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BIOCHEMICAL MECHANISMS OF ETOPOSIDE; UPSHOT OF CELL DEATH

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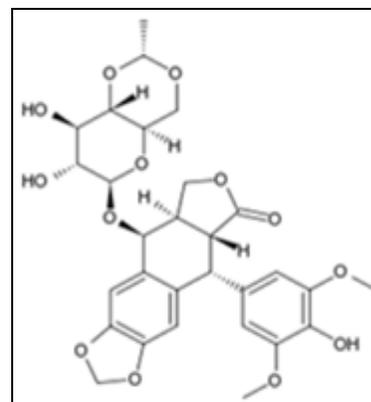
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ABSTRACT: Etoposide is an antineoplastic agent employed for healing various malignancies in human. It is a semi-synthetic derivative of podophyllotoxin and was introduced in 1971. The primary mode of action of etoposide is the inhibition of the function of the enzyme topoisomerase II. It stabilizes DNA-enzyme complex, and accumulation of this complex enhances the probability of double-stranded DNA breaks. However, etoposide can also cause DNA lesions in cancerous cells. This genotoxicity and antineoplastic activity of drug at the same time serve as the impetus for intensive research. Etoposide treatment causes permanent DNA breaks pertaining to its significant role in cell death and growth arrest in tumor cells. This review attempts to provide the deep knowledge of its possible biochemical pathways which are triggered in response to DNA breaks and leads to apoptosis and cell cycle arrest as well transcriptional activation of various death receptors. Molecular mechanism of drug efflux and activation of repairing proteins which can cause resistance in its action are also discussed. This current picture of known facts serve as a gateway to design more effective and efficient antitumor drugs, with new therapeutic strategies, combinations and modulation followed by the visualization of its biochemical mechanisms.

INTRODUCTION: Etoposide is an anti-cancer drug commonly known as VP-16 with its brand names Toposar VePesid, Lastet and NSC 141540. Etoposide has the chemical formula $C_{29}H_{32}O_{13}$. Its scientific name is 4'-demethyl-epipodophyllotoxin 9-[4, 6-O-(*R*)-ethylidene-beta-D-glucopyranoside], 4'-(dihydrogen phosphate) and scientific number 33419-42-0.

Etoposide is classified as a "plant alkaloids" which make up a group of chemotherapy medications. These plant alkaloid drugs are segregated into three sub-divisions based on the nature of plant they derived.

Vinca alkaloids derive from the Periwinkle plant. Taxanes from the Pacific yew tree. Epipodophyllotoxins taken from the May apple plant.



$C_{29}H_{32}O_{13}$

Synthesis:

Etoposide and teniposide make up the epipodophyllotoxin. It is a semi-synthetic derivative of podophyllotoxin and was first made in 1971 and approved by FDA in 1983. Podophyllotoxin (etoposide derivative) was first extracted from the rhizomes and dried roots of species of the genus *Podophyllin peltatum* (May apple). It was first treated with the reagent hydrogen bromide to produce 1-bromo-1-deoxyepipodophyllotoxin then bromine was

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replaced by a hydroxy group, resulting in 4-demethylepipodophyllotoxin. After that the 4-hydroxy group was coupled with 2, 3, 4, 6-tetra-*O*-acetyl- β -D-glucopyranose. The protecting group at the 4-hydroxy was removed by hydrogenolysis and the acyl groups by hydrolysis, and the cyclic *O*-4,6-acetal was formed by reaction with acetaldehyde dimethyl acetal¹.

Target Action of Etoposide:

The principal cytotoxic object for etoposide is the enzyme topoisomerase II. Etoposide, an antineoplastic agent kills the cell by stabilizing enzyme-cleaved DNA complex. Topoisomerase is the enzyme which controls the topological structure of DNA. This enzyme involved in the temporary cutting, winding and unwinding of DNA. For this purpose it is critically important at several different steps in transcription, replication and chromosomal structure. Topoisomerases II cut both strands of a double-stranded DNA molecule, get ahead another duplex through the cut, and then reseal it again by the process that utilizes ATP. Etoposide interferes

with the religation of DNA which causes stabilization of DNA–topoisomerase complex leads to the stabilization of cleavage complex. This accumulation generates permanent DNA strand breaks triggering recombination, mutagenesis, repair pathways and chromosomal translocations ultimately causing DNA strand break (Fig. 1).

When these breaks are permanent in the cell, then initiate death pathways and eventually inhibit DNA synthesis. Etoposide is cell cycle dependent and phase specific drug and essentially affects S and G2 phases of cell cycle. This grounds for significant errors in DNA synthesis at the premitotic phase of cell division and can lead to death of cancerous cell. This cytotoxicity lies in its capability to form a stable complex between topoisomerase II and DNA ultimately inducing intensive DNA damage. This confirms the effect of etoposide on topoisomerase II by converting it into a cellular toxin and inducing crumbling of genome leading to cell cycle arrest and apoptosis².

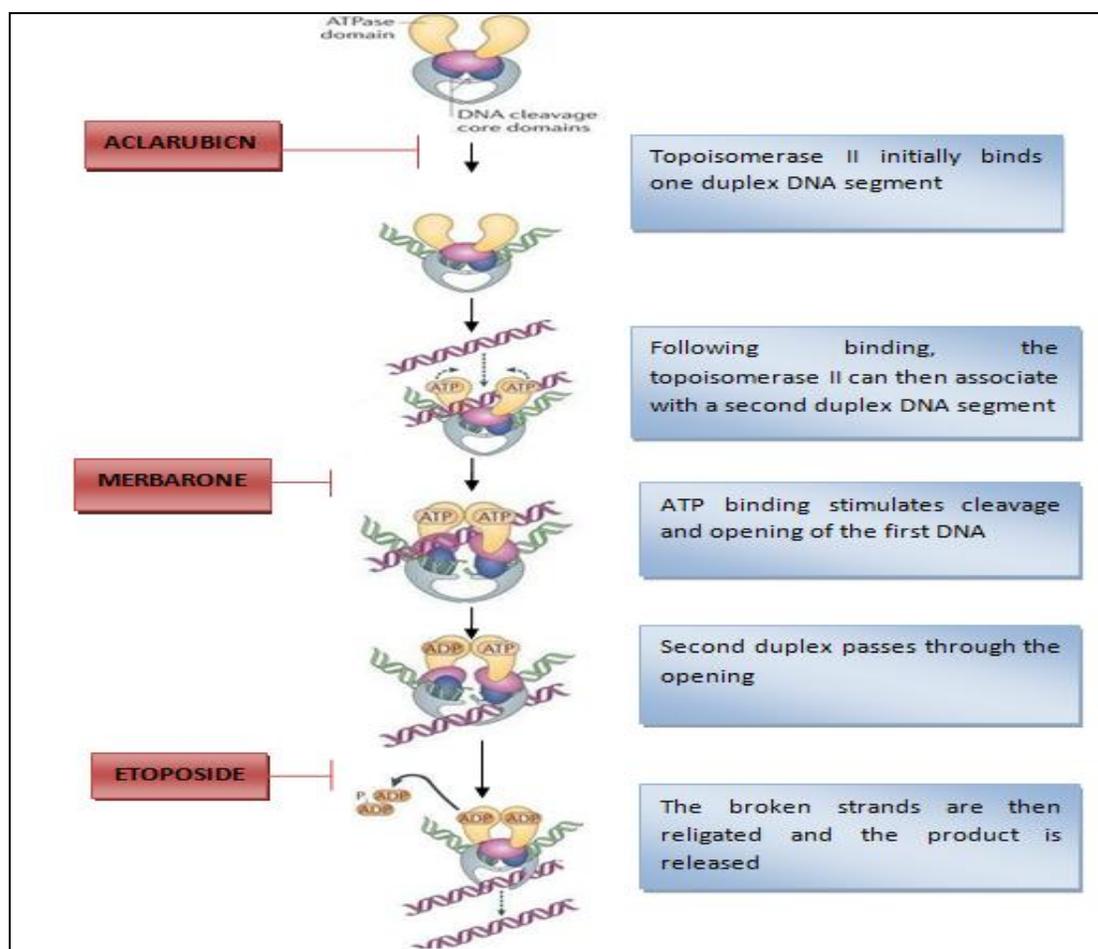


FIG. 1: CATALYTIC ACTIVITY OF TOPOISOMERASE II AND INHIBITORY ACTION OF ETOPOSIDE

Classification on the Basis of Functions:

Topoisomerase inhibitors belong to a class of drugs that blocks topoisomerases which may kill cancer cells. On the basis of their function they are classified in to three major groups.

- Topoisomerase II inhibitor compounds which interfere enzyme-DNA binding, the initial step of the cycle. These intercalating compounds include Aclarubicin and Suramin.
- These compounds form enzyme-cleaved DNA complex (cleavable complex) by stabilizing DNA topoisomerase II complexes and are termed as non intercalator compounds. Etoposide, Teniposide and Saintopin are common drugs of this group.
- These are also non intercalator compounds that do not induce formation of cleavable complex by blocking ATP activity which results in the inhibition of topoisomerase II-catalyzed DNA cleavage. This group includes Merbarone and ICRF-187^{3,4}.

Indications:

Etoposide is used as FDA approved drug for treating testicular cancer and lung cancer however it is also used in prostate, bladder, stomach and uterine cancers, mycosis fungoides, Hodgkin's and non-Hodgkin's lymphoma, Wilm's tumor, Kaposi's sarcoma, Ewing's sarcoma, rhabdomyosarcoma and neuroblastoma. Its high-dose therapy may also be recommended in bone marrow transplant settings.

Therapeutic Combinations:

Oral etoposide has been reported as sole agent in different treatments. It is effective in combination therapies with other anti-cancer drugs to show some degree of success.

- Etoposide in combination with ifosfamide shows synergistic effect towards malignant tumors of childhood⁵.
- In hormone- refractory prostate carcinoma combination of oral cyclophosphamide and oral etoposide is used⁶.

- DICE (dexamethasone, ifosfamide, cisplatin, etoposide) combination is used as infusional chemotherapy for refractory non-Hodgkin's lymphoma⁷.

Therapeutic Combinations in Lung Cancer:

- In combination with cisplatin it is also indicated for refractory small cell lung carcinoma^{8,9}.
- The combination etoposide/ifosfamide or cisplatin/ifosfamide is effective in treating non-small cell lung cancer. Combination of etoposide with ifosfamide is finer than combination with cisplatin owing to its better tolerability¹⁰.
- Combinations of paclitaxel and etoposide have also been reported in treatment of lung cancer¹¹.
- Clinical trials of etoposide, cyclophosphamide, and vincristine for small-cell lung cancer appear to be an effective treatment¹².

Common Side Effects of Etoposide:**Thrombocytopenia:**

Etoposide can reduce the platelet count by decreasing its production, which help in blood clotting. When platelets count is low it cause bleeding or bruising, such as nose bleeds, blood spots and rashes on the skin.

Leukopenia:

Low white blood count higher the risk of getting infections. White blood cells are important for inducing the immunity.

Alopecia:

This usually starts in 3–4 weeks after executing therapy, although it may happen earlier. Hair fall occurs completely.

Anorexia:

Etoposide can also cause metallic taste during chemotherapy and it lasts for several months. Normal taste usually comes back after treatment.

Fatigue:

Feeling exhausted is the most common associated side effect of chemotherapy, especially at the end of the therapy and remains for some weeks after treatment.

Less common side effect:**Anemia:**

Etoposide can lessen the number of RBC's red blood cells ultimately causing anemia. This may make feel breathless and tired.

Nausea Vomiting Diarrhea:

Nausea and vomiting may begin after the treatment and last for up to five days. Etoposide can also cause diarrhea. This can easily controlled by medications.

Skin changes:

Itchy skin rashes may occur and skin may be darken butgenerally comes to normal in few months after the treatment.

Sleeplessness, headaches and confusion:

These effects occur only in high-dose treatment and are very rare.

Allergic reaction:

Allergic reaction includes high temperature, shivering, dizziness, a headache, reddening of the face, breathlessness, anxiety, and urticaria.

Adverse Reactions:**Acute leukemia:**

Etoposide can cause second cancer usually acute leukemia that develops years later with etoposide treatment.

