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ACCUMULATION OF ARSENIC IN RADISH (*RAPHANUS SATIVUS* L.), AND THEIR EFFECTS ON GROWTH AND ANTIOXIDANT ACTIVITIES

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ABSTRACT: This study investigated the effects of arsenic on radish in the pot (green house experiment), result revealed that length, FW and DW of plant (EDP and leaf) was significantly ($P < 0.05$) affected at higher concentration of As (≥ 40 ppm) treatment. Chl-a and Chl-b was significantly affected at >40 ppm As treated plant compared with C*. TSS and Proline content in plant showed, increasing trend at increasing concentration of As treated plant. Ca, P and K content in plant was more after 30 DAS followed by 60DAS, P was significantly ($P < 0.05$) more affected from 10 to 80ppm of As treated plant as compared to Ca and K. APX and CAT activities of As treated plant leaf showed increasing trends from C* and DAS. About 70 μg of As /g of DW of EDP accumulate after 60DAS at ≥ 40 to 80ppm, it was more than leaf. As uptake by the EDP was significantly more in 60DAS ($R^2 = 0.810$) than 30DAS ($R^2 = 0.753$) as compared with C*. BCFs was >1 for the As treated plant at >40 and 50ppm after 60DAS.

INTRODUCTION: The consumption of arsenic contaminated water is the main path for its transportation into the environment and biological systems^{5, 33, 34}. Arsenic in the environment is causing significant global health problems which have once again brought this element to the attention of the broad public³³. The United States Environmental Protection Agency (USEPA), the International Agency for Research on Cancer (IARC), and the World Health Organization (WHO) have all identified inorganic arsenic compounds as human carcinogens³³.

As a non-essential element for living organisms, with propensity to adversely affect the human health²³ and the functioning of ecosystems¹⁰, inorganic arsenic calculated as arsenite and arsenate comprise 96% of the total arsenic in vegetables^{19, 20}. In aerobic soils, arsenate (AsV) is the most stable and dominant species and uptake of arsenate by plants has been studied extensively^{18, 19, 20}.

In an anaerobic environment under reducing conditions, arsenite (AsIII) is the dominant species. Both inorganic arsenic species are highly toxic to plants³⁴. Arsenate is a phosphate analog, and therefore it can compete with phosphate in the cytoplasm, replacing phosphate in ATP, leading to the disruption of energy flows in cells^{10, 14, 32}. On the other hand, AsIII is highly toxic to plants because it reacts with sulfhydryl groups (-SH) in enzymes, their cofactors and tissue proteins²⁵.

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<p style="font-size: x-small;">DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.5(8).3536-43</p>	

Translocation from roots to shoots unlike phosphorus (P), arsenic (As) has generally low mobility with respect to translocation from roots to shoots in plants except hyperaccumulators¹². The inefficient root to shoot translocation is also reflected in the generally low ratios of shoot As to root As concentrations in plants supplied with inorganic As in tomato³, Ramirez¹⁴, reported that the arsenic accumulate in vegetables of different families, and arsenic uptake trends were Asteraceae>Brassicaceae>Amaranthaceae> Cucurbitaceae>Liliaceae>Solanaceae> Fabaceae family.

Metal toxicity causes oxidative stress, which can take place possibly by generating reactive oxygen species (ROS)^{9,21, 22}. To scavenge ROS and avoid oxidative damage, plants possess the antioxidative enzymes superoxide dismutase (SOD), catalase (CAT) and peroxidase (POD), glutathione peroxidase, ascorbate peroxidase and glutathione reductase, as well as nonenzyme antioxidants such as ascorbic acid and glutathione^{21, 22, 26}. Ballia district (Central Ganga plain, area: 2981 sq.km) is situated at the eastern part of the Uttar Pradesh¹⁵. The climatic condition of district Ballia is sub humid climate, grassland vegetation and annual rainfall ranged from 983 - 864.8 mm. More than permissible limit of arsenic (10ppb; WHO 2010) were found in shallow in Ballia district and its main region is geogenic problem^{15, 24}. Root vegetables viz radish, carrot, turnip etc. were mostly grown in Ballia district. Such vegetables are might be rich source of arsenic for human being through their food intake and cause several disorder like liver cancer²³.

Therefore, the present study was based on the analysis of arsenic in soil, and their effects on growth, antioxidant activities and uptake in radish plant.

MATERIALS AND METHODS:

Study sites and Set-up of Pot experiment: Fifty soil samples (15 Km apart from each) were collected randomly, from different agricultural field of district Ballia (25.76° N, 84.14° E), UP-India. Chemical analysis of the soil showed that the ranged of organic matter, total N, pH and As concentration were 2 to 4.65%, 0.1 to 1.26%, 6.6 to

8.8 and 0.002 to 145 mg kg⁻¹, respectively. For the pot experiment, surface (0–20 cm) soil samples (Agricultural field: As content 0.002 mg kg⁻¹) which were ground to pass through a 4.0mm mesh and were air-dried, then mixed with basal fertilizers, at ratios of 100 mgNkg⁻¹ dry weight (DW) soil as NH₄NO₃ and 30 mg P kg⁻¹ and 80 mg kg⁻¹ as K₂HPO₄³⁰. Eleven pots were prepared, namely C (control) and treatments (As concentrations: 10, 20, 30,40,50,60,70,80,90 and 100 mg kg⁻¹ DW soil). Each treatment was carried out in six replicates.

