



Received on 12 January, 2012; received in revised form 24 February, 2012; accepted 20 April, 2012

FORMULATION AND EVALUATION OF CURCUMIN LOADED MAGNETIC NANOPARTICLES FOR CANCER THERAPY

T. Silambarasi, S. Latha*, M. Thambidurai and P. Selvamani

Department of Pharmaceutical Technology, Anna University of Technology, Tiruchirappalli - 620024, Tamil Nadu, India

ABSTRACT

Keywords:

Curcumin,
magnetic nanoparticles,
cancer,
drug release kinetics

Correspondence to Author:

S. Latha

Department of Pharmaceutical
Technology, Anna University of
Technology, Tiruchirappalli - 620024,
Tamil Nadu, India

The conventional chemotherapeutic agents in oncology drug discovery still exhibit poor specificity in reaching tumor site and often restricted by dose-limiting toxicity. The combination of developing drug formulation by utilizing both controlled release technology and drug targeting technology may provide a more efficient and less harmful solution to conquer the limitations found in conventional chemotherapy. In this study, the anticancer drug curcumin was encapsulated in a polymeric magnetic nanoparticle which was synthesized with polymers β -cyclodextrin cross linked with epichlorhydrin, hydrophobically modified dextran byoleoylchloride and magnetite as magnetic material. Particle size, surface morphology, zeta potential and magnetic measurements were used to characterize the developed drug formulations. The developed drug-iron conjugated nanoparticles were found to be within the size range of 100nm with excellent negative surface charge ($>-30\text{eV}$) and spherical in shape. The magnetic susceptibility and magnetization curve substantiate the super paramagnetic property of the developed drug formulation. Furthermore, the drug content and encapsulation efficiency found was directly proportional to epichlorhydrin β -cyclodextrin concentration in the developed formulation. The *in-vitro* release profile of curcumin loaded magnetic nanoparticles exhibited biphasic initial release first 24 hours and release extended upto 72 hours. The drug release kinetics indicated that drug release from drug formulations were best explained by Higuchi's equation, as these plots showed the highest linearity but a close relationship was also noted with first-order kinetics.

INTRODUCTION: Curcumin, a major polyphenolic pigment of the turmeric root [*Curcuma longa* L., Family: *Zingiberaceae*] found widely cultivated in several tropical parts of Asia. Turmeric commonly finds use as a spice in Indian cooking, a cosmetic agent for skin care and a traditional Indian and Chinese medicine. It has low intrinsic toxicity but a wide range of pharmacological activities including antitumor, antioxidant, anti-amyloid and anti-inflammatory properties¹.

Several clinical trials dealing with cancer have stated the pharmacokinetics, safety and efficacy of curcumin in humans. However, clinical applications of Curcumin have been hampered by its extremely poor water solubility i.e., less than $1\ \mu\text{g}/\text{ml}$ ². Many methods have been developed for improve solubility of poor water soluble drugs but they induce severe side effects such as neurotoxicity, nephrotoxicity and hypersensitivity etc.

To evade this difficulty, considerable efforts have been devoted to the research and development of alternative methods to improve drug solubility without the use of organic solvents and other relatively harmful excipients.

The development of drug nanocarriers for poorly soluble pharmaceuticals seems to be more promising. The therapeutic application of hydrophobic, poorly water-soluble agents is associated with some serious problems, such as poor absorption and low bioavailability. Colloidal drug delivery systems such as liposomes, micelles and nanoparticles have been intensively examined for their use in tumor therapy. The effectiveness of drug delivery systems can be attributed to their physical and chemical properties.

Recent investigations have been trying to develop targeted therapeutic systems by using external forces, including magnetic fields, ultrasound, electric fields, temperature, light and mechanical forces to concentrate drugs within tumor sites. In these systems, the drug is localized at a specific targeted area by externally generated magnetic forces⁷. The magnetic particles tagged with drug molecules are targeted to specific sites of the body by application of external magnetic fields where the drug molecules were gradually released, thus improving the therapeutic efficiency of the drugs by lowering the collateral toxic side effects on the associated healthy cells or tissues.

Cyclodextrins (CDs) are ring-shaped structures composed of glucose units connected by α -d1, 4 bonds forming a polar interior (cavity) and a polar exterior. The cavity gives the CD its unique ability to include other molecules or part of them depending on the size and polarity leading to, e.g. enhanced stability and solubility. CDs, in polymeric form may present some advantages in many cases such as higher solubility than the native CDs, lower toxicity or cooperative binding effects. The CD polymers which have been extensively studied are random cross-linked epichlorhydrin-CD polymers which can be tailored to yield either water-soluble or in-soluble polymers³.

