

ABSTRACT

Wheat belongs to the genus *Triticum* of the grass family, Poaceae. This genus is originated in tropical South west Asia, where it occurs in wild as well as in cultivated forms. Man has depended upon the wheat plant for thousands of years. Wheat (the *Triticum* spp.) is cultivated worldwide. The grain production varies from year to year and hence the grains should be stored strategically from years of overproduction for the use in year of under production. Also grain must be stored for several other reasons such as point of production is not the point of consumption and the time of production is not the time of consumption. Stored grains can have losses in both quantity and quality. Grain quality after harvest is influenced by a wide variety of biotic and abiotic factors and has been studied as a stored grain ecosystem. Losses occur when the grain is attacked by microorganism and other organisms including insects, mites, rodents and birds. The grain losses found in quantity and quality; can be in the form of depletion in seed viability, hardness, color, size and shape, grain weight and various biochemical parameters viz., protein, carbohydrate and vitamins under post harvest storages. The storage fungi damage the grains in several ways; they reduce the germinability, produce undesirable odor and kernel discoloration, decrease the food value and also produce toxins injurious to the health of consumers.

INTRODUCTION: Stored grain can have losses in both quantity and quality. The loss occurs during post harvest storage of wheat grains due to the biotic and abiotic factors. The post-harvest losses of wheat are estimated about 8 per cent of production. Several million colonies of storage fungi have been reported from a gram of dust collected from grain elevators and warehouses ¹. The post harvest loss of wheat grain has been found to be highest during storage. Grain quality after harvest is influenced by a wide variety of biotic and abiotic factors and has been studied as a stored grain ecosystem. Stored grains can have losses in both quantity and quality. Losses occur when the grain is attacked by microorganisms and other organisms including insects, mites, rodents and birds ²⁻¹¹.

Experiments Conducted and Outcomes:

Experiment 1: Characterization of grain hardness of stored wheat grains (Hardness tester): Two Indian wheat grain varieties viz., U. P. 262 and H. D. 1982 were selected and samples were taken from five different sites of Allahabad at four different time periods starting from 12 months of storage (P1) to a maximum of 30 months with time interval of 6 months each (P2 to P4). The important parameter like hardness was carefully observed and determined during every six months for two years. It was observed that the deterioration of the stored wheat grain occurred with time duration which leads to the morphological change of stored wheat grains. The samples collected from the local farmer of Naini and Karchana has shown only negligible change in shape as compared to the other samples collected from the wholesale markets and Govt. godown, Naini.

Experiment 2: Determination of seed viability of stored wheat grains by sprouting test: The germination studies show that viability of the stored wheat grain reduces on storage. The study

has shown that the U.P. 262 has much more sprouting capability as compared to H.D. 1982. The sprouting capacity of both the varieties analyzed showed a decrease to about 70% after three years of storage. After 3 years it takes from days to weeks and in some cases even months for sprouting to be complete.

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Experiment 3: Evaluation of the infestation of stored wheat grains (Dilution plate technique): All the experiments on infestation of epidermal and sub-epidermal region were done very carefully systematically. The mycoflora of stored wheat grains predominantly consisted of ubiquitous Aspergillus, mould genera Alternaria, Cladosporium, Fusarium, Mucor, Rhizopus and their Penicillium because of possibly omnipresence, capacity to grow on all possible substrates and a wide range of temperature and humidity. The most frequent species observed in the stored wheat grains of Aspergillus were A. niger, A. fumigatus, Alternaria alternata, Fusarium moniliformis, Rhizopus arrhizus and a few Pencillium species. Among these the frequency of A. niger was highest which has the capacity to produce mycotoxin which can contaminate and cause spoilage.

Experiment 4: Quantification of wheat grain spoilage on the basis of biochemical parameters (HPLC test): The biochemical study of stored wheat grain was done and it was observed that deterioration of protein and carbohydrates takes place. The deterioration or loss of protein and carbohydrate content is very much apparent in every six months. It was observed that during the study, about 3-4% loss of protein and carbohydrate was prevalent, this can be due to genotypic nature and biotic and abiotic factors also. The percentage of protein loss leads to future infection and infestation by other organisms.

DISCUSSION: There are several reports related to hardness is a whole grain characteristic defined as the grain resistance to applied deformation is largely determined by the endosperm properties considering its proportion in grain and its hardness reduces with the time duration ¹²⁻¹⁶.

The result obtained in the investigation showed that the stored grain loses its viability with time. This can possibly be due to the mycotoxins produced by fungi. Similar results of post harvested wheat grains having reduced sprouting under storage has been obtained by other scientists ¹⁷⁻²³.

Reports from various countries show that this species is found to be the most common post harvest fungi. The presence of fungi in stored wheat grains in Italy has been reported by many scientists ²⁴⁻²⁶. The investigation done on biochemical losses as continuous deterioration has previously shown in many reports in the post harvest stored wheat grains ^{27, 28}.