Influx of Etoposide:

Etoposide cellular uptake is diffusion dependent. Not any specialized transporter apparently takes part in etoposide transport while considerable saturated secretion occurs through intestinal tissues. Transport Once it enters in the cell, it has cytotoxic effect. Etoposide phosphate, is not diffusible in the cell. Unless alkaline phosphatase cleaves its phosphate group and make it cytotoxic agent, etoposide. Amphipathic nature of etoposide allows its association with hydrophobic domains of membrane proteins or membrane lipid¹³.

Metabolism:

Etoposide metabolism is arbitrated by CYP3A4 and CYP3A5, both transcriptionally controlled by NR1I2. Further, etoposide can also be converted into the O-demethylated metabolites and, can be catalyzed by prostaglandin synthases or myeloperoxidase. These metabolites share same strength of hampering topoisomerase II and are more reactive than their parent drug. Inactivation of parent drug and metabolites is induced by glutathione and glucuronide conjugation. ABCC1, ABCC3 and ABCB1 are involved in efflux of conjugated or unconjugated forms of etoposide and represent the mechanisms of drug resistance. Etoposide glucuronide acts as a good substrate for both ABCC2 and ABCC3 and the elimination of this metabolite from the liver is almost completely dependent on these efflux pumps¹⁴.

Efflux of Etoposide:

A family of transmembrane proteins, ATP-binding cassette (ABC) transporters, is involved in transportation of a wide array of substrates across biological membranes through an ATP dependent reaction. ATP-binding cassette (ABC) multidrug transporters, for instance P-glycoprotein (P-gp, ABCB1) and multi-drug resistance proteins (ABCC2 and ABCC3), may have an important impact on cancer chemotherapy. From recent data it has been proved that ABCC1 is a good transporter for etoposide as a substrate. Furthermore, it has also been seen that both ABCC2 and ABCC3 can moderately transport etoposide. ABCB1 transports various substrates across the cell membrane including chemotherapeutic agents such as etoposide and it is an important transporter for etoposide.

Transporters of Etoposide:**MDR1 (Multi-Drug Resistance1):**

The (ABCB1) gene is located on chromosome 7q21.12 which encodes P-gp and used as a transporter in many hydrophobic substrates and anti-tumor drugs including etoposide, mainly expressed in organs such as intestine and liver involved in the excretion. In lung, P-gp express at the apical side of ciliated collecting ducts or ciliated epithelial cells, and also expressed on the apical and sideways surfaces of serous cells of

bronchial glands but not in mucus-secreting cells. The MDR (ABCB) has two distinct sites, H site and R site, for drug binding and transport that interacts in a positively supportive manner. The two sites have more or less identical affinity for etoposide. These transporters are present in the lung so are involved in treating small cell lung cancer which is mediated by etoposide.

Main transporters of etoposide are following:

1. Multidrug resistance protein 1 (ABCB1)
2. Multidrug resistance-associated protein 6 (ABCC6)
3. Canalicular multispecific organic anion transporter 2 (ABCC3)
4. Multidrug resistance-associated protein 1 (ABCC1)
5. Multidrug resistance-associated protein 7 (ABCC10)
6. Canalicular multispecific organic anion transporter 1 (ABCC2)
7. ATP-binding cassette sub-family G member 2 (ABCG2)

ABC transporters can be expressed in tumor cells where they can actively efflux a broad spectrum of different anticancer drugs and also contribute to multidrug resistance. These efflux pumps have broad and substantially overlapping substrate specificities¹⁵.

Drug Active Sites:

The drug is composed of a polycyclic ring system (rings A-B-C-D rings), a glycosidic moiety at C4 position, and a pendant ring (E-ring) at the C1 position. A, B, and E-rings of etoposide by substituents appear to be referred in the interaction between etoposide and human topoisomerase II α in the binary enzyme-drug complex. The protons of the A-ring, the H5 and H8 protons of the B-ring, as well as the H2 and H6 protons and the 3' and 5' methoxyl protons of the pendent E-ring interact with both enzymes in the binary protein-ligand complexes.

Substituents on E-ring of etoposide are necessary for drug action against topoisomerase II. E-ring is intimately associated with the protein in the binary complex. Since the E-ring has free rotation about the 1'-linkage to the C-ring, the H2' and H6' protons, as well as the 3'- and 5'-methoxyl groups. Although the 3'-methoxyl, 5'-methoxyl, and 4'-hydroxyl moieties of the E-ring all are important for etoposide task. The glycosidic moiety of etoposide, which is attached to the C-ring at the C4 position, does not interact with topoisomerase II α in the binary enzyme-drug complex. D-ring of etoposide also does not appear to contact topoisomerase II α in complex. Alterations in the D-ring may affect etoposide activity toward topoisomerase II α in the ternary enzyme drug-DNA complex^{16,17}.

DNA enzyme complex, the most likely target of Etoposide:

Topoisomerase II use to generate transient double-stranded breaks in the backbone of nucleic acid. In order to sustain the integrity of the cleaved genetic material throughout drug activity, this enzyme forms proteinaceous bridges that extend the nucleic acid break. This bridge is affixed by covalent phosphotyrosyl bonds, customary between the active site of the homodimeric enzyme and the newly created 5' termini of DNA. Because of this cleavage complex (topoisomerase II-cleaved DNA complex) is normally short-lived intermediate in the catalytic cyclic activity of enzyme, which is endured by the cell. However, by the treatment of topoisomerase inhibitor drugs, when cleavage complexes present in high amount, become possibly toxic, endorsing illegitimate recombinations, frame shift mutations, permanent double-stranded DNA breaks and cell death. The cytotoxic prospective of topoisomerase II inhibitors has now been utilized clinically by the expansion of antitumor drugs. Rapidly proliferating cells are most susceptible to these agents because containing high concentrations of topoisomerase II in aggressive malignancies.

Etoposide induced scission reveals the ability to inhibit religation at specific sequences (rather than direct drug DNA) interactions that mediate cleavage complex formation. These conclusions are reliable with the recently proposed "positional

poison model” for the action of engendering to DNA lesions by topoisomerase II-targeted anticancer drug (Fig. 2). Finally it is proposed that the primary pathway for the non-covalent enzyme-

drug and DNA ternary complex formation is through etoposide-topoisomerase II interactions leads to DNA fragmentation and apoptosis¹⁸.

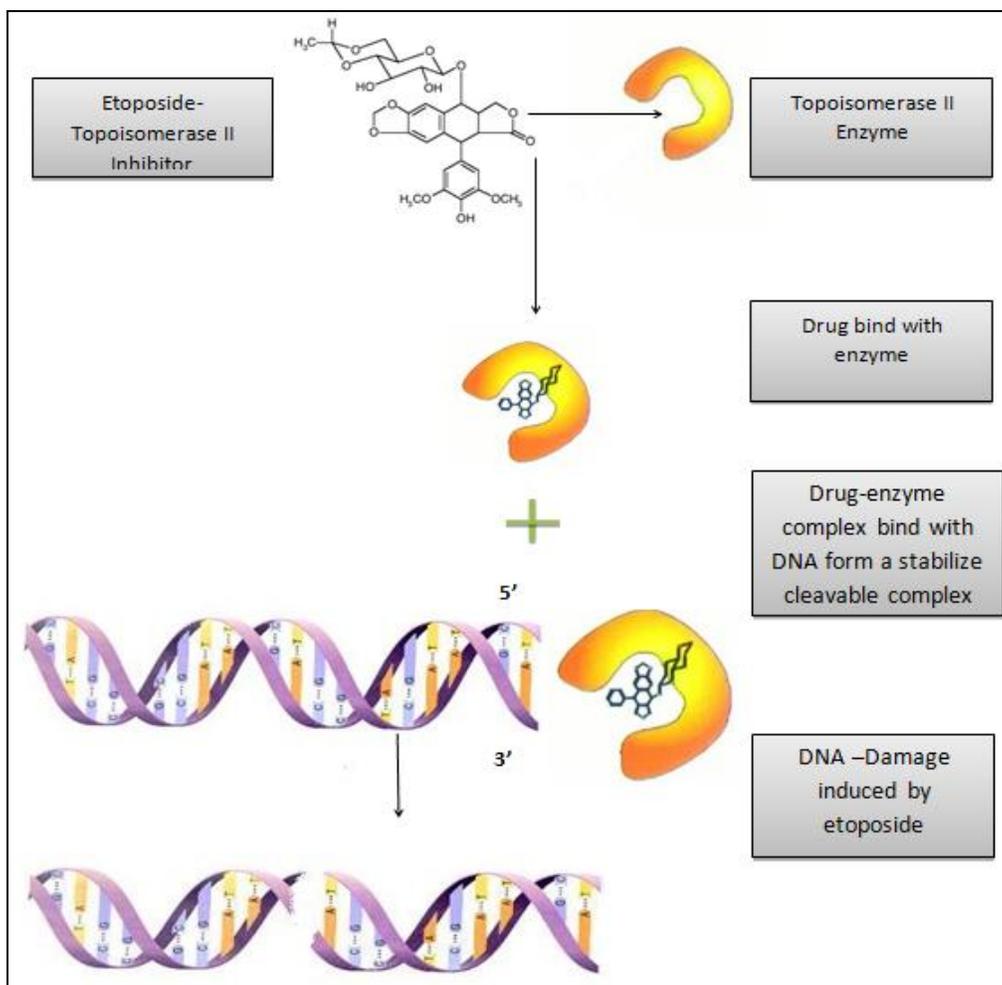


FIG. 2: DRUG BINDING AND STABILIZATION OF CLEAVABLE COMPLEX

Biochemical Mechanism of Action of Etoposide: ATR mediated pathway in response to single strand break:

Etoposide acts after Topo II mediated DNA cleavage, in which Topo II enzyme covalently binds to DNA- 5' termini, whereas DNA-3' termini remains free cause the formation of lesions. These lesions develop in single-strand DNA gaps and double-strand DNA breaks. This causes the stimulation of ATR (Ataxia telangiectasia Rad3-related) pathway which is chiefly involved in the reaction to the halting of replication forks by etoposide, results in the development of extended single-stranded DNA regions¹⁹. Etoposide mediated ATR pathway entails the activity of the Ataxia telangiectasia Rad3-related (ATR) checkpoint kinase. Etoposide-induced DNA damage happens

behind the replication fork and this damaged DNA is further transformed into extended single-stranded DNA region (ssDNA) which is coated with RPA (Replication protein A)²⁰. ATR recognition of RPA-coated single strand DNA depends on ATR-interacting protein (ATRIP). ATR signaling is reliant on co-localization of the ATR-ATRIP complex with the Rad9-Rad1-Hus1 (9-1-1) complex, a heterotrimeric ring-shaped molecule. 9-1-1 complex carries a critical activator, TOPBP1 to ATR. TOPBP1 is obligatory for ATR activation that can arouse ATR activity²¹.

Once ATR complex is amassed with DNA lesion prompts signals to synchronize cell cycle, and processes of repair and replication are commenced. ATR phosphorylates CHK1 on Ser317 and Ser345,

which is a trustworthy marker of CHK1 activation. Once CHK1 is phosphorylated, it release from chromatin material and further phosphorylates its substrates. Activated CHK1, phosphorylates Cdc25A protein phosphatase which uphold its ubiquitin-arbitrated proteolysis. Loss of Cdc25A leads to S-phase and G2-phases arrest in the of the cell division cycle. ATR signaling is critical for regulating replication either by the regulation of CHK1-CDC25 pathway^{22, 23}. ATR is principally a replication stress response and it can also be activated by double strand DNA break (**Fig. 3**). ATR is triggered more gradually and chiefly in S and G2 phases of cell cycle. This steady ATR activation at double-strand break site is a necessity for CDK-dependent DSB ends resection, which exposes a large single-stranded region. It can also

be proposed that DSBs guides to ATM stimulation and the repair of DSBs can turn out RPA-coated ssDNA that elicits ATR pathway²². It may occur at TOPBP1 level by its phosphorylation through ATM. Phosphorylated TOPBP1 is more proficient activator of ATR²¹.

ATR is one of the kinases which involves in the phosphorylation of Chk1 in reaction to genomic stress. P53 is also phosphorylated by Chk1 and this phosphorylation prompts downstream activation of p53 and its target genes i.e., Bax and PUMA (p53-upregulated modulator of apoptosis) ultimately leading to apoptosis, while transcriptional activation of p21 and 14-3-3 causes cell cycle arrest in G1 phase²⁴.

