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HEPATOPROTECTIVE ACTIVITY OF *APONOGETON NATANS* (LINN.) ENGL. AND KRAUSE - AN IMPORTANT FOLKLORE MEDICINE

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ABSTRACT: The liver plays a significant role in metabolism and detoxification. *Aponogeton natans* (Linn.) Engl. and Krause is a important ingredient in preparation of an important Ayurvedic preparation Useerasava. The screening for hepatoprotective activity against carbon tetrachloride induced hepatotoxicity was followed as per standard procedure. Animals received CCl₄ (2 ml/kg, 1:1 in olive oil, i.p.) to induce hepatotoxicity. Group I served as normal control and received the vehicle alone. Group II served as toxic control, Group III, IV, V, VI received 200 mg/kg of petroleum ether, benzene, chloroform, methanol extracts respectively and Group VII was treated with silymarin. The groups received the pre-treatment of *A. natans* extracts ANME and ANCE at dose levels of 200 mg/kg body weight significantly controlled the change in the biochemical parameters. The methanol extract (ANME) and chloroform extract (ANCE) at dose levels of 200 mg/kg exhibited significant increase respectively in the serum total protein level as compared to toxic control group and the effect was compared with the standard group treated with silymarin (Sily-100). The LPO, SOD, Catalase (CAT) and GPx activities in the toxic control group depleted significantly as compared to the normal control group. The antioxidants activity increased by ANME, ANCE and standard drug silymarin. Treatment with *A. natans* methanol extract increased the level of biochemical parameters activities when compared to toxic control. *A. natans* methanol extract was found to significantly increase the level of antioxidants levels. The level of LPO which was elevated brought to normal. The above results were further supported by histopathological evidences.

INTRODUCTION: The liver is plays a significant role in metabolism and detoxification of exogenous toxins and therapeutic agents¹. A number of natural and synthetic agents act as hepatotoxins and produce a variety of liver diseases. There are number of herbal extracts, which are reported to have antihepatotoxic activity².

Aponogeton natans (Linn.) Engl. and Krause. belongs to Aponogetonaceae family. *Aponogeton natans* (Linn.) Engl. and Krause occurs in plains and marshy places in Asia, Australia, India and Srilanka. Traditionally leaf pastes are taken with hot water to cure cuts and wounds³. Fresh tuber paste is boiled with 200 ml of coconut oil and applied on hair before bath for three days to cure fungal infection⁴.

Aponogeton natans (Linn.) Engl. and Krause is an ingredient in preparation of an important Ayurvedic preparation Useerasava. This asava is useful in haemothermia, anaemia, impurity of blood and diabetes⁵. A perusal of existing reports reveals that the no detailed hepatoprotective study had been

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done earlier. The study has been planned to investigate the hepatoprotective activity of various extracts of *Aponogeton natans* (Linn.) Engl. and Krause. leaf with leafstalks against CCl₄ induced liver toxicity in experimental animals.

MATERIALS AND METHODS:

Plant Material: Fresh parts of *Aponogeton natans* (Linn.) Engl. and Krause. were collected from Salipur, Cuttack, Odisha, India which was identified and authenticated by Professor P. Jayaraman, PARC, Chennai. The voucher specimen was given the No. PARC/2009/398.

Preparation of Extract: The air dried powdered leaves with leafstalks was loaded into soxhlet apparatus and was subjected to extraction for about 72 hours with petroleum ether (60 - 80 °C), benzene, chloroform and methanol successively. The solvents were distilled off after extraction and the extracts were concentrated under reduced pressure. The extracts were kept in refrigerator until tested⁶. The four extracts were then subjected to phytochemical analysis. For the pharmacological tests, the extracts were dissolved in 1% Tween-80 in normal saline solution to prepare 200 mg/kg concentrations.

Experimental Animals: Wistar albino rats of both the sex weighing 140 - 160g were used for the toxicological studies. The animals were kept in standard conditions of day and night cycles at 22 °C in polypropylene cages. The animals were given on standard pellet water *ad libitum*. The rats were acclimatized to laboratory conditions by housing them in propylene cages prior to the experiments for one week. The experiment was conducted in Institute of Pharmacy and Technology, Salipur, Cuttack and CPCSEA recognized local Ethical Committee approved the protocol bearing No. 19/IAEC-IPT/13.

