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## ISOLATION, PURIFICATION AND CHARACTERIZATION OF ANTIBACTERIAL PROTEINS FROM INDUCED LARVAE OF ANTHEREAE MYLITTA DRURY (DABA TV)

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### **Keywords:**

Antibacterial proteins, Anthereae mylitta drury, Pseudomonas aeruginosa AC-3, Pseudomonas DAS - 01

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**ABSTRACT:** Antibacterial proteins are evolutionarily ancient, effect or molecules of innate immune defense and are ubiquitously found in all kingdoms. The present study deals with the induction of antibacterial proteins in the haemolymph of *Anthereae mylitta* drury (Daba TV) by injecting non-pathogenic *Pseudomonas* DAS-01. Purified protein fraction obtained on ammonium sulphate precipitation followed by ion exchange chromatography had shown antibacterial activity against *Pseudomonas aeruginosa* AC-3 and *Pseudomonas* DAS - 01. Flow through obtained in ion exchange chromatography found effective in inhibiting the growth of both *P. aeruginosa* AC-3 and *Pseudomonas* DAS - 01. In induced larvae, purified antibacterial peptides obtained by ion exchange chromatography on SDS-PAGE had shown a molecular weight of 16kDa and 24kDa which have antibacterial activity and were not observed in control.

**INTRODUCTION:** Anthereae mylitta drury a lepidopteran insect of the Saturniidae family produces tasar silk of commercial importance. The environment and various diseases of silkworm are major constraints in tasar culture which reduced the crop yield and made the silk industry unreliable <sup>1</sup>. In Anthereae mylitta drury the bacterial infections (flacherie) lead to rectal protrusion, sealing of anal lips and leading to chain type excreta. The causative organisms for this flacherie include Streptococcus sp., Staphylococcus sp., Bacillus thuringiensis, Serratia marcescence, Pseudomonas sp., etc. Antibacterial peptides are evolutionarily conserved and have both hydrophobic and hydrophilic sides that enable the molecules to be soluble in aqueous environments yet also enter lipid - rich membranes<sup>2</sup>.



These peptides kill target cells through diverse mechanisms once they reach the target microbial membrane. They possess a wide array of biological mechanisms from direct killing of invading pathogens to modulation of immunity and other biological responses of the host  $^3$ . These antimicrobial peptides are the first line of host defense in various species, have a mass of  $\leq 10 \text{ kDa}$  and are readily synthesized and efficiently diffuse at the point of pathogen entry or infection, hence form an invaluable component of the innate immune system  $^{4,5}$ .

The two stages to fight an infection involves, removing the bacteria from circulation by phagocytosis, nodule formation and de novo synthesis of RNA and specific proteins which give rise to increasing antibacterial activity in haemolymph <sup>6</sup>. Various works has been carried out on induction of immunity in insects <sup>7 - 9</sup>. Some peptides that are well characterized in insects include mastoparan, poneratoxin, cecropin, moricin and melittin <sup>10</sup>. Earlier, antibacterial peptides have been isolated and purified from insects of different

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orders, *viz.*, dipteran, isopteran, lepidopteran and coleopteran <sup>3, 11, 12, 13</sup>. Given the amphoteric character of proteins the pH of the solution is important in the determination of the type of ion exchanger used. Immunoglobulins, although they can be purified by either cation or anion exchange chromatography, are most frequently purified by anion exchange with DEAE resins <sup>14</sup>.

Present study has taken up to induce synthesis of antibacterial proteins in *Anthereae mylitta* drury (Daba TV) and also to isolate, purify and characterize these proteins and to find their efficacy in controlling bacterial disease caused by *Pseudomonas* sp.

### **MATERIALS AND METHODS:**

Culture Medium and Maintenance of Micro Organisms: In the present experiment the bacteria used for challenging Anthereae mylitta drury (Daba TV) were P. aeruginosa AC-3 and nonpathogenic Pseudomonas DAS - 01. Bacterial colonies were grown on LB agar plate at 37 °C overnight in an orbital shaker at 300 rpm and 37 °C temperature. The overnight grown culture was centrifuged (REMI centrifuge) next day at 4 °C and 5000 rpm, and the pellet was resuspended in sterile saline. Before injecting, the number of Pseudomonas cells counted by serial dilution method, made a dosage of 1 X 10<sup>6</sup> cells/ml and stored for further study.