The tested topsoil samples were mixed thoroughly with Na₂HAsO₄·7H₂O²² at the above-mentioned concentrations, filled into plastic pots (20 cm in diameter, 15 cm in height, 2.5 kg air-dried soil per pot) and equilibrated for 28 days²². Seeds of radish were surface sterilized in 1% (w/v) sodium hypochloride for 3 min, washed several times with sterilized distilled water (SDW), and soaked in SDW overnight²⁶. Twenty soaked seeds were sowed directly into the 0.8% agar plate and incubated for three days in dark room^{26, 28}. Three similar sizes of sprouted seed were placed in the each treatment pots in the greenhouse with natural light (10–12h; photoperiod) and temperature (16–30°C). The tested soils were watered to reach 60% of the water-holding capacity and this level was maintained by watering daily throughout the experiment. The plants were harvested after 15, 30, 45 and 60 DAS (days after showing) for further analysis.

Physico-chemical analysis of plant: Edible part (EDP), Leaf (L) of the plant were examine for their length, fresh weight (FW), dry biomass (DW) were examined after 15, 30, 45 and 60 DAS. Leaf chlorophyll a (Chl-a) and b (Chl-b) were estimated by spectrophotometrically (A_{663nm} and A_{645nm}) by method¹. The content of glucose, fructose and sucrose was determined by the ferrocyanide method⁶. Proline content (µg mg⁻¹ FW) was determined according to Bates² in the 0.5 gm fresh weight (FW) of leaf tissue at 520nm. Analysis of ions leafs of plant were harvested and dried in an oven at 68°C for 48 h and the dried leaves were ground and digested in an acid mixture (HNO₃:H₂SO₄:HClO₄ = 10:1:3) for determination of K⁺ and Ca⁺ according¹⁶. Leaf phosphate content was measured according⁴.

Total soluble sugars (TSS) were estimated by earlier described method²⁶, 100 mg of fresh leaves were placed in a test tube containing 5 ml 2.5 N HCl, boiled for 3 h then cooled to room temperature. The solution was neutralized with solid sodium carbonate until effervescence stopped. The final volume was made up to 100 ml with distilled water, centrifuged at 1000 g for 10 min and the supernatant (0.5–1.0 ml) collected for TSS analysis using the anthrone method described above.

Antioxidant enzyme activity of the plant: Catalase (CAT; EC 1.11.1.6) and APX activity (EC 1.11.1.11) was analyzed by the method earlier described (26) in fresh leaf tissue (0.5 g) was homogenized with liquid N₂ in QB buffer [100 mm potassium phosphate, pH 7.8, 1 mm EDTA, 1% Triton X-100, 15% glycerol¹¹] to assess ascorbic acid peroxidase (APX) and catalase (CAT) activities. Crude homogenate was centrifuged at 15,000 g for 20 min at 4°C, and the supernatant used for determination of CAT, and APX activity in U_{mg}⁻¹protein. Protein content was determined with the Bradford method using BSA as standard earlier described²⁶.

Plant and soil metal analysis: The plants were immersed in a 0.01M HCl solution to remove any external cadmium and rinsed with deionized (DI) water for 1 min. Subsequently, the plants were separated into three parts: root, stem and leaf. After that, they were dried at 100°C for 10 min, then at 70°C in an oven until completely dry. The plant and soil samples were digested with a solution of 3:1 HNO₃:HClO₄ (v/v).

The concentrations of As were determined using the atomic absorption spectrophotometer by the method³². Bio-concentration factor (BCF) was calculated by earlier report¹⁴, BCF is the ratio of the metal concentration of the edible portion of the vegetable (dry weight) to the metal concentration of the soil [$BCF = MC_{\text{Radish}} / MC_{\text{soil}}$].

Statistical analysis: The data obtained were subjected to ANOVA, and means were compared with Duncan's multiple range test. All statistical analyses were conducted using SPSS (Version 14; IBM, Armonk, NY, USA).

RESULTS: Growth of redish plant was decreasing with increasing concentration of arsenic in pot experiment as compared with control (C*), Length of edible part (EDP) and leaf (L) was significantly ($P < 0.05$) decreasing from As treatment of 40 and 100ppm, 30 to 100ppm respectively after 30DAS, while length of EDP and L was decreasing from at 30 and 10ppm respectively after 60DAS and plant survived upto 90ppm and 80ppm after 30 and 60DAS respectively (**Table 1**). Influence of As concentration on fresh weight of the plant was not significantly ($P < 0.05$) affected upto 40ppm treatment after 30DAS, while after 60DAS significantly effect was observed from 10 to 80ppm (Table 1).

Dry biomass of the plant (EDP and L) was significantly reduced at >20ppm and >10ppm after 30 and 60DAS respectively. Dry biomass reduction in EDP of the plant 5 to 42% and 26 to 56% was observed after 30 and 60DAS respectively, upto 80ppm as compared with control (Table 1). Chlorophyll a (Chl-a) was more as compared with chlorophyll b (Chl-b) in the plant leaf, and Chl-a and Chl-b was not significantly affected upto 40ppm treated plant compared with C* (Table 1). Fructose and sucrose content in EDP of the plant was decrease with increased concentration of As, maximum reduction was observed at >40ppm in both 30 and 60DAS (**Table 2**).

