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CHARACTERIZATION AND BIOLOGICAL EVALUATION OF CRUDE POLYSACCHARIDE FROM *VITEX NEGUNDO* LINN. LEAVES

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ABSTRACT: Naturally occurring non-starch polysaccharides are known for their therapeutic value. This paper describes isolation, characterization and evaluation of glucose lowering potential of one such plant polysaccharide isolated from the leaves of the plant Vitex negundo. It was isolated and characterized using various analytical techniques and biochemical tests which confirmed the presence of carbohydrates and elements such as C, H and O. It demonstrated a reduction in glucose concentration in-vitro, however non-significant in comparison to standards. In-vivo, it was given per orally in fasting male C57 mice at 10, 30 and 100 mg/kg single dose post 30 min of which serum glucose was recorded. Glucose load was given 30 min after administering crude polysaccharide after which serum glucose concentrations were estimated at 15, 30, 60 and 120 min. Obtained results suggest a non-significant reduction in AUC as well as individual glucose levels at all the time points in crude polysaccharide treated animals in comparison to vehicle control in fasting as well as glucose loaded conditions. Thus, the crude polysaccharide isolated from the leaves of V. negundo doesn't have a significant antihyperglycemic activity in single dose when given via per oral route in normal fasting as well as glucose loaded mice. It may have antihyperglycemic action on repeated dosing in glucose loaded diabetic animals, which needs further investigation. However, this paper is first to study anti-hyperglycemic potential of crude polysaccharide from leaves of V. negundo.

INTRODUCTION: Many phytoconstituents have been known for their therapeutic potential *in-vitro* as well as *in-vivo*. Plant polysaccharides are one of them which have been reported for their anti-inflammatory, anti-cancer, anti-hyperglycemic and anti-oxidant properties in various experimental models ^{1, 2, 3}.

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Vitex negundo is a plant from Verbanaceae family which is a woody aromatic shrub growing to a small tree having tri or penta-foliate leaves with quadrangular branches and bluish-purple flowers ⁴, ⁵. Entire plant has been reported in different systems of medicines for various activities however, seeds, roots, leaves and bark are mainly used. Traditionally, *V. negundo* has been used as anti-oxidant, analgesic, anti-inflammatory, as a vermifuge and in arthritis ⁴ and leaves are reported for antibacterial, antitumor, astringent, febrifuge, sedative, tonic and vermifuge ⁶. Plant mainly contains flavanoids, flavone glycosides, volatile oil, triterpenes, tannins and polysaccharides ^{6, 7}. Mature fresh leaves of *V. negundo* were confirmed for anti-inflammatory, analgesic and antihistaminic activities in various inflammation models⁸. Moreover, water extract of *V. negundo* showing test for polysaccharide positive, demonstrated antiinflammatory activity in formalin induced inflammation⁷. Aqueous and ethanolic leaf extracts of *V. negundo* (500 mg/kg) have also demonstrated significant antidiabetic potential in alloxan induced diabetes model of rats⁹. Methanolic extract of the *V. negundo* leaves exhibited glucose lowering potential post glucose challenge in fasting mice¹⁰.

Type II diabetes is a chronic metabolic disorder resulting from defects in insulin action and secretion. It is associated with other diseases such as obesity, hypertension, hyperlipidemia (increased VLDL, triglycerides and decreased HDL cholesterol), and cardiovascular disease, which is together known as 'Metabolic Syndrome'. It is characterized by fasting and postprandial hyperglycemia and if not treated timely, can cause microvascular and macrovascular long-term complications, such as nephropathy, neuropathy, retinopathy, and atherosclerosis. Risk factors for diabetes include - obesity, sedentary lifestyles and diets rich in fats 11, 12. Insulin resistance is decreased responsiveness of target tissues (skeletal muscles, adipose tissues, liver) to normal circulating levels of insulin. As a result of insulin resistance, there is reduced glucose utilization, which leads to hyperglycemic conditions 13 .