Anthereae mylitta Drury (Daba TV) Rearing: During the month of April 150 Daba TV coccons were collected from the forest patches of Jakaram, Warangal District, accomadated in wire mesh cages of size 2ftX2ftX2ft. The emerged moths were tested for microsporidiasis <sup>15</sup>. The eggs from healthy moth were incubated. The hatched larvae were reared in the field. Third instar larvae of 18gm weight each were divided into two batches containing 50 larvae each. One batch was injected with a dose of 2µl of non-pathogenic Pseudomonas DAS - 01 from stored culture medium on the dorsal segment by Hamilton micro syringe whereas for control at the same position, 2µl Ringer's solution was injected. Haemolymph was collected from both the batches in the test tubes by cutting the proleg. 1 mg of Thiourea was added to the test tubes to prevent melanisation. After centrifugation at 15000 rpm for 15 min the supernatant was collected and stored at - 20 °C for further analysis.

Antibacterial Assay: Antibacterial assay was done by diffusion method <sup>16</sup>. LB media is inoculated with 0.1 ml (10<sup>6</sup> cells/ml) of two bacterial strains. Sterile paper discs soaked in *Pseudomonas* DAS-01 induced and control Daba TV haemolymph were inserted in the media present in separate plates. The plates were incubated at 35 °C and inhibitory activity was determined after 48 hrs by zone of inhibition.

**Protein Estimation:** Haemolymph proteins of induced and control, flow through, wash and elute was estimated by <sup>17</sup>.

**Protein Purification:** The protein was purified by ammonium sulphate precipitation and ion exchange chromatography using DEAE - Sepharose.

**Ammonium Sulphate Fractionation:** Concentration required for crude supernatant was found to be 0 - 30% saturation. The precipitate obtained was further dialyzed against 0.01 M phosphate buffer (pH 7.5).

Ion Exchange Chromatography: 20ml dialysate obtained from ammonium sulphate precipitation (0 - 30% fraction) was subjected to ion exchange chromatography using DEAE-Sephroase. DEAE sepharose was packed in Bio-Rad econo column (42.5 X 2.5 cm) equilibrated with 0.01 M phosphate buffer pH 7.5 with a flow rate of 1.5mL/min which was set up in an FPLC system (Bio-Rad). The bound fraction was eluted by a linear gradient of 0 - 1 M NaCl in 0.01 M phosphate buffer. Flow through, wash and elute peak fractions were pooled and dialyzed against the phosphate buffer pH 7.5 (0.01 M). The dialyzed fractions were studied for their effectiveness against bacteria by means of zone of inhibition <sup>18</sup>.

Antibacterial Assay with Dialysed Fractions: For this 10 healthy 3<sup>rd</sup> instar larvae of Daba TV were injected with 5µl flow through, and ten were injected with 5µl Ringer solution. After half an hour, they were challenged with 2µl of overnight grown culture of *P. aeruginosa* AC-3 and *Pseudomonas* DAS - 01 and fed on tender leaves of *Terminalia arjuna*. Haemolymph collected from control and flow through injected larvae was tested for antibacterial activity by inhibition zone assay method.

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**SDS** - **PAGE:** Haemolymph of flow through injected larvae and control was subjected to 15% SDS PAGE <sup>6</sup>. Haemolymph samples diluted to sample buffer of mercaptoethanol and heated to 100 °C for 3 min before application.

**RESULTS:** Present study focuses on induction of immunity in *Anthereae mylitta* drury (Daba TV) by

injection of live non-pathogenic strain of *Pseudomonas* DAS - 01. Haemolymph collected from immunized third instar Daba TV has revealed a clear zone formation of size 2.3 mm and 2.4 mm against *P. aeruginosa* AC3 and Pseudomonas DAS - 01 in plate growth inhibition assay whereas untreated larvae (Control) were found to be devoid of significant zone of inhibition (**Fig. 1** and **2**).