TSS and Proline content was almost not significantly affected up to 40ppm, while increasing trend was observed upto 40ppm for TSS and decreasing trends were >40 to 80ppm. Increasing concentration of proline content was observed for all the As treated plant as compared with C* (table 2). Calcium (Ca), phosphorous (P) and potassium (K) content in plant part (EDP and L) was significantly ($p < 0.05$) affected with increasing As concentration in pot as compared with C* (**Table 3**). Ca, P and K content in plant was more after 30 DAS followed by 60DAS, P was significantly ($P < 0.05$) more affected from 10 to 80ppm As treated plant as compared to Ca and K. Ca and P content was more in EDP as compared with leaf of all the As treated plant and C* after both 30 and 60DAS, while K content was more in leaf then EDP (Table 3).

TABLE 1: INFLUENCE OF ARSENIC ON GROWTH PARAMETERS OF EDIBLE PART (E), LEAF (L) AND LEAF CHLOROPHYLL A & B OF THE RADISH PLANT IN POT-EXPERIMENT (GREEN HOUSE EXPERIMENT) AFTER 30 AND 60 DAYS

Treatment	Length (cm)				Fresh weight (g)				Dry weight (g)				Chl-a		Chl-b	
	30DAS		60DAS		30 DAS		60 DAS		30 DAS		60 DAS		30 D	60 D	30 D	60 D
	E	L	E	L	E	L	E	L	E	L	E	L	L	L	L	L
C*	12 ^a	8 ^a	16 ^a	11 ^a	48 ^a	22 ^a	112 ^a	63 ^a	10.7 ^a	5 ^a	22 ^a	11.2 ^a	54 ^a	67 ^a	35 ^a	43 ^a
10 ppm	11 ^a	7 ^{ab}	16 ^a	9 ^b	47 ^a	25 ^a	92 ^b	43 ^b	10.2 ^a	4.5 ^{ab}	16.2 ^{ab}	8 ^b	54 ^a	58 ^{ab}	34 ^{ab}	41 ^a
20 ppm	10 ^a	7 ^b	16 ^a	9 ^b	48 ^a	28 ^a	67 ^{bc}	41 ^b	9.7 ^a	4.1 ^b	15 ^b	7.6 ^b	51 ^a	50 ^{ab}	22 ^b	32 ^{ab}
30 ppm	9 ^a	6.3 ^c	13 ^b	6.8 ^{bc}	48 ^a	22 ^a	57 ^{bc}	38 ^b	9.0 ^b	4.0 ^b	12.9 ^{bc}	7 ^b	43 ^a	50 ^{ab}	20 ^b	30 ^b
40 ppm	8.2 ^b	5.1 ^{bc}	12 ^b	6 ^c	47 ^a	20 ^a	55 ^{bcd}	37 ^c	8.8 ^b	3.5 ^{bc}	12 ^b	6.2 ^{bc}	36 ^{ab}	47 ^b	21 ^b	21 ^{bc}
50 ppm	7 ^{bc}	5 ^c	11 ^b	5.8 ^{bc}	43 ^b	20 ^a	51 ^{cd}	34 ^{cd}	8.2 ^{ab}	3.1 ^{bc}	11.7 ^{bc}	6 ^c	30 ^b	37 ^{bc}	20 ^b	21 ^{bc}
60 ppm	6.1 ^c	4.2 ^d	9 ^{bc}	6 ^{abc}	37 ^{bc}	17 ^b	44 ^d	37 ^{cd}	6.9 ^{bc}	3.0 ^c	10.3 ^c	5.7 ^{cd}	21 ^{bc}	25 ^c	19 ^{bc}	20 ^{bc}
70 ppm	5 ^d	4.6 ^d	7 ^{bc}	5.3 ^{cd}	32 ^c	13 ^{bc}	40 ^{de}	33 ^{cd}	6.5 ^{bc}	2.9 ^{cd}	10 ^c	5.0 ^d	18 ^c	22 ^{cd}	20 ^{bc}	19 ^{bc}
80 ppm	3.6 ^{de}	4.1 ^{de}	4.2 ^d	4.6 ^d	22 ^{cd}	11 ^c	38 ^e	31 ^d	6.1 ^{cd}	2.8 ^{de}	9.5 ^c	4.1 ^{bcd}	17 ^c	13 ^d	17 ^c	16 ^c
90 ppm	2.3 ^e	3 ^e	ND	ND	17 ^d	10 ^c	ND	ND	5.3 ^d	1.7 ^e	ND	ND	9 ^d	ND	8 ^{cd}	ND
100 ppm	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND

Data are only of 30DAS and 60DAS, Figures followed by different letters in a same line are significantly different at $P < 0.05$, $n = 6$. C* = Control (0 ppm Arsenic in pot), DAS or D = Days after showing, E = Edible part of the Plant, L = Leaf of the plant, Chl-a = Chlorophyll-a and Chl-b = Chlorophyll-b.

