IJPSR (2012), Vol. 3, Issue 09





INTERNATIONAL JOURNAL OF PHARMACEUTICAL SCIENCES AND RESEARCH



Received on 29 May, 2012; received in revised form 18 July, 2012; accepted 29 August, 2012

ANTI-GLYCATION EFFECT OF WHITTON ROOT (EULOPHIA NUDA) IN-VITRO CONDITION

Dinesh Prasad Yadav*1, Ram Charan Chhipa 1 and Balvinder Singh 2

Department of Biochemistry, Suresh Gyan Vihar University ¹, Mahal Jagatpura, Jaipur, Rajasthan, India Faculty of Science, Swamy Vivekanand Mahavidyalaya ², Sahajahanpur, Uttar Pradesh, India

Keywords:

AGEs, Plant extract, TCA treatment, IgG Glycation

Correspondence to Author:

Dinesh Prasad Yadav

Department of Biochemistry, Suresh Gyan Vihar University, Mahal Jagatpura, Jaipur, Rajasthan, India

E-mail: ddineshh2004@gmail.com

ABBREVIATIONS:

AGEs; Advanced Glycation End- Products , NIDDM; Non insulin dependent diabetes mellitus, BSA; Bovine serum albumin, PBS; Phosphate buffer saline; TCA, Tri-chloro acetic acid, PF; Plant Filtrate

ABSTRACT

Non enzymatic glycation takes place when elevated levels of reduced sugars react with amino groups of proteins and is called as advanced glycation end products (AGEs) are responsible for Diabetes Mellitus. Hydroalcoholic extract of Whitton root (Eulophia nuda) was tested for in-vitro inhibition of nonenzymatic glycation of Immunoglobin G. Plant extracts have their own importance and now being studied extensively due to having little or no side effects in all aspects of life sciences from botany to medicine in biochemical research. In present study Whitton root was selected and used to check the glycation inhibitory activity in-vitro condition. Various combinations of glucose, protein and Whitton root extracts were made under in vitro conditions and their activity was monitored with Trichloro acetic acid treatment method at 350 nm. Glycated products/ AGEs were more with high glucose and high protein concentration and these were decreased by highest concentration of Whitton root extract i.e. 30 mg/mL or 300 μL. Lower concentrations of plant extract produced either no or least response against advanced glycation end products (AGEs).

INTRODUCTION: Nonenzymatic glycation of protein by reducing sugars is a complicated pathway of various modifications that lead to formation of advanced glycation end products (AGEs). Stable Amadori products are formed by organic reactions with the Schiff base which will further undergo a series of reactions through dicarbonyl intermediates to form AGEs. It is also assumed to be formed by sequential glycation and oxidation reactions called glycoxidation. The accumulation of the reaction products of protein glycation in living organisms leads to structural and functional modifications of tissue proteins ¹.

The role of Advanced glycated end-product proved to be significantly responsible in diabetic microvascular complications of retinopathy, neuropathy and nephropathy as well as their role in the accelerated hyperlipidemia ². There is evidence that glycosylation leads to biochemical modification of Immunoglobulin, and other macromolecules and is cause for pathogenesis of diabetic complications ³.

Elevated glucose level is etiologic agent for the pathogenesis of diabetic complications by increasing protein glycation and formation of advanced glycation end products (AGEs).



Even in the absence of Diabetes Mellitus the formation of AGEs progressively increases with normal aging. However, they are formed at more paced levels in diabetes. AGEs are not only markers but also important causative factors for the pathogenesis of diabetes, Hyperlipidemia, ketoacidosis, cataract, diabetic nephropathy, and neuropathy.

Thus, the design and discovery of inhibitors of AGEs formation is proved to be a boon for the prevention of diabetic or other pathogenic complications. (4) Aminoguanidine, a hydrazine-like small molecule, is the first AGEs inhibitor explored in clinical trials. However, the drug was not used commercially due to its side effects in phase III clinical trials in patients associated with diabetes, although related to some extent to the confiscation of pyridoxal, resulting in vitamin B_6 deficiency 5 .