Carbon Tetrachloride Induced Hepatotoxicity

Model: The screening for hepatoprotective activity against carbon tetrachloride induced hepatotoxicity was followed as per standard procedure⁷⁻⁹. The rats were divided into seven groups with six rats in each. Group I served as normal control and received the vehicle alone (Sterile distilled water, 2 ml p.o.) for 5 days. Group II served as toxic control animals received CCl₄ (2 ml/kg, 1:1 in olive oil, i.p.) on the 3rd and 4th day. Group III, IV, V, VI

received 200 mg/kg of petroleum ether, benzene, chloroform, methanol extracts respectively and Group VII was treated with silymarin (10 mg/kg) for 5 days and on the 3rd and 4th day CCl₄ (2 ml/kg, 1:1 in olive oil, i.p.) was given 1 h after the treatment of the extract. The animals were sacrificed 48 h after the last injection of CCl₄ under mild ether anaesthesia. The blood was collected and allowed to stand for 30 min at 37 °C and centrifuged to separate the serum to estimate various biochemical parameters.

Histopathological Study of Liver and Kidney:

The histopathological study was carried out for liver and kidney tissue. The liver was transferred to 4% formalin solution and processed for histopathological studies following the standard procedure described by¹⁴. The microtome sections were taken and stained with hematoxylin and eosin. The section thus obtained was scanned in Carl-Zeiss microscope with photographic facility and photomicrographs were taken.

Statistical Analysis: The results are expressed as mean ± SD. The difference between experimental groups was compared using one way ANOVA followed by Tukey's multiple comparison column test.

RESULTS AND DISCUSSION: The results observed in pre-treatment of *A. natans* extracts with respect to induction of hepatotoxicity using CCl₄ are given in (Table 1). A marked reduction in total protein levels was observed in the group treated with CCl₄ (5.033 ± 0.307, P < 0.001) when compared to the normal control group. The group treated with CCl₄ significantly increased the level of SGOT, SGPT, ALP, total bilirubin. Rats treated with CCl₄ (toxic control) developed significant liver damage and it was well indicated by elevated levels of specific hepatic enzymes like SGOT (298.01 ± 2.553, P < 0.001), SGPT (482.95 ± 1.742, P < 0.001), ALP (251.98 ± 2.564, P < 0.001) and total bilirubin (3.335±0.254, P < 0.001) in serum.

The groups received the pre-treatment of *A.natans* extracts methanol extract (ANME) and chloroform extract (ANCE) at dose levels of 200 mg/kg body weight significantly controlled the change in the biochemical parameters. The methanol extract (ANME) and chloroform extract (ANCE) at dose levels of 200 mg/kg exhibited significant increase

9.01 ± 0.222 mg/dL, 7.98 ± 0.177 mg/dL, (P < 0.01, P < 0.05) respectively in the serum total protein level as compared to toxic control group and the effect was compared with the standard group 9.61 ± 0.560 mg/dL (P < 0.001) treated with silymarin (Sily-100). The SGOT, SGPT, ALP and total bilirubin level decreased in drug treated groups to significant level. The SGOT 137.60 ± 3.984, 84.28 ± 3.052 (P < 0.001), SGPT 238.25 ± 1.089, 132.18 ± 1.2 (P < 0.001), ALP 204.31 ± 2.053, 139.42 ± 1.1 (P < 0.001) and total bilirubin 1.90 ± 0.143, 1.28 ± 0.2 (P < 0.001) levels decreased significantly in methanol extract (ANME) and standard drug silymarin group as compared to toxic control group.

The SGOT 277.43 ± 3.782 (P < 0.01), SGPT 472.08 ± 2.850 (P < 0.05), ALP 239.68 ± 1.202 (P < 0.05) and total bilirubin 2.03 ± 0.207 (P < 0.01) levels significantly decreased in chloroform extract (ANCE) as compared to toxic control group. Administration of petroleum ether extract (ANPE) and benzene extract (ANBE) did not display effect of increase in the serum enzyme levels as compared to toxic control group.

