(Research Article)

E-ISSN: 0975-8232; P-ISSN: 2320-5148



PHARMACEUTICAL SCIENCES



Received on 05 October, 2017; received in revised form, 10 June, 2018; accepted, 12 June, 2018; published 01 July, 2018

EFFECT OF METHYL PARATHION (AN ORGANOPHOSPHATE) ON BIOCHEMICAL CONTENTS OF FRESH WATER CAT FISH HETEROPNEUSTES FOSSILIS (BLOCH)

T. Bheem Rao, K. Thirupathi and Y. Venkaiah *

Department of Zoology, Kakatiya University, Waranagal - 506009, Telangana, India.

Keywords:

H. fossilis,
Proteins, Carbohydrates,
Ninhydrine positive substances,
Methyl parathion, Organophosphate

Correspondence to Author: Dr. Venkaiah Yanamala

Assistant Professor, Department of Zoology, Kakatiya University, Waranagal -506009, Telangana, India.

E-mail: venkaiahyanamala07@gmail.com

ABSTRACT: The present work was planned to study the effect of methyl parathion an organophosphate compound (OP) on biochemical contents of fresh water cat fish *Heteropneustis fossilis*. The exposure of fish to the sublethal concentrations of the toxicant methyl parathion was investigated and the variations were observed in biochemical contents in different tissues of the fish *i.e.* gill, liver, intestine, muscle and brain. The quantitative variations were observed in proteins, carbohydrates and ninhydrine positive substances at different time intervals i.e. 24, 48, 72 and 96 h. The results revealed that the components of proteins, carbohydrates and ninhydrine positive substances were found to be decreased significantly at different time intervals of methyl parathion exposure to different tissues of fish compared to control. The maximum decrease in proteins followed by ninhydrine positive substances (free amino acids) and carbohydrates was observed at 72 h and 96 h compared to 24 h and 48 h in different tissues of the fish H. fossils. Thus, our present investigation reports that the changes observed were depending on period of exposure of fish to the concentration of methyl parathion.

INTRODUCTION: Pesticides are used worldwide in aquaculture and agriculture to control the insects, pests and other vectors ¹, which ultimately find their way into aquatic habitats like rivers, lakes and ponds. The environmental quality is determined by the assessing to toxicity of different chemicals to fish and other aquatic organisms. They ultimately enter the organisms through food webs and also through contact in water ^{2, 3}. Most of the chemicals used as pesticides are acutely toxic to many nontarget organisms such as invertebrates, birds, mammals and fishes especially those inhibiting the marine environment ^{4, 5, 6}.



DOI:

10.13040/IJPSR.0975-8232.9(7).2869-74

Article can be accessed online on: www.ijpsr.com

DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.9(7).2869-74

Some of the pesticides have been reported to persists the environment and tend to bio-accumulation in organisms ⁷. It has been reported that pesticides can be actively toxic to fish ^{8, 9, 10, 11, 12, 13, 14}. Pesticide toxicity to fish has been investigated in several studies ^{15, 16}. Usage of pesticides in the ecosystem leads to development of various types of morphological, physiological, biochemical and behavioral changes in individual ¹⁷.

Hence it is necessary to study the immediate effect of pesticides on fish which forms a part of human diet. Among these pesticides, organophosphate compounds (OP's) are commonly used insecticides, which maintain less toxicity, persistence and also rapid biodegradability in the environment ¹⁸. Methyl Parathion (0, 0 -di methyl-0-4 nitro phenyl phosphoro thioate)-Bayer (Germany) is a synthetic Organophosporous pesticide, known to be toxic to fish ¹⁹ and insects ²⁰ and applied abundantly in agriculture ²¹.

Fishes are very sensitive to a variety of toxicants in water, which uptake and accumulate many toxicants such as pesticides ^{22, 23}. However pesticide exposure causes severe alternations in the tissue biochemistry of fishes. In general the toxic effects will be more when two (or) more toxicants act together in a synergistic manner ^{24, 25, 26, 27, 28}. The present study has been carried out to investigate the effect of methyl parathion (an Organophosphate) on biochemical contents of fresh water cat fish *Heteropneustes fossilis* (Bloch).