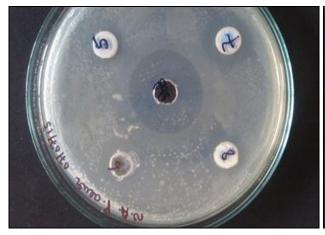


FIG. 1: ANTIBACTERIAL ACTIVITY SHOWN BY INDUCED AND CONTROL LARVAE 7- Induced on *Pseudomonas aureus*, 8 - Induced on Pseudomonas DAS, 5, 6 - Control.

8

FIG. 2: NO ANTIMICROBIAL ACTIVITY SHOWN BY CONTROL LARVAE INDUCED WITH RINGER SOLUTION 5, 6, 7, 8 - No Effect

Results also show that, the fractions collected from ion exchange chromatography also shown antibacterial activity. Flow through has shown highest activity with a clear zone formation of size 2.5 mm and 2.6 mm against *P. aeruginosa* AC3 and *Pseudomonas* DAS-01 (**Fig. 3**). Lepidopteran antibacterial peptides had strong inhibition against various bacteria <sup>19</sup>. It is known that insects

inoculated with live non-pathogenic or killed bacteria can acquire resistance to subsequent challenge by bacterial pathogen <sup>20</sup>. It was found that in Drosophila, response to microbial infections mounts a multifaceted immune response involving humoral reactions that culminate in destruction of invading organisms by lytic peptides <sup>21</sup>.

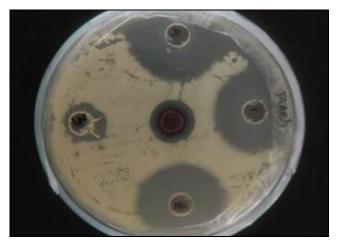


FIG. 3: ANTIBACTERIAL ACTIVITY SHOWN BY FRACTIONS COLLECTED FROM ION EXCHANGE CHROMATOGRAPHY 1- Flow through on *Pseudomonas aureus*, 2, 4 – Flow through on *Pseudomonas* DAS - 01, 3-

Estimation of haemolymph proteins after injecting non-pathogenic strain of *Pseudomonas* DAS - 01 purification by ammonium sulphate precipitation, and Ringer's solution (control) in third instar Daba TV resulted in 28.6mg/ml, 24.8mg/ml respectively.

non-pathogenic strain of *Pseudomonas* DAS - 01 and Ringer's solution (control) in third instar Daba TV resulted in 28.6mg/ml, 24.8mg/ml respectively. It was found that protein concentration in injected haemolymph was higher by 3.8mg/ml than its control counterpart.

Purification of antibacterial proteins by ion exchange chromatography resulted in peaks like flow through, wash and elution (**Fig. 4**). **Table 1** shows antibacterial activity of flow through, wash and elution. The flow through injected Daba TV larvae was found effective against both *P. aeruginosa* AC-3 and *Pseudomonas* DAS - 01 with maximum zone of inhibition followed by wash and least activity was recorded in elute.

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Present results also show that flow through obtained from ion exchange chromatography contains maximum protein followed by wash and elute (**Table 1**). This difference can be attributed to induced proteins formed against bacteria for self-defence. Studies on insect haemolymph revealed the presence of a variety of proteins in response to

TABLE 1: ANTIBACTERIAL ACTIVITY OF HAEMOLYMPH OF BACTERIAL INJECTED, CONTROL AND FRACTIONS COLLECTED FROM ION EXCHANGE CHROMATOGRAPHY

S. no.	Source	Protein concentration	Zone of inhibition for	Zone of inhibition for
		(mg/ml)	P. aureginosa (mm)	<b>DAS - 01 (mm)</b>
1	Haemolymph (injected)	28.6±1.4	2.3	2.4
2	Haemolymph (Control)	24.8±1.6	0	0
4	Flow through	$32.4\pm1.2$	2.5 (high activity)	2.6 (high activity)
5	Wash	$8.4\pm0.4$	0.8 (moderate)	0.6 (moderate)
6	Elute	$6.2\pm0.2$	0.4 (low activity)	0.4 (low activity)