The result of antioxidant enzymes like LPO, SOD, Catalase and GPx in CCl₄ induced hepatotoxicity are given in (Table 2). The LPO, SOD, Catalase (CAT) and GPx activities in the toxin control group depleted significantly (P < 0.001 respectively) as compared to the normal control group. LPO, SOD, Catalase and GPx activity increased at both methanol extract (ANME), chloroform extract (ANCE) and standard drug silymarin.

LPO 14.48 ± 0.875, 11.23 ± 0.833 (P < 0.001), SOD 30.95 ± 0.688, 31.29 ± 0.819 (P < 0.001), Catalase 48.91 ± 1.195, 55.35 ± 1.826 (P < 0.001) and GPx 35.77 ± 0.62, 39.68 ± 0.51 (P < 0.001) activity increased significantly in methanol extract (ANME) and silymarin respectively as compared to toxic control group. The LPO 21.15 ± 1.100 (P < 0.05), SOD 24.97 ± .0425 (P < 0.05), Catalase 39.21 ± 1.901 (P < 0.05) and GPx 30.50 ± 0.45 (P < 0.05) activity increased significantly in ANCE as compared to toxic control group. Administration of petroleum ether (ANPE) and benzene (ANBE) did not display effect of increase in the LPO, SOD, Catalase and GPx activity as compared to toxic control group.

TABLE 1: EFFECT OF A. NATANS EXTRACTS ON BIOCHEMICAL PARAMETERS IN CCl₄ INDUCED HEPATOTOXICITY IN RATS

Groups	Treatment	Dose	Level of biochemical parameters				
			SGOT (U/L)	SGPT (U/L)	ALP (U/L)	Total Bilirubin (mg/dL)	Total Protein (mg/dL)
Group I	Normal Control	2ml vehicle	49.33 ± 2.454	94.98 ± 2.630	126.95 ± 1.331	0.836 ± 0.147	10.20 ± 0.295
Group II	Toxic Control	2ml vehicle	298.01 ± 2.553 ^d	482.95 ± 1.742 ^d	251.98 ± 2.564 ^d	3.335 ± 0.254 ^d	5.033 ± 0.307 ^d
Group III	ANPE	200 mg/kg	289.53 ± 2.958 ^d	478.13 ± 3.822 ^d	248.23 ± 4.067 ^d	2.730 ± 0.160 ^d	6.30 ± 0.933 ^e
Group IV	ANBE	200 mg/kg	288.01 ± 3.804 ^d	475.15 ± 3.861 ^d	244.95 ± 3.970 ^d	2.80 ± 0.258 ^d	6.45 ± 1.269 ^e
Group V	ANCE	200 mg/kg	277.43 ± 3.782 ^{bd}	472.08 ± 2.850 ^{cd}	239.68 ± 1.202 ^{cd}	2.03 ± 0.207 ^{be}	7.98 ± 0.177 ^c
Group VI	ANME	200 mg/kg	137.60 ± 3.984 ^{ad}	238.25 ± 1.089 ^{ad}	204.31 ± 2.053 ^{ad}	1.90 ± 0.143 ^{af}	9.01 ± 0.222 ^b
Group VII	Silymarin	10 mg/kg	84.28 ± 3.052 ^{ad}	132.18 ± 1.259 ^{ad}	139.42 ± 1.170 ^{ad}	1.28 ± 0.245 ^a	9.61 ± 0.560 ^a

Values are mean ± SEM (n=6) one-way ANOVA followed by Tukey's multiple comparison column test. Where, a=P≤0.001, b=P≤0.01, c=P≤0.05 vs Toxic Control d=P≤0.001, e=P≤0.01, f=P≤0.05 vs Control

