IJPSR (2016), Vol. 7, Issue 6



INTERNATIONAL JOURNAL



Received on 22 January, 2016; received in revised form, 29 March, 2016; accepted, 03 April, 2016; published 01 June, 2016

BACTERIOLOGICAL QUALITY OF CHLOROQUINE SYRUPS SOLD IN CALABAR MUNICIPALITY, NIGERIA

E. A. Okpo, C. I. Mboto and B. E. Agbo*

Department of Microbiology, Faculty of Biological Sciences University of Calabar, P. M. B. 1115, Calabar, Nigeria.

Key words:

Bacteriological, Syrup, Turbidity, Identification And Colony Forming Units.

Correspondence to Author: B. E. Agbo

Department of Microbiology, Faculty of Biological Sciences University of Calabar, P. M. B. 1115, Calabar, Nigeria.

Email: profbaseadeone@gmail.com

ABSTRACT: Bacteriological qualities of three different brands of chloroquine syrup (EZR, DMR and MBR) commonly sold in Calabar municipality, Nigeria were evaluated using standard microbiological techniques. Each brand was evaluated in duplicates and examined visually for possible abnormalities such as colour, turbidity and un-usual odour. This was followed by determination of viable cell count and identification of bacterial isolates. Chloroquine syrups sample EZR and MBR had mean bacterial count of 1.0x103cfu/ml respectively and DMR had 3.0x103cfu/ml. The isolates were staphylococcus aureus (47%), Escherichia coli (38%) and Pseudomonas aeruginosa (15%). All strain of S. aureus was sensitive to gentamycin (CN), ciprofloxacin (CPX), septin (SXT), Erythromycin (E) and resistance to Taravid (OFX), Amoxacillin (AM) and Ampicillin (AP). Escherichia coli strains isolated showed sensitivity to ciprofloxacin (CPX), Taravid (OFX) and septin (SXT) and was resistance to gentamycin (CN), Amoxacillin (AM) and Ampicillin (AP). P. aeroginosa strains were sensitive to Ciprofloxacin (CPX) and Taravid (OFX), and was resistant to Gentamycin (CN). The colony forming unit (cfu) for the sample DMR found in the study is below World Health Organization (WHO) standard. Thus, this finding is a major health concern.

INTRODUCTION: Chloroquine remains one of the drugs of choice for the treatment of malaria infection which frequently lead to death in children in many developing countries of the world including Nigeria ⁵⁻²⁰. Different brands of chloroquine syrups for children are available in the market. These brands are often produced with variations in manufacturing processed and possibly with different excipients which result in different bio availabilities ⁶.

QUICK RESPONSE CODE			
	DOI: 10.13040/IJPSR.0975-8232.7(6).2586-90		
	Article can be accessed online on: www.ijpsr.com		
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.7 (6).2586-90			

Syrups are aqueous preparations characterized by sweet taste, a viscous consistency and serve as vehicles to convey medications. This ensure palatability and easy administration ⁸.

Bacterial contamination of pharmaceutical products is rapidly becoming a matter of public health concern globally. Pharmaceutical products are often susceptible to contamination by a variety of microorganisms during manufacturing and Due to consumption the ubiquity of microorganisms specific control measures must be adopted to avoid microbial contamination. Air is a major source of contamination. During the use of products, contamination with pharmaceutical microorganisms irrespective of their harmful status can bring about physical and chemical changes of the product ². Barid ³ observed that contaminating

microbes may bring about the conversion of syrup formulation into the toxic metabolites. Similarly, Altah *et al.*,¹ recognized such products as microbiologically unsafe for consumption and may pose potential health hazards to patients, as well as constitute wastage and may yield serious economic losses to the manufacturers. Khanfar *et al.*,⁴ preservatives observed that are used for maintaining the quality of syrup formulations, however some of these may be a source of contamination as they may contain microbes.

Some studies have revealed an increase in the number of infection cause by contaminated nonsterile preparations. These infection are as a result of contamination by bacteria during the preparation of the syrups. Contamination can evolve from the atmosphere where the syrups is being prepared or from the water used in the preparation. More so, it can evolve from the active ingredient being contaminated before it is used for the preparation¹⁰.

In Nigeria, most non-sterile pharmaceutical products have been reported to be contaminated by microorganisms mostly bacteria ⁹⁻¹¹. This may be influenced by the environment and quality of the raw materials used during formulation. Some disease outbreaks have been associated with the use of heavily contaminated raw materials of natural origin ¹². Some studies have revealed that incidence of microflora in syrups is indicated by the nature of ingredients (whether natural or synthetic), the quality of the vehicle and the care and attitude of the personnel involved in the handling ^{13, 21}.

