



Received on 05 November, 2016; received in revised form, 03 January, 2017; accepted, 02 February, 2017; published 01 June, 2017

AMELIORATED ACTIVITY OF *MURRAYA KOENIGII* LEAVES CHLOROFORM EXTRACT (MKCE) AGAINST LEAD INDUCED HEPATIC DYSFUNCTIONS IN MICE

Rohan S. Phatak^{*1} and Somnath M. Matule²

Directorate of Research¹, Department of Pharmacology², Krishna Institute of Medical Sciences Deemed University, Karad - 415110 Maharashtra, India.

Keywords:

Hepatic Function Parameters,
Lead, Liver, *Murraya koenigii*

Correspondence to Author: Rohan Sharadanand Phatak

Junior Research Officer,
Krishna Institute of Medical
Sciences Deemed University,
NH-4, Near Dhebewadi Road,
Malkapur - 415539, Karad, India.


Email: phatak.rohan1983@gmail.com

ABSTRACT: Lead is a well-known hepatotoxic and all-pervading heavy metal. Ingestion of lead induces structural dysfunctions, physiological distortion and biochemical impairment in the liver. The present study was aimed to ascertain the ameliorated effect of chloroform extract of *Murraya* leaves against the deleterious effects of lead on hepatic function parameters. *Murraya koenigii* Chloroform Extract (MKCE) was prepared by maceration. Male albino Swiss mice were divided into three groups of six each. Group I (Control group) served as normal diet and water *ad libitum* as control group, Group II (lead treated group) received intraperitoneal injection of lead acetate (15mg/kg) daily once whereas Group III (MKCE with lead-treated group) received MKCE (50 mg/kg) and injection of lead acetate intraperitoneally (15mg/kg) daily once time. Experimental study continued for consecutive 7 days. On the 8th day, liver were desiccated from all animals; homogenized to collect supernatant into vials. Serum ALT, AST, ALP, TB, TP and ALB levels were estimated. Significant rise in levels of ALT, AST, ALP, TB were observed along with decreased level of TP and ALB in lead-intoxicated mice. Hepatic enzyme levels were significantly reduced ($p < 0.001$) and augmented levels of protein and albumin in group III. Results in the study revealed significant hepatoprotective activity of MKCE against lead intoxication in mice by restoring all hepatic function markers to near normal level.

INTRODUCTION: Liver is a major organ in the body and plays a vital role in the physiological functions, secretion of bile, metabolism and detoxification of xenobiotics and/or drugs¹. Xenobiotics like carbon tetrachloride², fat soluble chemical as industrial pollutant, drugs like acetaminophen, isoniazid, rifampicin³, and ingestion of heavy metals in drinking water like lead, arsenic, mercury, copper and cadmium⁴ are serious environmental pollutants and also examples of hepatotoxic agents.

In the metabolic pathway, transformation reactions (oxidation, reduction or hydrolysis) followed by conjugation converts xenobiotics into suitable forms for elimination from the body. However it requires a multiple transformation reactions for eliminating xenobiotics, hepatotoxic drugs, heavy metals but their intermediates induce hepatotoxicity also. Therefore liver is most influenced by xenobiotics and becomes susceptible to xenobiotic-induced injury/toxicity³.

A large number of medicinal plants and naturally occurring products like silymarin, resveratrol, curcumin and ginkgo with high efficacies and either low or no toxicity³ have been reported as potent therapeutics for exerting hepatoprotection in liver damage. One of these medicinal plants, *Murraya koenigii* L. (Rutaceae) is also reported as antioxidant⁵ and hepatoprotective agent⁶.