TABLE 2: EFFECT OF A. NATANS EXTRACTS ON ANTI-OXIDANT PARAMETERS IN CCl₄ INDUCED HEPATOTOXICITY IN RATS

Groups	Treatment	Dose	Level of antioxidant parameters			
			LPO	SOD	Catalase	GPx
Group I	Normal control	2 ml vehicle	11.05 ± 0.43	35.62 ± 0.792	63.21 ± 0.084	49.11 ±
Group II	Toxic control	2 ml vehicle	25.40 ± 0.912 ^d	20.86 ± 0.705 ^d	31.88 ± 1.183 ^d	23.78 ± 0.53 ^d
Group III	ANPE	200 mg/kg	22.77 ± 0.749 ^d	21.53 ± 0.708 ^d	35.26 ± 1.357 ^d	24.44 ± 0.51 ^d
Group IV	ANBE	200 mg/kg	22.70 ± 0.978 ^d	22.85 ± 1.110 ^d	34.75 ± 2.029 ^d	27.15 ± 0.48 ^d
Group V	ANCE	200 mg/kg	21.15 ± 1.100 ^{cd}	24.97 ± .0425 ^{cd}	39.21 ± 1.901 ^{cd}	30.50 ± 0.45 ^{cd}
Group VI	ANME	200 mg/kg	14.48 ± 0.875 ^a	30.95 ± 0.688 ^a	48.91 ± 1.195 ^{ad}	35.77 ± 0.62 ^{ad}
Group VII	Silymarin	10 mg/kg	11.23 ± 0.833 ^a	31.29 ± 0.819 ^a	55.35 ± 1.826 ^{af}	39.68 ± 0.51 ^{ad}

Values are mean ± SEM (n=6) one-way ANOVA followed by Tukey's multiple comparison column test. Where, a=P≤0.001, b=P≤0.01, c=P≤0.05 vs Toxic Control d=P≤0.001, e=P≤0.01, f=P≤0.05 vs Control

Hepatotoxicity induced by CCl₄ is the most commonly used model system for the screening of hepatoprotective activity of plant extracts/drugs. The total protein and albumin levels decreased due

to the hepatotoxin intoxication. The reduction is attributed to the damage produced and localized in the endoplasmic reticulum which results in the loss of P - 450 leading to its functional failure with a decrease in protein synthesis and accumulation of triglycerides.

In the present study, CCl₄ intoxication reduced the serum total protein and albumin levels. The pre-treatment of methanol extract (ANME) restored the total protein and albumin levels. The rise in protein and albumin level suggests the stabilization of endoplasmic reticulum to protein synthesis¹⁰. The liver marker enzymes (AST, ALT and ALP) are cytoplasmic in nature; upon liver injury these enzymes enter into the circulatory system due to altered permeability of membrane. In this study, significant increase in AST and ALT levels in the serum was observed after administration of CCl₄. ALP level also increased after CCl₄ administration. The increased levels of these enzymes significantly decreased by pretreatment with methanol extract (ANME). Reduction in the levels of AST, ALT and ALP towards the normal value is an indication of stabilization of plasma membrane as well as repair of hepatic tissue damages caused by CCl₄¹¹.

Many studies have demonstrated that the hepatoprotective effect of plant extracts may be related to its antioxidant capacity to scavenge reactive oxygen species. CCl₄ intoxication reduced the total protein level in liver homogenate, which restored significantly with the pre-treatment of methanol extract (ANME). Liver cells have antioxidant defense system consisting of antioxidants named as GSH and antioxidant enzymes mainly catalase and GPx to protect its own cells against oxidative stress condition, which causes destruction of cell elements and finally cell death¹².

GSH is mainly distributed among living cells and is tangled in many biological and cellular functions, substituting in a role as an indispensable intracellular reducing agent for maintenance of intracellular redox status. It is also the most indispensable biomolecule protecting against chemically generated cytotoxicity, by playing a role in the destruction of reactive agents by conjugation and hydroperoxide reduction, or by direct quenching of free radicals. CCl₄ intoxication

slightly reduced the level of GSH, which was significantly restored in methanol extract (ANME) treated (higher dose) rats. Trichloromethyl peroxy radical, the metabolic product of CCl₄ binds covalently to the macromolecules and causes peroxidative degradation of cellular membrane leading to the necrosis of hepatocytes. The hepatic antioxidant enzymatic activity of catalase and GPx significantly decreased in CCl₄ intoxicated rats as compared with control rats.

