IJPSR (2014), Vol. 5, Issue 5





Received on 13 December, 2013; received in revised form, 16 February, 2014; accepted, 10 March, 2014; published 01 May, 2014

COMPARISON OF GC-MS ANALYSIS OF PHYTOCHEMICALS IN THE ETHANOLIC EXTRACTS OF *MARCHANTIA LINEARIS* LEHM & LINDENB. AND *MARCHANTIA POLYMORPHA* L. (BRYOPHYTA)

Remya Krishnan and K. Murugan K*

Plant Biochemistry and Molecular biology Laboratory, Department of Botany, University College, Trivandrum 695 034, Kerala, India

Keywords:

GC-MS analysis, Bioactive compounds, *Marchantia linearis, M. polymorpha*, Ethanol extract.

Correspondence to Author:

Dr. Murugan K

Associate Professor, Plant Biochemistry and Molecular biology Laboratory, Department of Botany, University College, Trivandrum 695 034, Kerala, India

Email: harimurukan@gmail.com

ABSTRACT: Bryophytes have unique aroma, taste and possess bioactive potential compounds with medicinal properties. Chemical studies of the bryophytes were limited in India due to the small size and expertise in identification of the species. Many of the bryophytes are used as folklore medicine by local tribes. The solvent extracts results poor yield leads to insufficient for testing biological activity. In vitro culture and suitable chemical synthesis was an alternative undertaken to overcome this issue. In the present study, the bioactive phytochemicals of Marchantia linearis and *M. polymorpha* have been analyzed using GC-MS. The phytochemical composition of the whole thallus ethanol extract was investigated using Perkin-Elmer Gas Chromatography - Mass Spectroscopy. GC-MS analysis of M. linearis and M. polymorpha ethanol extracts revealed the presence of the GC-MS chromatogram of the many peaks .The major compounds in M. linearis and M. polymorpha are n-hexadecanoic acid (28.13% and 18.72% respectively) followed by 1,2 benzene dicarboxylic acid (17.63% and 8.93%) respectively). The medicinal potentiality of the liverworts may be due to the bioactive compounds present in the ethanolic extracts. Further studies are warranted to isolate and purify the lead molecules to evaluate its biological potentialities.

INTRODUCTION: Bryophytes are distributed along the tropical and temperate ecosystems and their ecological significances were well documented ¹. Although, bryophytes prefer damp or moist environments, they are resistant to pest and pathogenic attacks which suggest that they contain a pool of phytochemicals which are antimicrobials.



Bryophytes comprises three groups liverworts (6,000 species), the hornworts (300 species), and mosses (15,000 species). This group represents the second largest group of land plants and pioneer in the evolution of land plants². Bryophytes are with small image and are characterized by dominant gametophyte followed by a reduced or well sporophyte remains developed parasitic on gametophyte ³. Jonathan Shaw *et al* 4 documented this group in many forest ecosystems and forms rich diversity along moist environments, wetland, and mountain ecosystems. Further, bryophytes are ideal environmental indicators and are important contributors to the nutrient and biogeochemical cycling.

International Journal of Pharmaceutical Sciences and Research

Relatively, little is known about the secondary metabolites of bryophytes particularly at structural level and the information is very scattered ⁵. The major limitations for this are the difficulties of proper identification, the limited number of the similar species available for subsequent analyses due to their inconspicuous position in the field and the instrumentation sophistication required.

However, during the last several years more than 400 lead molecules were isolated and structurally elucidated ⁶. The major compounds are flavonoids, biflavonoids, terpenes, terpenoids, like diterpenoids, triterpenoids, lipophilic mono and disesquiterpenoids ⁵.

Characteristic scents of liverworts were due to the aromatic bibenzyls, benzoates, cinnamates or naphtalenes. Most of the phytochemicals in the bryophytes are biologically active substances. Phylogenetically, economically and ecologically these phytochemicals are likely involved in defense and protection against pest and microbial infections. Compared to the vascular plants, bryophytes have no thick walled cells to provide protection from abiotic or biotic stress.

Again, most bryophytes grow on forest floor in a close proximity to several biodegrading wastes and soil borne organisms. Hence, a defense against fungi or bacteria is vital for surviving in the moist, dampy habitat. Microbicidal activity against bacteria and fungi was reported from mosses⁷ and the liverwort *Marchantia linearis*⁸. Since bryophytes are not well known to the common man, even to conservation biologists, it is necessary to highlight their role in nature and their therapeutical value.