<p>QUICK RESPONSE CODE</p> 	<p>DOI: 10.13040/IJPSR.0975-8232.8(6).2619-23</p>
<p>Article can be accessed online on: www.ijpsr.com</p>	
<p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.8(6).2619-23</p>	

It is a small shrub, about 2.5 meters in height widely used as a spice having its characteristic flavour and aroma⁷. Besides it has been reported for various pharmacological activities like antimicrobial⁸, antidiabetic⁹, anti-inflammatory¹⁰ and antinociceptive¹¹, anticancer¹² and caring effect in haematological indices¹³. The present study was aimed to ascertain the ameliorated effect of chloroform extract of *Murraya koenigii* leaves against the deleterious effects of lead on hepatic function parameters.

MATERIAL AND METHODS:

Collection of *Murraya koenigii* leaves: Fresh *Murraya koenigii* were purchased from the local market in the City of Karad (Western Maharashtra) in the month of March 2015 and was validated/authenticated them from the Department of Botany, Yashwantrao Chavan College of Sciences, Karad.

Preparation of chloroform extract of *Murraya koenigii* leaves: Chloroform extract of *Murraya koenigii* leaves was macerated by adding about 70 g of shade dried leaves of *Murraya koenigii* in 2.5 l of chloroform. The collected extract was obtained 6% of yield from the crude leaves and stored in suitable container as *Murraya koenigii* Chloroform Extract (MKCE). According to earlier report, the LD₅₀ value for the aqueous extract of leaves of *Murraya koenigii* was found to be 150mg/kg i.p. in rats^{5,6}. Doses of lead acetate (15 mg/kg i. p) and MKCE (50 mg/kg p. o) were used in our previous study¹³.

Experimental Animals: Male Swiss albino mice (n=6 in each group) weighing between 25-30 g were used in the study. Animals were obtained from the Animal House, Krishna Institute of Medical Sciences, Karad, India. The animals were maintained under standard husbandry conditions at room temperature, light: dark cycle for an acclimatization period of 15 days. Experiment was compiled with the guidelines of Committee for the Purpose of Control And Supervision of Experiments on Animals (CPCSEA) for animal experimentation of laboratory and Institutional Animal Ethics Committee (Reg. No. 255/PO/ 2000/bc/CPCSEA) KIMS, Karad approved for the study.

Experimental Design: Male Swiss albino mice were randomly divided into three groups with each consisting of six animals and was continued at once daily for the consecutive seven days.

Group-I (Normal): Mice received normal diet and water only *ad libitum*

Group-II (Pb treated): Mice received Lead acetate (15 mg/kg i.p)

Group-III (Pb + MKCE treated): Mice received MKCE (50 mg/kg p.o) + lead acetate (15 mg/kg i.p))

On 8th day, mice were anaesthetized under ether and sacrificed by cervical decapitation.

Liver tissues were collected with vials containing saline and stored at 4°C for further analyzing hepatic function parameters.

Blood lead analysis: Blood lead levels in mice were analyzed by atomic absorption spectrometry in accordance to our previous study¹³.

Tissue homogenate: Ten percent liver homogenate in 0.15 M KCl was homogenized in glass mortar pestle and centrifuged in cold (0-4 °C) at 12000 rpm for 30 min. The obtained supernatant was added into eppendorf tubes, labeled and stored at -20 °C and assayed all parameters.

Assessment of liver function:

Aspartate Pyruvate (AST) and Alanine Pyruvate (ALT): It was tested by a combined method of Mohun and Cook (1957) and Reitman and Frankel (1957)^{14,15}.

Alkaline Phosphatase (ALP):

It was analyzed by method of Kind and King (1954)¹⁶.

Total Bilirubin (TB): It was determined by method of Mallay and Evelyn (1937)¹⁷.

Total Protein (TP): It was estimated by method of Lowry *et al.* (1951)¹⁸.

Albumin (ALB): It was measured in the method by Kingsley and Frankel (1939)¹⁹.

Statistical analysis: Data were expressed as the mean \pm S.E.M (n=6). Statistical analysis was done using analysis of one way Analysis of Variance (ANOVA) followed by Dunnett's test and all readings were considered significant at $p < 0.05$.