Dextran (mainly α -1, 6-polyglucose with some α -1, 4 branching) has been particularly popular owing to its clinical approval for use as a plasma expander. Some of hydrophobically modified dextran can form nanoscale

particulate system and few of them shows promise carriers for lipophilic drug delivery. The association of hydrophobically modified polycationic dextran derivatives with oppositely charged surfactants directs to aqueous solutions with complex phase behavior and self-assembling morphologies in solution⁴. The spontaneous formation of hydrophobic cores in aqueous solution, exhibit a great potential in drug/gene delivery research and for other biomedical applications.

In this present work, the solubility of the curcumin was effectively improved by making inclusion complex within beta cyclodextrin cavities. Also, the drug release was extended by use of hydrophobically modified dextran and the targeting can be facilitated by the functionalizing the complex with the magnetic material.

MATERIALS AND METHODS: Dextran 40, β Cyclodextrin and oleoylchloride were purchased from M/s. Sigma Aldrich Chemicals Ltd, Bangalore. Epichlorhydrin purchased from M/s. Spectrochem, Mumbai. Gift sample of Curcumin was obtained from M/s. Natural remedies Bangalore. All other chemicals and reagents used were of analytical grade obtained locally.

Synthesis of β -cyclodextrin Copolymer: Cyclodextrin copolymer (Ep β -CD) was prepared by cross linking β Cyclodextrin (β -CD) with Epichlorhydrin (EP) under strong alkaline conditions. 5 g of anhydrous β -CD was dissolved in 8ml NaOH 33% (w/w) solution under mechanical stirring overnight. Then, 4.07 g of Epichlorhydrin was added to the solution. The solution was heated to 30°C. In order to obtain a high molecular weight polymer, the reaction was stopped in the vicinity of the gelation point by addition of acetone. The obtained aqueous phase was heated at 50°C overnight, neutralized with 6N HCl, filtered and then recovered³.

Synthesis of Oleoyldextran: 100 ml of dimethyl formamide contain 1gm of lithium chloride was prepared and 4gm of dextran (Mw = 40000 g/mol) was solublized by means of magnetic stirring with reflux. Then 390mg oleoylchloride and 0.031ml of pyridine were mixed and stand for 3hours at 50°C to obtain clear solution.

Finally oleoyldextran was precipitated by the addition of isopropyl alcohol. The precipitate was separated, solubilized in deionized water, purified by dialysis and freeze dried.

Identification of Synthesized Compounds: In order to confirm the synthesized compounds, the FT-IR spectra of the starting materials were compared with the spectrum of synthesized compounds. Here the starting polymer β -cyclodextrin FT-IR spectrum was compared with resulting product Epichlorhydrin- β -Cyclodextrin copolymer. Similarly the FT-IR spectrum of dextran (Mw40000) was compared with oleoyldextran FT-IR spectrum³.

Compatibility studies of Drug Excipients: Compatibility studies were carried out to study the possible interactions between drug and other inactive ingredients. The polymers oleoyldextran and epichlorhydrin- β -Cyclodextrin copolymer and drug

TABLE 1: FORMULATION OF POLYMER CONCENTRATIONS

Formulation Code	Polymer Ratio (Ep β CD: MD)	Ep β CD (mg)	Oleoyldextran (mg)	Drug (curcumin) feeding (mg)
F1	1:1	10	20	5
F2	1:2	15	15	5
F3	2:1	20	10	5

Characterization of Nanoparticles:

Size and Size distribution: The magnetic nanoparticles were dispersed in deionized water and subjected to particle size, Size distribution and polydispersity measurement in (Malvern Particle Size Analyzer MS2000).

Measurement of Surface Charge: Zeta potential measurement of formulated drug magnetic nanoparticles suspension with deionized water was performed with Malvern Zetasizer Nano ZS (MAL 000967).

Particulate morphology: The morphology of the nanoparticles was observed by field emission scanning electron microscopy (FE-SEM; Hitachi S-4700, Japan). A drop of diluted nanoparticles suspension was placed on a 400 mesh carbon-coated copper grid. After drying, the samples were sputter-coated with a gold-palladium alloy and analyzed at an electron voltage of 5 kV.

(curcumin) were studied using FT-IR 410 pc spectrometer and the compatibility studies between drug and polymers carried out. Drug and KBr were compressed under 10 tonnes pressure in a hydraulic press to form a transparent pellet. The pellet was scanned from 4000 to 400cm⁻¹ in FT-IR spectrometer⁶.

Formulation of Magnetic Nano Assemblies: The Epichlorhydrin- β -Cyclodextrin copolymer and oleoyldextran were diluted with nanopure water. Both were mixed with 5mg curcumin and allow to sonicated for 15 minutes. To the above mixture 10 ml aqueous solution of 0.0240 g FeCl₃.6H₂O and 0.0342 g of FeCl₂.H₂O for 30minutes at 40°C. Three different polymers proportions were chosen was mentioned in **Table 1**. Finally, ammonia solution was added drop by drop to above mixture. The dark brownish color drug loaded magnetic nanoparticles was collected by centrifugation at 12000 rpm.