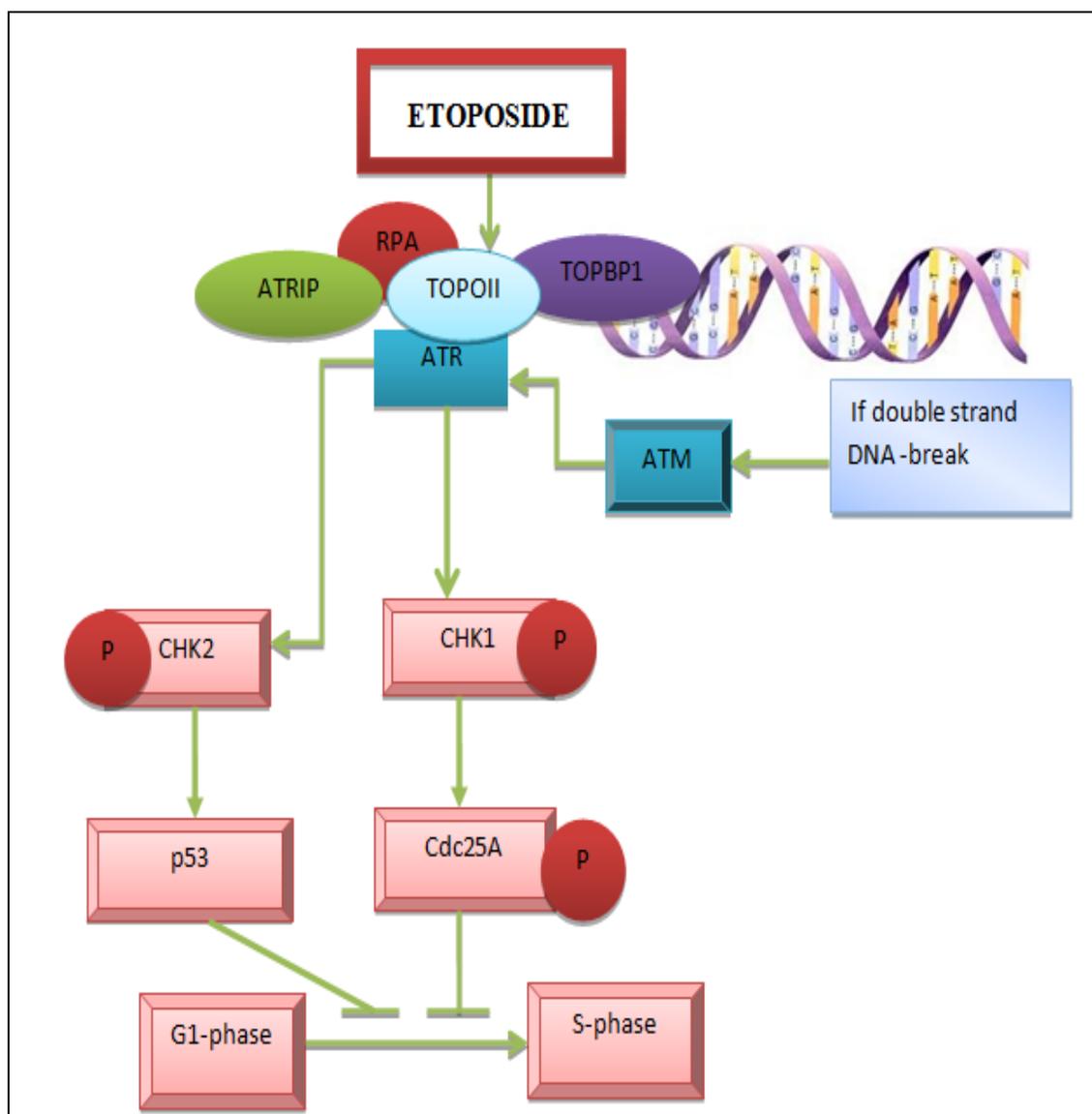


FIG. 3: ATR ACTIVATION BY ETOPOSIDE

ATM mediated pathway in response to double strand break:

Ataxia telangiectasia mutated (ATM) manganese dependent serine-threonine protein kinase is a tumor suppressor protein, signals the presence of DNA double strand break in mammalian cell. DNA damage results in ionizing radiations, replication error or chemicals. Chemotherapeutic drug etoposide also cause DNA damage. Etoposide, a topo II inhibitor drug stabilize cleavable complex (enzyme-DNA complex) cause DNA double strand break, leads to the activation of ATM, a key player in multiple signaling cascade in response to double strand DNA break and acts as a master switch and regulator in cell cycle progression. After etoposide treatment, ATM response to cell cycle checkpoints factors (chk factors) DNA repair or apoptosis (**Fig. 4**). ATM is activated by MDC complex include three proteins the MRE11 (meiotic recombination-11), Rad50 and NBS1 (Nijmegen's breakage syndrome). ATM first phosphorylates NBS1 on several residues in response to DNA damage and MRE11/Rad50/NBS1 complex is needed for chk2 activation. The phosphorylation of p53 is controlled by CHK2 or ATM directly phosphorylates p53. When ATM phosphorylates serine 15 it activates p53 and N-terminal for chk2 activation²⁵.

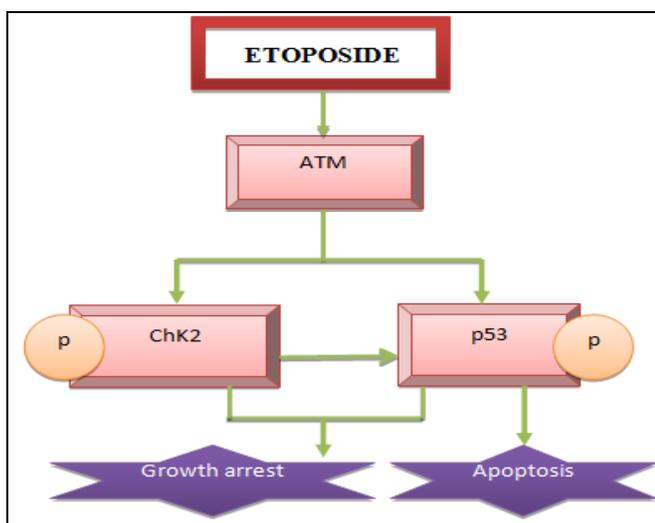


FIG. 4: MECHANISM OF ATM IN GROWTH ARREST AND APOPTOSIS

P53, dependent functions:

P53 is tumor suppressor and have key function in the cellular response towards genomic toxicity²⁶. Etoposide cause the phosphorylation of p53 within 6 hrs. A stress signal is conveyed to the p53 protein

by post-translational modifications and answers the activation of the p53 protein as a transcriptional factor that instigates a program of cell cycle arrest and cellular senescence. Further p53-responsive and interactive proteins are produced by transcriptional network of p53 that interrelate with an outsized number of other signal transduction pathways and account for a numerous positive and negative auto-regulatory feedback loops. Many factors regulate p53 activation and its downstream response depends on cellular environment. Etoposide treatment is liable to a speedy and widespread initiation of cell death and additional increase in p53 and PUMA expressions as well as Bax processing. Transcriptional activation of gene encoding p21 and 14-3-3 leads to G1 cycle arrest^{27, 28}.

P53 and Growth Arrest:

Transcriptional activation of p53 can inhibit both G1 to S and G2 to M phase of cell cycle. P53 mediated transcriptional induction of p21 has a foremost role in mediating G1 arrest. In cell cycle progression, p21 can inhibit Cdk2 which requires for S phase, resulting in the buildup of hypophosphorylated Rb (retinoblastoma) directing to G1 to S phase arrest (**Fig. 5**). The M phase progression entails Cdc2 which can be hindered by several proteins like p21, GADD45 or 14-3.3. A direct transcriptional repression of cdc2/cyclin B by p53 also results in G2 to M phase arrest. P53 controls the expression of these inhibitory proteins to provoke growth arrest²⁹.

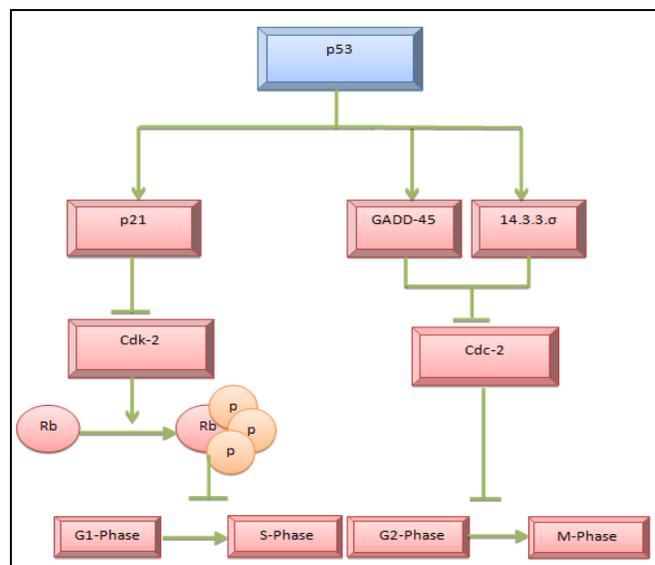


FIG. 5: P53 ROLE IN GROWTH ARREST

P53 and apoptosis:

Etoposide induce apoptosis in p53 dependent-pathway, activated in response to diverse stress stimuli under the action of ATM²⁶. p53, a transcription factor mediates its diverse functions by two routes to apoptosis either by "intrinsic or mitochondrial" cell death pathway or "extrinsic cell death pathway".

P53 Induced Intrinsic pathway:

Intrinsic apoptotic pathway is triggered, through application of the DNA-damaging topoisomerase inhibitors etoposide³⁰. Phosphorylation of p53 results in the up-regulation of proteins in the nucleus induce expression or transcriptional activation of proapoptotic Bcl-2 family proteins (BAX, NOXA, PUMA, BID, CD95, APAF-1, DR5, p53AIP1) implicated in apoptosis^{31, 32, 33}. The intrinsic apoptotic pathway is dictated by the expression of Bcl-2 family proteins, directs the release of cytochrome c from the mitochondria leads to apoptosis through caspase cascade (**Fig. 6**). NOXA and PUMA genes are direct transcriptional

targets of p53 and have critical role for p53-induced apoptosis. Increased expression of PUMA promotes mitochondrial translocation and multimerization of Bax³⁴.

Bcl-2 family comprises anti-apoptotic (pro-survival) and pro-apoptotic proteins. Bcl-2 family includes: anti apoptotic proteins, whose members are most structurally similar to Bcl-2, such as Bcl-X_L, and pro-apoptotic proteins Bax, Bak and BH3-only proteins^{35, 36}. Inhibition of proapoptotic proteins through antiapoptotic Bcl-2 members is prevented by activated BH3-only proteins. Further a conformational change in Bax and Bak proteins occur which direct their insertion into mitochondrial membrane through oligomerization and pores formation. This whole leads to release of proapoptotic factors from mitochondria into cytochrome c and a series of events is initiated in which binding of cytochrome c with Apaf1 occurs and apoptosome is formed that consequently triggers caspase cascade causing cell death³⁷.