TABLE 2: EVALUATION OF FRUCTOSE, SUCROSE, TOTAL SOLUBLE SUGARS AND PROLINE CONTENT OF THE RADISH PLANT PART (EDP OR LEAF) UNDER DIFFERENT CONCENTRATION (PPM) OF ARSENIC IN POT EXPERIMENT, AFTER 30 AND 60 DAS (DAYS AFTER SHOWING)

Treatment	Fructose		Sucrose		Total soluble sugars				Proline ($\mu\text{g/g}$ of FW)			
	30 DAS		60 DAS		30 DAS		60 DAS		30 DAS		60 DAS	
	EDP	Leaf	EDP	Leaf	EDP	Leaf	EDP	Leaf	EDP	Leaf	EDP	Leaf
C*	18±2.1	22±2.3	23±2.5	32±1.7	105±5	86±6	111±11	101±9.1	19±2.1	17±1.5	18±2.1	22±1.6
10 ppm	18±1.4	20±2.3	22±1.4	23±2.5	102±7.1	89±6.1	112±8.5	95±8.6	21±2.0	24±1.7	20±2.0	24±1.3
20 ppm	17±2	20±2.1	22±1.8	22±2.4	107±8	91±5.3	115±9.5	78±7.5	20±1.8	22±2.2	24±2.6	28±2.2
30 ppm	15±1.5	18±1.5	21±2.2	21±1.6	118±12	90±4.5	119±8	72±9	25±3.1	29±2.1	26±2.8	38±2.8
40 ppm	14±1.2	15±1.6	16±2.1	20±1.2	93±5.8	73±3.9	98±10	70±4.6	26±2.8	38±3.2	26±2.2	59±4.2
50 ppm	14±0.9	14±1.2	13±1.5	15±1.4	85±7.1	62±5.2	91±6	53±3.5	27±2.2	39±3.5	28±3.1	61±3.5
60 ppm	12±1.2	12±1.1	12±1	12±1.2	71±2.8	54±6.1	84±4.9	41±4.2	28±2.1	46±3.2	31±2.5	64±4.6
70 ppm	8±0.7	11±1.2	7±0.8	10±1.5	22±3.1	44±4.6	52±5.1	37±1.5	27±1.5	47±2.9	32±2.1	65±3.1
80 ppm	4±0.5	10±1.0	6±1.5	9±1.2	18±1.9	36±5.2	22±3.5	28±3.2	21±1.9	43±3.7	30±1.7	43±2.5
90 ppm	3±0.7	ND	2±0.5	ND	16±2.2	14±2.1	ND	11±0.9	18±1.3	33±2.8	ND	ND
100 ppm	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND

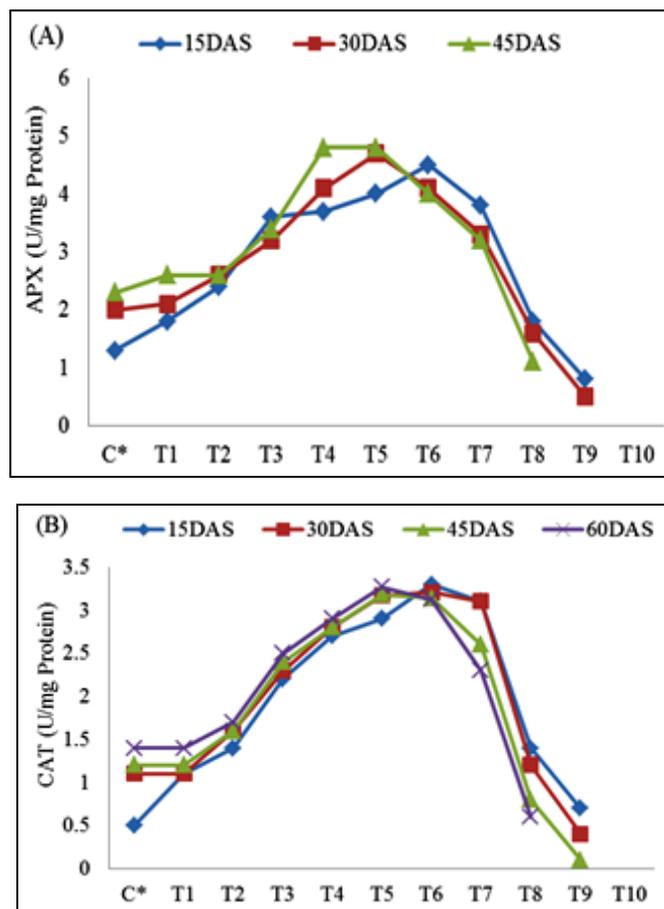
Data are only of 30DAS and 60DAS, data are expressed as mean \pm SD, $P < 0.05$, $n = 6$. C* = Control (0 ppm Arsenic in pot), DAS = Days after showing, EDP = Edible part of the Plant, FW = Fresh weight, ND = Not detected, Fructose = (mg /Kg of FW), sucrose = (mg /Kg of FW) and TSS = (mg /Kg of FW).