MATERIALS AND METHODS: In the present study, fresh water fish *H. fossilis* was exposed to different concentrations of an organophosphate pesticide methyl parathion and the variations in biochemical constituents of the fish were studied. The technical grade pesticide methyl parathion 50% E.C was taken for this present study.

The fresh water fish *H. fossilis* (ranging in weight 70 to 100 gm and in length 25 cm to 35 cm) were brought from local fresh water tanks located within the radius of 15 km from the Kakatiya University, Warangal, Telangana State, India to the laboratory in well aerated polythene bag and acclimatized to the ambient room temperature $(28 \pm 2 \, ^{\circ}\text{C})$ in large plastic containers. During the period of acclimatization they were fed with oil cake mixed with rice flour. The period of acclimatization lasted for 15 days.

The healthy fishes were grouped into five batches containing six and each were exposed to different concentrations of pesticide methyl parathion to calculate the medium lethal concentration LC_{50} value using Probit analysis method ²⁹. The methyl parathion was dissolved in ethanol and diluted with water to the required concentrations. The fishes (five groups) were exposed to the sub lethal concentration (0.1g - 0.5g) of methyl parathion for 24, 48, 72 and 96 h respectively. Another group was maintained as control without pesticide.

At the end of each exposure period, fishes were scarified and the tissues such as gill, liver, intestine, muscle and brain were dissected out and stored on ice-jacketed container for biochemical studies. The tissues were weighed to the nearest milligram and processed for further analysis.

The tissues were homogenized (10%) in 10% Tri Chloro Acetic Acid (TCA) centrifuged at 2000 rpm for 15 min and clear supernatant and sediment was used for the analysis of total proteins, carbohydrates and ninhydrine positive substances (FAA). The protein sediment and supernatant (TCA precipitated and soluble proteins) was dissolved in 1N NaOH and protein content was determined through the method 30 described by Schacterle and Pollack (1973) 31. Ninhydrine positive substances were estimated by the method of Lee and Takahashi (1966) 32 and the total carbohydrate content in the tissues were estimated by the method of Carroll *et al.*, $(1956)^{33}$.

Statistical Analysis: Statistical analysis was performed by one-way analysis of variance ANOVA to compare the results between the tissue components.

RESULTS: The results obtained from the quantitative estimates on the effect of Methyl parathion on biochemical contents of various tissues of fresh water cat fish *H. fossilis* are presented in **Table 1, 2, 3, 4** and **Fig. 1, 2, 3, 4** respectively. In this study when the fish tissues *i.e.* gill, liver, intestine, muscle and brain after treatment with different concentrations of the test chemical methyl parathion at different time intervals a drastic reduction was observed total biochemical content of different tissues of *H. fossilis* compared to control. The results presented shows that the protein content was significantly decreased in the gill of fish exposed to methyl parathion (**Table 1, 2** and **Fig. 1, 2**).

TABLE 1: TCA SOLUBLE PROTEINS IN VARIOUS TISSUES OF H. FOSSILIS EXPOSED TO VARYING PERIODS OF SUB-LETHAL CONCENTRATION OF METHYL PARATHION

1 EXIODS OF SUB-LETHAL CONCENTRATION OF WEITHTL TAXATIHON						
Dose / Tissue	Control	24 h	48 h	72 h	96 h	
Gill	4.99 ± 0.22	$4.368 \pm 0.26**$	$3.10 \pm 0.26***$	$1.83 \pm 0.48**$	0.88 ± 0.50	
Liver	6.10 ± 0.22	4.22 ± 0.45	3.55 ± 0.45	$2.05 \pm 0.50**$	1.05 ± 0.45	
Intestine	3.42 ± 0.17	1.65 ± 0.33	1.33 ± 0.34	$0.83 \pm 0.82**$	0.66 ± 0.38	
Muscle	4.49 ± 0.11	4.05 ± 0.28	3.50 ± 0.28	$1.44 \pm 0.54**$	0.88 ± 0.41	
Brain	2.49 ± 0.21	1.94 ± 0.33	1.21 ± 0.33	0.71 ± 0.24	$0.33 \pm 0.15**$	