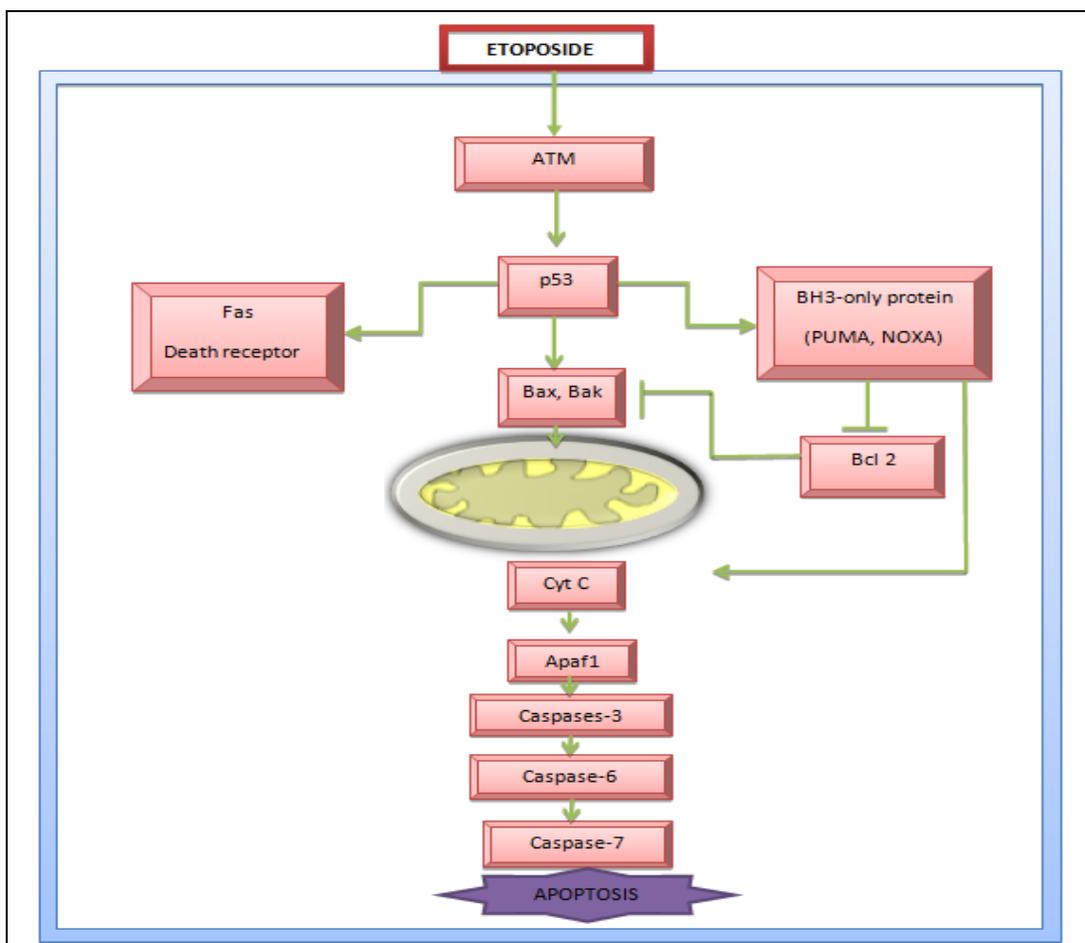


FIG. 6: INTRINSIC APOPTOTIC PATHWAY BY P53

P53 Induced Extrinsic pathway:

Etoposide through its genotoxic function can up-regulate p53 and turn on death receptor pathway. P53 activates various genes by transcription, involved in cell death as Fas receptor³⁸. Etoposide may rapidly up-regulate death receptor Fas or Fas ligand by this means directly triggering the apoptotic pathway³⁹. Upon an apoptotic stimulus, stem helices are allowed by Fas death domains to interact and stabilize the open form. FADD (Fas-associated death domain) can then bind to the Fas molecules; this consequence is procedural interlinked DISC (death-inducible signaling complex) formation. This allows activation of caspase 8, via its oligomerisation causes the induction of apoptosis⁴⁰. DISC places the molecules of procaspase-8 in a spatial orientation favorable for their activation^{41, 42}. c-FLIP has a high affinity to bind to the DISC and is structurally similar to caspase-8 so it is an apoptotic inhibitor⁴³.

Caspase8 mediated apoptotic pathway:

Outer mitochondrial membrane localized caspase 8 slices cytosolic procaspase 3 directly and turns on it. Once caspase-3 is activated during etoposide-induced apoptosis, MST1 will be cleaved and activated. MST1 (mammalian STE20-like kinase 1) is a serine/threonine kinase, over-expression of which brings about apoptotic morphological changes such as chromatin condensation. The cleaved N-terminal kinase domain of MST1 translocates into the nucleus and mediates the phosphorylation of histone H2AX, and the phosphorylated H2AX then recruits caspase-activated DNase to mediate DNA fragmentation. During the procedure of etoposide-induced apoptosis, the breakage of MST1 robustly comes with strong H2AX phosphorylation at Ser-139 recruits caspase-activated DNase to mediate DNA fragmentation⁴⁴. The caspase 3 cleaves DFF45 (DNA fragmentation factor 45), dissociates from DFF40, inducing oligomerization of DFF40 (a heterodimeric factor of DFF40) that has DNase activity.

Caspase 8 mediated intrinsic pathway:

Etoposide induced Caspase 8 pathway is activated through FADD/TRADD-dependent mechanisms

not only trigger extrinsic apoptotic pathway via caspase 3, also recruit mitochondrial apoptotic signaling^{45, 46}.

Activated caspase 8, by the DISC cleavage, directly activates caspase 3 as well as breaks the pro-apoptotic Bcl-2 family member Bid, forming the truncated form of Bid, t-Bid. t-Bid then triggers apoptotic pathway by translocation in to outer mitochondrial membrane (OMM) and promotes oligomerization of Bax or Bak, results the release of cytochrome *c* and other apoptogenic proteins from the intermembrane space (IMS) of the mitochondria into the cytosol⁴⁷. This step can be antagonized by the anti-apoptotic Bcl-2 family members. The released cytochrome *c*, interacts with Apaf-1 (Apoptotic protease-activating factor 1) and, in the presence of (d)ATP, gets attached to adapter protein Apaf-1 and activates it, as a consequence caspase 9 are recruited, causing the formation of apoptosome, stimulation of caspase 9, and subsequent execution of caspases, caspase 3, 6 or 7 which propagate further caspase processing events, leads to cell death⁴⁸.

Caspase8 induced mitochondrial fragmentation:

Caspase 8 target Bid, a direct activator of integral polytopic ER membrane protein Bap31, the cytosolic tail of which is cleaved by caspase-8 to produce proapoptotic Bap20^{45, 46}. It can lead to severe ER stress by provoking Ca²⁺ release and enhanced mitochondrial fission. In addition to this, p20 also interferes with Bap31-mediated protein trafficking and causes a speedy conduction of ER calcium signals. These Ca²⁺ signals stimulate DRP1 (dynamin-related protein) dependent organelle fission and cause the release of cytochrome *c*. Anti-apoptotic members of Bcl-2 family fundamentally targets ER calcium homeostasis and Bap31/procaspase-8 complexes⁴⁹.

These evidences suggest that signaling pathway via etoposide treatment activate p53 which stimulate caspase 8 causes the cleavage of BAP31 to BAP20 on the ER. ER causes the Ca²⁺ release uptake by mitochondria (**Fig. 7**). When Ca²⁺ level exceed mitochondrial fission occurs, eventually contribution to apoptosis by interrupting mitochondrial integrity and function^{50, 51}.

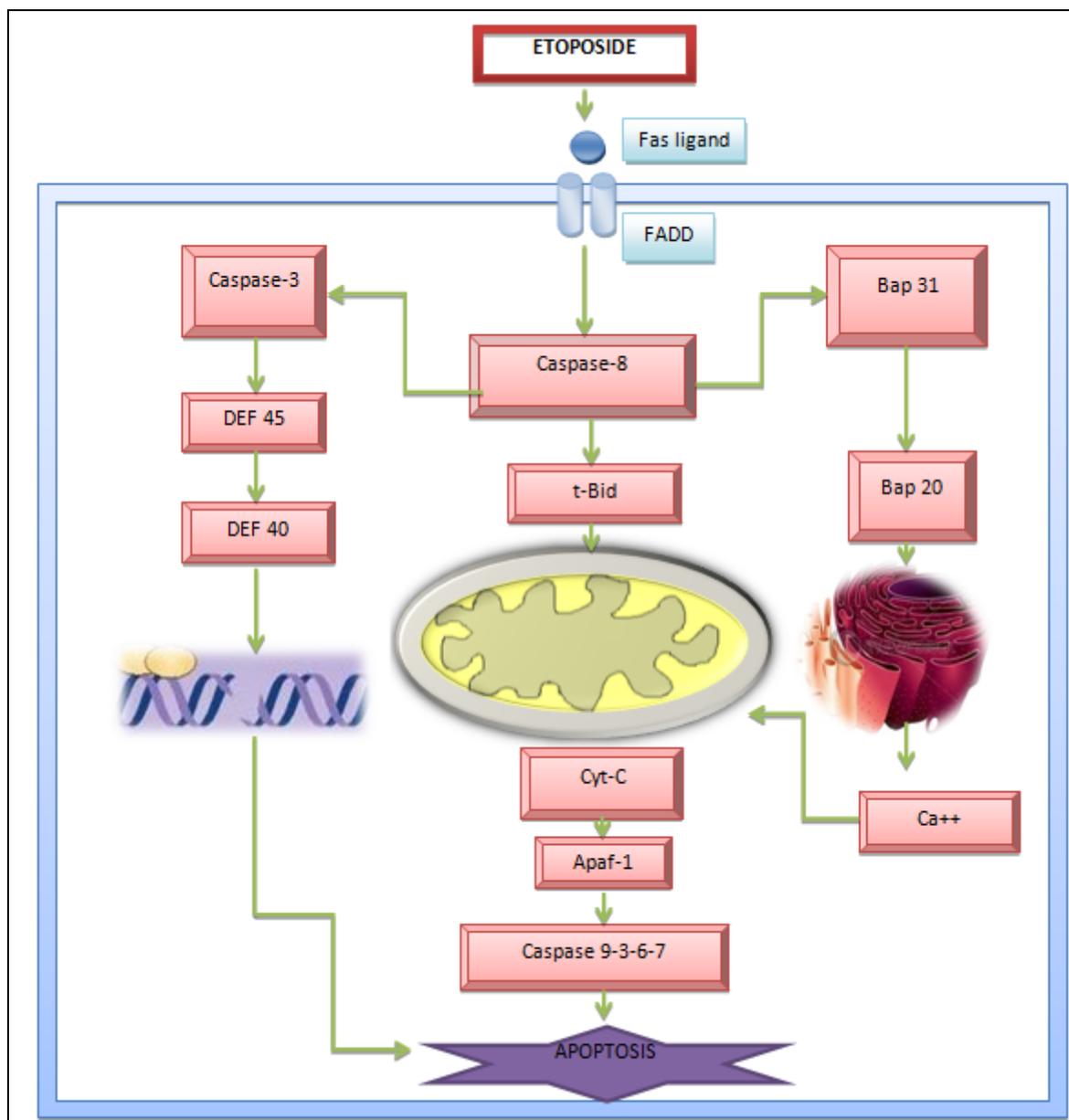


FIG. 7: ROLE OF CASPASE 8 IN APOPTOSIS

Role of NF- κ B:

Etoposide causes DNA damage, treated by the main DNA damage sensor p53 which is transactivated by p53 provoked protein with death domain (PIDD) and activates NF- κ B. Upon DNA damage NF- κ B shifts to the nucleus and activate transcription of DR4 receptor gene. However, it is observed that a dynamic induction of the p53-dependent phosphorylation of FADD at Ser194 brings a crucial change in sensitization to DNA damage by augmenting MEKK1 pathway. The phosphorylation of p65 is carried out through p53 dependent death receptor expression. Other receptors like DR3, DR4 and DR5 are also stimulated by etoposide. Cells can express a

copiously functional relation among p53 and p65 by Fas receptor assembly which are highly receptive to etoposide. Fas receptor has ability to activate p65 in FasL-independent manner and caspase-dependent apoptosis as well p65 induce apoptotic death by caspase-independent manner^{38, 52}. Tumor necrosis related apoptosis inducing ligand (TRAIL/Apo-2L) that attaches to death receptor-4 (DR4, TRAIL-R1) and death receptor-5 (DR5, TRAIL-R2) is a protein that persuades cell death in a multiple type of cancers rather than normal non-transformed cells. Etoposide augmented DR4 protein levels escorts to apoptosis with the TRAIL induced cell death pathway. Etoposide express DR5 gene through the

collaboration of transcription factors NF- κ B and p53 by binding with DR5. The DR4 protein resides on the plasma membrane; upon attachment with TRAIL it activates caspase-8 and forms the DISC (death inducing signaling complex) to executed death signaling program ensuing apoptosis.