Magnetism measurement: The hysteresis loops of iron oxide magnetic nanoparticles, drug loaded magnetic nanoparticles were measured with vibrating sample magnetometer (VSM, Lakeshore, Model 7410 at 83 and 300K) for the electromotive force induced by magnetic particles in which the particles vibrated at a constant frequency and uniform magnetic field. The Magnetic susceptibility of the formulated magnetic nanoparticles was determined using Fugro magnetic susceptibility meter.

Drug loading and Encapsulation efficiency: Determination of drug loading efficiency and encapsulation efficiency of curcumin loaded magnetic nanoparticles were carried out extracting curcumin out from nanoparticle. Briefly, 2 mg of curcumin loaded magnetic nanoparticles were mixed with 50 ml methanol and allowed 20minutes under sonication. The resulting mixture was subjected to centrifuge at 5000 rpm for 10 minutes. The supernatant was assayed by UV-Vis Spectrometer (Shimadzu 1800) at 420 nm. The drug loading efficiency (L.E.) and drug encapsulation efficiency (E.E.) were determined as follows.

Drug Loading Efficiency (%w/w)=

$$\frac{\text{Mass of drug in Nanoparticles}}{\text{Mass of Nanoparticles}} \times 100$$

Encapsulation Efficiency (%w/w)=

$$\frac{\text{Mass of drug in Nanoparticles}}{\text{Mass of Nanoparticles}} \times 100$$

In-vitro Drug Release Profile: Dialysis bags (cut off size of 12–14 kDa) were filled with a predetermined amount of each formulation (2mg) and put into 40 ml of phosphate buffer solution (pH 7.4) used as receptor phase. The receptor phase was stirred and thermal controlled at 37°C. At fixed time interval 2 ml of the receptor phase were withdrawn and substituted with fresh buffer. The drug release was assayed spectrophotometrically at 420 nm. The cumulative percentage amount of drug release was calculated and graph plotted against time.

RESULTS:

FT-IR spectrum of Polymers:

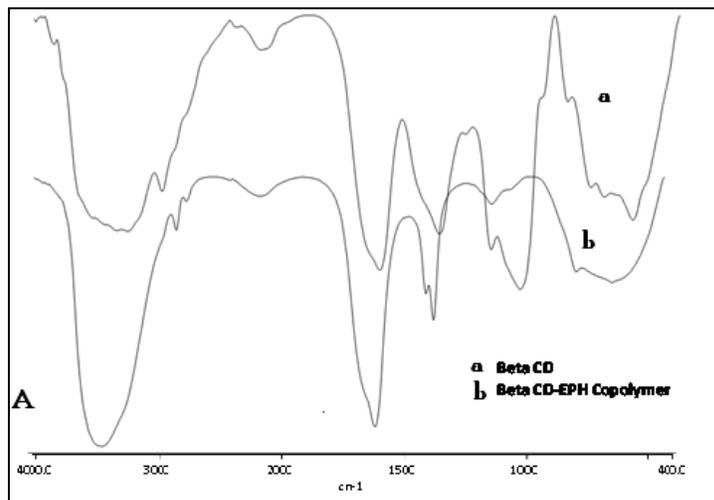


FIGURE 1A: FT-IR SPECTRUM OF β-CD And Epβ-CD COPOLYMER

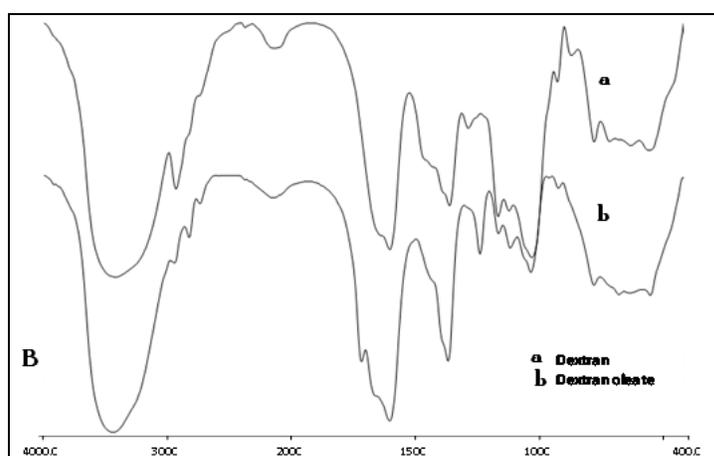
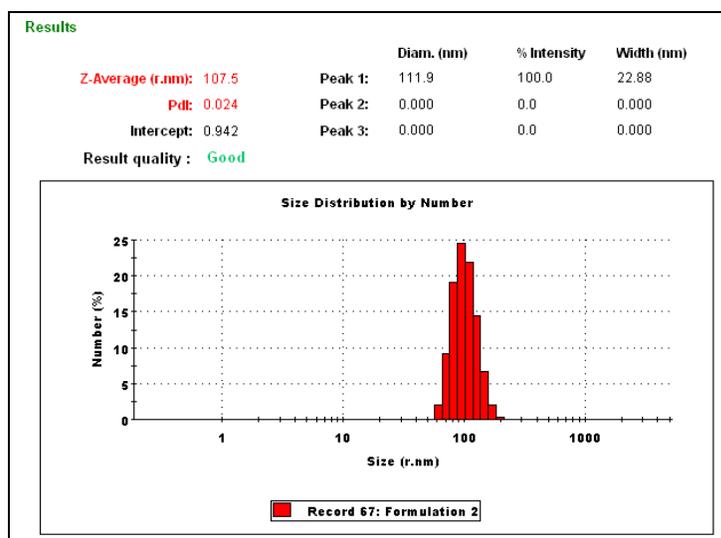
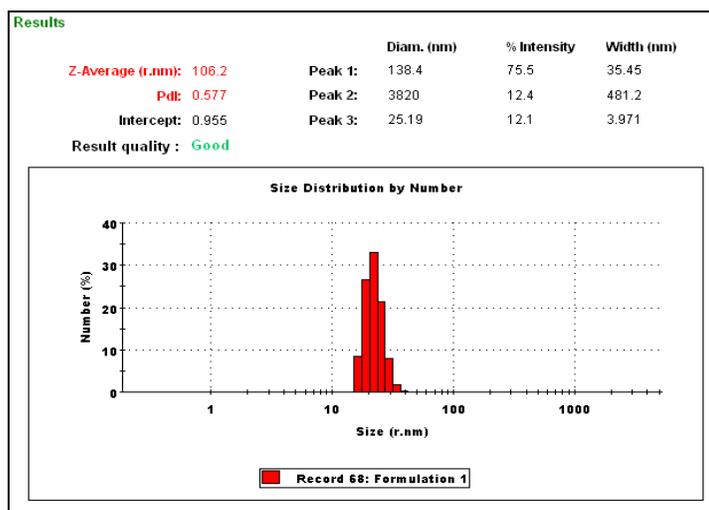


FIGURE 1B: FT-IR SPECTRUM OF DEXTRAN AND OLEOYLDEXTRAN

TABLE 2: CHARACTERISTICS OF CURCUMIN LOADED MAGNETIC NANOPARTICLES

Formulation Code	Particle Size (nm)	Poly dispersity Index	Zeta Potential (mV)	Conductivity (S/cm)	Magnetic Susceptibility
F1	106.2	0.577	-34.6	0.453	-33X10 ⁻⁶
F2	107.5	0.024	-34.1	0.332	-29X10 ⁻⁶
F3	101.0	0.030	-37.3	0.297	-28X10 ⁻⁶

Particle Size Analysis of Curcumin Magnetic Nanoparticles and Polydispersity:



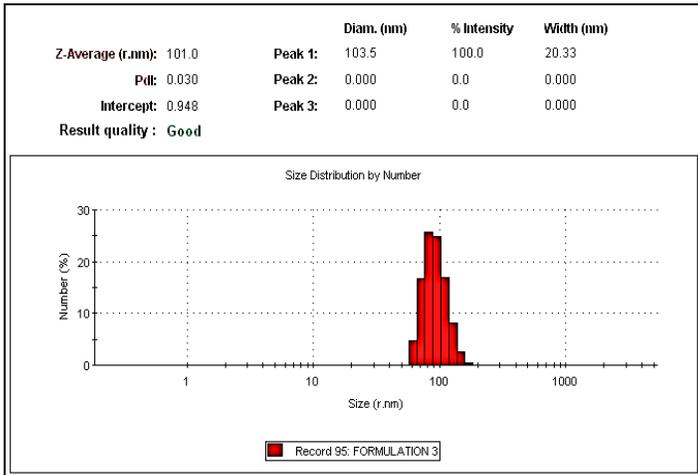


FIGURE 2: PARTICLE SIZE ANALYSIS OF CURCUMIN MAGNETIC NANOPARTICLES FORMULATIONS

Zeta potential of Curcumin Magnetic Nanoparticles and Magnetism measurement:

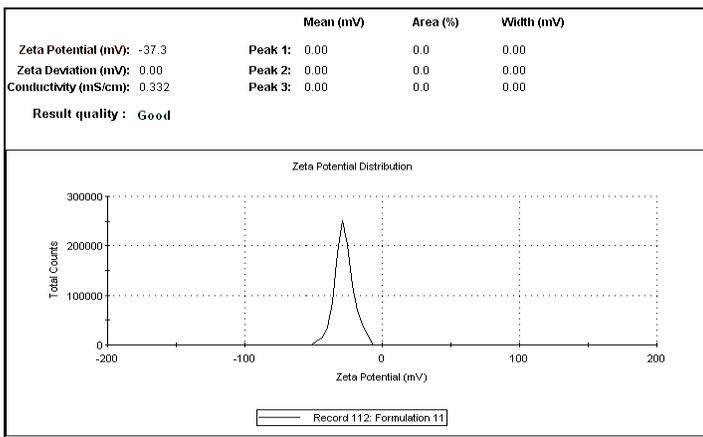


FIGURE 3: ZETAPOTENTIAL DISTRIBUTION

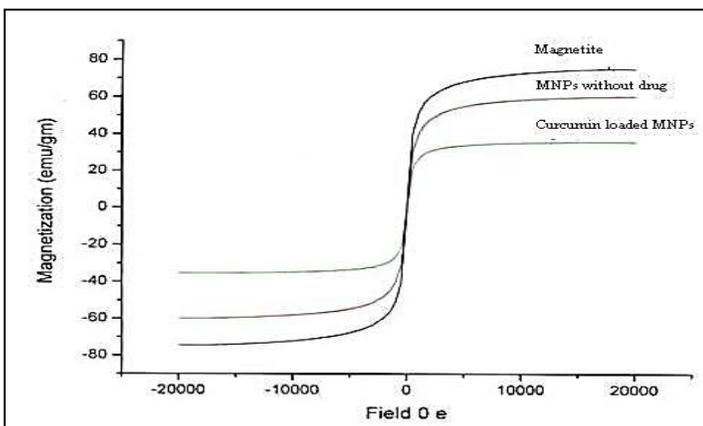


FIGURE 4: MAGNETIZATION HYSTERESIS LOOPS OF MAGNETITE, MNPs WITHOUT DRUG AND CURCUMIN LOADED MNPs

TABLE 3: EFFECT OF DIFFERENT POLYMER PROPORTION ON ENCAPSULATION EFFICIENCY, DRUG LOADING EFFICIENCY AND *IN-VITRO* DRUG RELEASE

Formulation code	Feed drug (mg)	Encapsulation efficiency (%)	Drug loading efficiency (%)	<i>In-vitro</i> release after 96 hours (%)
F1	5	42.48±0.69	7.22±0.17	72.32±0.77
F2	5	72.30±0.49	10.3±0.34	83.41±0.67
F3	5	29.54±0.98	4.87±0.1	57.38±0.44

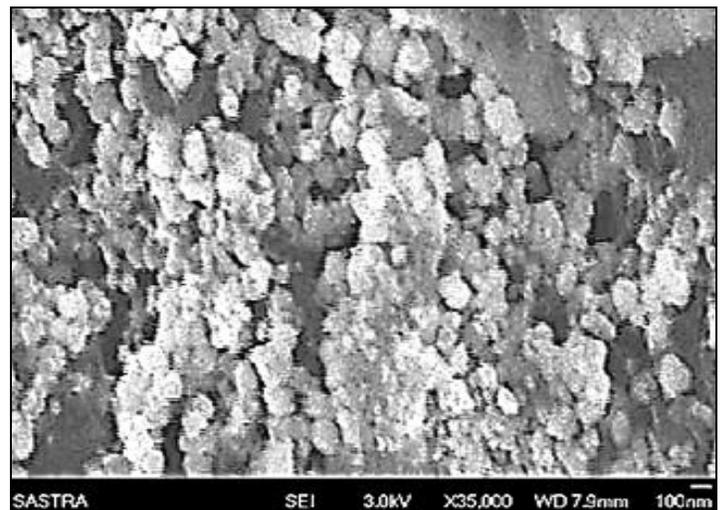
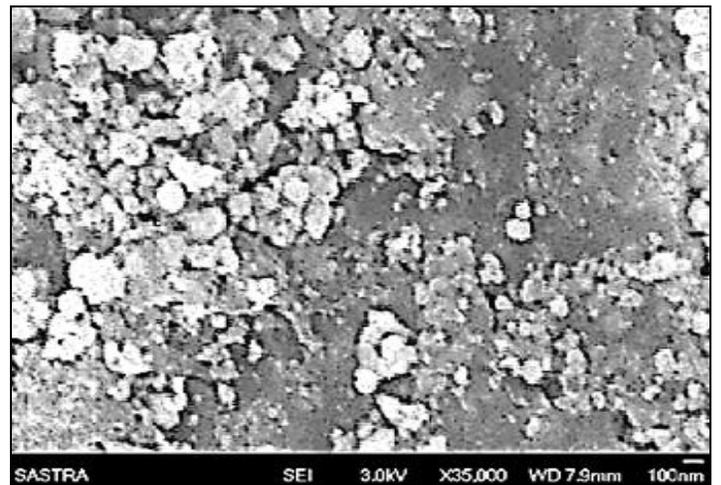
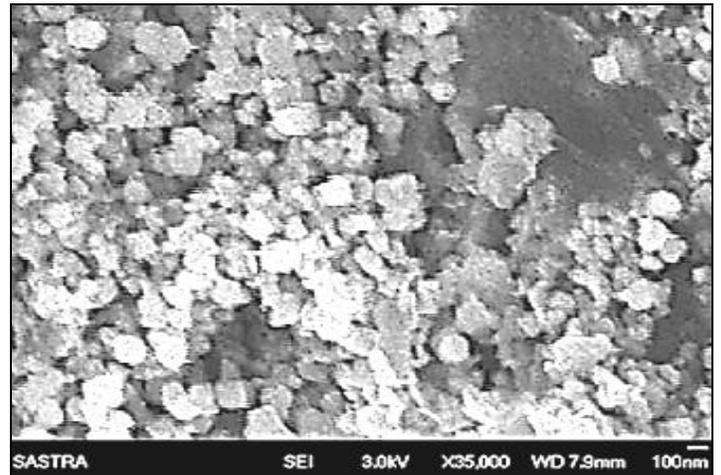