RESULTS: The current study was estimated the hepatoprotection ability of MKCE either to reduce

the deleterious effects of lead or to preserve the hepatic physiologic mechanisms distorted by lead. All these examinations indicate a hepatoprotective potential of the extract. Elevation of levels in hepatic markers is indicated liver damage as shown in **Table 1**.

TABLE 1: EFFECTS OF MKCE ON HEPATIC PARAMETERS IN LEAD-INTOXICATED MICE

Groups	I	II	III
Parameters	Normal	Pb treated	MKCE + Pb treated
AST (U/I)	90.81 \pm 0.72	170.11 \pm 4.86 ^{##}	105.31 \pm 1.91 ^{**}
ALT (U/I)	48.78 \pm 1.05	81.45 \pm 3.18 ^{##}	62.45 \pm 2.00 ^{**}
ALP (U/I)	101.88 \pm 0.84	300.22 \pm 1.82 ^{##}	158.58 \pm 9.62 ^{**}
TB (mg %)	0.049 \pm 0.002	0.43 \pm 0.06 ^{##}	0.043 \pm 0.01 ^{NS}
TP (mg %)	6.44 \pm 0.21	0.63 \pm 0.16 ^{##}	5.67 \pm 0.26 [*]
ALB (mg %)	2.12 \pm 0.02	0.45 \pm 0.06 ^{##}	1.32 \pm 0.15 ^{**}

Data represents the mean \pm SEM of six mice. AST: aspartate pyruvate, ALT: alanine pyruvate, ALP: alkaline phosphatase, BIL: total bilirubin, TP: total protein; ALB: albumin, * $P < 0.05$ compared to control, ** $P < 0.001$ compared to control, ## $P < 0.001$ compared to control, ^{NS}: non-significant

A significant rise of ALT, AST, ALP levels is shown in lead-intoxicated group II. Treatment with chloroform extract of *Murraya koenigii* L. has shown attenuated levels of hepatic enzyme ($p < 0.001$) in group III. Total protein and albumin levels were significantly decreased in group II ($p < 0.001$) while increased levels of TP ($p < 0.05$) and ALB ($p < 0.001$) in MKCE treated group when compared to control group. It is indicated the hepatoprotective activity of MKCE against lead induced hepatic damage.

DISCUSSION: Lead is a ubiquitous heavy metal and well-known hepatotoxic agent. Ingestion of lead induces structural dysfunctions, physiological distortion and biochemical impairment in the liver²⁰. In the assessment of liver damage by lead, the determination of hepatic enzyme like AST, ALT, and ALP levels is mostly used. Generally necrosis or membrane damage of tissue caused by diseases like viral hepatitis, cardiac infarction, injury to muscle releases a high level of hepatic enzyme into circulation²¹. Elevated level of serum hepatic enzymes is an indicative of liver injury due to cellular leakage and loss of functional integrity of cell membrane of liver²². Hepatic enzyme AST plays vital role in catalyzing conversion of alanine into pyruvate and glutamate. Therefore ALT parameter is more specific to the liver and for detecting liver injury²¹. Our study is also in agreement with other studies regarding to elevated

levels of AST, ALT, and ALP in lead treated group and reversed to some degree by use of some plant extracts such as *Ocimum sanctum*²³, oil of *Sesamum indicum*²⁴, *Moringa oleifera*²⁵, *Pongamia Pinnata*²⁶, *Coriandrum sativum*²⁷, and natural antioxidants like Curcumin²⁸, Hesperetin²⁹.

Decrease in protein content in mice treated with lead might be related to decrease hepatic DNA and RNA in hepatocytes²⁶ so it may result decreased capacity to synthesize protein. The beneficial effects of MKCE may offer protection by stabilizing the cell membrane in hepatic damage induced by lead which is in conformity with study by Zongping *et al.* (2013)²⁹.