Moreover, many isolated crude polysaccharides from different parts of various plants have been reported for their anti-diabetic activity viz. crude polysaccharides isolated from the root of Liriope spicata var. prolifera demonstrated hypoglycemic and hypolipidemic activities in normal and STZ induced type II diabetes mellitus ¹⁴; crude polysaccharides from endodermis of Citrus paradisi demonstrated inhibitory effects on α glucosidase and α -amylase enzymes ¹⁵; galactomannans isolated from Trigonella foenumgraecum L. seeds showed anti-hyperglycemic activity in alloxan induced diabetes in mice in both acute and chronic studies ¹⁶; a sulphated polysaccharide isolated from Saccharina japonica has been reported for hypoglycemic activity in alloxan 17 induced diabetic rats and mice and polysaccharide from lotus plumule (Nelumbo

nucifera Gaertn) has been reported for promoting insulin release and ameliorating lipid profile and glucose intolerance in non-obese diabetic mice ¹⁸. Polysaccharides isolated from *Lycium barbarum* fruits have also been reported for their anti-diabetic activity *via* promoting glucose uptake in skeletal muscle and liver through GLUT4 transporter as well as increasing insulin signalling and sensitivity through PI3K/Akt/Nrf2 phosphorylation and activation of PI3K- and p38MAPK pathway ¹⁹.

Considering the anti-diabetic potential of V. *negundo* leaves as well as various naturally occurring polysaccharides based on above reports, this paper aimed at isolating the crude polysaccharide from the leaves of V. negundo as per the reported method ²⁰, characterizing it and evaluating its anti-hyperglycemic potential *in-vitro* using HepG2 cell line and *in-vivo* in normal male C57 mice post IPGTT (intraperitoneal glucose tolerance test). Anti-diabetic mechanisms of a plant extract / phytoconstituent can be attributed due to its insulin sensitizing, mimetic, secretogogue property or via inhibition of intestinal digestion and/or absorption of carbohydrates^{2,3}. Main targets of insulin include liver, skeletal muscle and fat cells. Hence, in this paper, a preliminary attempt has been made to evaluate anti-hyperglycemic activity of the crude polysaccharide of V. negundo leaves.

MATERIALS AND METHODS:

Isolation of Crude Polysaccharide from V. negundo Leaves (Modified as Reported by Wang et al., 2013): ²¹ Fresh, mature leaves of V. negundo were collected from the botanical garden of K. B. Institute of Pharmaceutical Education & Research, Gandhinagar, Gujarat, India and were confirmed for the identity by the Department of Pharmacognosy, K. B. Institute of Pharmaceutical Education & Research, Gandhinagar, Gujarat, India (PH/HERB/14/001). Collected leaves were shade dried and powdered for storage in dry and air tight container. Shade dried leaf powder was treated with methanol till the solution turned colourless. This colourless dried leaf powder was then extracted with 3 parts of distilled water on a water bath for 2 h each time. The pooled water extract post filtration was allowed to cool and precipitated with 70% methanol until maximum precipitation was achieved.

Precipitated polysaccharides were separated by centrifugation, repeatedly washed with acetone, dried in oven (just to evaporate moisture not exceeding 40 - 50 °C) and stored in an air tight vial till use. Crude polysaccharide yield obtained with this method was ~ 1.5% w/w.

Characterization of the Crude Polysaccharide Isolated from *V. negundo* Leaves:

Physicochemical Properties: Colour, texture, melting point, pH and solubility were evaluated for crude polysaccharide from *V. negundo* leaves were analysed.

Biochemical Tests: Presence of polysaccharides was confirmed using reported chemical tests for proteins, tannins, alkaloids and reducing/non-reducing sugars ²².

FT-IR Spectroscopy (400 - 4000 cm⁻¹) was performed to obtain fingerprints as well as identify functional groups and type of linkages present in crude polysaccharide ¹⁷ isolated from different batches of powdered *V. negundo* leaves. The sample was run on the Bruker instrument (3000 Hyperion Microscope with Vertex 80 FTIR System).

HRLCMS was carried out to obtain an approximate molecular weight and molecular formula of the crude polysaccharide isolated from *V. negundo* leaves ¹⁶. The sample was run on the Agilent instrument with ESI -positive mode ionization technique followed by QTOF detection. Elemental analysis was carried out to estimate the type and percentage of elements present in crude polysaccharide isolated from *V. negundo* leaves. The sample was run on various MICRO V2.03, Elemental analysis system GmbH.