The declined enzymatic action would result in an increased steady-state level of oxidants, adding to cell injury. The catalase level was increased by administration of methanol extract (ANME) to CCl₄ intoxicated rats suggesting that it has the capacity to restore the enzyme action towards normalizing in CCl₄ injured liver.

However, administration of methanol extract to CCl₄ intoxicated rats had less effect in hepatic GPx activity and relative liver weight as compared to the CCl₄ treated toxin control group. This result suggests that methanol extract (ANME) markedly inhibited CCl₄ induced liver damage by elevated hepatic antioxidant enzymatic system such as catalase and GSH. The rise in marker enzymes level in CCl₄ treated animals has been attributed to damaged structural integrity of the liver¹³.

Administration of the methanol extract (ANME) preserved the structural integrity of the hepatocellular membrane as evidenced from attenuation of the marker enzymes level when compared to CCl₄ treated animals. It was further verified by the histopathological judgement of the liver tissue.

Histopathological Study of Liver in CCl₄ Induced Hepatotoxic Rats: The histopathological feature, as shown in (Fig. 1) indicated the normal liver lobular architecture and cell structure of the liver in the normal control animals. There were no pathological changes observed in normal control animals. In CCl₄ treated animals, Some centrilobular hepatocyte necrosis, microvesicular fatty change and extensive fatty change were observed on the midzonal or entire lobe there was a vacuolar degeneration of hepatocytes around central vein with moderate to severe hepatocyte necrosis due to CCl₄ toxicity (Fig. 2).

The histological observations also supported results obtained from the serum enzyme levels. Histological appearance of rat liver treated with *A. natans* petroleum ether extract (ANPE) and CCl_4 showed extensive necrosis and inflammation around central veins with bridging necrosis (**Fig. 3**) and histological appearance of rat liver treated with *A. natans* benzene extract (ANBE) and CCl_4 showed inflammation and necrosis around central veins, the cellular degeneration was more pronounced with mild necrosis and fibrosis (**Fig. 4**).

Histological appearance of rat liver treated with *A. natans* chloroform extract (ANCE) and CCl_4 showed mild to moderate portal inflammation. It displayed developmental progress with

receding of fatty changes and necrosis (**Fig. 5**) and histological appearance of rat liver treated with *A. natans* methanol extract (ANME) and CCl_4 revealed tremendous progress with the disappearance of fatty deposition and necrosis. It showed only mild portal inflammation in methanol extract (ANME) treated animals, with lesser vacuolar degeneration and hepatic necrosis (**Fig. 6**). Similar changes were also observed in the silymarin treated animals.

Histological appearance of liver tissue of rats treated with CCl_4 and silymarin sections showed good recovery with absence of necrosis and fatty depositions. The central vein has minimal portal inflammation (**Fig. 7**).