Although the exact mechanism of lead toxicity is not completely cleared but cumulative data showed that oxidative stress plays an essential role in its toxicity²⁷. Thus in further studies, many antioxidant enzyme parameters like superoxide dismutase, catalase, reduced glutathione, glutathione peroxidase, glutathione-s-transferase, total antioxidant capacity, thiobarbituric acid reactive substances should be investigated to elaborate free radical scavenging effects of MKCE in mice due to its presence of flavonoids and phenolics.

CONCLUSION: Results of the study indicated that MKCE exerted significant hepatoprotective activity against lead intoxication in mice by restoring all possible hepatic function markers to near normal level. Further phytochemical investigations are needed to characterize the active hepatoprotective principle of MKCE.

CONFLICT OF INTEREST: None declared.

ETHICAL APPROVAL: Institutional Animal Ethics Committee.

ACKNOWLEDGEMENTS: Authors are thankful to Dr. C. C. Khanwelkar, Professor and Head, Department of Pharmacology and Dr. A. V. Sontakke, Professor and Head and staff of Department of Biochemistry, Krishna Institute of Medical Sciences for their constant support and providing facilities in the project.

REFERENCES:

1. Shaikh AS, Haq SA, Baig MA, Ansari JA. Antioxidant and hepatoprotective activity of *Murraya koenigii* L. against CCl₄-induced hepatotoxicity in rats. IJAPMBS 2015; 3(1): 1-6.
2. Boll M, Weber LW, Becker E, Stampfl A. Mechanism of carbon tetrachloride-induced hepatotoxicity. Hepatocellular damage by reactive carbon tetrachloride metabolites. Z Naturforsch C 2001; 56(7-8):649-59.
3. Singh D, Cho WC, Upadhyay G. Drug-Induced Liver Toxicity and Prevention by Herbal Antioxidants: An Overview. Front Physiol 2016; 6: 363.
4. Al-Attar AM. Vitamin E attenuates liver injury induced by exposure to lead, mercury, cadmium and copper in albino mice. Saudi J Biol Sci 2011; 18(4):395-401.
5. Ghosh D, Firdaus SB, Mitra E, Dey M, et al. Protective effect of aqueous leaf extract of *Murraya koenigii* against lead induced oxidative stress in rat liver, heart and kidney: a dose response study. Asian J Pharm Clin Res 2012; 5(Suppl 4):54-58.
6. Ghosh D, Firdaus SB, Mitra E, Dey M, et al. Hepatoprotective activity of aqueous leaf extract of *Murraya koenigii* against lead-induced hepatotoxicity in male wistar rat. Int J Pharm Pharm Sci 2013; 5(1):285-295.
7. Muthulinggam Nishan, Partiban Subramanian. *Murraya koenigii* (curry leave) - A review on its potential. Int J PharmTech Res 2014-2015; 7(4): 566-572.
8. Malwal M, Sarin R. Antimicrobial efficacy of *Murraya koenigii* (Linn.) Spreng. root extracts, Indian J Nat Prod Res 2011; 2(1): 48-51.
9. Yadav S, Vats V, Dhunnou Y, Grover JK. Hypoglycemic and antihyperglycemic activity of *Murraya koenigii* leaves in diabetic rats. J Ethnopharmacol 2002; 82(2-3):111-6.
10. Darvekar VM, Patil VR, Choudhari AB. Anti-inflammatory activity of *Murraya koenigii* Spreng on experimental animals. J Nat Prod Plant Resour 2011; 1 (1): 65-69.
11. Patil RA, Langade PM, Dighade PB, Hiray YA. Antinociceptive activity of acute and chronic administration of *Murraya koenigii* L. leaves in experimental animal models. Indian J Pharmacol 2012; 44(1):15-9.
12. Noolu B, Ajumeera R, Chauhan A, Nagalla B, Manchala R, Ismail A. *Murraya koenigii* leaf extract inhibits proteasome activity and induces cell death in breast cancer cells. BMC Complement Altern Med 2013; 13:7.