In-vitro Glucose Uptake Activity of Crude Polysaccharide from V. negundo Leaves Using HepG2 Cell Line: ²³ HepG2 is a mouse liver cancer line which is well reported model to test insulin mimetic activity of herbal extracts / isolated phytochemicals. It gives idea about mechanistic aspect of the test substance such as involvement of membrane transporters (such as GLUT2 in liver cells) or liver enzymes which lead to glucose utilization. Experimental design to evaluate the glucose uptake potential of the crude polysaccharide from V. negundo leaves was

slightly modified as reported by Zhang *et al.*, 2008 ²³. HepG2 cells were maintained in DMEM culture medium containing 10% FBS, 2mM glutamine, 4.5 g/l (a high glucose condition) and antibiotics at 37°C, 5% CO₂. Once the cells achieved 80 - 90% confluency, they were detached from the culture flask with a solution of 0.25% trypsin and 1mM EDTA. Trypsin digestion was stopped by a complete media (DMEM with high glucose equivalent to 25 mM glucose) and plated in a density of 10,000 cells/well in a 96 well plate. 24 h post plating, they were incubated with different concentrations of PS (crude polysaccharide of *V. negundo* leaves), Metformin and Insulin under standard conditions.

Appropriate blanks and standards were used. Concentration of glucose left in the media was estimated colorimetrically post incubation at 24 and 48 h using the commercially available glucose estimation kits. % Reduction in glucose in media was calculated as {(absorbance of media from untreated cells – absorbance of media from treated cells) / (absorbance of media from untreated cells)} \times 100. 1 to 7 represents concentrations of the drugs in ascending order *viz*. 9.09nM, 27.27nM, 90nM, 272.72 nM, 909.09 nM, 2727.27 nM and 9090.9 nM respectively for standards and same values in ng/ml for PS.

In-vivo Anti-Hyperglycemic Activity of Crude Polysaccharide from *V. negundo* Leaves in C57 Mice: C57 mice are well reported animal model for type II diabetes ²⁴ (Srinivasan and Ramarao, 2007). Male C57 mice of 8-10 weeks age were taken for the study. Animals were divided into five groups and kept on overnight fasting before treatment. They were labeled as follows:

Group I (n=6) Vehicle control, treated with deionized water, P.O.

Group II (n=6) Crude Polysaccharide (10 mg/kg), P.O.

Group III (n=6) Crude Polysaccharide (30 mg/kg), P.O.

Group IV (n=6) Crude Polysaccharide (100 mg/kg), P.O.

Animals were approved by the IAEC (Registration number: KBIPER/2011/221) and were housed and acclimatized for one week in standard polypropylene cages (six animals/cage), maintained

under controlled room temperature (22 ± 2 °C) and humidity (55 \pm 5%) in 12 h /12 h light-dark cycle. Fasting blood collection was done and polysaccharide fraction was administered by oral route at 10, 30 and 100 mg/kg doses in overnight fasted mice. Vehicle system used to suspend the drug was 1% Tween 80: 1% PEG400: 0.5% CMC in deionized water. Glucose load (1.5 g/kg/10 ml) was injected intraperitoneally after 30 min of polysachharide administration. Blood was collected at 0, 15, 30, 60 and 120 min, after glucose load. Serum was separated and analyzed for glucose levels. Serum glucose was measured using Agappe Diagnostic Ltd., Kit, using Biotek Synergy Spectrophotometer.

RESULTS:

Physicochemical Properties of the Isolated Crude Polysaccharide from V. *negundo* **Leaves:** Isolated crude polysaccharide from V. *negundo* leaves appeared dark brown in colour with amorphous texture and hygroscopic property. Its pH when solubilized in water was found to be 6 to 6.5. Its melting point was recorded as 293° to 296°C. It is solubilized in hot water with sonication while partially soluble in solvents such as methanol, ethanol and DMSO.

Biochemical Tests for Confirmation of Polysaccharides: Biochemical tests confirmed the presence of carbohydrate and other possible constituents were found to be absent **Table 1**.