FIGURE 5: SEM MICROGRAPHS OF CURCUMIN MAGNETIC NANOPARTICLES

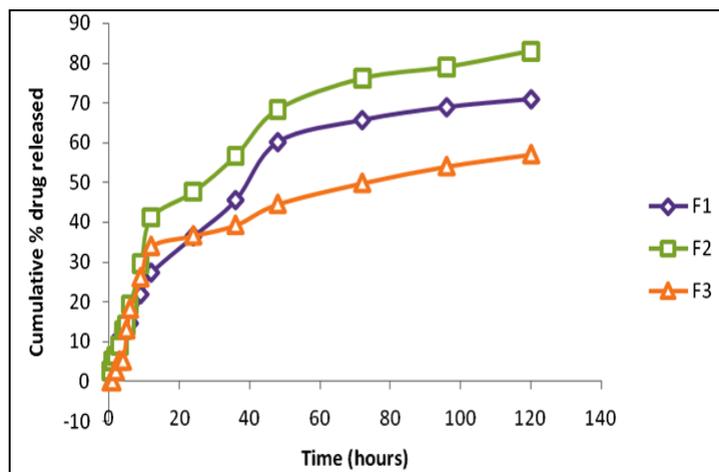


FIGURE 6: *IN-VITRO* DRUG RELEASE PROFILE OF CURCUMIN LOADED MAGNETIC NANOPARTICLES

DISCUSSIONS:

Confirmation of Synthesized Polymers: The differences between the FT-IR spectrums (**Figure 1 a**) suggest cross-linkage of β -CD in Ep β -CD copolymer. The peaks of C-H stretching ($2890\text{--}2880\text{ cm}^{-1}$) and bending ($1480\text{--}1280\text{ cm}^{-1}$) from normal alkanes, CH_2Cl rocking and wagging band ($1190\text{--}1070\text{ cm}^{-1}$) and the C-Cl broad band ($700\text{--}420\text{ cm}^{-1}$) confirm the formation of polymer with the addition of ECH. Meanwhile, the similarities between the spectra of β -CD and β -CDEP copolymer also indicate that the basic structural units are preserved in the polymer. From the FTIR spectrum (**Figure 1b**) of dextran and oleoyldextran the formation of carbonyl group confirmed by C=O stretching 1738 cm^{-1} . C=O stretching ($1735\text{--}1750\text{ cm}^{-1}$) functional region for ester groups. Hence the esterification occurred between dextran and oleoylchloride and the formation of oleoyldextran was confirmed.

Compatibility studies: The FT-IR spectrum of individual polymers, drug curcumin, magnetite and mixture of all ingredients were compared and no significant drug excipient interactions were noted. The results confirmed the components used in the formulations were compatible.

Formulation of Magnetic Nanoparticles: The cross linked Ep β -CD copolymer restrain number of cavities, hydrophobic oleoyl side chain of oleoyldextran form inclusion complex with cyclodextrin cavities and dextran encircle over drug loaded cyclodextrin cavities. The hydrophobic drug curcumin fluently form inclusion complex within remaining cyclodextrin cavities.

The targeting material nano sized magnetite covered by the self-assemblies.

Particle size: The particle size of the formulations was around 100nm with low polydispersity which illustrated from particle size histograms (**Figure 2**). Smaller particles have higher surface area/volume ratio, which makes it easier for the encapsulated drug to be released from the particles via diffusion and surface erosion and also have the added advantage for the drug-loaded nanoparticles to penetrate into, and permeate through the physiological drug barriers. The smaller particles will have greater ease of entry and durability in the tumors. In contrast, larger particles have large cores, which allow more amount of drug to be encapsulated per particle and allows slow drug release. Thus, control of particle size provides a means of tuning drug release rates.

Particulate morphology: The particles were analyzed by SEM to obtain further information of their surface morphology. This can possibly result in a smaller size than measured by zetasizer since the particles have been dried. Pictures of the particles are shown in **Figure 5**. The particles were appear to form cluster like aggregates and spherical in nature. The SEM images also substantiate the particle size within nanoscale dimensions.