The outputs thus provide a considerable potential for biotechnological and biopharmaceutical applications. The Kani tribes of South Kerala use *Marchantia linearis* for curing many skin borne allergies and tumors ⁹.

With this background the present study was aimed to identify and compare the phytochemicals present in *M. linearis and M. polymorpha* using GC-MS analysis.

MATERIALS AND METHODS:

Plant material: Fresh thallus of *Marchantia linearis* and *M. polymorpha* were collected from Kallar river floor of Ponmudi hills, Kerala, India (**Fig. 1 a & b**). Taxonomic identity was confirmed by comparing with authenticated herbarium specimen at Department of Botany Herbaria, University of Calicut, Kerala. Voucher specimens of the plants are kept in the herbarium of the institute.



FIGURE 1A & B: THALLUS OF MARCHANTIA LINEARIS AND MARCHANTIA POLYMORPHA

Extraction procedure: The shade dried powder of the thallus of *Marchantia linearis* and *M. polymorpha* (50 g) were separately extracted with ethanol (500 ml, 48 h) at temperature between 60- 65° C by using Soxhlet extractor. The extracts were filtered in hot condition and concentrated in the vacuum under reduced pressure and dried in desiccators.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

The extracts contain both variable polarity from highly polar to non-polar components and 0.2 μ L sample of the solution was subjected to GC-MS for analysis for identifying the various phytochemicals.

GC-MS analysis: The GC-MS analyses of extracts were performed using a Perkin Elmer GC-MS equipped with a VF-5 MS fused silica capillary column (30 m x 0.25 mm i.d, film thickness 0.25 μ m). GC-MS spectroscopic detection, an electron ionization system with ionization energy of 70 eV was used. Pure helium gas was employed as a carrier gas at a constant flow rate of 1 mL/min. Mass transfer line and injector temperatures were set at 250°C. The oven temperature was programmed at 60°C for 2 min then increased to 300 °C for 6 min at the rate of 10°C/min. The samples were injected in split mode as 10:1. **Identification of phytochemicals:** Interpretation on Mass-Spectrum GC-MS was carried out with reference to the software database of libraries like WILEY08, NIST08 and NIST08s. The name, molecular weight and percentage of unknown compounds were evaluated by software.

RESULTS AND DISCUSSION: In the present study, the GC-MS analysis of the ethanolic extracts of *M. linearis* and *M. polymorpha* showed the presence of seven and five compounds respectively (**Fig. 2a & b**).

The identified compounds of the thallus of M. *linearis*, their retention indices, compound, percentage composition, chemical structure and activities are given in **Table 1**.



FIGURE 2A & B: GC MS CHROMATOGRAM OF ETHANOLIC EXTRACTS OF *M. LINEARIS* AND *M. POLYMORPHA*

			Marchantia linearis	Marchantia polymorpha
Sl. No.	Retention time	Compound name	Peak area %	Peak area %
1	182367	Phenol 2,4-bis (1,1-dimethyl ethyl)	16.81	-
2	23.392	Hexadecanoic acid	28.13	18.72
3	23.667	1-heptacosanol	3.54	-
4	25.150	Octadecanoic acid	10.72	-
5	25.458	Eicosyl heptaflurobutyrate	5.35	-
6	27.100	Isochiapin B	2.53	-
7	28.025	1,2-benzene dicarboxylic acid	17.63	8.93
8	24.950	9-octadecanoic acid	-	25.34
9	27.500	Dichloro acetic acid 1-adamantyl methyl ester15.21	-	15.21
10		Ethyl adamantane	-	11.51

TABLE 1: GC-MS CHROMATOGRAM ANALYSIS OF ETHANOLIC EXTRACTS OF *MARCHANTIA LINEARIS* AND *MARCHANTIA POLYMORPHA* SHOWING MAJOR PHYTOCHEMICALS

Biological activities of the compound are based on Phytochemical and Ethnobotanical Databases of Duke. The results in *M. linearis* showed the presence of hexadecanoic acid (28.13%), phenol, 2,4-bis(1,1-dimethyl ethyl) (16.81%), 1heptacosanol (3.54%), octadecanoic acid (10.72%), (2.53%), and 1,2-benzene dicarboxylic acid (17.63%). The spectrum profile of GC-MS confirmed the presence of 7 major components with retention time 18.267, 23.392, 23.667, 25.150, 25.458, 27.100 and 28.025 respectively (**Fig. 2a & b**) the individual fragmentation of the components is illustrated in (**Fig. 3 a-g**).