TABLE 3: EVALUATION OF CALCIUM, PHOSPHORUS AND POTASSIUM CONTENT OF THE RADISH PLANT PARTS (EDP / LEAF) UNDER DIFFERENT CONCENTRATION (PPM) OF ARSENIC IN POT EXPERIMENT, AFTER 30 AND 60 DAS (DAYS AFTER SHOWING)

Treatment	Ca				P				K			
	30 DAS		60 DAS		30 DAS		60 DAS		30 DAS		60 DAS	
	EDP	Leaf	EDP	Leaf	EDP	Leaf	EDP	Leaf	EDP	Leaf	EDP	Leaf
C*	22±3.1 ^a	13±2.2 ^a	28±2.6 ^a	27±3.5 ^a	22±3.1 ^a	19±1.5 ^b	31±4.2 ^{ab}	21±1.3 ^a	144±25 ^a	148±11 ^a	151±12 ^a	189±23 ^a
10 ppm	18±2.5 ^a	12±1.8 ^a	27±2.1 ^a	20±2.1 ^b	22±2.5 ^b	11±1.1 ^{ab}	21±3.5 ^a	15±1.6 ^b	132±21 ^a	142±24 ^a	141±17 ^b	178±21 ^{ab}
20 ppm	18±1.2 ^a	8±1.2 ^{ab}	21±1.5 ^b	18±1.2 ^{bc}	21±2.1 ^b	10±0.9 ^{bc}	20±2.8 ^a	14±1.2 ^{bc}	131±18 ^b	134±17 ^a	121±12 ^c	164±18 ^b
30 ppm	18±1.5 ^a	7.3±1.4 ^{ab}	18±1.2 ^b	11±1.8 ^c	17±0.9 ^{bc}	9.4±0.7 ^c	16±1.5 ^{bc}	13±2.1 ^{bc}	105±21 ^{bc}	117±10 ^b	111±9 ^{bc}	124±22 ^{ab}
40 ppm	15±1.8 ^{ab}	6.2±1.4 ^{bc}	15±1.6 ^b	9.2±1.5 ^c	15±1.4 ^c	8.1±0.5 ^c	11±1.2 ^c	12±1.6 ^{bc}	83±14 ^c	91±4 ^{bc}	93±11 ^{cd}	82±14 ^{bc}
50 ppm	14±1.0 ^{ab}	5±0.8 ^c	13±1.7 ^b	8.4±1.9 ^c	13±1.2 ^c	8.1±1.2 ^c	10±1.1 ^c	6±0.7 ^{abc}	56±10 ^{dc}	69±3.1 ^c	56±5 ^d	42±2.1 ^{cd}
60 ppm	12±0.9 ^b	4.2±1.1 ^{ac}	10±0.7 ^c	8±1.2 ^c	12±1.1 ^{dc}	8±0.7 ^c	10±1.7 ^c	4.1±0.3 ^d	44±6 ^{dc}	61±4.5 ^{cd}	34±3.1 ^e	39±3.5 ^d
70 ppm	11±1.3 ^c	4.2±0.6 ^d	10±0.8 ^c	6±1.1 ^d	11±0.9 ^c	7±1.1 ^c	9±0.9 ^d	4.1±0.8 ^d	40±2 ^{cd}	52±3.6 ^d	30±2.5 ^{de}	22±1.1 ^{de}
80 ppm	9±1.1 ^d	2.1±0.8 ^{bcd}	6±1.2 ^{bc}	4±0.6 ^{cd}	4±0.7 ^{de}	5.1±1.3 ^d	8±0.5 ^{de}	2.7±0.5 ^e	13±1.3 ^e	19±1.1 ^e	14±1.1 ^{de}	11±0.4 ^e
90 ppm	8±0.6 ^d	1.9±0.4 ^d	ND	ND	2±0.3 ^c	1.8±0.8 ^e	ND	ND	6±0.3 ^e	8±1.2 ^{de}	ND	ND
100 ppm	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND

Data are only of 30DAS and 60DAS, data are expressed as mean \pm SD. Figures followed by different letters in a same line are significantly different at $P < 0.05$, $n = 6$. C* = Control (0 ppm Arsenic in pot), DAS = Days after showing, EDP = Edible part of the Plant and ND = Not detected

APX and CAT activities of As treated plant leaf showed increasing trends from C* to 40ppm and C* to 50ppm respectively after 15, 30, 45 and 60DAS (Figure 1). Maximum APX activity (4.82 U mg protein⁻¹) was observed at 40ppm As treated plant leaf (Figure 1A), while maximum CAT activity 3.31 U mg protein⁻¹) was observed at 50ppm As treated plant leaf after 60DAS (Figure 1B). About 70 µg of arsenic /g of dry weight of EDP accumulate after 60DAS at ≥ 40 to 80ppm As treated plant, more As uptake was observed in EDP than leaf (Figure 2).



ARSENIC TREATMENT IN POT EXPERIMENT

FIGURE 1: ANTIOXIDANT ACTIVITY IN LEAF OF RADISH PLANT UNDER DIFFERENT CONCENTRATION (PPM) OF ARSENIC [C*=no arsenic, T1=10, T2=20 T3=30, T4=40, T5=50, T6=60, T7=70, T8=80, T9=90, T10=100ppm treatment, after 15, 30, 45 and 60 DAS. D or DAS=Days after showing. (A)=APX activity (U/mg protein), (B)=CAT activity (U/mg protein).