Surface charge: Surface charge reflects the electrical potential of particles and is influenced by the composition of the particle and the medium in which it is dispersed. Nanoparticles with a zeta potential above $\pm 30\text{ mV}$ have been shown to be stable in colloidal dispersion, as the surface charge prevents aggregation of the particles. The curcumin loaded magnetic nanoparticles had negative charge within normal value -32.3mV (**Figure 3 & 4**).

Magnetization measurement: Magnetic hysteresis loops of pure iron oxide and the drug loaded magnetic nanoparticles were determined and it shows the samples were super paramagnetic in nature. The magnetization value of Fe_3O_4 , magnetic nanoparticles without drug and curcumin loaded magnetic nanoparticles were 72.93emu/g , 58.23emu/g and 37.75emu/g respectively. The saturation of magnetization of curcumin loaded magnetic nanoparticles was smaller than that of bulk magnetite,

but both particles had similar properties that were close to the paramagnetic behavior. The magnetic susceptibility reported in **Table 2** which evident a clear magnetic response moreover the nanoparticles can readily be moved and collected with an external magnetic field.

Drug loading and Encapsulation efficiency: The drug loading and the encapsulation efficiency of curcumin-loaded in magnetic nanoparticles are shown in **Table 3**. The encapsulation efficiency of the formulations tested in this study 42.48%, 72.30% and 29.54%. The encapsulation efficiency and drug content increases with the beta cyclodextrin concentration increase in the formulation. A successful nano delivery system should have a high drug-loading capacity, thereby reducing the quantity of matrix materials for administration. The drug loading efficiency was determined as 7.22%, 10.3% and 4.87% for final formulation.

Based on the parameters shown in Table 2, the curcumin loaded magnetic nanoparticle formulation was optimized. The physiochemical characteristics such particle size, polydispersity, zeta potential, conductivity and magnetic susceptibility were not shown any considerable variance for all three formulations. The SEM images furthermore evident the all formulations were spherical in nature. However encapsulation efficiency and drug content were substantially improved with increasing concentration of Cyclodextrin copolymer (Ep β -CD); because of high amount of hydrophobic drug was encapsulated in β -Cyclodextrin cavities by forming inclusion complex.

***In-vitro* drug release of Curcumin loaded Magnetic Nanoparticles:** The *in-vitro* release behavior of curcumin loaded magnetic nanoparticles exhibit biphasic pattern initial release for first 24 hours and the release slowly continued upto 96 hours. The release of the drug from magnetic nano carrier is restricted by concentration of oleoyldextran in the formulation. The cumulative percentage of drug release after 96 hours from the formulations was found to be 72.32%, 83.41% and 57.38%. It clearly proven that the cumulative percentage of drug release was noticeable decreased with increasing concentration of oleoyldextran in formulation it is due to gelling property of oleoyldextran (**figure 6**).

Furthermore, the formulation which contains 2:1 proportion of Ep β -CD and oleoyldextran exhibits improved drug release profile in extended manner upto 4 days.

CONCLUSION: Magnetic nano particulate system with bio degradable polymers Ep β -CD copolymer and oleoyldextran were formulated and the anticancer drug curcumin was effectively loaded. The characteristics of the formulation comply with normal nanosized nanoparticulate system. The *in-vitro* release concluded the anti-cancer drug curcumin loaded magnetic nanoparticles successful in targeting the drug molecule as well as controlling release from the formulation. The results have demonstrated this magnetic nano particulate system applied for all site specific hydrophobic drugs.

REFERENCES:

1. Aggarwal BB, Kumar A, Bharti AC. Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res.* 2003; 23:363-398.
2. Maheshwari RK, Singh AK, Gaddipati J, Srimal RC. Multiple biological activities of curcumin: a short review. *Life Sci.* 2006; 78:2081-2087.
3. Renard E, Deratani A, Volet G, Sébille C. Characterization of water soluble high molecular weight β -cyclodextrin-epichlorohydrin polymers: *Eur. Polym. J.* 1997; 33:49-57.
4. Heath P, Wanida C, Timothy G, Jon D. Structural and magnetic properties of nanoscale iron oxide particles synthesized in the presence of dextran or polyvinyl alcohol. *J. MagnMagn Mater.* 2001; 225:41-46.
5. Jing Z, Xi Guang C, Yan Yan L, Cheng Sheng L. Self-assembled nanoparticles based on hydrophobically modified chitosan as carriers for doxorubicin Nano: *Nanotech Biol and Med.* 2007; 3:258-265.
6. Jianshu Li, Huining X, Jiehua Li, YinPing Z. Drug carrier systems based on water-soluble cationic β -cyclodextrin polymer: *Int. J. Pharm.* 2004; 278: 329-342.
7. Kiso Y, Suzuki Y, Watanabe N, Oshima Y, Hikino H. Antihepatotoxic principles of *Curcuma longa* rhizomes. *Planta Med.* 1983; 49:185-187.
8. Maeda H. The enhanced permeability and retention (EPR) effect in tumor vasculature: the key role of tumor-selective macromolecular drug targeting. *Adv Enzyme Regul.* 2001; 41:189-207.
9. Gref R, Amiel C, Molinard K, Daoud-Mahammed S, Sebille B, Gillet B, Beloeil J.C, Ringard C, Rosilio V, Poupaert J, Couvreur P. New self-assembled nanogels based on host-guest interactions: characterization and drug loading. *J. Control Rel.* 2006; 111:316-324.
10. Larsen KL. Large cyclodextrins: *J. Incl. PhenomMacrocycl Chem.* 2002; 43:1-13.9.
11. Zharov VP, Galitovskaya EN, Johnson C, Kelly T. Synergistic enhancement of selective nano photothermolysis with gold nano clusters: potential for cancer therapy, *Lasers in Surgery and Medicine.* 2005; 37:219-226.