The values are expressed as mean $mg/g \pm SE$ of wet weight of tissue; n=6; statistically significant value to respective control value *P < 0.001, **P < 0.005

TABLE 2: TCA PRECIPITATED PROTEINS IN VARIOUS TISSUES OF *H. FOSSILIS* EXPOSED TO VARYING PERIODS OF SUB LETHAL CONCENTRATION OF METHYL PARATHION

Dose / Tissue	Control	24 h	48 h	72 h	96 h
Gill	16.11 ± 0.25	$13.38 \pm 0.98**$	$9.77 \pm 0.98**$	$7.72 \pm 0.55**$	4.05 ± 0.31 *
Liver	14.38 ± 0.31	$11.05 \pm 0.28*$	$7.82 \pm 0.28*$	6.05 ± 0.85	$3.32 \pm 0.90**$
Intestine	11.21 ± 0.59	10.99 ± 0.61 **	$8.99 \pm 0.61**$	$7.94 \pm 0.39**$	$6.05 \pm 0.72**$
Muscle	10.66 ± 0.52	9.38 ± 0.52	8.16 ± 0.52	$6.16 \pm 0.73**$	$3.94 \pm 0.14*$
Brain	6.49 ± 0.37	$5.38 \pm 0.42**$	$3.99 \pm 0.42**$	$2.44 \pm 0.40**$	$0.88 \pm 0.33***$

The values are expressed as mean $mg/g \pm SE$ of wet weight of tissue; n=6; statistically significant value to respective control value *P < 0.001, **P < 0.05, ***P < 0.005

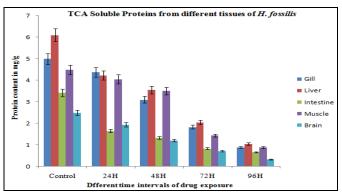


FIG. 1: TCA SOLUBLE PROTEINS IN VARIOUS TISSUES OF *H. FOSSILIS* AND THE VALUES ARE EXPRESSED AS MEAN ± SE mg/gm OF WET WEIGHT OF TISSUE

It was observed that the TCA soluble proteins were decreased and is more pronounced in the gill at 48, 72, 96 h exposure gill, liver, intestine, muscle and brain estimated greater reduction in TCA soluble proteins (**Table 1** and **Fig. 1**). Whereas in TCA precipitated protein contents were significantly decreased at 24, 48, 72 and 96 h of exposure of methyl parathion (**Table 2** and **Fig. 2**).

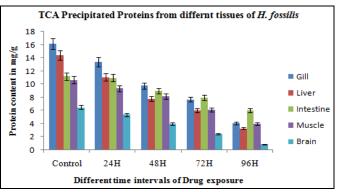


FIG. 2: TCA PRECIPITATED PROTEINS IN VARIOUS TISSUES OF *H. FOSSILIS* AND THE VALUES ARE EXPRESSED AS MEAN ± SE mg/gm OF WET WEIGHT OF TISSUE

While the Carbohydrates were more in brain compared to other tissues in control and also highly decreased in brain and liver at 96 h (**Table 3** and **Fig. 3**). The Free amino acids were found be highest in control muscle and gradually decreased in brain (**Table 4** and **Fig. 4**) from 24 h, 48 h, 72 h, 96 h.