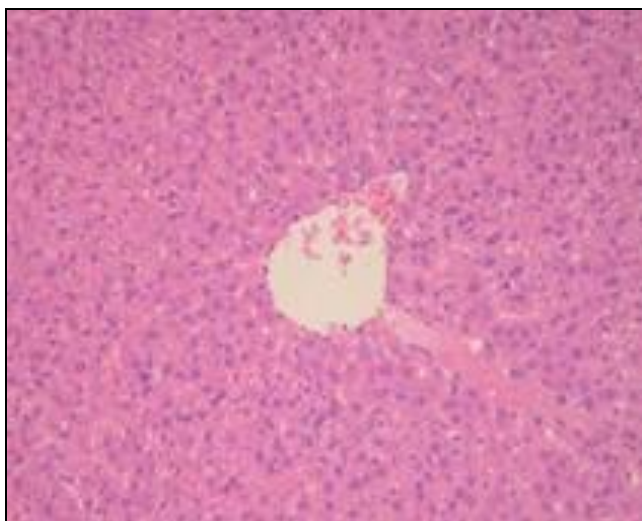


FIG. 1: HISTOLOGICAL APPEARANCE OF NORMAL RAT LIVER

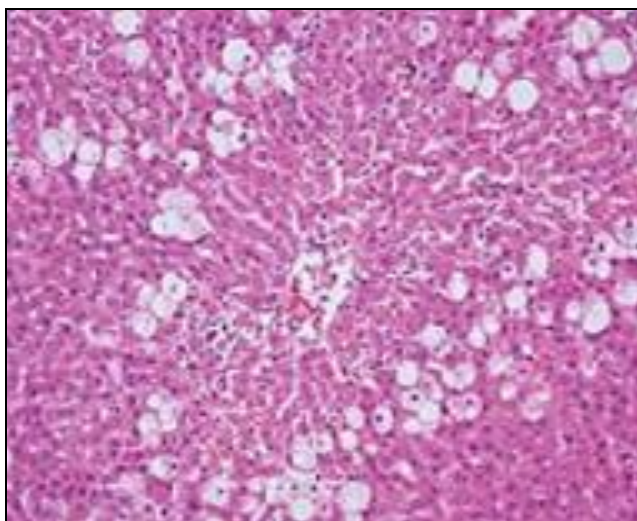


FIG. 2: HISTOLOGICAL APPEARANCE OF RAT LIVER TREATED WITH CCl_4

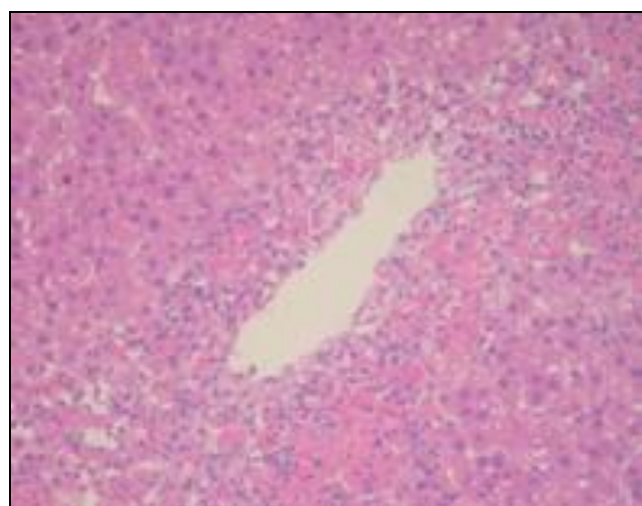


FIG. 3: HISTOLOGICAL APPEARANCE OF RAT LIVER TREATED WITH *A. NATANS* PETROLIUM ETHER EXTRACT AND CCl_4

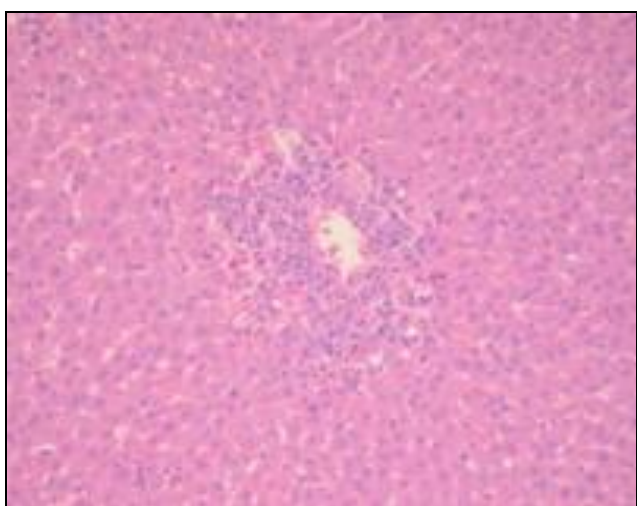


FIG. 4: HISTOLOGICAL APPEARANCE OF RAT LIVER TREATED WITH *A. NATANS* BENZENE EXTRACT AND CCl_4

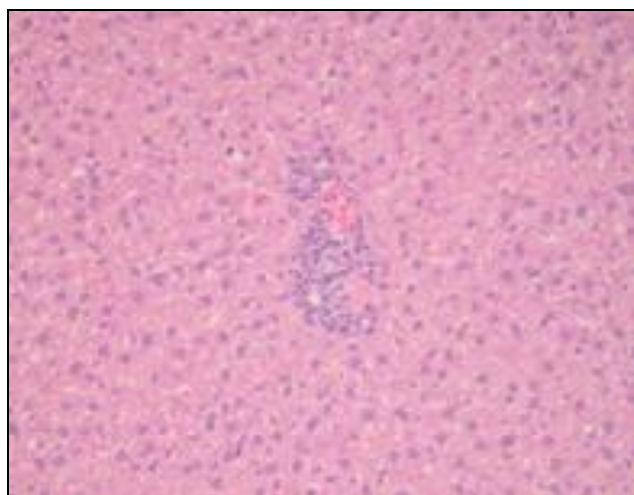


FIG. 5: HISTOLOGICAL APPEARANCE OF RAT LIVER TREATED WITH A. NATANS CHLOROFORM EXTRACT AND CCl₄

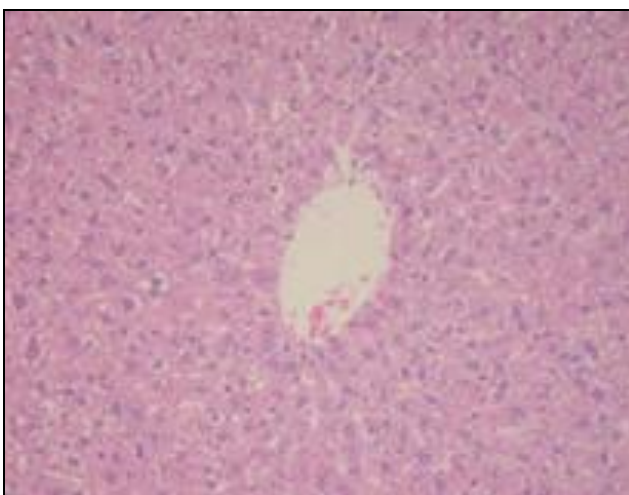


FIG. 6: HISTOLOGICAL APPEARANCE OF RAT LIVER TREATED WITH A. NATANS METHANOL EXTRACT AND CCl₄

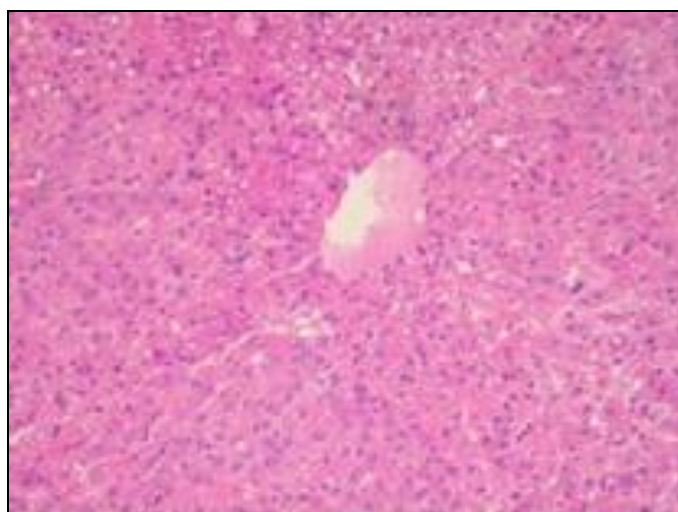


FIG. 7: HISTOLOGICAL APPEARANCE OF RAT LIVER TREATED WITH SILYMARIN AND CCl₄

CONCLUSION: The results obtained from CCl₄ induced hepatotoxic model in rats indicated that after treatment of toxicants there was significant rise in SGOT, SGPT, ALP and total bilirubin and total protein levels. Treatment with *A. natans* methanol extract increased the level of SGOT, SGPT, ALP, total bilirubin and total protein activities when compared to toxic control. In case of antioxidant enzyme, *A. natans* methanol extract was observed that it significantly increase the level of SOD, CAT and GPX. The level of LPO, which was elevated due to hepato toxicant, was brought to normal. Histopathological evidences further supported the above results. In conclusion, the results of this study demonstrated that *A. natans methanol* extract is effective for the prevention of CCl₄ induced hepatic damage in rats and therefore

it could be used as a hepatoprotective agent. The protective effects against liver damage may be attributed due to the free radical scavenging effect, inhibition of lipid peroxidation, and increased antioxidant activity¹⁵.