Ceramide Production by Etoposide:

Ceramide, a tumor-suppressor lipid is generated, when cells come across stimuli of apoptotic signaling, for instance radiation (UV and γ -irradiation), death receptors (CD95 and TNF receptor), or subjected to genotoxic chemotherapy. Etoposide causes accumulation of ceramide derived from SM hydrolysis under the action of SMase, functions as an important mediator of the apoptosis of glial cell induced by etoposide⁵³.

Ceramide up-regulate thioredoxin-interacting protein (Txnip) which is a tumor suppressor gene, so, etoposide, by this way is involved in provoking apoptosis. Txnip; also termed as vitamin D3 up-regulated protein 1, endogenously hampers thioredoxin, which is a small redoxprotein. Thioredoxin associates with ASK1 and encourages ASK1 degradation and ubiquitination to restrain ASK1-arbitrated apoptosis by inhibiting ASK1 kinase activity. Elevated Txnip levels results in ASK1 instigation.

MAPK related apoptotic pathway:

ASK1 activation:

ASK1 after being phosphorylated and oligomerization with threonine residues in its activation loop is dissociated from thioredoxin. Txnip induction by ceramide, cause an increase dephosphorylation at the inhibitory residue Ser83 and phosphorylation of ASK1 at activation residue Thr845. Thioredoxin blocks activity of apoptosis signal-regulating kinase 1 (ASK1) and also decrease the consequent ASK1-dependent apoptosis. Txnip eradicates this effect, therefore, Txnip possesses proapoptotic role and is a stress-responsive protein. Txnip signaling regulate ASK1, p38 MAPK- and JNK-mediated pathways, as well as endoplasmic reticulum (ER) stress death signaling pathways. ASK1 is the member of MAPKKK family. Ceramide induces apoptosis through phosphorylation of p38 MAPK and JNK⁵⁴.

Activation of MEKK1:

Etoposide causes DNA damage and activates MEKK1 (Mitogen-activated protein kinase) and NF- κ B a protein. MEK kinase 1 (MEKK1) induces apoptosis through both caspase amplification and transcriptional activation or regulation. MEKK1 activates through apoptotic pathway leads to the cleavage by caspase 3-like protease into the 91kDa kinase domain in reaction to DNA damaging or genotoxic mediators such as etoposide⁵⁵.

Fas-induced activation of MEKK1 via caspase 3 or related proteases slices MEKK1. FADD phosphorylation contributes to the activation of MEKK1 pathway at Ser194, can up-regulate MEKK1 pathway. In apoptosis, caspase 3 and related proteases break MEKK1 at the peptide bond after D68 and sets it free from a compartment, triton-insoluble cytoplasmic complex. MEKK1 involved in pro-survival and pro-apoptotic pathways is an activator of JNKc-Jun N-terminal kinase.

MEKK1 activates the (JNK); as well as it turns on IKK causing the degradation of I κ B and the stimulation of NF- κ B. MEKK1 phosphorylates the inhibitor I κ B leads to its breakdown proceeding with activation of NF- κ B.

JNK Activation:

The regulation of JNK is done by multiple MAPKKK' proteins, involving MEKK1, MEKK4 and ASK1, that is regulated by Rac1 and GTPases. The MAPKK, MKK4/7, dually phosphorylates JNK within the Thr¹³⁸-Pro-Tyr¹⁸⁵ motif (pTppY) in the catalytic core of active JNK. JNK is also induced by the PKC δ activation is needed for the unrelenting activation of JNK and the inactivation of ERK. PKC δ is activated through caspase dependent manner eventually leading the stimulation of JNK and inhibition of ERK. ERK1 and ERK2 has critical role in mitogenic signaling and oncogenesis. There is also evidence that activation of ERKs stimulates protection against apoptosis mediated factors (**Fig. 8**). On the other hand JNK activation is significant and sometimes ample to provoke apoptosis⁵⁶.

JNK prompts extrinsic apoptotic signaling through up regulation of death receptors and their ligands.

JNK persuades apoptosis by diverse mechanisms, including cytochrome release and needs the pro-apoptotic (Bcl-2) family proteins, Bax (Bcl-2 associated x protein) and Bak1 (BCL-2 antagonist killer1). Active JNK robustly connected to

transcription-dependent apoptotic signaling, chiefly through the commencement of c-Jun and other AP-1 proteins such as JunD, JunB and ATF-2 and may influence apoptosis by up regulating apoptosis-related genes⁵⁷.

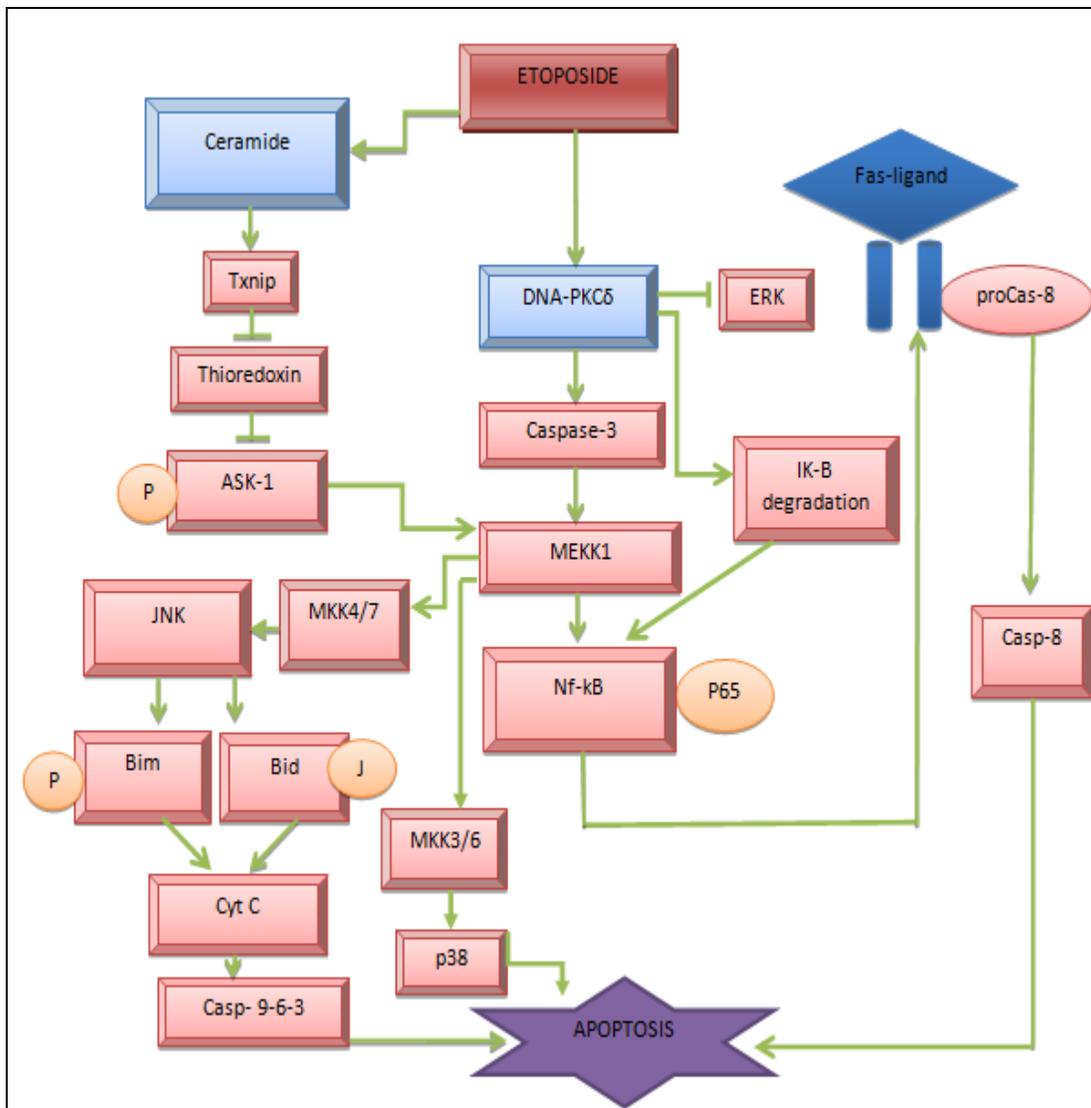


FIG. 8: ACTIVATION OF MAPK FAMILY PROTEINS VIA DNA-PK'S AND CERAMIDE

c-Abl pathway:

Cellular responses to genotoxic stresses after etoposide treatment including the activation of c-Abl, JNK and p53 expression. Apoptosis resulted in the degradation and activation of c-Abl after etoposide treatment, activates c-Abl kinases. The activated c-Abl plays a role in amplification of apoptosis induced by etoposide^{58, 59, 60}.

The proto-oncoprotein c-Abl is a member of the Src family of non-receptor tyrosine kinases. c-Abl is ubiquitously expressed in the cells and concentrates

both in the cytoplasm and nucleus where it is engaged in definite roles. c-Abl protein includes diverse domains which shows high affinity for phosphorylated tyrosine like Src homology 2 (SH2) domain. SH3 (Src78homology 3) domain binds to proline-rich domains containing PxxP motif as well as c-Abl SH3 binding proteins proved fruitful in regard to characterize its immediate downstream targets proteins, such as DNA-PK, ATM, the Abl family proteins and transcriptional factor p73. The exact mechanism of c-Abl activation is not known however on the basis of data it was seen that its

activity relies on ataxia-telangiectasia-mutated (ATM) gene. Atm is a nuclear member of a family of phosphatidylinositol-3-kinase like enzymes which phosphorylates c-Abl by binding at serine 465 residue. It is also reported that with regard to DNA-PK it used to activate a similar sequence of events⁶¹.

Caspases cleave c-Abl into at least three fragments. Different caspases like Caspase 8, caspase 10 and the effector caspase 3 cleave c-Abl, and sequential cleavage depends on the action of different caspases. c-Abl may be involved in the G1-S checkpoint with an intact kinase domain. G1 arrest mechanism by c-Abl is unclear, however several observations shown that c-Abl binds with p53. c-Abl-p53 complex up regulates p21 in the DNA damage signaling pathway that causes G1 arrest. c-Abl encourages the functions of p53 by enhancing the expression of the target gene p21 and by enhancing its transcriptional functions. Indirect job of c-Abl in G1 arrest now obvious from that c-Abl can control the p53 level by inhibiting Mdm2-mediated degradation of p53 and supporting p53 accumulation⁶².

p73 Mediated Growth Arrest:

p53 and its homologue p73 have highly significant sequences and functional similarities. p73 is an important developmental gene and also expected as a tumour suppressor. However, p73 activation after DNA damage induces by a way that is distinct from

that of p53. p73, a member of the p53 gene family, which is phosphorylated by c-Abl. P73, a high sequence homologous of p53 induces both cell cycle arrest G1 and apoptosis (Fig. 9). Unlike p53, it can directly interact with c-Abl. The induction of G1-arrest by c-Abl remains unclear. The verity that p73 is able to encourage p21 expression, leads to G1 growth arrest^{63, 64}.

p73 Mediated Apoptosis:

Molecular mechanisms through which p73 induces apoptosis are remain unclear. p73, confined in the nucleus and remains in nucleus during apoptosis, indicates that p73 has indirect effect on Bax translocation. p73 can also directly transactivate PUMA and this direct effect of PUMA on Bax conformation and mitochondrial relocation propose that there is a molecular association between mitochondrial apoptotic pathway and p73^{65, 66}. p73 mediated direct transactivation of PUMA occur rapidly and stronger than direct transactivation of Bax. PUMA used to translocates Bax into mitochondria and provides the fast conformational alterations of Bax from an inactive to an active state which can causes Bax mitochondrial translocation and release of cytochrome c induced by p73 mediated apoptosis⁶⁶. Evidence provides several possible downstream effectors activation which are the transcription factors p73 and p53 and also the stress-activated kinases, p38 and JNK⁶⁷.