TABLE 3: CARBOHYDRATES CONTENT IN VARIOUS TISSUES OF *H. FOSSILIS* EXPOSED TO VARYING PERIODS OF SUB LETHAL CONCENTRATION OF METHYL PARATHION

Dose / Tissue	Control	24 h	48 h	72 h	96 h
Gill	3.60 ± 0.39	$3.47 \pm 0.45**$	1.94 ± 0.54 **	$2.08 \pm 0.25**$	1.24 ± 0.30
Liver	6.38 ± 0.59	$3.88 \pm 0.58***$	1.24 ± 0.581	1.38 ± 0.13	1.10 ± 0.27
Intestine	4.85 ± 0.41	3.46 ± 0.56	2.63 ± 0.561	1.52 ± 0.45	0.96 ± 0.37
Muscle	2.36 ± 0.55	1.80 ± 0.28	1.38 ± 0.29	0.96 ± 0.35	0.83 ± 0.13
Brain	7.22 ± 0.99	$4.30 \pm 0.87**$	$1.52 \pm 0.87**$	1.10 ± 0.46	0.83 ± 0.17

The values are expressed as mean $mg/g \pm SE$ of wet weight of tissue; n=6; statistically significant value to respective control value *P < 0.001, **P < 0.005, ***P < 0.005

TABLE 4: FREE AMINO ACIDS/NINHYDRINE POSITIVE SUBSTANCES IN VARIOUS TISSUES OF *H. FOSSILIS* EXPOSED TO VARYING PERIODS OF SUB LETHAL CONCENTRATION OF METHYL PARATHION

Tissue / Dose	Control	24 h	48 h	72 h	96 h
Gill	3.09 ± 0.39	2.66 ± 0.35	1.83 ± 0.36	1.27 ± 0.35	$0.60 \pm 0.22**$
Liver	2.88 ± 0.44	$2.49 \pm 0.18**$	$1.94 \pm 0.187**$	1.38 ± 0.26	$0.83 \pm 0.18**$
Intestine	2.66 ± 0.35	2.71 ± 0.34	1.99 ± 0.35	1.38 ± 0.38	0.71 ± 0.25
Muscle	3.83 ± 0.39	3.83 ± 0.50	3.05 ± 0.50	$1.86 \pm 0.52**$	$0.44 \pm 0.33***$
Brain	2.49 ± 0.16	2.16 ± 0.17	1.83 ± 0.17	1.66 ± 0.32	$0.38 \pm 0.26***$

The values are expressed as mean $mg/g \pm SE$ of wet weight of tissue; n=6; Statistically significant value to respective control value *P < 0.001, **P < 0.005, ***P < 0.005

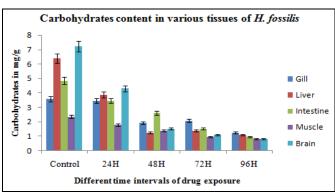


FIG. 3: CARBOHYDRATE CONTENT IN VARIOUS TISSUES OF *H. FOSSILIS* AND THE VALUES ARE EXPRESSED AS MEAN ± SE mg/gm OF WET WEIGHT OF TISSUE

DISCUSSION: Organophosphates are non-persistent in nature and their degradation is rapid ³⁴. These are short lived and highly toxic when compared to a number of other pesticides for example organophosphate pesticide is found to be more toxic to fish than organochlorine ^{35, 36, 37}. Therefore, organophosphates can affect the organisms exposed even for a very short period.

The degree of toxicity produced by the poisonous substances is dose dependent upon environmental conditions such as temperature, pH, O_2 concentration and presence of residual molecules ³⁸. Due to Methyl parathion in toxification, the normal functioning of cells with the resultant the fundamental alterations in biochemical mechanisms in test fish was noticed. Similar results were also noticed in freshwater Labeo rohita upon chronic exposure to the pesticide methyl parathion ³⁹. In our present investigation, the effect of methyl parathion on biochemical constituents of fresh water fish H. fossilis showed a considerable variation in different tissues of fish H. fossilis 40. Findings were correlated on pesticide treated mollusks.

In the present study methyl parathion caused reduction in the total structural and soluble protein content in various tissues of *H. fossilis i.e.*, gill, liver, intestine, and muscle brain were found to be declined during the exposure period of 24, 48, 72 and 96 h. The studies on Methyl parathion toxicity cause metabolic dysfunction in fish 41, 42, 43. Carbohydrates are less sensitive as compared to proteins towards the OP compounds. The results of the present findings showed a significant decrease in carbohydrate content in all the tissues.