Process of treatment of purified human IgG with glucose induced its glycation is time and pH dependent. Kenedy *et al.* 1994 ⁶ proved that the level of glycated IgG, IgM, and IgA increased in diabetic patients.

In 1912, Louis Camille Maillard explained the browning of proteins in food and called it as Maillard reaction. This is also known as non-enzymatic glycation of proteins, or a process which links chronic hyperglycemia to a series of physiopathological alterations considered important in the development of chronic complications of different diseases like diabetes and hyperlipidemia ⁷.

The further rearrangement of these glycated proteins give rise to a stable Amadori product that degrades into a variety of compounds which, more reactive than the sugars from which they are derived ⁸. These propagators again form yellow-brown, often fluorescent (some are non fluorescent), irreversible compounds, usually called Advanced Glycation End-Products (AGEs) or Maillard products. Candidate active AGE compounds include *N*-(carboxymethyl)-Llysine (CML) pyrraline, pentosidine and their crosslink's ⁹.

Medicinal plants have been used for decades to treat disease in subcontinent. The vast use of herbal remedies and healthcare preparations, described in ancient Holy texts gave evidence for the occurrence of natural products with medicinal properties. In fact,

wild plants produce a multi range of bioactive molecules, making them enriched source of various types of medicines ¹⁰. Higher plants, as sources of medicinal compounds, have continued to play a significant role in the maintenance of human health since ancient times ¹¹.

Over 50% of all modern clinical drugs are of natural product origin ¹² and natural products play an important role in drug designing programs in the pharmaceutical industry ¹³.

It is known that medicinal plants have little or no side effects. Metformin is the only ethical drug approved for the treatment of non insulin dependent diabetes mellitus (NIDDM) patients, ¹⁴ which is derived from a medicinal plant *Galega officialis* and historically used for treatment of diabetes ¹⁵. There are many anti-diabetic plants, which might provide useful sources for the development of drugs, in the treatment of diabetes mellitus. The literature on medicinal plants with hypoglycemic activity is vast, as many of these plants were used for many centuries and sometimes as regular constituents of the diet.

Synthetic inhibitors and inhibitors from plant extracts have their own importance and now are studied extensively. There are reports of some natural substances isolated from plants with AGE-inhibitory effects. One such compound is curcumin isolated from *Curcuma longa* (Turmeric), commonly known as Haldi. Ginger (*Zingiber officinale* Rosc.) is another spice useful for diabetic therapy ¹⁶. When type 2 diabetic rats are fed ginger, they show hypoglycemic activity, thus improving their diabetic condition. ⁽¹⁷.

Now it is the need of time to develop some new compounds either from plants or synthetically to control diabetes and other age accelerating diseases. As plants have fewer side effects so these should be preferred to study.

In this study, Whitton root (*Eulophia nuda*) extract were used to study its effect on glycation and Maillard products. The major objective of this study was to investigate the effect of Whitton root (*Eulophia nuda*) as inhibitor of AGE or Maillard reaction under *in vitro* conditions and measure its activity against AGE production or inhibition.

MATERIALS AND METHODS:

Preparation of Whitton root (*Eulophia nuda*) extract: Dried and ground Whitton root rhizomes and fruit (10g) was extracted by hydroalcoholic method with 60 mL of 50% ethanol at 37°C for 10 days and then filtered and stored at 4°C for further experiment.

Sample Recovery: Rotary Evaporator was used for recovery of sample by ethanol evaporation. Obtained dried and ground sample was dissolved in 25 mL of phosphate buffer saline and stored at 4°C for further use.

In vitro glycation inhibition with Whitton root (Eulophia nuda) extract:

Conditions and concentrations selection for Whitton root (*Eulophia nuda*) extract: Glycation inhibition with Whitton root extract was measured with different concentrations of protein (BSA) and glucose (two of each) and three concentrations or volumes of Whitton root extract. These are given in **table 1**.

