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PHARMACOLOGICAL PERCEPTION OF PEPTIDES FROM MARINE SPONGE: A REVIEW

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ABSTRACT

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Marine sponges, the sessile invertebrates of the *Phylum porifera* are invaluable tool in current research. They remain as a goldmine to chemist and pharmacologist due to its defensive weapons, the secondary metabolites. Endogenous peptides from marine sponges and associated microorganisms are promising lead compound for drug development. Some of the compounds are under clinical trials. These peptides can act against variety of diseases in humans including bacterial, fungal, protozoan, HIV, inflammatory and even tumor. This review focuses on sponge symbiotic association with other organisms, significance of peptides as secondary metabolites and its pharmacological effects by highlighting its role as anti-bacterial, anti- fungal, anti- HIV and anti- tumor agents. Sponge-microbial associations are found to be very specific in the production of particular bioactive compounds.

INTRODUCTION: More than two decades ago marine invertebrates have provided key structure and compounds that proved their potential in several fields, particularly as new therapeutic agent for a variety of diseases. Among marine invertebrates, Porifera (sponges) remain the most prolific phylum, concerning novel pharmacologically active compounds¹.

Pharmaceutical interest in sponges was aroused in the early 1950's by the discovery of a number of unknown nucleosides: spongothymidine and spongouridine in the marine sponge *Cryptotheca crypta*². Sponges can provide potential drugs against many major world-wide occurring diseases. Of the 18,000 marine natural products described, they are responsible for more than 5300 different products and every year hundreds of new compounds are being discovered³. The antitumor natural product patent registrations in recent years over 75% are from sponges⁴.

One among the well-established sector in the research of marine natural products is the search of bioactive peptides from marine sponges. The bioactive peptides are protein fragments which have a positive impact on the functions and conditions of living beings.

However, they are produced only in limited quantities by living species including marine sponges⁵. With respect to the diversity of the secondary metabolites produced marine sponges' remains as 'gold mine' to chemists and found their way in to pharmacological applications.



Marine Sponge Secondary Metabolites: With the variety of species inhabit the world's oceans, the intense concentration of species coexisting in these limited extent habitats necessarily makes them highly competitive and complex⁶. As a result of this intense competition, a high percentage of species have evolved chemical compounds to defend.

These chemical adaptations⁷ generally take the form of so-called "secondary metabolites. Marine invertebrates which are abundant in the Indo-Pacific regions are rich in secondary metabolites and are becoming targets of continuing search for bioactive compounds⁸. Researchers found that sponges have highest diversity of defensive chemical weapons *i.e.*, secondary metabolites^{9, 10} to repel and deter predators¹¹, compete for space with other sessile species¹² and for communication and protection against infection.

Among marine organisms, the largest number of secondary metabolites isolated since 1965 have come from sponges¹³. The chemical diversity of sponge substances is remarkable. Sponge derived compounds are mostly nitrogen-containing ones and also non-nitrogenous compounds¹⁴. In addition to the unusual nucleosides, other classes of substances such as bioactive terpenes, sterols, cyclic peptides, alkaloids, fatty acids, peroxides, and amino acid derivatives (which are frequently halogenated) have been described from sponges or from their associated microorganisms^{15, 16, 17, 18, 19}.

The main biological activities of those sponge metabolites have been cytotoxic and antimicrobial while other activities (anti-inflammatory, immunosuppressive, neurosuppressive, and antifouling activities) have been limited, suggesting the need for an evaluation of anti-cancer and anti-infective agents^{20, 21}. The studies on sponge chemical ecology include three different aspects. First, diversity of chemical compounds produced by sponges; second, potential functions of these metabolites in nature and finally, the strategies for their use for human benefit²².