The most serious problems of bacterial contamination of syrups is where there is no obvious signs of spoilage hence, it is usually advisable to have knowledge of the bacterial content of all drugs and medicines whether there are required to be sterile or non-sterile ^{13, 21}. Ibrahim *et al.*,⁸ have revealed that chloroquine is the most commonly administered syrups within most Nigerian households. Neglecting it wide spread used, few studies, if any have evaluated their bacteriological quality. This study is therefore aimed at evaluating the bacteriological quality of the most commonly sold syrups in Calabar municipality.

MATERIALS AND METHODS:

Collection of Samples:

Three different brands of chloroquine syrups (EMR, DMR and MBR) were purchased in duplicate from different drugs outlets in Calabar Municipality, Nigeria.

Determination of pH:

The pH of different brands of chloroquine syrups was determined using Mettler Toledo's pH meter, produced by Wincom Company Limited, China. An aliquot of the syrups was dispense into test tube and the sensitive part of the pH meter was dipped inside the syrup for 10 seconds, followed by reading of the scale on the meter. The summary of the reading were noted.

Enumeration of Microorganisms:

Bacteria count was carried out using Nutrient agar, Mac Conkey agar and Tryticase soy agar following the method as described by clinical and laboratory standard institute (CLSI).

The bacterial isolates were characterized according to the method described by Bergey's manual of determinative bacteriology ¹⁵ in which the following reactions were examined: Gram's staining reaction, catalase test, motility test, carbohydrate utilization test, coagulase test, methyl red test, indole test, Vogesproskauer test, citrate test, urease test and oxidase test.

Antimicrobial Sensitivity Testing (Disk Diffusion Method):

Antimicrobial susceptibility test was carried out using Mueller-Hinton Agar following the method described by CLSI. Antibiotics evaluated include Gentamycin (CN), Ciprofloxacin (CPX), Taravid (OFX), Septrin (SXT), Erythromycin (E), Anoxacillin (AM) and Ampicillin (PN).

RESULT:

The macroscopic characteristic of examined samples of chloroquine syrups is represented in **Table 1**. The table shows that there were no abnormalities, nor unpleasant odour in the syrup, though sample DMR was turbid but there was no suggestive of spoilage. **Table 2** shows the mean viable bacterial count of analyzed syrup. In this table, the mean count for sample EMR and MBR was 1.0×10^3 cfu/ml respectively and that of DMR was 3.0×10^3 cfu/ml. The means count of sample EMR and MBR has conform to the standard set by NAFDAC as their colony forming unit did not exceed 1.0×10^3 cfu/ml. DMR shows a high level of contamination as it has colony forming unit up to 3.0×10^3 cfu/ml which is above the standard microbiological specification for the certification of syrups. **Fig. 1** shows that percentage occurrence of bacterial isolates in the syrups studied. In this figure, *staphylococcus aureus* has the highest frequency of occurrence (47%), followed by *Escherichia coli* (38%) and *Pseudomonas aeruguiosa*(15%). The result of the susceptibility pattern of the isolates is represented in **Table 3**.

TABLE 1	: MACROSCOPIC	CHARACTERISTIC	OF EXAMINE	SAMPLES OF	CHLOROOUINE

Syrups Code	Colour	Turbidity	Un-usual odour
EMR	Yellow	Clear	No Un-usual odour
DMR	Yellow	Turbid	No Un-usual odour
MBR	Yellow	Clear	No Un-usual odour

 TABLE 2: TOTAL VIABLE BACTERIAL COUNT OF SAMPLED SYRUPS ACCORDING TO BRAND

Syrups Code	Colonial Count (cfu/ml)				
	MacConkey agar	Nutrient agar (cfu/ml) Trypticas soy		Mean count	
	(cfu/ml)		agar (cfu/ml)		
EMR	$1.0 \mathrm{x} 10^3$	1.0×10^{3}	NG	$1.0 \ge 10^3$	
DMR	2.0×10^3	3.0×10^3	3.0×10^3	3.0×10^3	
MBR	1.0×10^3	2.0×10^3	1.0×10^3	$1.0 \ge 10^3$	

Key:

NG: No growth



FIG.1: PERCENTAGE OCCURRENCE OF BACTERIA ISOLATES IN WXAMINED CHOLOQUINE SYRUP SAMPLE

TABLE 3: ANTIBIOTICS SUSCEPTIBILITY PROFILE OF BACTERIA ISOLATED FROM SAMPLED SYRUPS

Antiboitic	Disc potency (µg)	S. aureus	Esch. coli	P. aeruginosa
Gentamycin (CN)	10	+	-	-
Ciprofloxacin (CPX)	10	+	+	+
Taravid (OFX)	10	-	+	+
Septrin (SXT)	30	+	+	NA
Erythromycin (E)	10	+	NA	NA
Amoxacillin (AM)	30	-	-	NA
Ampicillin (PN)	30	-	-	NA

Key: + Sensitive, - Resistant, NA - Not Applicable

DISCUSSION: Studies have revealed several inadequacies surrounding the production of many pharmaceutical products including chloroquine

syrup ⁹. In this study, the finding of a high bacterial load in all the three samples evaluated may be suggestive of the use of contaminated raw materials

or the introduction of contaminant during production or poor storage.

In a study conducted by Daniyam et al., ¹² on the microbiological examination of non-sterile products, the presence of S. aureus although ubiquitous in the environment is undesirable because of their spoilage potentials, and their presence in a product suggest poor environmental hygiene during processing or the use of heavily contaminated raw materials. On the other hand, Escherichia coli are found in the respiratory, intestinal and urinogenital tract of human. Since E. *coli* can be transmitted faecal-orally, the personnel may be a major contributory factor to this type of contaminant. Consequently, compliance with aseptic technique and personnel hygiene during preparation of pharmaceutical product may minimize microbial cross contamination. This obviously will prevent spoilage of the product and possible detrimental effect for patients.

The detection of *S. aureus* in the sampled syrup is of major health significant. This is because *S. aureus* secrete toxin which contribute to gastrointestinal distress ^{14, 17}. More so, the high number of *S. aureus* in these preparations suggest that they are able to tolerate the presence of preservatives in such products. In an unrelated study carried out by Takon and Antai ⁹, *S. Aureus* was a prominent isolate of the spoiled pharmaceutical product evaluated. The presence of *Escherichia coli* is a good indicator of faecal contamination resulting from water supply used in preparation of the syrups.

Furthermore, the isolation of *Escherichia coli*, *S. aureus*, and *P. aeruginosa* in these products indicate a possible health risk. The possible adverse effect on health and the spoilage potentials of these contaminants highlights the need to reduce the degree of contamination of such products by establishing official guideline such as Good Manufacturing Practice (GMP) and ensuring compliance through regular monitoring of nonsterile pharmaceutical products. The result of this study is at variant with NAFDAC specification. However, some of them (EMR and MBR) with a low mean cell count of 1.0×10^3 cfu/ml respectively conformed to the NAFDAC standard.

Generally *P.aeruginosa* is inherently resistance to most common antibiotic accept carbinicilin, colistinesulphate and Gentimycine. However in this study carbinciline and colistinesulphate were not evaluated.

The susceptibility pattern of each bacterial isolate to antimicrobials agents showed that *S. aureus* was sensitive to Gentamycin (CN), Ciprofloxacin (CPX), Septrin (SXT), Erythromycin (E), and resistance to Taravid (OFX), and Amoxacillin (AM). *E. coli* was sensitive to ciprofloxacin (CPX), Taravid (OFX), Septine (SXT) and resistance to Gentamycin (CN), Amoxacillin (AM) and Ampicillin (AP). *P. aeruginosa* was sensitive to Ciprofloxacin (CPX) and Tavavid (OFX), and resistance to Gentamycin (CN).

The result reveals that chloroquine syrup sample DMR falls below NAFDAC Standard and makes need for public health intervention.

Poor storage on shelf's that may exposed the drugs to direct sun light and high temperature may be a major contributory factor to amplify the bacterial load from a few colonies at the inception to contamination.

In conclusion, adherence to standard manufacturing practice is one of the method that can be used to prevent contamination of chloroquine syrups.

Most of the pharmaceutical company rely on their bore hold water which is never evaluated by third party and this could be a major source of contamination. Some studies reveals that waters use in production of non-sterile pharmaceutical product is a major source of contamination ^{3, 8, 10, 21}.