TABLE 1: BIOCHEMICAL TESTS DONE USINGCRUDE POLYSACCHARIDE SAMPLE FROM V.NEGUNDO LEAVES

Test	Result	Inference
FeCl ₃ Test	Negative	Absence of flavanoids
		and tannins
Dragendorff's	Negative	Absence of alkaloids
Test		
Biuret's Test	Negative	Absence of proteins
Fehling's Test	Negative	Absence of reducing
		sugars
Benedict's Test	Negative	Absence of reducing
		sugars
Tollen's Test	Negative	Absence of reducing
		sugars
Barfoed's Test	Positive	Indicates presence of
		non-reducing sugars
Iodine test	No change in	Absence of
	colour of	polysaccharides such
	iodine	as starch, glycogen and
	solution	dextran
Molisch's Test	s Test Positive Presence o	
		polysaccharides

Spectral Interpretation of the Isolated Crude Polysaccharide from V. negundo Leaves:

FT-IR Spectral Interpretation: ²⁵ FT-IR spectra over a range of 400 - 4000 cm⁻¹ indicated a similar pattern of spectra for all the batches of the crude polysaccharide isolated (all figures not shown here) at different time points, suggesting stability and non-degrading nature of compound when stored in cool and dry place in airtight containers. Moreover, "finger print region" which is generally exhibited by carbohydrates at maximum absorbance of 1200-950 cm⁻¹ was also evident from the spectra. Additionally, OH stretching from carboxylic acids at 3500 - 2200 cm⁻¹, aliphatic CH stretching at ~2918 cm⁻¹, carbonyl stretching at ~1600 cm⁻¹, C-O-C stretching from ethers and esters at 1200 cm⁻¹. C-O-H stretching vibrations at ~ 1000 cm⁻¹. confirmed the presence of polysaccharides and various functional groups present in the same Fig. 1.



FIG. 1: FT-IR SPECTRA OF THE CRUDE POLYSAC-CHARIDE FRACTION FROM V. NEGUNDO LEAVES

HRLCMS Interpretation: Molecular formula and mass of the isolated polysaccharide was found to be $C_{24}H_{48}O_{23}$ and 705.12 respectively **Fig. 2**.



FIG. 2: MASS SPECTRA OF THE CRUDE POLYSAC CHARIDE FRACTION FROM V. NEGUNDO LEAVES

Elemental analysis of crude polysaccharide isolated from the leaves of *V. negundo* showed 66.52%, 23.8% and 7.1% of carbon, hydrogen and oxygen respectively.

% Change in Glucose Uptake in Crude Polysaccharide Treated HepG2 Cells: % Reduction in media glucose in treated cells was observed in all test groups, however it was nonsignificant and not concentration dependent **Table 2**.

TABLE 2: % REDUCTION IN GLUCOSE UPTAKE FROM MEDIA IN CELLS TREATED WITH VARIOUS CONCENTRATIONS OF METFORMIN (M1 TO M5), CRUDE POLYSACCHARIDE OF *V. NEGUNDO* LEAVES (PS1 TO PS7) AND MARKETED INSULIN PREPARATION (TORRENT PHARMACEUTICALS; 11 TO 17); N = 2 IN CASES THE READING WERE NOT OBTAINED DUE TO CONTAMINATION ISSUE; p NOT LESS THAN 0.05, ONE WAY ANOVA TEST FOLLOWED BY BONFERRONI'S MULTIPLE COMPARISON TEST

Groups	% reduction of glucose in culture media at 24 h			cose in culture media at 24 h % reduction of glucose in culture media		
	Mean	SEM	Ν	Mean	SEM	Ν
M1	9.705213	5.955302	3	5.436165	0.6134306	3
M2	14.07857	2.90989	3	11.12104	3.729574	3
M3	5.053996	2.429784	3	8.102098	2.849001	3
M4	9.849979	0.8148567	3	6.866554	4.438274	3
M5	17.26344	5.47238	3	21.13835	5.562269	3
PS1	14.57358	1.274974	3	4.232713	1.248665	3
PS2	26.11289	2.928025	3	14.08956	6.491202	3
PS3	20.37593	10.85751	2	7.323866	0.7117562	2
PS4	10.34732	2.445087	3	6.866554	4.438274	3
PS5	11.32567	2.956045	2	10.77262	5.680295	2
PS6	33.24616	13.7247	3	9.259704	1.462801	3
PS7	29.64567	20.0279	3	8.279751	6.749648	2
I1	12.89942	3.101388	3	8.704971	1.153193	3
I2	19.02165	8.668576	3	6.644202	0.2474506	3
I3	11.50312	4.636819	3	12.98009	1.9324	2
I4	10.02043	1.310252	3	6.204082	2.170014	3
15	18.90257	3.762309	3	23.48336	7.074167	3
I6	8.136127	0.9721043	3	31.13847	11.87979	2
I7	8.00537	3.103146	2	28.66279	6.735893	2