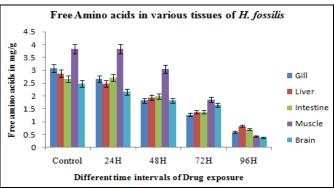


FIG. 4: NINHYDRINE POSITIVE SUBSTANCES IN VARIOUS TISSUES OF *H. FOSSILIS* AND THE VALUES ARE EXPRESSED AS MEAN ± SE mg/gm OF WET WEIGHT OF TISSUE

The decrease in carbohydrates and ninhydrine positive substances (Free amino acids) was observed in various tissues of *H. fossilis* compared to control. The decrease in carbohydrate content is significant and may result in impairment of carbohydrate metabolism due to toxic effects 44, 45, 46

CONCLUSION: From the present study it is concluded that the Methyl parathion exposure has a strong potential to alter the biochemical constituents in various tissues of *H. fossilis*. Further research has to be focused to evaluate the effect of alternative pesticides and related chemicals to reduce the adverse effects on physiological and biochemical aspects of fish. Therefore, the use of pesticides in the field may be a threat to a human, fauna and flora of the environment.

ACKNOWLEDGEMENT: The author expresses their deep gratitude to the Head of the Department of Zoology, Kakatiya University for providing logistic Support.

CONFLICT OF INTEREST: The authors declare that there is no conflict of interest that would prejudice the impartiality of this scientific work.

REFERENCES:

- 1. Yonar EM, Yonar SM, Sener M and Silici US and dusukcan M: Protective role of propolis in chlorpyrifosinduced changes in the haemotological parameters and oxidative antioxidative status of *Cyprinus carpio*. Food Chem. Toxicol. 2012; 50: 2703-2708.
- Pritchard JB: Environmental Health Perspectives Journal Article Aquatic Toxicology: Past, Present, and Prospects, 1993; 100: 249-257.
- 3. Tak AM, Bhat FA, Jan U and Shah M: Changes in carbohydrate metabolism in *Cyprinus carpio* Var.

- communis during short term exposure to dimethoate. RJPBCS. March April 2014; 5(2): 14-66.
- 4. Suryavanshi U, Sreepada RA, Ansari ZA, Nigam S and Badesab S: A study on biochemical changesin the penaeid shrimp, *Metapenaeus monoceros* (Fabricius) following exposure to sublethal doses of organochlorine pesticide (endosulfan). Chemosphere. 2009; 77: 1540-1550.
- Shoaib N, Siddiqui PJA and Ali A: Acute toxic effects of organophosphate pesticides on Killi fish (*Aphanius dispar*) juveniles. Pak. J. Zool. 2012; 44(2): 569-572.
- Selvakumar S, Geraldine P, Shanji S and Jayakumar T: Stress or-specific induction of heat shock protein 70 in the freshwater prawn *Macrobrachium malcolmsonii* (H. Milne Edwards) exposed to the pesticides endosulfan and carbaryl. Pestic.Biochem. Physiol. 2005; 82: 125-132.
- Jayashree R and Vasudevan N: Persistence and distribution of endosulfan under field condition. Environ. Monit. Assess. 2007; 131: 475-487.
- Gurusamy K and Ramadoss V: Impact of DDT on oxygen consumption and opercular activity of *Lepidocephalichthys thermalis*. J. Ecotoxicol. Environ. Monit 2000; 10(4): 239-248.
- Moore A and Waring CP: The effect of a synthetic pyrethroid pesticide on some aspect of reproduction in Atlantic salmon (salmosalar). Aquat. Toxicol., 2001; 52: 1-12
- Prasad M, Bandyopadhyay K, Ajit and Aditya: Xenobiotic impact on sensitivity in *Anabas testudineus* (Bloch). J. Ecobiol., 2002; 14(2): 117-124.
- 11. Srivastava S, Sudha S and Keerty S: Effect of carbaryl on glucose content in the brain of *Heteropneustes fossilis*. J. Ecotoxicol. Environ. Monit., 2002; 12(3): 205-208.
- 12. Eder KJ, Leutenegger CM, Wilson BW and Werner I: Molecular and cellular biomarker responses to pesticide exposure in juvenile chinook salmon (*Oncorhynchus tshawytscha*). M. Environ. Res., 2004; 58(2): 809-813.
- Sheikh N and Yeragi SG: Effect of Rogor 30E (Organophosphate) on muscle protein in the fresh water fish Lepidocephale cthyesthermalis. J. Ecotoxicol. Environ. Monit 2004; 14(3): 233-235.
- Visvanathan P, Maruthanayagam C and Govindaraju M: Effect of malathion and endosulfan on biochemical changes in Channa punctatus. J. Ecotoxicol. Environ. Monit. 2009; 19(3): 251-257.
- Sarkar B, Chatterjee A, Adhikari S and Ayyappan S: Carbofuran and Cypermethrin induced histopathological alterations in the liver of *Labeo rohita* (Ham.) and its recovery. Journal of Applied Ichthyology 2005; 21: 131-135.
- Shoaib N, Ahmed P and Siddiqui: Impact of organophosphate pesticides, methyl parathion and chloropyrifoson some tissue enzymes in fish (*Ahpanius dispur*). Indian Journal of Geo-Marine Sciences 2016; 45(7): 869-874.
- 17. Jeyapriya SP, Venkatesh P and Suresh N: Acute and subchronic effect of monocrotophos on haematological indices in *Catla catla* (Hamilton). International Journal of Pure and Applied Zoology 2013; 1(3): 235-240.
- Singh RN, Pandey PK, Singh NN and Dass VK: Acute toxicity and behavioral responses of common carp *Cyprinus carprio* (Linn.) to an Organophosphate (Dimethoate). World Journal of Zoology 2010; 5(3): 183-188.
- Macek KJ and Mc Allester WJ: Insecticide Susceptibility of some common fish family Representatives. Trans. Am. Fish. Soc., 1970; 99: 20-27.