FIGURE 3: A, B, C, D, E, F & G: THE INDIVIDUAL FRAGMENTATION OF THE COMPONENTS OF GC MS CHROMATOGRAM OF *M. LINEARIS*

Similarly *M. polymorpha* displayed only 5 major compounds such as hexadecanoic acid (18.72%), 9-octadecanoic acid (25.34%), dichloroacetic acid 1- adamantyl methyl ester (15. 21%), 1,2-benzene dicarboxylic acid (8.93%) and 1-ethyl adamantine (11.51%). The corresponding retention times were

23.342, 24.950, 27.500, 28.033 and 28.400 respectively (**Fig. 4 a-e**). Hexadecanoic acid and 1, 2-benzene dicarboxylic acid is the common phytochemicals between the species but with different percentages.





FIGURE 4: A, B, C, D & E: THE INDIVIDUAL FRAGMENTATION OF THE COMPONENTS OF GC MS CHROMATOGRAM OF *M. POLYMORPHA*

Aparna *et al*¹⁰ attempted the structural and kinetics studies of n-hexadecanoic acid, is an inhibitor of phospholipase A(2) and confirmed as an antiinflammatory compound. Phenol, 2, 4-bis (1, 1dimethyl ethyl) derivatives are function as antioxidant.1-heptacosanol has the role of antioxidant and antimicrobial¹¹. Isochiapin B has been identified by Chinese medicine to treat arthritis, tonsillitis, and other ailments. 1-ethyl adamantine was employed against viral fever and also to treat Parkinson's disease.

include Derivatives adapalene, amantadine, dopamantin, karmantadin, memantine. rimantadine, saxagliptin, tromantadine and vildagliptin as antiviral agents against HIV12. In addition to these play various pharmacological actions like anti-inflammatory, antibacterial, antihelminthic activi-ties for sesquiterpene lactones and anti-arthritic, pesti-cide, antitumor, cancer anti-histaminic, hepatoprotective, preventive, hypocholesterolemic and anti-¬inflammation activities for fatty acids ester.

GC-MS analysis of the present study was more commendable than that of *Eupatorium odoratum*¹¹; *Trichilia connaroides* (Meliaceae) ¹³; *Epaltes divaricate* ¹⁴; *Cassia italic* ¹⁵ and *Fagonia longispina* ¹⁶. Selvamangai and Bhaskar ¹⁷ carried GC-MS analysis in *Eupatorium triplinerve* International Journal of Pharmaceutical Sciences and Research

revealed the following components such as hexadecanoic acid (14.65%), 2,6,10-trimethyl, 14ethylene14pentadecne(9.84%),bicycle[4.1.0]heptan es (2.38%), decanoic acid (3.86%), 1-undecanol (7.82%),1-hexyl-1-nitrocyclohexane (2.09%).1.14-tetradecanediol (6.78%), octadecanoic acid 2-hydroxy-3-[(9E)-9-(19.18%)and octa decenoyloxy] propyl (9E)-9-octadecenoate (8.79%).

The percentages of the compounds are comparatively lower than *Marchantia* species.

There is growing importance in analyzing the secondary metabolites and their therapeutic potentialities. *Marchantia* is used in Chinese medicine however there are no reports on the thorough phytochemical analysis of the plant. This is the first report of the presence of some of the unique molecule resolved by GC-MS analysis and their medical utilities.

CONCLUSION: GC-MS analysis of phytochemicals in the ethanol extracts of thallus of *M.linearis and M.polymorpha* revealed 7 and 5 major compounds. The presence of various bioactive compounds justifies the use of the *Marchantia* species for curing various ailments by Chinese traditional practitioners.

Therefore, GC-MS method is a direct and accurate analytical method for identification of phytochemicals in the plant extracts. However, isolation of individual phytochemical constituents and subjecting it to medicinal property will definitely give promising results.