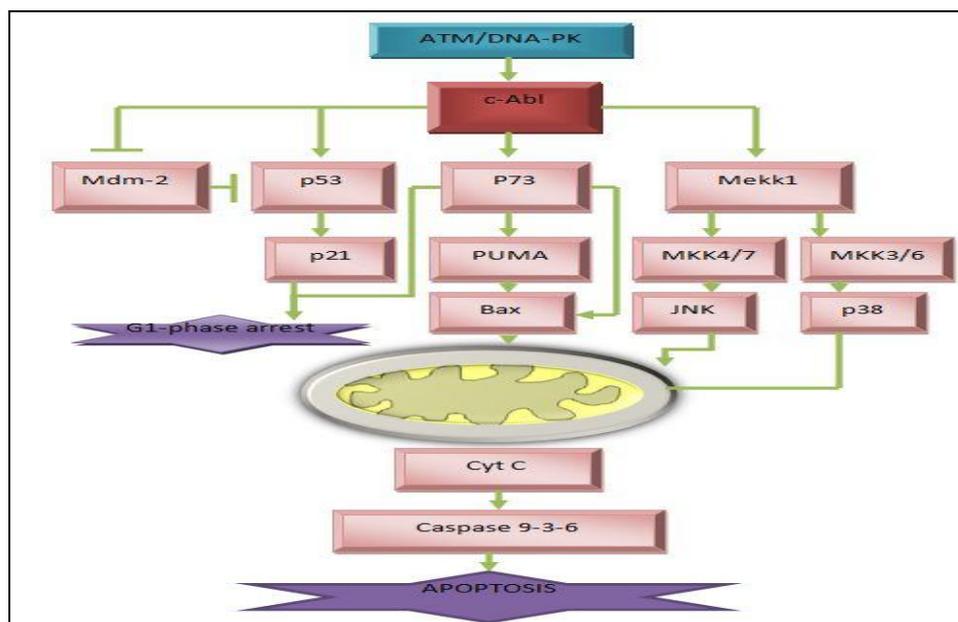


FIG. 9: cABL ACTIVATION AND ROLE OF p73 ISOFORM

Molecular Basis of Etoposide Resistance:

Ras/Raf/MEK/ERK pathway and cell survival:

After growth factor stimulation of the appropriate receptor, results in the packing of membrane-bound Ras with GTP. Ras can be activated by growth factor receptor tyrosine kinases [GFRTK]. Ras/GTP then engages Raf to membrane where it is activated. Both Raf and Ras are proto-oncogenes, and brings about the deregulation of the ERK signaling that entails anti-apoptotic pathway. In more than 30% of common cancers the deregulation of ERK pathways occur, therefore, Raf kinases are a striking target for novel treatments with aim to reverse the deregulation process (Fig. 10). Raf-1 acts as deactivator of

apoptosis via its target MEK/ERK that are capable to inhibit activated caspases; through stimulation of the anti-apoptotic factor NF-kB. NF-kB controls the expression of IAPs, phosphorylates Bad and cause it to relocate into cytosol from mitochondria, in this manner shielding the mitochondria⁶⁸. Activated ERK can be translocated to the nucleus and phosphorylate additional transcription factors like CREB, Elk-1, Fos and globin transcription factor 1 (Gata-1) and others which can bind with the promoters of many genes, including growth factor and cytokine genes. These genes are vital in promoting growth and averting apoptosis of multiple cell types⁶⁹.

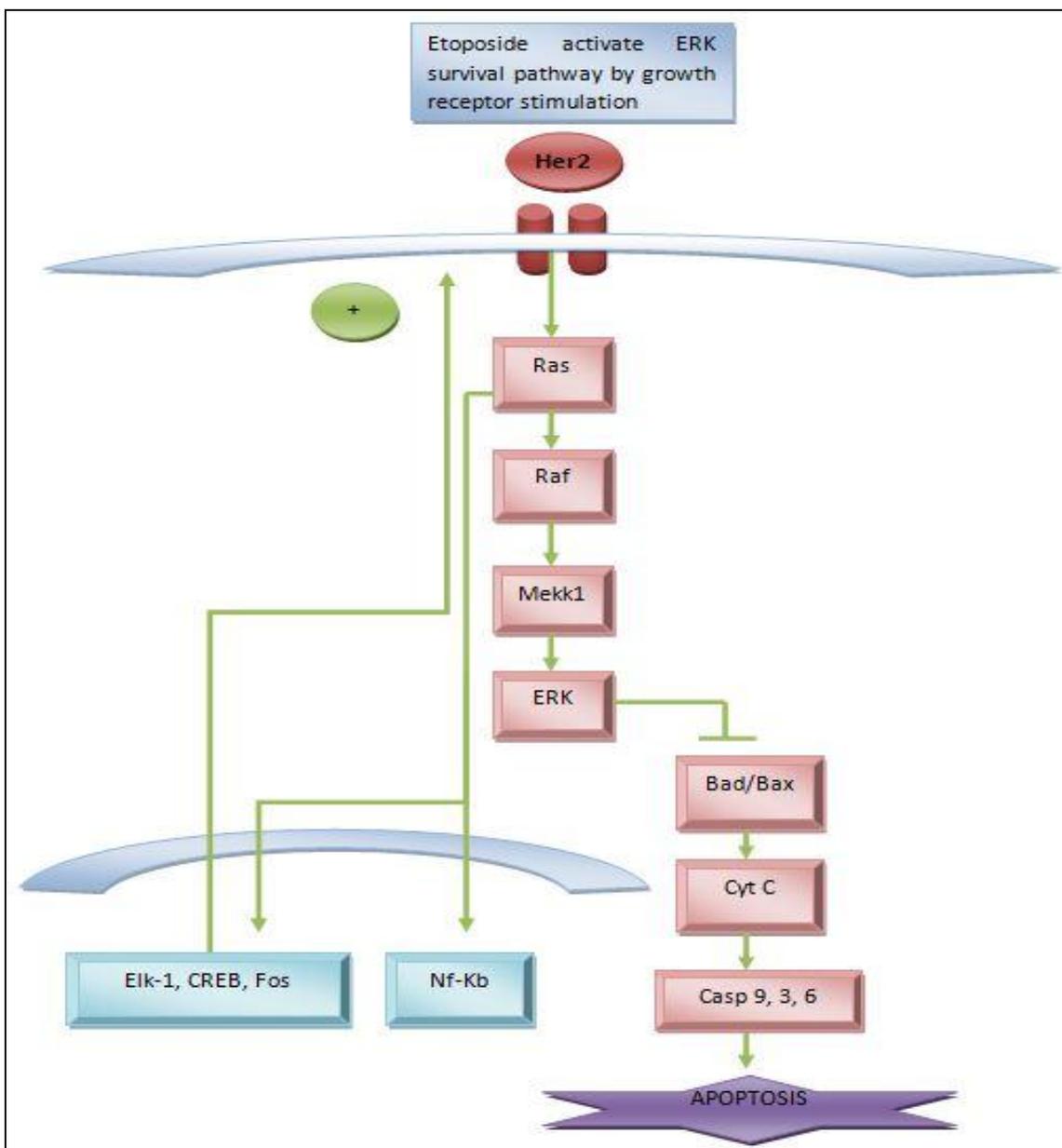


FIG. 10: ERK SURVIVAL PATHWAY

Role of AKT in cell survival:

The main hindrance to booming treatment of cancer is the resistance in chemotherapy. Etoposide can stimulate PI3K (phosphoinositide-3 kinase) and Akt, activities in concentration and time-dependent manner⁷⁰. Akt is a Serine/threonine kinase protein that is engaged as a mediator in many physiological responses such as stimulation of cell proliferation and inhibition of apoptosis. Three mammalian isoforms of Akt are currently known that are Akt1/PKB-Alpha, Akt2/PKB-Beta and Akt3/PKB-Gamma.

Growth factor receptor mediates signal transduction pathway that has been involved in granting resistance to chemotherapy on cancer cells via Akt apoptotic resistant pathway. HER2 (Human Epidermal Growth Factor Receptor 2) emerged to have a higher phosphorylation level of Akt. HER2 cause Akt activation through PI3K which associates with an augmented resistance of cells to several chemotherapeutic agents like etoposide^{71, 72}. PI-3K is recruited to the phosphotyrosine residues and converts PIP2 into PIP3. PI3K can also be activated by Ras. The actions of Akt in the cell are diverse and numerous, but all eventually result in cell-proliferation and cell survival.

Akt hampers apoptosis by inhibiting the formation of the BAD/BclXL complex and phosphorylate the BAD component of this complex. In its antiapoptotic role Akt triggers IKK, which finally activates NF- κ B and cell survival (Fig. 11). Another interaction of Akt can be seen with death protease caspase 9, activity of which is decreased by direct inhibition through Akt. YAP, the target of Akt is another transcriptional factor that combines with 14-3-3 proteins and causes cytoplasmic localization and nuclear export. Akt is a major survival protein, and phosphorylates p21; a member of CDK inhibitors family, involved in cell cycle arrest and curtails cell proliferation. Akt also phosphorylates Ser 166 on MDM which then enters into nucleus to target p53 for degradation. Akt also activates mTOR which promotes cell proliferation by the synthesis of protein^{73, 74}.

Phosphorylation of XIAP (X linked inhibitor of apoptosis) through Akt hinders its autoubiquitination thus condensing apoptosis. The activity of Akt is impeded negatively by PTEN (Phosphatase and Tensin Homolog), SHIP (SH2-Containing Inositol Phosphatase) and CTMP (Carboxyl-Terminal Modulator Protein).