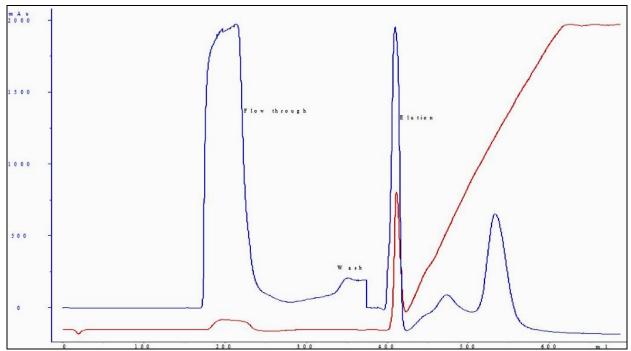


FIG. 4: PURIFICATION OF ANTIBACTERIAL PROTEINS BY ION EXCHANGE CHROMATOGRAPHY Peak 1: Flow through, Peak 2: Wash, Peak 3: Elution.

3rd instar (Daba TV) inoculated with this active effective flow through fraction and the control with Ringer solution. The haemolymph obtained from these larvae when subjected to SDS - PAGE had shown various bands. But two peptidesin the low

molecular weight region were observed only in the flow through fraction injected larvae. These bands were detected between 16 and 24 kDa low molecular weight region (**Fig. 5**).

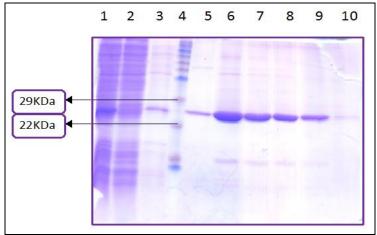


FIG. 5: SDS-PAGE OF LARVAL HAEMOLYMPH INJECTED WITH LIVE BACTERIA AND RINGER SOLUTION 1, 2 - control, 4- marker 3, 5, 6, 7, 8, 9, 10 - induced antibacterial proteins.

These proteins are effective in fighting against Gram-negative bacteria such as P. aeruginosa AC-3, and Pseudomonas DAS - 01, which was concluded by inhibition zone assay method. Lysozyme of 11 - 28 kDa protein, directly attacks bacteria by hydrolysing their peptidoglycan layer of the cell wall <sup>23</sup>. Cecropins and attacins are small molecular weight proteins ranging from 4 to 10 kDa and are effective against both Gram-positive and Gram-negative bacteria <sup>24</sup>. Antimicrobial peptides (AMPs) are reported to be small molecular weight proteins with the molecular sizes in the range of 7 to 12 kDa <sup>25, 2</sup>. Protactins 1 and 3 purified and characterized from larval haemolymph of Protaetia brevitarsis, have molecular masses of 7.5 and 12 kDa, respectively while protactin 2 was determined to be 9kDa 13.

**DISCUSSION:** The spread of flacherie in insects can be limited by synthesizing antimicrobial peptides in its body. They have pattern recognition proteins that can bind to the external surface of bacteria or other pathogens <sup>26</sup>.

In this study the non-induced and the induced haemolymph of *Anthereae mylitta* larvae was screened for the antimicrobial activity against *P. aeruginosa* AC-3 and pseudomonas DAS - 01. The non - induced haemolymph did not show inhibitory activity whereas induced haemolymph had shown inhibitory activity. Defensive peptides or proteins play a crucial role in insect humoral immune response against invading microorganisms <sup>27, 28</sup>. Insect species injected with bacteria into haemocoel, elicit the synthesis of peptides and proteins, which are individually or cooperatively

active against the foreign microorganisms and induction is a common process in many insect species <sup>29</sup>. *Anthereae mylitta* drury (Daba TV) injected with purified antimicrobial fraction (flow through) resulted into proteins which were active against tested bacterial strains.

Antibacterial assay resulted in maximum zone of inhibition confirms that peptides are produced to combat bacterial infection. Low molecular weight for these peptides is in correlation with molecular weight of lysozyme. However, constitutive and inducible proteins may be present in haemolymph of some insects and may act as signalling molecules such as lysozyme <sup>30, 31</sup>.

It may be assumed that the proteins identified in this study might play an important role in their self-defense against bacterial infection in *Anthereae mylitta* drury. However, further studies are needed to study the combined effect of proteins. Many bacteria have already developed resistance to conventional antibiotics like Ampicillin, penicillin *etc.*, It is also believed that antimicrobial peptides will be assumed in the near future as an alternative for the classical antibiotics <sup>30</sup>. The advantages of antimicrobial peptides are many *viz*, selectivity, fast killing, broad antimicrobial spectra and lack of resistance development <sup>32</sup>.