REFERNCES:

- 1. De Brito Valente E, Katia Cavalcanti Porto, Cid José Passos Bastos and Jana Ballejos-Loyola: Diversity and distribution of the bryophyte flora in montane forests in the Chapada Diamantina region of Brazil. Acta Botanica Brasilica 2013; 27(3): 506-518.
- Laura Kļaviņa, Oskars Bikovens, Iveta Steinberga, Viktorija Maksimova and Linda Eglite :Characterization of chemical composition of some bryophytes common in Latvia. Environmental and Experimental Biology 2012; 10: 27–34.
- Cox CJ, Goffinet B, Wickett NJ, Boles SB and Shaw AJ: Moss diversity: A molecular phylogenetic analysis of genera. Phytotaxa 2010; 175-195.
- 4. Jonathan Shaw A, Peter Szovenyi and Blanka Shaw: Bryophyte diversity and evolution: windows into the early evolution of land plants. American Journal of Botany 2011; 98(3): 352–369.
- 5. Alam: Some Indian bryophytes known for their biologically active compounds .International Journal of Applied biology and Pharmaceutical technology 2012; 3(2): 239-246.
- Alam A: Some Indian bryophytes known for their biologically active compounds. International Journal of Applied Biology and Pharmaceutical Technology 2012; 3(2): 239-246.
- Manoj GS and Murugan K: Phenolic profile, antimicrobial and antioxidant potentialities of methanolic extract of a liverwort, *Plagiochila beddomei* Steph. Indian journal of Natural Products and Resources 2012; 3(2): 173-183.
- 8. Murugan K and Remya Krishnan: Phytochemical analysis, *in vitro* antifungal activity and mode of action of ethanolic extract of *Marchantia linearis* Lehm & Lindenb. A

bryophyte. World Journal of Pharmacy and Pharmaceutical Sciences 2013; 2(5): 3650-3666.

- 9. Ramesh M and Manju CN: Ethnobryological notes from Western Ghats, India, The Bryologist 2009; 112(3): 532.
- Aparna V, Dileep KV, Mandal PK, Karthe P, Sadasivan C and Haridas M: Anti-inflammatory property of nhexadecanoic acid: structural evidence and kinetic assessment. Chemical Biology and Drug Design 2012; 80(3): 434-9.
- 11. Venkata Raman B, Samuel La, Pardha Saradhi M, Narashimha Rao B, Naga Vamsi Krishna A, Sudhakar M and Radhakrishnan TM: Antibacterial, antioxidant activity and GC-MS analysis of *Eupatorium odoratum*. Asian Journal of Pharmaceutical and Clinical Research 2012; 5(2): 99-106.
- 12. Moses Owolabi1a S, Akintayo Ogundajo, Kamil Yusuf O, Labunmi Lajide, Heather Villanueva E, Jessika Tuten A and William Setzer : Chemical Composition and Bioactivity of the Essential Oil of *Chromolaena odorata* from Nigeria. Records of natural products 2010; 4(1): 72-78.
- Senthilkumar N, Murugesan S and Vijayalakshm KB: GC-MS-MS analysis of *Trichilia connaroides* (Wight & Arn.) Bentv (Meliaceae): A tree of ethnobotanical records. Asian Journal of Plant Science and Research 2012; 2(2):193-197.
- 14. Amala K, Saraswathy A and Amerjothy S: GC-MS analysis of *N*-Hexane extract of *Epaltes divaricata* (L.). Cass 2013; 2 (1): 33-35.
- Sermakkani M and Thangapandian V: GC-MS Analysis of Cassia italica leaf methanol extract. Asian Journal of Pharmaceutical and Clinical Research 2012; 5(2): 90-94.
- 16. Hamidi N, Lazouni HA, Moussaoui A, Ziane Laid and Amal S: GC-MS analysis of ethanol extract from the aerial parts of *Fagonia longispina* (Family Zygophyllaceae) Asian Journal of Natural & Applied Sciences 2012; 1(2): 136-142.
- 17. Selvamangai G and Bhaskar A: GC-MS analysis of phytocomponents in the methanolic extract of *Eupatorium triplinerve*. Asian Pacific Journal of Tropical Biomedicine 2012: 1329-1332.

How to cite this article:

Krishnan R and Murugan K: Comparison of Gc-Ms Analysis of Phytochemicals In The Ethanolic Extracts of *Marchantia Linearis* Lehm & Lindenb. and *Marchantia Polymorpha* L. (Bryophyta) *Int J Pharm Sci Res* 2014; 5(5): 1981-87. doi: 10.13040/IJPSR.0975-8232.5 (5).1981-87.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)