Sponge Microbial Symbiosis: As marine sponges have been considered as a rich reservoir of bioactive compounds, a conceptual progress occurred focusing on sponges, highlighting the symbiosis of sponge-

associated microorganisms (bacteria and fungi) in the host epibiotic defense²³. Symbionts include archaea, bacteria, cyanobacteria, and microalgae. Bacteria associated with marine sponge *Theonella swinhoei* include unicellular cyanobacteria, unicellular bacteria and filamentous bacteria²⁴. Many sponges contain large amounts of bacteria within their tissues, sometimes occupying 40 to 60% of the total biomass (equivalent to 108 to 1010 bacteria per gram)²⁵.

Therefore, it has often been proposed that associated bacteria might be the actual producers of many sponge-derived natural products²⁶. Fungi associated with marine sponges are also known to produce many bioactive agents²⁷. The members of the class Demospongiae are the richest producer of pharmacologically significant bioactive compounds in association with microbes²⁸.

In some cases, these microorganisms and not sponge cells are the likely source of the secondary metabolites of interest^{29, 30, 31}. It is reported that sponge hosted diverse microorganisms are metabolically very active in their respective host and it was demonstrated by comparing 16S rRNA gene derived sequence³². The host (sponge) synthesizes bioactive compounds that provide protection against attacking microorganisms or eukaryotes, *e.g.*, acetylenic compounds³³. The symbiotic bacteria or fungi produce secondary metabolites that act as antibiotics, *e.g.*, cribrastatin³⁴, or as cytostatic agents, *e.g.* sorbicillactone-A³⁵.

Functionally, these compounds act only as defense molecules. Another functional class of secondary metabolites of sponges and their associated microorganisms play a dual role: they are involved in defense as well as in the activation of pathways crucial for self defense⁹. The occurrence of structurally similar substances in unrelated sponges, particularly those which were otherwise known exclusively from microorganisms, led to the hypothesis that such substances were of microbial origin including some already in drug trials³⁶.

An antibacillus compound, which was chemically identified as the peptide antibiotic andrimid was detected in the extract of the sponge *Hyatella* sp. A bacterial isolate M22-1, belonging to the genus *Vibrio* was also isolated from the homogenate of the same

sponge. The bacterium when cultured in marine agar also produced the same compound. This suggests that the origin of andrimid in the sponge is from the bacterium³⁶. An epibiotic bacterial strain *Pseudoalteromonas maricaloris* KMM 636T, isolated from the Great Barrier Reef sponge *Fascaplysinopsis reticulata* was the source of two brominated chromopeptides such as bromoalterochromide A and bromoalterochromide A. They showed moderate cytotoxicity to the eggs of the sea urchin *Strongylocentrotus intermedius*^{37, 38}. Leucamide A closely resembles the compound albeit, which is found frequently in cyanobacteria.

Scanning electromicrographs of *Leucetta microraphis* revealed the presence of microbial symbionts, including cyanobacteria in the tissue of the sponge *Leucetta microraphis*. The sponge-derived leucamide A might, therefore be produced by cyanobacteria associated with it and not by the invertebrate itself³⁹. The fungus *Aspergillus versicolor*, isolated from *Petrosia* sp. (Jeju Island, Korea) yielded three known polyketides such as decumbenones A, B and versiol, and the cytotoxic lipopeptide fellutamide C³⁰.

Marine Sponge Peptides: Marine sponges are synthesizing a wide variety of peptidic and organic molecules with biological activities. Natural peptides have been invaluable tools for pharmacological and biochemical investigations of a wide range of physiological functions⁴⁰. Cyclic dipeptides (also known as diketopiperazines, DKPs) are known to have antibiotic, antiviral and antitumour properties, and are

a relatively unexplored class of bioactive peptides that may hold great promise for the future⁴¹.

Lithistid sponges are renowned among marine organisms for their ability to produce a diverse array of biologically active metabolites⁴², including novel peptides characterized by a high proportion of D and/or *N*-methylated amino acids. The similarity between lithistid peptides and those from microorganisms leads to the speculation that lithistid peptides might arise from symbiotic microbes⁴³.