As uptake by the EDP was significantly more in 60DAS ($R^2 = 0.810$) than 30DAS ($R^2 = 0.753$) as compared with control. More BCFs (>1) was observed for the plant at 40 (BCF=1.3) and 50ppm (BCF=1.08) after 30DAS, while > 1 BCF were

recorded for all treatment after 60DAS. Maximum BCF=1.73 was recorded at 40ppm treated plant after 60DAS.

DISCUSSION: Radish is the commonly cultivated vegetable crop of the India and it is showing in most of the part of the arsenic infested zone of the Ballia district-Uttar Pradesh. Report of the Central Team¹⁵ on Arsenic mitigation in rural drinking water sources in Ballia (2011) showed, maximum ground water aquifers have ≥ 50 ppb level of arsenic in Ballia district, and ground water is commonly use for drinking as well as irrigation practices in Ballia district²⁴. The main reason for high level of arsenic in Ballia district is geogenic sources²⁴.

Due to irrigation practices in agriculture field, arsenic present in the soil. In the present study, Agricultural field soil samples ($N=50$) showed arsenic level (range 0.002 to 145 mg kg⁻¹) in Ballia district. Presence of arsenic metal in the soil allow to enter in the tropic level and damaging the ecosystem, in this connection the present study, Radish plant were shown in pot augmented with different concentration of arsenic (10 to 100ppm) and result revealed that plant was not survive at > 80ppm As treatment after 60DAS (Table 1), due to metal toxicity^{9,10,14}.

Plant growth was significantly ($P < 0.05$) affected in the pot experiment under increased concentration of arsenic (Table 1) after 60DAS as compared with 30DAS and control. Length, Fresh weight and Dry weight of the treated plant were not statically significant ($P < 0.05$) affected at lower concentration of As after 30DAS, due to tolerance behavior of the plant^{10, 29}, while at higher concentration of As treated plant showed significant reduction in their growth^{10, 29, 31,32}. This highlights that although soil arsenic concentration is an important variable, there are other factors that may influence plant uptake⁸. One is arsenic speciation, which is known to affect plant uptake^{7, 10, 20}. The second influence includes soil characteristics such as pH, organic matter, clay content, water regime and nutrient balance (i.e. phosphate)^{5, 9}. Chlorophyll -b content in treated leaf plant was significantly affected under As treatment as compared with chlorophyll-a (Table 1).

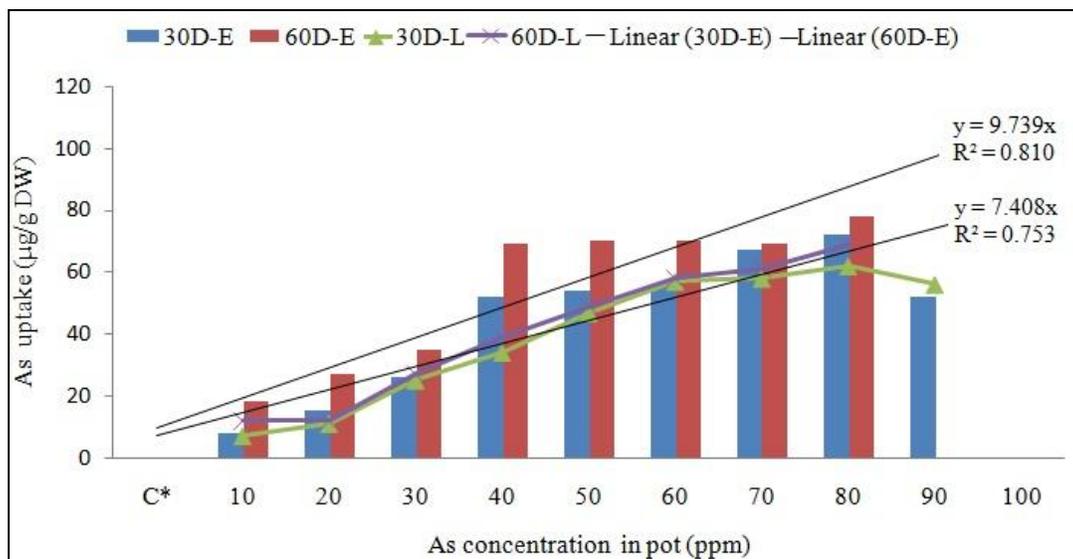


FIGURE 2: Arsenic uptake by the radish plant (Leaf and EDP: Edible part) at different treatment of arsenic concentration in pot after 30 and 60DAS. DAS or D=Days after showing, E=Edible part of the plant and L=leaf of the plant.

In general, under plant stress alter the nutrient imbalance²⁶, chlorophyll content was reduced at increased concentration of ion or metals²⁷. In the present study, Fructose and sucrose content in EDP of the plant was decrease with increased concentration of As, maximum reduction was observed at >40ppm in both 30 and 60DAS (Table 2). TSS and Proline content was almost not significantly affected up to 40ppm, while increasing trend was observed upto 40ppm for TSS and decreasing trends were >40 to 80ppm. Increasing concentration of proline content was observed for all the As treated plant as compared with C* (Table 2). Decreased fructose and sucrose, Increased TSS content of plants under stress is another important defense strategy to cope with stress^{26, 27, 28}.