**CONCLUSION:** Thus in conclusion some inducible antibacterial peptides are present in *Anthereae mylitta* drury (Daba TV). These peptides can be used as blueprints for the design of new antimicrobial agents. Antimicrobial peptides based therapies will form attractive candidates as

alternative antibiotic treatments, since they offer several potential advantages over currently used classes of drugs.

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### **REFERENCES:**

- Reddy RM, Hansda G, Ojha NG and Suryanarayana N: Heterobeltiosis in F1 hybrids of wild and domesticated ecoraces of tropical tasar silkworm *Antheraea mylitta* Drury. Sericologia 2008; 49: 189-200. Doi: 10.5829/idosi.aje.2014.7.4.8697.
- Biljana M and Havard J: Peptides and Peptidomimetics for Antimicrobial Drug Design. Pharmaceuticals 2015; 8: 366-415. Doi:10.3390/ph8030366.
- David R and Pascale C: How bacterial pathogens colonise their hosts and invade deeper tissues: Microbes and Infection 2015; 17(3): 173-183. http://doi.org/10.101.6/jmicinf. 2015.01.004.
- Rethnapriya E and Ravichandran S: Bioactive peptides from the grapsid crab grapsusstrigosus Sakai. Asia journal of pharmaceutical and clinical research 2014; 7(5): 305-308.
- Guangshun W: Human Antimicrobial Peptides and Proteins. Pharmaceuticals 2014; 7(5): 545-594. Doi.10-3390/ph7050545.
- Laemmli UK: Cleavage of Structural Proteins during the Assembly of the Head of bacteriophage T4. Nature 1970; 227: 680–685.
- 7. Leonardo FL, Natalia MR, Agusto UF, Matheus CGV, Fernanda LC, Valquiria B, Denise F and Celia CR: Humoral and cellular immune responses induced by the urease-derived peptide Jaburetox in the model organism *Rhodnius prolixus*. Parasites and Vectors 2016; 9: 412. https://dx. doi.org/10.1186/s13071-016-1710-3.
- 8. Daniel Ardia R, Jacob Gantz E, Brent C, Schneider and Stefanie S: Costs of immunity in insects: an induced immune response increases metabolic rate and decreases antimicrobial activity. Functional ecology 2012; 26(3): 732-739. DOI.10.1111/j.1365-2435.2012.01989.x.
- Robert K, Badrul A and Ulrich T: Damage signals in the insect immune response. Frontiers in plant science 2014; 5: 1-11. Doi:10.3389/fpls.2014.00342.
- Kerridge A, Lappin-Scott H and Stevens JR: Antibacterial properties of larval secretions of the blowfly, *Lucilia* sericata. Medical and veterinary entomology 2005; 19: 333-337.
- Rabeeth M, Muthukumar M, Balaji Rajkumar M and Kumar KH: Isolation, evaluation and purification of antibacterial peptides from rhinoceros beetle, *Oryctesr hinoceros* (L).GERF Bulletin of biosciences 2012; 3(2): 16-22.
- 12. Hui YY, Chowdhury M, Ya- Dong H and Xiao QY: Insect Antimicrobial Peptides and their Applications.

Applications of microbial biotechnology 2014; 98(13): 5807-5822. DOI.10.1007/s00253-014-5792-6.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