TABLE 1: CONCENTRATION OF DIFFERENT COMPONENTS USED TO STUDY GLYCATION INHIBITION UNDER *IN VITRO* CONDITIONS WITH WHITTON ROOT (*EULOPHIA NUDA*) EXTRACT

S. No.	Components of reaction	Concentration
1	B: Buffer (PBS)	0.1 <i>M</i>
2	P1 : Protein (BSA)	15 mg/Ml
3	P2 : Protein (BSA)	7.5 mg/mL
4	G1 : Glucose	25.0 m <i>M</i>
5	G2 : Glucose	5.5 m <i>M</i>
6	PF1 : Plant Filtrate	300 μL
7	PF2 : Plant Filtrate	150 μL
8	PF3 : Plant Filtrate	75 μL

*In vitro g*lycation of BSA with Whitton root (*Eulophia nuda*) extract: Glucose, BSA with or without inhibitor (plant extracts in PBS pH 7.4) were prepared and their mixture was incubated at 37°C and 50°C for 5 weeks. During this, samples were drawn for glycation inhibition activity after 1st, 3rd and 5th week of incubation. The samples kept at 4°C until analysis.

Trichloracetic acid (TCA) method for Maillard reaction inhibitory activity of Whitton root (*Eulophia nuda*) extract: Matsuura *et al*, 2002, ¹⁸ described this method and known to be as TCA treatment which was followed with some modification.

RESULTS AND DISCUSSIONS: Whitton root rhizomes and fruit were dried in hot air oven at 37°C. After drying, ground to powder form and kept in 50% ethanol for extraction. As it was cheaper and less time consuming, Rotary Evaporator was used to recover the sample. After 1st, 3rd and 5th week of incubation samples were drawn for glycation inhibition activity.

Effect of Whitton root extraction on Maillard Reaction:

TABLE 2: DIFFERENT COMBINATIONS OF REACTION

''	TABLE 2: DIFFERENT CONDINATIONS OF REACTION		
	S. No.	Different Combinations of reaction	
	1	G1+P1	
	2	G1+P1+ PF1	
	3	G1+P1 + PF2	
	4	G1+P1 + PF3	
	5	G1+P2	
	6	G1+P2+ PF1	
	7	G1+P2 + PF2	
	8	G1+P2 + PF3	
	9	G2+P1	
	10	G2+P1+ PF1	
	11	G2+P1 + PF2	
	12	G2+P1 + PF3	
	13	G2 + P2	
	14	G2+P2+ PF1	
	15	G2+P2 + PF2	
	16	G2+P2 + PF3	

Glucose concentration 25.0 mM and 5.5 mM were selected as it was cleared that 25.0 mM is a hyperglycaemic condition and 5.5 mM is a normal glucose level 19. Bovine serum albumin (BSA) was used as protein for glycation and its concentration was 15 mg/mL and 7.5 mg/mL. Whitton root extracts (300 μ L, 150 μ L and 75 μ L) in phosphate buffer were prepared. Each extract along with glucose and BSA were incubated at 37°C and 50°C for five weeks to monitored glycation and Millard reaction inhibitory activity.

TCA treatment method described by Matsuura *et al.* 2002 was adopted with some modifications. The absorbance change based on Schiff base formation was measured by spectrophotometer at 350 nm. This gave the apparent inhibitory activity. Real inhibitory activity was estimated by subtracting the quenching effect from the apparent inhibitory activity. The results obtained by this method are explained below.

Maillard reaction inhibitory activity by whitton root (*Eulophia nuda*) extract: The results obtained from whitton root (*Eulophia nuda*) extract after TCA treatment at 37°C and 50°C. At temperature 37°C (**Figure 1**), G1 P1 produced more Maillard products 0.1 after 5th week, while G2 P2 produced minimum products 0.03 after 1st week at same temperature. G1 P2 and G2 P1 produce moderate level of products.