Proline accumulation in plants is an indicator of general stress tolerance or salinity tolerance as it mediates osmotic adjustment and protects intracellular macromolecules from dehydration and also serves as a hydroxyl radical scavenger²¹. In the present study, calcium, P and K content in plant was more after 30 DAS followed by 60DAS, while P was significantly ($P < 0.05$) more affected from 10 to 80ppm As treated plant as compared to Ca and K. Influence of As in cell leads to nutrient imbalance⁹. It has been shown that plants take up arsenate, the dominant species in oxic environments, via the phosphate transport system since the phosphate ion is similar to the arsenate

ion⁵. Antioxidant activities of the traded plant leaf showed their defense mechanism against reactive oxygen species (ROS), which has been produce under stress condition^{21, 26}. In the present study, APX and CAT activities of As treated plant leaf showed increasing trends from C* to 40ppm and C* to 50ppm respectively after 15, 30, 45 and 60DAS (Figure 1). In the present study, about 70 µg of As /g of DW of EDP accumulate after 60DAS at ≥ 40 to 80ppm As treated plant, more As uptake was observed in EDP than leaf (Figure 2).

It has been shown that arsenic in soil is the major source for arsenic uptake by crops (8). Similar reports are available for accumulation of As in rice, beetroot, lettuce, potato and radish etc.^{34, 14, 19} and the amount of As in the edible parts of these vegetables can reach relatively high levels of As, for example more than 30 mg As kg⁻¹ (DW) in radish^{19, 20}, which could pose a problem for heavy consumers of particular vegetables cultivated in As contaminated soils. In wild-type *A. thaliana*, 73 and 2.6% of P and As, respectively, taken up by roots was distributed to the shoots¹².

Generally, arsenic is primarily found in the roots rather than in the shoots of plants. Rofkar & Dwyer,¹⁷ reported more than 15 times higher As concentration in *Spartina pectinata*, and 28-75 times higher As concentration in rice were found in the roots than in the shoots (13). In the present study, As uptake by the EDP was significantly

more in 60DAS ($R^2 = 0.810$) than 30DAS ($R^2 = 0.753$) as compared with control. The value of $R^2 \approx 1$ indicate the linear relation of As uptake in plant and As concentration in soil, result revealed (Figure 2) maximum linear correlation found in the case of maturity of the plant in EDP. More BCFs (>1) was observed for the plant at 40 (BCF=1.3) and 50ppm (BCF=1.08) after 30DAS, while > 1 BCF were recorded for all treatment after 60DAS. Maximum BCF=1.73 was recorded at 40ppm treated plant after 60DAS. Plants with BCF values ≥ 1 are often classified as hyperaccumulators²⁹. The Asteraceae and Brassicaceae families have been identified to have members classified as hyperaccumulator plants²⁹.

Therefore, the result revealed from BCF calculation in radish plant act as hyperaccumulator for arsenic after their maturity. Hence, arsenic reaches to the human being through trophic level beyond the recommended permissible level.

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REFERENCES:

1. Arnon DI: Copper enzymes in isolated chloroplasts. Polyphenoloxidase in *Beta vulgaris*. *Plant Physiology* 1949; 24:1–15.
2. Bates L S, Waldren R, Teare I D: Rapid determination of free proline for water stress studies. *Plant Soil* 1973;39: 205-207.
3. Burlo F, Guijarro I, Carbonell-Barrachina AA, Valero D, Martinez-Sanchez F: Arsenic species: effects on and accumulation by tomato plants. *J Agric Food Chemistry* 1999; 47(3): 1247–53.
4. Cottenie A: Soil and plant testing as a basis of fertilizer recommendations. *FAO Soils Bulletin*. 1980; 38:64–65.
5. Dixon HBF: The biological action of arsenic acids especially as phosphate analogues. *Adv Inorg Chemistry* 1997; 44:191–228.
6. Furnholmen A M, Winefordner JD, Knapp F W, Dennison R A: Potato sugars, quantitative analysis of glucose and fructose in potatoes. *J. Agric. Food Chem.* 1964; 12(2):109.
7. Hoque MA., Hoque MM, Ahmed M: Declining groundwater level and aquifer dewatering in Dhaka metropolitan area, Bangladesh: causes and quantification. *Hydrogeology Journal*. 2007; 15: 1523–1534.
8. Huang RQ, Gao SF, Wang WL, Staunton S, Wang G: Soil arsenic availability and the transfer of soil arsenic to crops in suburban areas in Fujian Province, southeast China. *Sci Total Environ*. 2006; 368:531–41.