12. Rao CV, Rivenson A, Simi B, Reddy BS. Chemoprevention of colon carcinogenesis by dietary curcumin: a naturally occurring plant phenolic compound. *Cancer Res.* 1995; 55:259-266.
13. Thomas H, Tim L, Brigitte H, Stephanie H. Functional Polymers Based on Dextran. *Adv Poly Science.* 2006; 205:199-291.
14. Loo C, Lowery A, Halas N, West J, Drezek R. Immunotargeted nanoshells for integrated cancer imaging and therapy. *Nano Letters* 2005; 5:709-711.
15. Magenheim B. A new in vitro technique for the evaluation of drug release profile from colloidal carriers –ultrafiltration technique at low pressure, *Int. J. Pharm.* 1993; 94:115-123.
16. Matsumura Y, Maeda H. A new concept for macromolecular therapeutics in cancer chemotherapy: mechanism of tumorotropic accumulation of proteins and the antitumor agent smancs. *Cancer Res.* 1986;46:63-87.
17. Mehrdad H, Amir A, Pedram R. Hydrogel nanoparticles in drug delivery. *Adv Drug Del. Rev.* 2008; 60: 1638-1649.
18. Panyam J, Labhasetwar V. Biodegradable nanoparticles for drug and gene delivery to cells and tissue. *Adv Drug Del Rev.* 2003; 55:329-347.
19. Panyam D, Williams A, Dash D, Leslie P, Labhasetwar V. Solid-state solubility influences encapsulation and release of hydrophobic drugs from PLGA/PLA nanoparticles, *J. Pharm. Sci.* 2004;93:1804-1814.
20. Serguei V, Vinogradov R, Arin D, Zeman E, Batrakova V, Alexander V, Kabanov P. Nanogel formulation for drug delivery of cytotoxic nucleoside analogs, *J Controlled Release.* 2005; 103: 143-157.
21. Sudimack J, Lee RJ: Targeted Drug Delivery via the folate receptor. *Adv Drug Delivery Rev.* 2000;41:147-162.
22. Text Book of Nanoparticulates as Drug Carriers, Vladimir P by Imperial College Press. 2006; 397-411.
23. The Handbook of Nanomedicine, Kewal K, Jain MD, Jain Pharma Biotech, Basel. 2008; 195-244.
24. Thomas H, Tim Liebert Brigitte H, Stephanie Horning Functional Polymers Based on Dextran. *Adv Polymer Science.* 2006; 205:199-291.
25. Wagner E, Programmed drug delivery: nanosystems for tumor targeting. 2007; 7:587-593.
26. Yokoyama M, Okano T, Sakurai Y, Ekimoto H, Shibasaki C, Kataoka K. Toxicity and antitumor activity against solid tumors of micelle-forming polymeric anticancer drug and its extremely long circulation in blood. *Cancer Res.* 1991; 51:3229-3236.
27. Zauner W, Farrow NA, Haines AM. In vitro uptake of polystyrene microsphere: effect of particle size, cell line and cell density, *J. Controlled Release.* 2001;71:39-51.
28. Zefeng X, Guobin W, Kaixiong T, Jianxing L. Preparation of magnetite-dextran microspheres by Ultrasonication: *J Magn and Magn Materials.* 2005;293: 182-186.
29. Zharov VP, Galitovskaya EN, Johnson C, Kelly T. Synergistic enhancement of selective nanophotothermolysis with gold nano clusters: potential for cancer therapy, *Laser Surgical Medicines.* 2005 ;37:219-222.
30. Hashizume H, Baluk P, Morikawa S, McLean JW, Thurston G, Roberge S, *et al.* Openings between defective endothelial cells explain tumor vessel leakiness. *Am J Pathol.* 2000; 156:1363-1380.