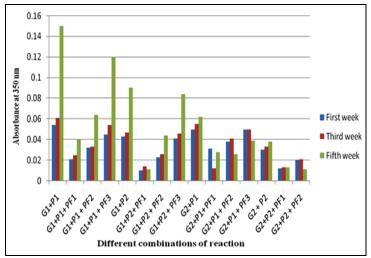


FIGURE 1: MALLARD REACTION INHIBITORY ACTIVITY BY WITH WHITTON ROOT (EULOPHIA NUDA) WITH TCA TREATMENT METHOD UNDER IN VITRO CONDITIONS MEASURED AT 37°C TEMPERATURE

Whitton root extract followed the pattern from PF1 to PF3 as PF1 (300 μ L) generated maximum response while PF 3 (75 μ L) generated least or no response at 37°C. BSA (15mg/mL, 7.5mg/mL), glucose (25.0m*M*, 5.5m*M*) with or without whitton root (*Eulophia nuda*) extract (300 μ L, 150 μ L and 75 μ L) in phosphate buffer (0.1M), pH 7.4 and mixture was incubated at 37°C for five weeks.

Samples were drawn for glycation inhibition activity after 1^{st} , 3^{rd} and 5^{th} week of incubation. Absorbance was recorded at 350nm. At 50°C (**Figure 2**) G1 P1 produced more Maillard products (0.10) after 5^{th} week, while G2 P2 produced minimum products 0.034 at same temperature after 1^{st} week of incubation. Here, it was also seen that G2 P2 generated maximum products at 3^{rd} week (0.047) in spite of 5^{th} week at 50° C. PF1 (300 μ L) generated maximum response while PF3 (75 μ L) generated least response at 50° C and minimum inhibitory activity was observed. At both temperatures PF2 (150 μ L) produced moderate response against Maillard products inhibition.

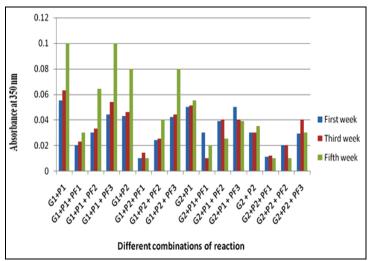


FIGURE 2: MALLARD REACTION INHIBITORY ACTIVITY BY WHITTON ROOT (*EULOPHIA NUDA*) WITH TCA TREATMENT METHOD UNDER *IN VITRO* CONDITIONS MEASURED AT 50°C TEMPERATURE

It was also observed from both figures that high temperature facilitate the production of Maillard products. Moreover, high temperature did not affect the activity of whitton root extract i.e. their trend towards inhibition is same. Onion and garlic have significant blood sugar lowering action. Our results are supported by Demerdash *et al.*2005 ²⁰.

CONCLUSIONS: Our studies concluded that whitton root has ability to inhibit Maillard products that ultimately lead to AGEs production. It was also cleared that 15mg/mL concentration of BSA was more active towards glycation of glucose. However, PF3 (75 μ L) concentration of whitton root extract was almost unable to produce any effect at 37°C. So it is suggested that more concentration of whitton root extract should be used to stop or decrease glycation in human body level in case of diabetes mellitus and persistent hyperglycemia.

ACKNOWLEDGEMENT: Authors are thankful to the management of SGV University, Jaipur for providing necessary facilities.

REFERENCES:

- Takeuchi, M., S. Yamagishi, M. Iwaki, K. Nakamura and T. Imaizumi. Advanced glycation end product (AGE) inhibitors and their therapeutic implications in diseases. *Int. J. Clin. Pharm. Res* (2004);24 (2): 95-101.
- 2. Singh, R., Barden, A., Mori, T., Beilin, L., (2001): Advanced glycation end-products: a review. *Diabetologia.*, 44: 129-46.
- Mendez, J.D.: Advanced glycosylation end products and chronic complications of diabetes mellitus. Gac Med Mex. 2003, 139: 49-55.