- Sebrin JI, Sukanya B and Jatin K: A Review on Antimicrobial Peptides from *Bombyx mori* L. and Their Application in Plant and Animal Disease Control. Journal of Advances in Biology and Biotechnology 2016; 9(3): 1-15. Doi:10.9734/JABB/2016/27539
- Grodzki AC and Berenstein: Antibody purification: ionexchange chromatography, Methods in molecular biology 2010; 588: 27-32. https://dx.doi.org/10.1007/978-1-59745-324-04.
- 15. Pasteur L: Etudessur la maladie des vers a soie. Gauthier-Villars, Paris. Tome I 1870; 322-327.
- 16. Bauer AW, Kirby WMM and Sherric JC: Antibiotic susceptibility testing by a single disc method. American journal of clinical pathology 1996; 45: 493-496.
- 17. Lowry H, Rosebrough NL, Far AL and Randal RJ: Protein measurement with Folin reagent. Journal of Biological Chemistry 1951; 193: 265-275.
- Sathyan N, Chaithanya ER, Anilkumar PR, Sruthy KS and Philip R: Comparison of the antimicrobial potential of the crude peptides from various groups of marine molluscs. International Journal of Research in Marine Sciences 2014; 3(2): 16–22.
- Peng R, Yang Z, Liu K, Yao H, Yang H, Cui Y and Hong H: Induction, selection and antibacterial activity of the antibacterial peptides from lepidopteran insect cultured cell lines. Frontiers of Biology in China 2008; 3(2): 203 -206
- Baster B, Alice LM, Boran A, Benjamin PJ and Nicole GM: Exposure to bacterial signals does not alter pea aphids survival upon a second challenge or investment in production of winged offspring. Plos One 2013; 8(8): 1-7. https://doi.org/10.1371/journal.pone.0073600.
- Rekha D, Smriti, Kumar S, Kumar A and Pradeep: Cationic Polymers and their Self-Assembly for Antibacterial Applications. Current topics in medicinal chemistry 2015; 15(13): 1179-1195.
- Gundeti R, Swetha SN, Rajalingam G and Nagarajarao P: Certain biochemical changes in haemolymph of eri silkworm, *Samia cynthiaricini* after inoculation with bacteria. International journalof applied science and biotechnology 2015; 3(2): 236-242. doi: 10.3126/ijasbt.v 3i2.12453.
- Jessica H and Laurel LL: Bacterial peptidoglycan degrading enzymes and their impact on host muropeptide detection. Journal of innate immunity 2009; 1: 88-97. doi.10.1159/000181181.
- 24. Peravali JB, Kotra SR, Sobha K, Nelson R, Rajesh KV and Pulicherla KK: Antimicrobial peptides: An effective alternative for antibiotic therapy. Mintage journal of pharmaceutical and medical sciences 2013; 2(2): 1-7.
- Vlisidou I and Wood W: Drosophila blood cells and their role in immune responses. FEBS Journal 2015; 282(8): 1368-82. doi: 10.1111/FEBS.13235.
- Ping L, Tingcai C, Shengkai J, Yugian W, Bohua F, Renwen L, Ping Z and Qing YX: PC, a Novel Oral Insecticidal Toxin from *Bacillus bombysepticus* involved in host lethality via APN and BtR-175. Scientific Reports, 2015; 5: 1-14. doi: 10.1038/srep11101.
- 27. Cytrynska M, Mak P, Barabas AZ, Suder P and Jakubowicz T: Purification and characterization of eight peptides from *Galleria mellonella* immune haemolymph. Peptides 2007; 28: 533 546. https://dx.doi.org/10.1016/j.peptides.2006.11.010.
- 28. Mak P, Zdybicka-Barabas A and Cytrynska MA: different repertoire of *Galleria mellonella* antimicrobial peptides in

- larvae challenged with bacteria and fungi. Developmental and Comparitive Immunology 2010; 34: 1129–1136. https://dx.doi.org/10.1016/j.dci.2010.06.005.
- 29. Vilcinskas A: Evolutionary plasticity of insect immunity. Journal of Insect Physiology 2013; 59: 123–129. Doi: 10.1016/j.jinsphys.2012.08.018.
- 30. Royet J, Gupta D and Dziarski R: Peptidoglycan recognition proteins: modulators of the microbiome and inflammation. Nature Reviews Immunology 2011; 11: 837–851. https://dx.doi.org/10.1038 /nri3089.
- 31. Bosco-Drayon V, Poidevin M, Boneca IG, Narbonne-Reveau K, Royet J and Charroux B: Peptidoglycan sensing by the receptor PGRP-LE in the Drosophila gut induces immune responses to infectious bacteria and tolerance to microbiota. Cell Host Microbe 2012; 12: 153–165. http://dx.doi.org/10.1016/j.chom.2012.06.002.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

32. Hancock RED and Sahl HG: Antimicrobial and host defense peptides as new anti-infective therapeutic agent. Nature biotechnology 2006; 24: 1551-1558. https://dx.doi.org/10.1038/nbt1267.

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