In-vivo Anti-Hyperglycemic Activity of Crude Polysaccharide from *V. negundo* Leaves in C57 Mice:

Effect of Single Dose Novel Crude Polysaccharide from V. negundo on Fasting Serum Glucose and AUC post IPGTT in Male C57 Mice: Novel crude polysaccharide isolated from *V. negundo* when given in a single oral doses of 10, 30 and 100 mg/kg in male C57 mice, decreased the fasting serum glucose levels as well as AUC120 min at all time points *viz.* 15, 30, 60 and 120 min post intraperitoneal glucose challenge but not at significant level in comparison to vehicle control **Fig. 3 & 4**.



FIG. 3: SERUM GLUCOSE LEVELS IN IPGTT OF MALE C57 MICE TREATED WITH SINGLE DOSE ORAL ADMINISTRATION OF POLYSACCHARIDE FRACTION. Values represent mean ± SEM, for n=6

Effect of Single Dose Novel Crude Polysaccharide from V. negundo on % Change in Fasting Serum Glucose vs. Baseline Post 30 min of Single Oral Dose in Male C57 Mice: % Change in glucose concentrations when calculated at 30 min vs baseline (normal fasting serum glucose



levels) post single oral dose of novel crude polysaccharide at 10, 30 and 100 mg/kg in male C57 mice, a non-significant reduction was observed in polysaccharide treated animals *vs.* vehicle control group **Fig. 5**.



FIG. 5: % CHANGE IN BASELINE CORRECTED SERUM GLUCOSE LEVELS AT 30 MIN AFTER SINGLE DOSE ORAL ADMINISTRATION OF POLYSACCHARIDE FRAC-TION IN C57 MICE. Values represent mean \pm SEM, for n = 6

CONCLUSION: Physicochemical characterization of the crude polysaccharide of V. negundo confirm the presence of carbohydrate having C, H and O elements only with molecular mass ~705.12. Further detail characterization is a separate project in itself and since our primary objective is to evaluate glucose lowering activity of the crude polysaccharide of V. negundo leaves, we have performed basic characterization. Preliminary invitro data suggest that crude polysaccharide treated HepG2 cells did show a reduction in glucose concentrations *i.e.* enhanced glucose uptake process but at non-concentration dependent and non-significant level. This suggests possible insulin sensitizing and glucose lowering potential of the crude polysaccharide which can be better evaluated in-vivo. Hence, further in C57 mice, no significant reduction in fasting serum glucose was observed post treatment with 10, 30 and 100 mg/kg PO of crude polysaccharide (PS) vs. vehicle control which means PS does not have a direct effect on blood glucose levels. Moreover, post glucose load, no significant reduction in serum glucose levels at time points 15, 30, 60 and 120 min was observed in PS treated groups vs. vehicle control which suggest that single dose of PS is not effective in lowering blood glucose levels. Further in-vivo experiments with repeated dosing are required in diabetic animals to explore efficacy and mechanistic aspect of crude polysaccharide of V. negundo leaves.

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CONFLICT OF INTEREST: There is no conflict of interest.

REFERENCES:

- 1. Dave DT and Shah GB: Pharmacological potential of naturally occurring non-starch polysaccharides (NSP). The Journal of Phytopharmacology 2015; 4(6): 299-310.
- 2. Thakur: Rasayana properties of Ayurvedic herbs: Are polysaccharides a major contributor. Carbohydrate Polymers 2012; 87: 3-15.
- 3. Ramberg JE, Nelson ED and Sinnott RA: Immunomodulatory dietary polysaccharides: a systematic review of literature. Nutrition Journal 2010; 9: 54.
- 4. Vishwanathan AS and Basavaraju R: A Review on *Vitex negundo* L. A Medicinally Important Plant; April-June 2010; EJBS 3(1): 30-42
- 5. Ahirrao RA, Patel MR and Pokal DM: Pharmacognostical studies of *Vitex negundo* leaves. Biological Forum An International Journal 2011. 3(1): 19-20.
- Ladda PL and Magdum CS: *Vitex negundo* Linn.: Ethnobotany, Phytochemistry and Pharmacology - A Review. International Journal of Advances in Pharmacy, Biology and Chemistry 2012; 1(1): 111-120.
- 7. Brindha: Studies on the chemical and medicinal value of *Vitex negundo* Linn. IJABR 2012; 2(2): 298-301.
- Dharmasiri MG, Jayakody JRAC and Gathena G: Anti inflammatory and analgesic activity of mature fresh leaves of *Vitex negundo*. Journal of Ethno pharmacology 2003; 87(2-3): 199-206.
- Prasanna RP, Sivakumar V and Riyazullah MS: Antidiabetic potential of aqueous and ethanolic extracts of *Vitex negundo*. International Journal of Pharmacognosy and Phytochemical Research 2012; 4(2): 38-40.
- 10. Villasenor IM and Lamadrid MA: Comparative antihyperglycemic potential of medicinal plants. Journal of Ethnopharmacology 2006. 104: 129-131.
- 11. Jain S and Saraf S: Type 2 diabetes mellitus-Its global prevalence and therapeutic strategies. Diabetes & Metabolic Syndrome: Clinical Research & Reviews 2010; 4: 48-56.
- 12. Holman RR: Type 2 diabetes mellitus in 2012: optimal management of T2DM remains elusive. Nat Rev Endocrinol 2013; 9(2): 67-68.
- 13. Sesti G: Pathophysiology of Insulin Resistance. Best Practice & Research Clinical Endocrinology & Metabolism 2006; 20(4): 665-679.
- 14. Chen: Anti-diabetic effects of water extract and crude polysaccharide from tuberous roots of *Liriope spicata* var. Prolifera in mice. Journal of Ethno-pharmacology 2009; 122: 205-209.
- 15. Liu G: Chemical compositions, α-glucosidase and αamylase inhibitory activities of crude polysaccharides from the endodermis of shaddock (*Citrus maxima*). Archives of Biological Sciences Belgrade 2012. 64(1): 71-76.
- 16. Kamble H, Kandharea AD, Bodhankar S, Mohan V and Thakurdesai P: Effect of low molecular weight galactomannans from fenugreek seeds on animal models of diabetes mellitus. Biomedicine & Aging Pathology 2013; 3: 145-151.
- 17. Wang: Hypoglycemic property of acidic polysaccharide extracted from *Saccharina japonica* and its potential mechanism. Carbohydrate Polymers 2013; 95: 143-147.
- 18. Liao and Lin: Lotus (*Nelumbo nucifera* Gaertn) plumule polysaccharide ameliorates pancreatic islets loss and serum lipid profiles in non-obese diabetic mice. Food and Chemical Toxicology 2013. 58: 416-422.
- 19. Cheng: An evidence-based update on the pharmacological activities and possible molecular targets of *Lycium*

23. Zhang Li, Hu J and Du G: Establishment of a cell-based

24. Srinivasan K and Ramarao P: Animal models in type 2

25. Yang: In-vitro antioxidant activities of sulfated poly-

Discov Ther 2008; 2(4): 229-233.

March 2007; 451-472.

49: 1031-7.

assay to screen insulin-like hypoglycemic drugs. Drug

diabetes research: An overview. Indian J Med Res 125,

saccharide fractions extracted from Corallina officinalis.

International Journal of Biological Macro-molecules 2011;

barbarum polysaccharides. Drug Design, Development and Therapy 2015: 9: 33-78.

- 20. Kwong C, Leung KN, Fungand KP and Mcroy Y: Immunomodulatory and Anti-tumour Polysaccharides from Medicinal Plants. The Journal of International Medical Research 1994; 22: 299-312.
- Wang: Multi-fingerprint and quality control analysis of tea polysaccharides. Carbohydrate Polymers 2013; 92: 583-590.
- 22. Kokate: Pharmacognosy 1988; 6: 12-28.

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