13. Phatak RS, Matule SM. Beneficial effects of *Murraya koenigii* leaves chloroform extract (MKCE) on erythrocyte, thrombocyte and leukocyte indices in lead-intoxicated mice. Biomed Pharmacol J 2016; 9(3): 1035-40.
14. Reitman S, Frankel A. Colorimetric method of determination of serum glutamate oxaloacetate and glutamic pyruvate transaminase. Am J Clin Pathol 1957; 28: 56-58.
15. Mohun AF, Cook LJ. Simple method for measuring serum level of glutamate-oxaloacetate and glutamate-pyruvate transaminases in laboratories. J Clin Pathol 1957; 10(4): 394-399.
16. Kind PR, King EJ. Estimation of plasma phosphatases by determination of hydrolyzed phenol with antipyrin. J Clin Pathol 1957; 7: 322-6.
17. Mallay HT, Evelyn KA. Estimation of serum bilirubin level with the photoelectric colorimeter. J Biol Chem 1937; 119: 481-4.
18. Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ. Protein measurement with Folin Phenol reagent. J Biol Chem 1951; 193: 265-75.
19. Kingsley SR, Frankel SJ. The determination of serum total protein, albumin and globulin by the Biuret reaction. J Biol Chem 1939; 131:197-200.
20. Ansar S, AlGhosoon HT, Hamed S. Evaluation of protective effect of rutin on lead acetate-induced testicular toxicity in Wistar rats. Toxin Reviews 2015; 34(4):195-9.
21. Palanivel MG, Raj Kapoor B, Kumar RS, Einstein JW, Kumar EP, Kumar MR et al. Hepatoprotective and antioxidant effect of *Pisonia aculeata* L. against CCl₄-induced hepatic damage in rats. Scientia pharmaceutica 2008; 76(2):203.
22. Drotman RB, Lawhorn GT. Serum enzymes as indicators of chemically induced liver damage. Drug Chem Toxicol 1978; 1(2):163-71.
23. Akilavalli N, Radhika J, Brindha P. Hepatoprotective activity of *Ocimum sanctum* Linn against lead induced toxicity in albino rats. Asian J Pharm Clin Res 2011; 4(2): 84-87.
24. Azab El-Saied Azab. Hepatoprotective effect of sesame oil against lead induced liver damage in albino mice: histological and biochemical studies. American J BioSci 2014; 2 (6-2): 1-11.
25. Onah CE, Meludu SC, Dioka CE, Onuegbu AJ, Onah CF, Ajaghaku DL, Nnodim JK et al. Amelioratory effect of methanolic leaf extract of *Moringa oleifera* on some liver and kidney function and oxidative stress markers in lead-intoxicated rats. Euro J Medicinal Plants 2016; 12(4): 1-12.
26. Anuradha R and Palaniyandi Krishnamoorthy. Impact of *Pongamia pinnata* extract on lead acetate mediated toxicity in rat liver. Inter J PharmTech Res 2012; 4(2): 878-882.
27. Kansal L, Sharma V, Sharma A, Lodi S, Sharma SH. Protective role of *Coriandrum sativum* (coriander) extracts against lead nitrate induced oxidative stress and tissue damage in the liver and kidney in male mice. Inter J Appl Biol Pharm Tech 2011; 2(3): 65-83.

28. Baxla SL, Gora RH, Kerketta P, Kumar N, Roy BK, Patra PH. Hepatoprotective effect of *Curcuma longa* against lead induced toxicity in Wistar rats. Vet World 2013; 6(9):664-7.

29. Zongping Liu, Jicang Wang, Zijun Yang, Huali Zhu. Antioxidative effects of hesperetin against lead acetate-induced oxidative stress in rats. Indian J Pharmacol 2013; 45(4): 395-98.

How to cite this article:

Phatak RS and Matule SM: Ameliorated activity of *Murraya koenigii* leaves chloroform extract (MKCE) against lead induced hepatic dysfunctions in mice. Int J Pharm Sci Res 2017; 8(6): 2619-23.doi: 10.13040/IJPSR.0975-8232.8(6).2619-23.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)