REFERENCES:

- 1. Altah A., Udokpon, A. and Ofum, M: Bacteriological quality of some pharmaceutical marketed by drug vendors. Journal of health science 2004; 11: 120-123.
- 2. Hossain M., Ara S. and Raphman MZ: Quantitative examination of aerobic bacteria and fungi in locally available antacid suspension and possible contamination by specific bacteria. Journal of biological Sciences 2004; 7(11): 2014-2017.
- Baird R: Microbial spoilage, infection risk and contamination control. In: Stephen P. Denyer, Norman A. Hodges, Sean P. Gorman (eds). Pharmaceutical microbiology. 7th Ed. Blackwell publishing company, Massachusett U.S.A. 2004; pp. 263-284

- 4. Khanfar M, Khalil R. and Abujafal EA: Evaluation of efficacy for different cough syrups manufactured by different pharmaceutical companies. International Journal of Pharmacology 2009; 5(5): 319-322.
- 5. Chen P, Patrick, GZ, and Chen, J: Chloroquine treatment of ARPE-19 cells lead to lysosome dilation and intracellular lipid accumulation; possible implications of lysosomal dysfunction in macular degeneration. *Cell and* Bioscience 2011; 1(10): 10-12.
- 6. Ibrahim MA and Idoka RO: The sorption of active ingredient from chloroquine phosphate syrup by a sample plastic container used by manufacturers in Nigeria. Nigerian Journal of Polymer Science and Technology 2000;1(1): 1-7.
- 7. Kim E, Wustenberg R. and Rusban A: Chloroquine activates the p53 pathway and reduced apoptosis in human glioma cells. Neuro-oncology 2010; 12(4): 389-400.
- 8. Ibrahim EC, Esimone CO, Ofoefule SI and Chah KF: Evaluation of the microbiological quality of some commercially available syrups and suspensions in Nigeria. Journal of phytomed therapy 2004; 7(1&2): 18-25.
- Takon IA and Antai SP: Microbial contamination of expired and unexpired aqueous and suspension drugs sold in some patent medicine store in Calabar. Nigerian Journal of Microbiology 2006;20(2): 109-110.
- 10. Mwanbete KD, Justin-Temu M and Fazleabbas SF: Microbiological assessment of commercially available quine syrups and water for injection in Daveses Salaam Tanzania. Trop J. Pharm. Res. 2009; 8 (5): 441-447.
- Emejuru MC, Ojiegbe GC, Azi S. and Nwosu NB: Microbiological load of selected oral liquid pharmaceuticals. Int. J. Community Res. 2013; 2(3): 39-45.

- Daniyan SY and Sangodere TA: Microbial assessment of some syrup sold in patent medicine stores in Minna Metropolis, Nigeria. Int. Res. J. Pharm. 2011; 2 (8):58-61.
- Parker MS: Oral dosage formulation. Int. J. Pharm. Tech. Prod. 2000; 6 (14): 18-25.
- Cheesebrough, M: District laboratory practice in tropical countries. New York: Cambridge University press. 2002; pp. 193-194.
- Bergey DH and John G: Bergey's manual of determinative bacteriology. Philadelphia; Lippincott Williams and Wilkins, 2000.
- Michaelides M, Stover NB, Francis PJ and Weleber RG: Retinal toxicity associated with hydroxyl-chloroquine and chloroquine: risk factors, screening and progression despite cessation of therapy. Arch. Ophthalmol, 2011; 129(1): 30-31.
- 17. Shaikh D, Lamshed TA. And Shaikh R: 1999. Microbial contamination of pharmaceutical preparations. Pakistan Journal of Pharmaceutical Science 2011; 1(1): 61-66.
- Nester MT, Baird R, and Anderson DG: Microbial examination of non-sterile product: Test for specified microorganisms. Pharm. Forum 2003; 29 (5): 1722-1733
- 19. Molina DK: Postmortem hydeoxly-chloroquine concentration in non-toxic cases. American journal of forensic medical pathology 2011; 6(3): 17-19.
- 20. Martin RE, Marchetti RV and Cowan BA: Chloroquine transport through the malaria parasites Chloroquine resistance transporter. Science 2009; 325 (5948):1680-1682.
- 21. Agbo BE and Mboto CI: Phytochemical and antibacterial evaluation of selected locally producedherbal medicines sold in Calabar, Nigeria. Archives of Applied Science Research 2012; 4(5):1974-1990

How to cite this article:

Okpo EA, Mboto CI and Agbo BE: Bacteriological Quality of Chloroquine Syrups Sold in Calabar Municipality, Nigeria. Int J Pharm Sci Res 2016; 7(6): 2586-90.doi: 10.13040/IJPSR.0975-8232.7(6).2586-90.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to ANDROID OS based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)