- Croft BA and Brown AW: Responses of arthropod natural enemies. Annual Review of Entomology 1975; 2: 285-335.
- Von Rumker R, Lawless EW and Meiners AF: Production, distribution, use, and environmental impact potential of selected pesticides. Office of Pesticide Programs, U.S. Environmental Protection Agency, Washington, D.C. 1974; 439.
- Herger W, Jung SJ and Peter H: Acute and prolonged toxicity to aquatic organisms of new and existing chemicals and pesticides. Chemosphere 1995; 31(2): 2707-2726
- 23. Bhuvaneshwari R, Mamtha N, Selvam P and Rajendran RB: Bioaccumulation of metals in muscle, liver and gills of six commercial fish species at Anaikarai dam of River Kaveri, South India. International Journal of Applied Biology and Pharmaceutical Technology 2012; 03: 08-14.
- 24. Kumar S and Singh M: Toxicity of dimethoate to a freshwater teleost *Catla catla*. Journal of Experimental Zoology, 2000; 31: 83-88.
- Tilak KS, Satyavardhan K and Thathaji PB: Biochemical changes induced by fenvalerate in the fresh water fish Channa punctatus. J. Ecotoxicol. Environ. Monit. 2003; 13: 261-270.
- Mathivanan R: Effects of sublethal concentration of quinalphos on selected respiratory and Biochemical parameters in the freshwater fish, *Oreochromis* mossambicus. J. Ecotoxicol. Environ. Moni., 2004; 14: 57-64
- 27. Shrivastava S and Singh S: Changes in protein content in the muscle of *Hetropneustes fossilis* exposed to carbaryl. J. Ecotoxicol. Environ. Monit. 2004; 14: 119-122.
- 28. Sujatha LB: Studies on the physiology, haematology and histopathology in the Indian Major Carp, *Catla catla* (Hamilton), as influenced by individual and synergistic toxic effects of a pesticide and two metallic compounds. Ph.D. thesis, University of Madra 2006
- Finney DJ: Probit Analysis, 3rd edition, London: Cambridge University Press 1971; 20.
- Lowry OH, Rosenbrough NJ and Randall RJ: Protein measurements with folin-phenol reagent. J. Biol. Chem., 1951; 193: 265-267.
- Schacterle GR and Pollack RL: A simplified method for the quantitative assay of small amounts of protein in biologic material. Analytical Biochemistry 1973; 51: 646-655
- 32. Lee YP and Takahashi T: An improved colorimetric determination of amino acids with the use of ninhydrin. Analytical Biochemistry 1966; 14: 71-77.
- Carroll NV, Longley RW and Roe JH: The determination of glycogen in liver and muscle by use of Anthrone Reagent. Journal of Biological Chemistry 1956; 220: 583-593.
- 34. Gul-e-Zehra N and Nafisa S: Acute Toxicity of Organophosphate Pesticides on Juveniles of the Marine Fish (*Oreochromis mossambicus*). INT. J. BIOL. BIOTECH. 2016; 13(3): 393-397.
- 35. Veeraiah KS: Toxicity and effect of chlorpyrifos to the fresh water fish *Labeo rohita*. Tilak Poll. Res. 2001; 20(3): 443-445.
- Deb N and Das S: Chlorpyrifos Toxicity in Fish: A Review. Current World Environment 2013; 8(1): 77-84.
- 37. Tilak KS, Veeraiah K and Kumari RGV: Toxicity and effect of chloropyriphos to the fresh water fish *Labeo rohita* (Hamilton). Neurol Research 2001; 20: 438-445.
- 38. Satyavardhan K: A Comparative Toxicity Evaluation and behavioral observations of fresh water fishes to