References:

- Chandra, S., Narang, M. and Srivastava, R.K. (1985). Studies on mycoflora of oilseeds in India. Interrelationship between mycoflora and seed. *J. Seed Sci. and Technol.* 13: 543-549
- Scott P. M. (1991). Possibilities of reduction or elimination of mycotoxins present in cereal grains. In: Cereal Grain: Mycotoxins, Fungi and Quality in Drying and Storage (J. Chelkowski, Ed.). Elsevier, Amsterdam. 529-572.
- Moss, M. O. (1991). Mycology of cereal grain and cereal products. In: Cereal Grain: Mycotoxins, Fungi and Quality in Drying and Storage (J. Chelkowski, Ed.). Elsevier, Amsterdam–London–New York–Tokyo. pp. 23-51.
- Lacey, L. and Magan, N. (1991). Fungi in cereal grains: their occurrence and water and temperature relationships. In: Cereal Grain: Mycotoxins, Fungi and Quality in Drying and Storage (J. Chelkowski, Ed.). Elsevier, Amsterdam–London– New York–Tokyo. pp. 77-118.
- Kuzmiene, G., Pazarauskiene, J. and Sirtautaite, S. (1991). Augalininkystės produktų laikymas ir perdirbimas. Vilnius: Mokslas. pp.381.

- Marin, S., Sanchis, V., Ramos, A.G. and Magan, N. (1998a). Environmental factors, interspecific interactions, and niche overlap between Fusarium moniliforme and F. proliferatum and Fusarium graminearum, Aspergillus and Penicillium spp. isolated from maize. Mycol. Res. 102: 831-837.
- Abramson, D. (1998). Mycotoxin formation and environmental factors. In: Mycotoxins in Agriculture and Food Safety (K. K. Sinha and D. Bhatnagar, eds.). New York: Marcel Dekker Inc. pp. 255-277.
- Magan, N., Hope, R., Cairns, V. and Aldred, D. (2003). Post-harvest fungal ecology: impact of fungal growth and mycotoxin accumulation in stored grain. European J. Plant Pathol. 109: 723-730.
- Krasauskas, A., Steponaviciene A., Railiene, M., Lugauskas, A., Raila, A. and Raudoniene V. (2005). Impact of environmental conditions on the spread of micromycetes in grain during its harvesting and storage. Bot. Lithuanica. 11: 101-109.
- Zvicevicius, E., Raila, A., Novosinskas, H. and Krasauskas,
 A. (2006). Mycotoxin producents in the grain layer. J. Ecologia. 3: 105-111.
- Neethirajan, S., Karunakaran, S., Jayas, D.S. and White, N.D.G. (2007). Detection techniques for stored-product insects in grain. *Int. Fd. Control* 18: 157-162.
- 12. Harper, J.N. and Peter, A.M. (1904). Protein content of wheat kernel. *Bull. Kentucky Agric. Exp. Stn.* 113.
- Jelinek, J. (1927). Apparatus for the determination of the hardnessof the kernel. *Proc. Int. Congress on flour* and bread, Prague, Czechoslovakia. ICC, Vienna.
- Newton R., Cook, W.H. and Malloch, J.G. (1927). The hardness of the wheat kernel in relation to protein content. J. Sci. Agric. 8: 205.
- 15. Pence, R.O. (1935). Hardness in wheat. *Am. Miller* 63: 20-28.
- 16. Greenaway, W.T. (1969). A wheat hardness index. *Cereal Sci. Today* 7: 272-274.
- 17. McCalla, T.M. and Norstadt, F.A. (1967). Influence of patulin on the growth of wheat plants. *Bact. Proc.* pp 17.
- Das, A.M. and Srivastava, D.N. (1969). Production and action of wheat seedling of toxic metabolites of Helminthosporium sativum Ann. Phytopathol. Soc. Japan 35: 275-285.
- 19. Misra, A.P. and Singh, T.B. (1969)..Inhibitory effects of the culture filterates from *Helminthosporium nodulosum* and *H. leucosylum* to seeds and seedlingsof *Eleusine coracana*. *J. Appl. Sci. India* 1: 19-21.
- 20. Ayachi, S.S. (1971). Mycoflora of wheat grain–IV: Toxin production of some frequently occurring fungi. Nat. Acad. Sci. Annual number (Abstract) 91.
- Roy, R.Y., Dwivedi, R.S. and Gupta, V.K. (1972). Effect of rhizosphere mycflora on the growth of *Trigonella* foenum-graecum Linn. Proc. Nat. Acad. Sci. India 42: 105-110.
- Srivastava, A.K. (1974). Some studies on mycoflora, especially fungi, associated with barley and millet in

ISSN: 0975-8232

- storage (Abstract). Proceedings of International storaging seeds: 34.
- 23. Christensen, C.M. and Kaufmann, H.H. (1969). Grain storage the role of fungi in quality loss. University of Minnesoto Press, Minn U.S.A.: 153.
- Wallace, H.A.H. and Sinha, R.N. (1981). Causal factors operative in distributional patterns and abundance of fungi: a multivariate study. In: The Fungal Community-Its Organisation and Role in Ecosystems (D.T. Wicklow and G.C. Carroll, Eds.). Marcell Dekker Inc., New York: 233-247.
- 25. Tsunoda, H. (1970). Microorganisms which deteriorates stored cereals and grains. In: Toxic microorganisms (M. Herzberg, Ed.). Published by UJNR Joint Panels on toxic

- microorganisms and the U.S. Department of Interior Washington D.C.
- Tsuruta, O. (1970). Microorganisms in stored grains.
 Training manual on storage and preservation of wheat grain: 307.
- 27. Greenblatt, G.A., Bettage, A.D. and Morris, C.F. (1994). Relationship between endosperm texture and the occurrence of friabilin and bound polar proteins. *Cereal Chem.* 72: 172-176.
- Morris, C.F., DeMacon, V.L. and Giroux, M.J. (1999a).
 Wheat grain hardness among chromosome 5D homozygous recombinant substitution lines using different methods of measurement. *Cereal Chem.* 76: 249-254.