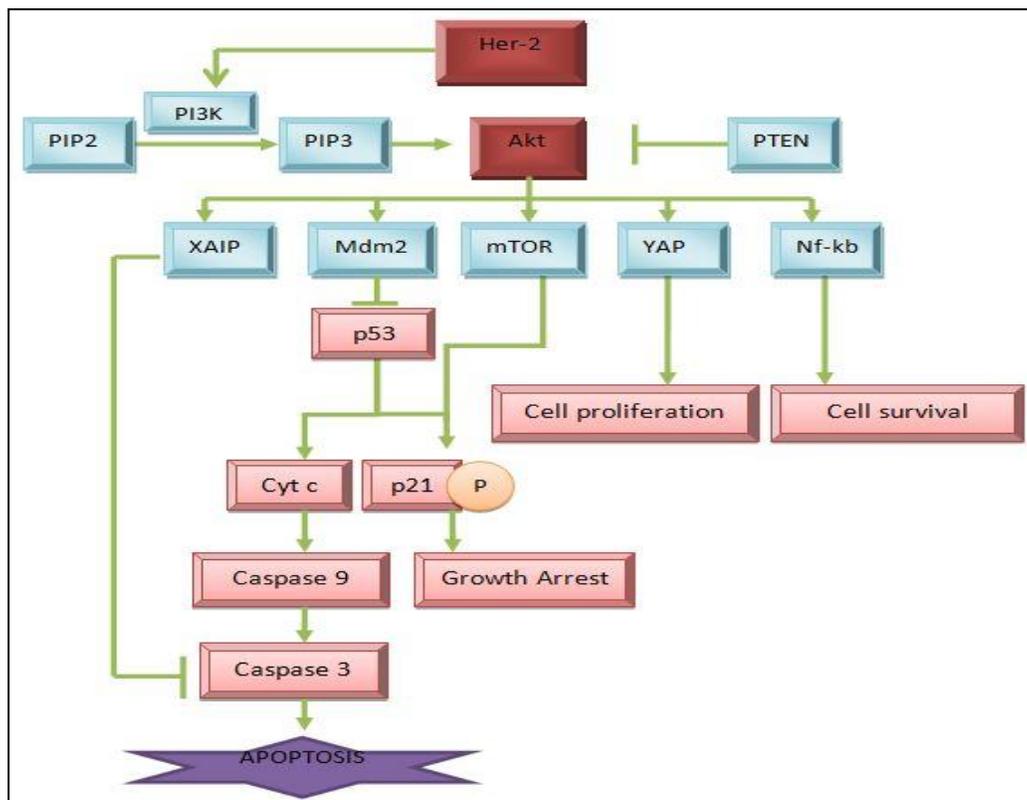


FIG. 11: AKT SURVIVAL PATHWAY

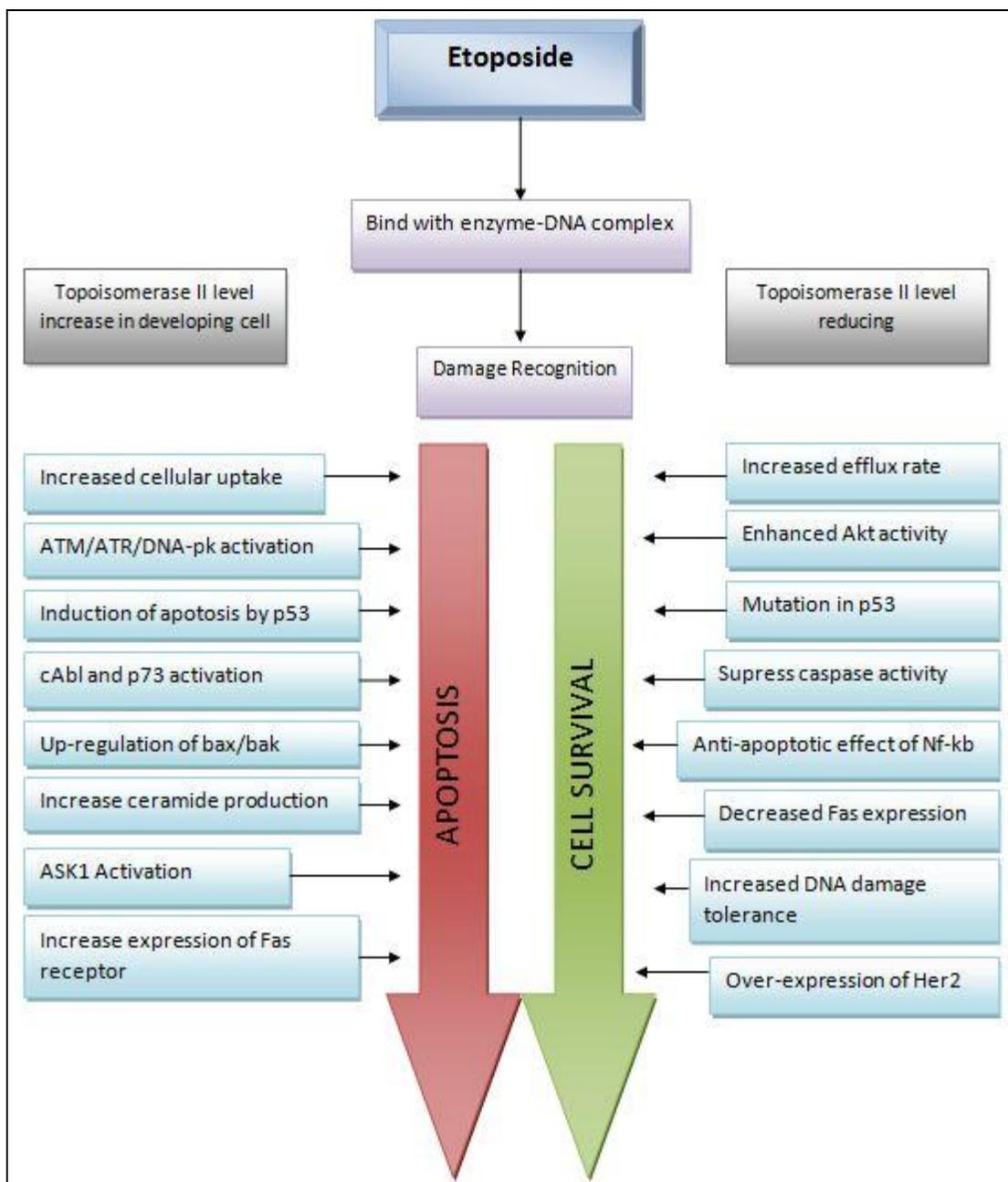


FIG. 12: CROSSTALK BETWEEN APOPTOTIC AND RESISTANT FACTORS

CONCLUSION: This review summarizes the molecular mechanism of etoposide, displaying its cytotoxic activity against a wide variety of tumors. Etoposide is a plant alkaloid derived from podophyllotoxin, and its mechanism of action purposed for its antineoplastic activity based on its interaction with topoisomerase II enzyme, causing DNA damage ultimately leading to cell death. In response to etoposide, DNA damage sensor proteins ATM, ATR, DNA-PKs activate signal transduction pathways. P53 dependent pathway causes growth arrest, apoptosis and induction of (Fas/TRAIL) death receptors. Increased production

of ceramide contributes in activation of ASK1/MEKK1/JNK/p38 which have diverse role in apoptosis.

Etoposide mode of action not only havemeticulous role in apoptosis, but also encumber the skill of drug to bringdown apoptosis. Down-regulation of apoptotic signaling pathway is fundamentallyan entireattribute of resistance. Mechanisms that haltpropagation of DNA damage signal to apoptoticapparatus include activation of PI3-K/Akt and Ras/Raf via Her2 pathway.

Future Perspective:

In order to ameliorate the potency and selectivity of anti-cancer drugs, novel strategies need to be developed. Despite the significant role of etoposide in tumors, acquired resistance mediated by DNA repair enzymes or proteins, may impede apoptosis in preclinical settings. Fortunately, the understanding of major biochemical pathways of etoposide has opened new direction of pharmacological manipulations in chemotherapy. Advanced modulation of key cellular pathways directed to circumvent etoposide resistance may be complementary way to develop alternative strategies. In future, cancer therapy by etoposide must be directed to look for adjuvant drugs that effect biochemical mechanism that can bypass drug resistance, increase its influx rate and transportation in cell, rather to exclusively search for a specific drug with targeting particular cellular constituents.

REFERENCES:

- Silverberg LJ, Dillon JL, Vemishetti P, Sleezer PD, Discordia RP, and Hartung KB: Efficient Synthesis of the Anticancer Drug Etoposide 4'-Phosphate: Use of Benzylic Ether-Protecting Groups on the Carbohydrate Segment. *Org. Proc. Res. Dev* 2000; 4: 34-42.
- Baldwin EL and Osheroff N: Etoposide, topoisomerase ii and cancer. *Current medicinal chemistry Anti-cancer agents* 2005; 5(4):363-372.
- Larsen KA, Escargueil EA, and Skladanowski A: Catalytic topoisomerase II inhibitors in cancer therapy. *PharmacolTher* 2003; 2:167-181.
- Chikamori KG, Grozav A, Kozuki T, Grabowski D, Ganapathi RK and Ganapathi M: DNA Topoisomerase II Enzymes as Molecular Targets for Cancer Chemotherapy. *Current Cancer Drug Targets* 2010; 10:758-771.
- Kung FH, Pratt CB, Vega RA, Jaffe N, Strother D, Schwenn M, Nitschke R, Homans AC, Holbrook CT, Golembe B, Bernstein M, Krischer JP: Ifosfamide/etoposide combination in the treatment of recurrent malignant solid tumors of childhood. A pediatric oncology group phase II study. *Cancer* 2006; 71: 1898-1903.
- Lorenzo GD, Autorino R, Laurentiis MD, Forestieri V, Romano C, Prudente A, Giugliano F, Imbimbo C, Mirone V and Placido SD: Thalidomide in combination with oral daily cyclophosphamide in patients with pretreated hormone refractory prostate cancer: A phase I clinical trial. *Cancer Biology and Therapy* 2007; 6(3):313-7.
- Coleman M, Leonard J, Shuster MW and Kaufman TP: 2001. DICE (dexamethasone, ifosfamide, cisplatin, etoposide) infusional chemotherapy for refractory or relapsed non-Hodgkin's lymphoma (NHL). *Eur J Haematol Suppl.* 2001; 64:41-5.
- Biagi JJ, Herbert KE, Smith C, Abdi E, Leahy M, Falkson C, Wolf M, Januszewicz H, Seymour JF, Richards K, Matthews JP, Dale B, Prince HM: A phase II study of dexamethasone, ifosfamide, cisplatin and etoposide (DICE) as salvage chemotherapy for patients with relapsed and refractory lymphoma. *Leukemia & Lymphoma* 2005; 46(2): 197-20.
- Kim HY and Mishima M: Second-line chemotherapy for small-Cell Lung Cancer (SCLC). *Cancer Treatment Review* 2011; 27(2): 143-150.
- Drings P, Abel U, Bülzebruck H, Stiefel P, Kleckow M, Manke HG: Experience with ifosfamide combinations (etoposide or DDP) in non-small cell lung cancer., *Cancer ChemotherPharmacol.* 1986; 18(2): S34-9.
- Perez EA, Geoffroy FJ, Hillman S, Johnson EA, Farr GH, Tazelaar HD, Hatfield AK, James E, Krook JE, Maillard JA, Levitt R, Marks RS. : Phase II study of oral etoposide and intravenous paclitaxel in extensive-stage small cell lung cancer. *Lung Cancer* 2004; 44(3):347-353.
- Comis RL: Clinical trials of cyclophosphamide, etoposide, and vincristine in the treatment of small-cell lung cancer. *SeminOncol.* 1986; 3:40-4.
- Kunta J, Yan J, Makhey VD and Sinko PJ: Active efflux kinetics of etoposide from rabbit small intestine and colon. *Biopharm Drug Dispos* 2000; 3:83-93.
- Lagas JS, Fan L, Wagenaar E, Vlaming MLH, Tellingena O, Beijnen JH and Schinkel AH: P-glycoprotein (P-gp/Abcb1), Abcc2, and Abcc3 Determine the Pharmacokinetics of Etoposide. *Clin Cancer Res* 2010; 16(1):130-40.
- Davidson AL, Dassa E, Orelle C and Chen J: Structure, Function, and Evolution of Bacterial ATP-Binding Cassette Systems. *American society of microbiology* 2008; 72(2): 317-364.
- Pitts SL, Jablonksy MJ, Duca M, Dauzonne D, Monneret C, Arimondo PB, Anklin C, Grave DE and Osheroff N: Contributions of the D-Ring to the Activity of Etoposide against Human Topoisomerase II α : Potential Interactions with DNA in the Ternary Enzyme-Drug-DNA Complex. *Biochemistry* 2011; 50(22): 5058-5066.
- Bender RP, Jablonksy MJ, Shadid M, Romaine I, Dunlap N, Anklin C, Graves DE and Osheroff N: Substituents on Etoposide that Interact with Human Topoisomerase II α in the Binary Enzyme-Drug Complex: Contributions to Etoposide Binding and Activity. *Biochemistry* 2008; 47(15): 4501-4509.
- Wilstermann AM, Bender RP, Godfrey M, Choi S, Anklin C, Berkowitz DV, Osheroff N and Graves DE: Topoisomerase II-Drug Interaction Domains: Identification of Substituents on Etoposide That Interact with the Enzyme *Biochemistry* 2007; 46(28): 8217-8225.
- Rossi R, Lidonnici MR, Soza S, Biamonti G and Montecucco A: The Dispersal of Replication Proteins after Etoposide Treatment Requires the Cooperation of Nbs1 with the Ataxia Telangiectasia Rad3-Related/Chk1 Pathway. *Cancer Res* 2006; 66(3):1675-83.
- Nam EA and Cortez D: ATR signalling: more than meeting at the fork. *Biochem J* 2011; 436(3):527-36.
- Jazayeri A, Falck J, Lukas C, Bartek J, Smith GC, Lukas J, and Jackson SP: ATM- and cell cycle-dependent regulation of atr in response to DNA double-strand breaks. *Nature cell biology* 2006; 8:37-45.
- Dierov J, Dierova R and Carroll M: BCR/ABL translocates to the nucleus and disrupts an ATR-dependent intra-S phase checkpoint. *Cancer Cell* 2005; 3:275-285.
- Pauklin S, Kristjuhan A, Maimets T and Jaks V: ARF and ATM/ATR cooperate in p53-mediated apoptosis upon oncogenic stress. *BiochemBiophys Res Commun* 2005; 334(2):386-94.
- Bhounik A, Takahashi S, Breitweiser W, Shiloh Y, Jones