E-ISSN: 0975-8232; P-ISSN: 2320-5148

- Fenvalerate. Middle-East Journal of Scientific Research 2013; 13(2): 133-136.
- 39. Thenmozhi C, Vignesh V, Thirumurugan R and Arun S: Impact of Malathion on Mortality and Biochemical changes of fresh water fish *Labeo rohita*. Iran J. Environ. Health Sci. Eng. 2011; 8: 325-332.
- Kabeer AI, Begum MD, Sivaiah S and Ramanarao KV: Sublethal toxicity of malathion on the proteases and free amino acid composition in the liver of Teleost, Tilapia mosambica (peters). Toxicol Lett. PMID: 6364457, 1984; 20(1): 59-62.
- 41. Tilak KS and Rao DK: Chlorpyrifos toxicity of freshwater fish. J.Aqua. Biol., 2003; 8(2): 161-166.
- 42. Venkatramana Sandhya GV, Rani PN and Moorthy PS: Impact of Malathion on the biochemical parameters of gobiid fish, *Glossogobius giuris* (Ham). J. Environ. Biol., 2006; 27(1): 119-122.

- 43. Pathak L, Singh DP and Kumar S: Effect of Malathion on haematological parameters of a fresh water Fish, *Labeo rohita*. Bionotes, 2009; 11: 68.
- 44. Thenmozhi C, Vignesh V, Thirumurugan R and Arun S: Impacts of malathion on mortality and biochemical changes of freshwater fish *Labeo rohita*. Iran. J. Environ. Health Sci. Eng. 2010; 8(4): 189-198.
- 45. Logaswamy S and Remia KM: Impact of Cypermethrin and Ekalux on respiratory and some biochemical activities of a fresh water fish, *Tilapia mossambica*. Curr. Biot., 2009; 3(1): 65-73.
- Susan AT, Shoba K, Veeraiah K and Tilak KS: Studies on biochemical changes in the tissues of *Labeo rohita* and *Cirrhinus mrigala* exposed to fenvalerate technical grade. J. Toxicol. Envt. Health Sci. 2010; 2(5): 53-62.

How to cite this article:

Rao TB, Thirupathi K and Venkaiah Y: Effect of methyl parathion (an organophosphate) on biochemical contents of fresh water cat fish *Heteropneustes fossilis* (bloch). Int J Pharm Sci & Res 2018; 9(7): 2869-74. doi: 10.13040/JJPSR.0975-8232.9(7).2869-74.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)