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CONFLICT OF INTEREST: Nil

REFERENCES:

1. Eldin HS, Gadir HA, Hassan and WA: Evaluation of the hepatoprotective activity of *Fagonia cretica* Linn. Journal of Pharmacognosy and Phytochemistry 2015; 3(3): 1-6.

2. Khan MA, Gupta A, Kumar S, Ahmad S, and Sastry JLN: Hepatoprotective activity of a new polyherbal formulation against paracetamol and D-galactosamine induced hepatic toxicity. *Journal of Pharmacy and Bioallied Sciences* 2015; 7(4): 246-249.
3. Britto JD and Mahesh R: Exploration of kani tribal botanical knowledge in agasthiyamalai biosphere reserve - South India. *Ethnobotanical Leaflets* 2007; 1: 1-10.
4. Jeyaprakash K, Ayyanar M, Geetha KN and Sekar T: Traditional uses of medicinal plants among the tribal people in Theni District (Western Ghats), Southern India. *Asian Pacific Journal of Tropical Biomedicine* 2011; 1: S20-S25.
5. http://www.bdu.ac.in/schools/biotechnology/industrial_biotechnology/sekardb/pdf/medicine/6.pdf
6. http://agritech.tnau.ac.in/horticulture/extraction_technique%20medicinal_plants.pdf
7. Shrivastava S and Gilhotra R: Hepatoprotective potential of polyherbal preparation against CCl₄ induced liver toxicity in rats. *International Journal of Pharmaceutical Sciences and Research* 2017; 8(3): 1498-1503.
8. Adewale OB, Adekeye AO, Akintayo CO, Onikanni A and Saheed S: Carbon tetrachloride (CCl₄) induced hepatic damage in experimental Sprague Dawley rats: Antioxidant potential of *Xylopiya aethiopicum*. *The Journal of Phytomedicine* 2014; 3(2): 118-123.
9. Ubhenin, AE, Igbe I, Adamude FA and Falodun A: Hepatoprotective effects of ethanol extract of *Caesalpinia bonduc* against Carbon tetrachloride induced hepatotoxicity in albino rats. *Journal of Applied Sciences and Environmental Management* 2016; 20 (2): 396-400.
10. Balogun FO and Ashafa AOT: Antioxidant and hepatoprotective activities of *Dicoma anomala* Sond. aqueous root extract against carbon tetrachloride induced liver damage in wistar rats. *Journal of Traditional Chinese Medicine* 2016; 36(4): 504-513.
11. Hashemi JM: *Hibiscus sabdariffa* calyx extract alleviate hepatotoxicity induced by carbon tetrachloride on male albino rats. *Nature and Science* 2014; 12(6): 111-120.
12. Wang J, Zhang Y, Liu R, Li X, Cui Y and Qu L: Geniposide protects against acute alcohol-induced liver injury in mice *via* up-regulating the expression of the main antioxidant enzymes. *Canadian Journal of Physiology Pharmacology* 2015; 93(4): 261-267
13. Juma KK, Joseph JNN and David MN: A review of the biochemical, hematological and histological modulations in acetaminophen induced hepatotoxicity and the potential of *Urtica Dioica* in the regeneration of the liver. *The Journal of Drug Metabolism and Toxicology* 2015; 6 (3): 1-7
14. Sadeghi Z, Akaberi A, Valizadeh J: *Otostegia persica* (Lamiaceae): A review on its ethnopharmacology, phytochemistry and pharmacology. *Avicenna Journal of Phytomedicine* 2014; 4: 79-88.
15. Siddiqui SZ, Ali S, Rehman A, Rubab K, Abbasi MA, Ajaib M and Z Rasool ZG: *Pyrus pashia*: A persuasive source of natural antioxidants. *Pakistan Journal of Pharmaceutical Sciences* 2015; 28(5):1763-72.

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