- Brwonlee M, Cerami A, Velassara H. Advanced glycosylation end product in tissue and the bicochemical basis of diabetic complications. New Eng Med 1998; 318 (20): 1315-21.
- Brownlee, M.,: Advanced protein glycosylation in diabetes and anging. Annu Rev Med. 1995, 146: 223-234
- Kenedy DM, Skillen AW, Self CH. Glycation of monoclonal antibodies impairs their ability to bind antigen. *ClinExpImmunol* 1994; 98(2):245-51.
- 7. Takeuchi, M., S. Yamagishi, M. Iwaki, K. Nakamura and T. Imaizumi. Advanced glycation end product (AGE) inhibitors and their therapeutic implications in diseases. *Int. J. Clin. Pharm. Res* 2004; 24 (2): 95-101.
- Wautier, J.L. and A.M. Schmidt. Protein Glycation: A Firm Link to Endothelial Cell Dysfunction. *Circulation Res* 2004; 95 (3): 233-245.
- Kaysen, G. The microinflammatory state in uremia: causes and potential consequences. J. Am. Soc. Nephrol 2001; 12:1549– 1557.
- Kala CP, Dhyani PP and Sajwan BS: Developing the medicinal plants sector in Northern India: Challenges and opportunities. *Journal of Ethnobiology and Ethnomedicine* 2, 2006: 32. DOI: 10.1186/1746-4269-2-32
- Farombi EO: African indigenous plants with chemotherapeutic potentials and biotechnological approach to the production of bioactive sporophylactic agents. *African Journal of Biotechecology* 2 (12), 2003: 662-671.ISSN:1684-5315
- Stuffness M and Douros J: Current status of the NCI plant and animal product program. *Journal of Natural Products* 45(1), 1982: 1-14. DOI: 10.1021/np50019a001

- Baker JT, Borris RP, Carté B, Cordell GA, Soejarto DD, Cragg GM, Gupta MP, Iwu MM, Madulid DR and Tyler VE: Natural product drug discovery and development: New perspective on international collaboration. *Journal of Natural Product* 58(9), 1995: 1325-1357.PMID:7494142.
- 14. Beisswenger, P.J., S.K. Howell, A.D. Touchette, L. Sundeep and B.S. Szwergold. Metformin Reduces Systemic Methylglyoxal Levels in Type 2 Diabetes. *Diabetes*, 1999; 48:198–202.
- Oubre, A.Y., T.J. Carlson, S.R. King and G.M. Reaven. From plant to patient, an Ethanomedical approach to the identification of new drugs for the treatment of NIDDM. *Diabetologia*, 1970; 40(5): 614-617.
- 16. Broadhurst, C.L., M.M. Polansky and R.A. Anderson. Insulin-like biological activity of culinary and medicinal plant aqueous extracts *in vitro*. *J. Agric. Food Chem.* 2000, 48(3):849-852.
- Kar, A., B.K. Choudhary and N.G. Bandyopadhyay. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. *J. Ethno-pharmacol.* 2003, 84 (1):105-108.
- 18. Matsuura, N., T. Aradate, C. Sasaki, H. Kojima, M. Ohara, J. Hasegawa and M. Ubukata. Screening system for the maillard reaction inhibitor from natural product extracts. *J. Health Sci.* 2002, 48(6): 520-526.
- Rahbar, S. and J.L. Figarola. Novel inhibitors of advanced glycation end products. *Arch. Biochem. Biophys.* 2003, 419: 63-79.
- Demerdash, F.M., M.I. Yousef and N.I. El-Naga. Biochemical study on the hypoglycemic effects of onion and garlic in alloxan-induced diabetic rats. Food ChemToxicol. 2005, 43(1):57-63.

Yadav D P *, Chhipa R C and Singh B: anti-glycation effect of whitton root (*eulophia nuda*) *in-vitro* condition Valsartan. *Int J Pharm Sci Res* 2012; Vol. 3(8): 3502-3506..