- N, and Ronai Z: ATM-dependent phosphorylation of ATF2 is required for the DNA damage response. *Mol Cell* 2005; 18(5):577-87.
25. Grandela C, Pera MF, Grimmond SM, Kolle G and Wolvetang EJ: p53 is required for etoposide-induced apoptosis of human embryonic stem cells. *Stem Cell Research* 2009; 1(2): 116-128.
 26. Valkov NI and Daniel M. Sullivan DM: Tumor p53 status and response to topoisomerase II inhibitors. *Drug Resist Updat*. 2003; 6(1):27-39.
 27. Valkov NI, Sullivan DM: Tumor p53 status and response to topoisomerase II inhibitors. *Drug Resist Updat* 2003; 6(1):27-39.
 28. Bar JK, Grelewski P, Noga L, Rabczyński J, Gryboś M and Jeleń M: The Association Between the p53/topoisomerase I and p53/topoisomerase II α Immunophenotypes and the Progression of Ovarian Carcinomas. *AdvClinExp Med* 2012; 21(1): 35–42.
 29. Deevya L, Narayanan DL, Rao N, Saladi RN and Joshua L. Fox JL: Review: Ultraviolet radiation and skin cancer. *International Journal of Dermatology* 2010; 49(9): 978-986.
 30. Grandela C, Pera MF, Grimmond SM, Kolle G and Wolvetang EJ: p53 is required for etoposide-induced apoptosis of human embryonic stem cells. *Stem Cell Research* 2009; 1(2): 116-128.
 31. Krautwald S, Ziegler E, Rölver L, Linkermann A, Keyser KA, Steen P, Wollert KC, Korf-Klingebiel M and Kunzendorf U: Effective Blockage of Both the Extrinsic and Intrinsic Pathways of Apoptosis in Mice by TAT-crmA. *J BiolChem* 2010; 285(26): 19997–20005.
 32. Cui H, Schroering A, Ding HF: p53 mediates DNA damaging drug-induced apoptosis through a caspase-9-dependent pathway in SH-SY5Y neuroblastoma cells 2002; *Mol Cancer Ther.* 1(9):679-86.
 33. Torkin R, Lavoie JF, Kaplan DR and Yeger H: Induction of caspase-dependent, p53-mediated apoptosis by apigenin in human neuroblastoma. *Mol Cancer Ther* 2005; 4(1):1-11.
 34. Soengas MS, Alarcón RM, Yoshida H, Giaccia AJ, Hakem R, Mak TW and Lowe SW: Apaf-1 and caspase-9 in p53-dependent apoptosis and tumor inhibition. *Science* 1999; 284(5411):156-9.
 35. VincetCryns V, Junyinguan J: Proteases to die for. *Genes & Dev.* 1998; 12:1551-1570.
 36. Haupt S, Berger M, Goldberg Z and YgaHaupt Y: Apoptosis - the p53 network. *J Cell Sci* 2003; 116(20):4077-85.
 37. Livesey KM, Kang R, Vernon P, Buchser W, Loughran P, Watkins SC, Zhang L, Manfredi JJ, Zeh HJ, Li L, Lotze IT and Tang D: p53/HMGB1 Complexes Regulate Autophagy and Apoptosis. *American Association for Cancer Research.* 2012; 72(8):1996-2005.
 38. Chipuk JE and Green DR: Dissecting p53-dependent apoptosis. *Cell Death Differ* 2006; 13(6):994-1002.
 39. Meley D, Spiller DG, White MR, McDowell H, Pizer B and See V: P53-mediated delayed nf-kappab activity enhances etoposide-induced cell death in medulloblastoma. *Cell death & disease* 2010; 1:41.
 40. Martínez-Delgado B, Cuadros M, Honrado E, Ruiz de la Parte A, Roncador G, Alves J, Castrillo JM, Rivas C, Benítez J and The Spanish Cooperative Group for the study of T-cell lymphomas⁵: Differential expression of NF- κ B pathway genes among peripheral T-cell lymphomas. *Leukemia* 2005; 19: 2254–2263.
 41. Scott FL, Stec B, Pop C, Dobaczewska MK, Lee JJ, Monosov E, Robinson H, Salvesen GS, Schwarzenbacher R and Ried SJ: The Fas–FADD death domain complex structure unravels signalling by receptor clustering. *Nature* 2008; 457(7232):1019-22.
 42. Lavrik I, Krueger A, I Schmitz I, Baumann S, Weyd H, Krammer PH and Kirchhoff S: 2003. The active caspase-8 heterotetramer is formed at the CD95 DISC. *Cell Death and Differentiation* 2003; 10, 144–145.
 43. Lavrik IN and Krammer PH: Regulation of CD95/Fas signaling at the DISC. *Cell Death and Differentiation* 2012; 19: 36–41.
 44. Fricker N, Beaudouin J, Richter P, Eils R, Krammer PH and Lavrik: INModel-based dissection of CD95 signaling dynamics reveals both a pro- and antiapoptotic role of c-FLIP. *Journal of cell biology* 2010; 190 (3): 377.
 45. Wen W, Zhu F, Zhang J, Keum Y, Zykova T, Yao K, Peng C, Zheng D, Cho Y, Ma W, Bode AM and Dong Z: MST1 Promotes Apoptosis through Phosphorylation of Histone H2AX. *J BiolChem* 2010; 285(50): 39108–39116.
 46. Ferreira E, Baldeiras I, Ferreira IL, Costa RO, Rego AC, Pereira CF and Oliveira CR: Mitochondrial- and Endoplasmic Reticulum-Associated Oxidative Stress in Alzheimer's Disease: From Pathogenesis to Biomarkers. *International Journal of Cell Biology* 2012; 23.
 47. Chandra D, Choy G, Den X, Bhatia B: Association of Active Caspase 8 with the Mitochondrial Membrane during Apoptosis: Potential Roles in Cleaving BAP31 and Caspase 3 and Mediating Mitochondrion-Endoplasmic Reticulum Cross Talk in Etoposide-Induced Cell Death. *Mol Cell Biol* 2004; 24(15):6592-607.
 48. Hao Z and Mak TW: Type I and Type II Pathways of Fas-mediated Apoptosis Are Differentially Controlled by XIAP. *J Mol Cell Biol* 2009; 2:63-64.
 49. Cullen SP and Martin SJ: Caspase activation pathways: some recent progress. *Cell Death and Differentiation* 2009; 16:935–938.
 50. Chandra D, Choy G, Den X, Bhatia B: Association of Active Caspase 8 with the Mitochondrial Membrane during Apoptosis: Potential Roles in Cleaving BAP31 and Caspase 3 and Mediating Mitochondrion-Endoplasmic Reticulum Cross Talk in Etoposide-Induced Cell Death. *Mol Cell Biol* 2004; 24(15):6592-607.
 51. Wang B, Nguyen M, Chang NC and Shore GC: Fis, Bap31 and the kiss of death between mitochondria and endoplasmic reticulum *EMBO J* 2011; 30(3): 451–452.
 52. Breckenridge DG, Marina Stojanovic M, W Marcellus RC, and Shore GC: Caspase cleavage product of BAP31 induces mitochondrial fission through endoplasmic reticulum calcium signals, enhancing cytochrome c release to the cytosol. *J Cell Biol* 2003; 160(7):1115-27.
 53. Iwasawa R, Mahul-Mellier AL, Datler C, Pazarentzos E and Grimm S: Fis1 and Bap31 bridge the mitochondria–ER interface to establish a platform for apoptosis induction. *The EMBO Journal* 2011; 30(3): 556-568.
 54. Ravi R, Bed A, Fuchs EJ, and Bedi A: CD95 (Fas)-induced Caspase-mediated Proteolysis of NF- κ B, 2012. *Cancer Res* 2012; 58: 882.
 55. Meley D, Spiller DG, White MR, McDowell H, Pizer B and See V: P53-mediated delayed nf-kappab activity enhances etoposide-induced cell death in medulloblastoma. *Cell death & disease* 2010; 1:41.
 56. Chen C, Lin C, Chang W, Huang W, Teng C, and Lin Y: Ceramide induces p38 MAPK and JNK activation through a mechanism involving a thioredoxin-interacting protein-mediated pathway. *Blood* 2007; 111(8):4365-74.
 57. Sawada M, Nakashima S, Banno Y, Yamakawa H, Hayashi K, K Takenaka K, Nishimura Y, NSakai N and Y Nozawa Y: Ordering of ceramide formation, caspase

58. activation, and Bax/Bcl-2 expression during etoposide-induced apoptosis in C6 glioma cells. *Cell Death Differ* 2000; 7(9):761-72.
59. Deak JC, Cross JV, Lewis M, Qian Y, Parrott LA, Distelhorst CW and Templeton DJ: Fas-induced proteolytic activation and intracellular redistribution of the stress-signaling kinase MEKK1., *Proc Natl Acad Sci U S A* 1998; 95(10): 5595-600.
60. Reyland ME, Barzen KA, Anderson SM, Quissell DO and Matassa AA: Activation of PKC is sufficient to induce an apoptotic program in salivary gland acinar cells. *Cell Death Differ*. 2000 Dec; 7(12):1200-9.
61. Clarke P and Tyler KL: 2009. Apoptosis in animal models of virus-induced disease. *Nat Rev Microbiol* 2009; 7(2):144-55.
62. Dan S, Naito M, Seimiya H, Kizaki A, Mashima T and Tsuruo T: Activation of c-Abl tyrosine kinase requires caspase activation and is not involved in JNK/SAPK activation during apoptosis of human monocytic leukemia U937 cells. *Oncogene* 1999; 18(6):1277-83.
63. Nicholson DW, Machuy N, Rajalingam K, T Rudel T and Machuy N: Requirement of caspase-mediated cleavage of c-Abl during stress-induced apoptosis. *Cell Death Differ* 2004; (3):290-300.
64. Lasfer M, Davenne L, Vadrot N, Alexia C, Sadjji-Ouatas Z, Bringuier A, Feldmann G, Pessayre D, Reyl-Desmars F: Protein kinase PKC delta and c-Abl are required for mitochondrial apoptosis induction by genotoxic stress in the absence of p53, p73 and Fas receptor *INSERM, U773. FEBS Letters* 2006; 580(11):2547-2552.
65. Shaul Y: c-Abl: activation and nuclear targets. *Cell Death Differ*. 2000; (3):290-300.
66. Nie Y, Li HH, Bula CM and Liu X: Stimulation of p53 DNA Binding by c-Abl requires the p53 C Terminus and Tetramerization. *Mol Cell Biol* 2000; 20(3):741-8.
67. Lohrum MAE and Vousden KH: Regulation and function of the p53-related proteins: same family, different rules. *Trends in Cell Biology* 2000; 10(5):197-202.
68. Lohrum MAE and Vousden KH: Regulation and function of the p53-related proteins: same family, different rules. *Trends in Cell Biology* 2000; 10(5):197-202.
69. Kim E and Um S: Thymine-DNA glycosylase interacts with and functions as a coactivator of p53 family proteins. *Biochemical and Biophysical Research Communications* 2008; 377(3): 838-842.
70. Ramadan S, Terrinoni A, Catani MV, Sayan AE, Knight RA, Mueller M, Krammer PH, Melino G and Candi E: p73 induces apoptosis by different mechanisms. *Biochemical and Biophysical Research Communications* 2005; 331(3): 713-717.
71. Melino G, Bernassola F, Rana, li M, Yee K, Zong WX, Corazzari M, Knight RA, Green DR, Thompson C and Vousden KH: p73 Induces Apoptosis via PUMA Transactivation and Bax Mitochondrial Translocation., *J Biol Chem* 2004; 279(9):8076-83.
72. Wang JYJ: Regulation of cell death by the Abl tyrosine kinase. *Oncogene* 2000; 19(49):5643-50.
73. Baccarini M: An old kinase on a new path: Raf and apoptosis. *Cell Death and Differentiation* 2002; 9: 783-785.
74. Steelman LS, Chappell WH, Abrams SL, Kempf CR, Long J, Laidler P, Mijatovic S, Maksimovic-Ivanic D, Stivala F, Mazzarino MC, Donia M, Fagone P, Malaponte G, Nicoletti F, Libra M, Milella M, Tafuri A, Bonati A, Bäsecke J, Cocco L, Evangelisti C, Martelli AM, Montalto G, Cervello M and McCubrey JA: Roles of the Raf/MEK/ERK and PI3K/PTEN/Akt/mTOR pathways in controlling growth and sensitivity to therapy-implications for cancer and aging. *Aging (Albany NY)* 2011; 3(3): 192-222.
75. Yu H, Ai Y, Liang-Liang Yu L, Zhou X, Liu J, Li J, Xu X, Liu S, Chen J, Liu F, Qi Y, Deng Q, Cao J, Liu S, Luo H, Yu J: Phosphoinositide 3-kinase/Akt pathway plays an important role in chemoresistance of gastric cancer cells against etoposide and doxorubicin induced cell death. *Int J Cancer* 2008; 122(2):433-43.

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