9. Kabata-Pendias A, Pendias H: Trace elements in soils and plants. 3rd edition. Florida: CRC Press; 2001. p. 225–33.
10. Meharg AA, Hartley-Whitaker J: Arsenic uptake and metabolism in arsenic resistant and nonresistant plant species. *New Phytologist* 2002; 154:29–43.
11. Ni M, Dehesh K, Tepperman JM, Quail PH: GT-2: in vivo transcriptional activation activity and definition of novel twin DNA binding domains with reciprocal target sequence selectivity. *Plant Cell*. 1996; 8: 1041–1059.
12. Quaghebeur M and Rengel Z: Arsenic uptake, translocation and speciation in *pho1* and *pho2* mutants of *Arabidopsis thaliana*. *Physiol. Plant* 2004;120: 280–286.
13. Rahman M A, Hasegawa H, Rahman M M, Rahman M A, and Miah M A M: Accumulation of arsenic in tissues of rice plant (*Oryza Sativa* L.) and its distributions in fractions of rice grains. *Chemosphere* 2007; 69: 942-948.
14. Ramirez-Andreotta MD, Brusseau ML, Artiola JF, Maier RM: A greenhouse and field-based study to determine the accumulation of arsenic in common homegrown vegetables grown in mining-affected soils. *Science of the Total Environment* 2013; 443: 299–306.
15. Report of the Central Team on Arsenic mitigation in rural drinking water sources in Ballia district, Uttar Pradesh State 14-17 September 2011, http://www.mdws.gov.in/hindi/sites/upload_files/ddwshindi/files/pdf/ballia-finalreport.pdf.
16. Richards LA: Diagnosis and importance of saline and alkali soils. *USDA Agricultural handbook No.60*. Washington, DC, 1954, USA.
17. Rofkar R J and Dwyer D F: Effects of light regime, temperature, and plant age on uptake of arsenic by *Spartina pectinata* and *Carex stricta*. *International Journal of Phytoremediation* 2011; 13:528-537.
18. Singh N, Ma LQ, Srivastava M, Rathinasabapathi B: Metabolic adaptations to arsenic-induced oxidative stress in *Pteris vittata* L. and *Pteris ensiformis* L. *Plant Science* 2006;170: 274–282.
19. Smith E, Juhasz AL, Weber J: Arsenic uptake and speciation in vegetables grown under greenhouse conditions. *Environ Geochem Health* 2009; 31:125–32.
20. Smith PG, Koch I, Reimer KJ: Uptake, transport and transformation of arsenate in radishes (*Raphanus sativus*). *Sci Total Environ* 2008; 390:188–97.
21. Sun RL, Zhou QX, Sun FH, CX Jin: Antioxidative defense and proline/phytochelatin accumulation in a newly discovered Cd-hyperaccumulator, *Solanum nigrum* L. *Environ. Exp. Botany* 2007; 60:468- 476.
22. Sun Y, Zhou Q: Effects of cadmium and arsenic on growth and metal accumulation of Cd-hyperaccumulator *Solanum nigrum* L. Chunyan Diao. *Bioresource Technology* 2008; 99: 1103–1110.
23. Tchounwou PB, Yedjou CG, Dorsey WC: Arsenic trioxide-induced transcriptional activation of stress genes and expression of related proteins in human liver carcinoma cells (HepG2). *Cell Mol Biol (Noisy-le-grand)* 2003;49 (7):1071-79.
24. Tripathi P.K. (2008): DISTRICT BROCHURE OF BALLIA DISTRICT, U.P., (A.A.P.: 2007-2008) http://cgwb.gov.in/District_Profile/UP/Ballia.pdf.
25. Tripathi RD, Srivastava S, Mishra S, Singh N, Tuli R, Gupta DK, Maathuis JM: Arsenic Hazards: Strategies for tolerance and remediation by plants. *Trends in Biotechnology* 2007; 25:158–165.
26. Upadhyay S K, Singh J S, Saxena AK and D P Singh: Impact of PGPR inoculation on growth and antioxidants status of wheat plant under saline condition. *Plant Biology* 2012; 4: 605-611.

27. Upadhyay SK, Ahmad M, Singh A: Ecological impact of weed (*Parthenium hysterophorus* L.) invasion in saline soil. International Journal of Scientific and Research Publications 2013;3(4):1-4.
28. Upadhyay SK, Singh JS, Singh DP: Exopolysaccharide-producing plant growth-promoting rhizobacteria salinity condition. Pedosphere 2011; 21:214–222.
29. Vithanage M, Dabrowska BB, Mukherjee AB, Bhattacharya P: Arsenic uptake by plants and possible phytoremediation applications: A brief overview. Environ Chem Lett 2011; 1–8.
30. Wang L, Zhou Q, Ding L, Y Suna: Effect of cadmium toxicity on nitrogen metabolism in leaves of *Solanum nigrum* L. as a newly found cadmium hyperaccumulator. Journal of Hazardous Materials 2008; 154:818–825.
31. Wei CY, Chen TB: Arsenic accumulation by two brake ferns growing on arsenic mine and their potential in phytoremediation. Chemosphere 2006; 63: 1048–1053.
32. Wei CY, Chen TB: The ecological and chemical characteristics of plants in the areas of high arsenic levels. Acta Phytoecologic Sinica 2002; 26: 695–700 (in Chinese).
33. World Health Organization. Exposure to arsenic: a major public health concern. Available: www.who.int/ipcs/features/arsenic.pdf 2010, [Accessed 19 June 2012].
34. Zhu YG, Williams PN, Meharg AA: Exposure to inorganic arsenic from rice: A global health issue? Environmental Pollution 2008; 154: 169–171.

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