Polytheonamide-B (pTB), a highly cytotoxic polypeptide, is one of the most unusual nonribosomal peptides from sponge origin. pTB is a linear 48-residue peptide with alternating D- and L- amino acids and contains a total of eight type of non proteinogenic amino acids. The strong cytotoxicity can be ascribed to its ability to form single molecule channels through biological membranes⁴⁴. Corticiamide A(1) and cyclocinamide B(2) represent the first peptides to be described from the genus *Corticium*⁴⁵.

A strain of *Penicillium brevicompactum* derived from the specimen of *Petrosia ficiformis* provided two new cyclopentadepsipeptides, petrosifungins A and B along with the known fungal metabolites brevianamide A, mycophenolic acid and asperphenamate. Since cyclodepsipeptides constitute new class of potential drugs, petrosifungins A and B, may serve as lead compounds for more pharmacologically potent and toxicologically safe derivatives⁴⁶.

TABLE 1: MARINE SPONGE DERIVED PEPTIDES

Substance	Class	Organism	Reference
Haliclonamides C, D and E	peptides	<i>Haliclona</i> genus	Sera et al., ⁴⁷
cyclo-(glycyl-L-prolyl-L- glutamyl)	cyclic peptides	<i>Suberites domuncula</i>	Mitova et al., ⁴⁸
mapacalcine; Mr 5 19,064 vastifica	dimeric peptide	<i>Cliona</i>	Morel et al., ⁴⁹
microsclerodermins C – E(3-4)	cyclic peptides	<i>Theonella</i> sp	Schmidt and Faulkner ⁵⁰
Koshikamide A1	linear peptide	<i>Theonella</i> sp	Fusetani et al., ⁵¹
Euryjanicin A(1)	cycloheptapeptide	<i>Prosuberites</i>	Vicente et al., ⁵²
Phakellistatins 15–18 (2–5), Phakellistatins 13 (1), hymenistatin 1 and hymenamides G, H, and J	cyclopeptides	<i>Phakellia fusca</i>	Zhang et al., ⁵³
stylisin 1(1) and stylisin 2 (2), phakellistatin 13 (3)	Cyclic heptapeptides	<i>Stylissa caribica</i>	Mohammed et al., ⁵⁴
stylissamides E (3) and F (4)	Cyclic heptapeptides	<i>Stylissa caribica</i>	Cychon and Kock, ⁵⁵
corticiamide A (1) and cyclocinamide B(2)	cyclic peptide	<i>Corticium</i> sp	Laird et al., ⁴⁵

Anti-Bacterial Peptides: In the year 2007 alone 961 new compounds were described from marine microorganisms reflecting an increase of 24% compared with the number of compounds reported for 2006. Discodermidins A-D have been isolated from *Discodermia kiiensis*. were proposed as antibacterial agents and two new cyclic peptides, cyclo-(glycyl-L-seryl-L-prolyl-L-glutamyl) and cyclo-(glycyl-L-prolyl-L-glutamyl), have been isolated from the cell extract of a *Ruegeria* strain associated with cell cultures of the sponge *Suberites domuncula* showed moderate activity against *B. subtilis*, with an MIC of 25 µg/ml and 50 µg/ml, respectively⁴⁸.

Cyclic lipopeptides are produced by different groups of bacteria, both Gram-positive and Gram-negative⁵⁵ and represent a unique class of bioactive microbial secondary metabolites⁵⁶. The antibacterial peptide activity from the marine sponge *Clathria Indica* among the various strains maximum diameter of (12 mm) zone of inhibition was recorded in *Escheirchia coli* strain and minimum zone of inhibition of (6mm) was observed in *Salmonella typhi*, *Proteus mirabilis* and *Lactobacillus vulgaris* strain⁵⁷.

On the basis of TLC observations on further confirmations with ¹H NMR peptide presented in between (6-8ppm fractions) subjected to studies by using chromatography technique. This resulted in the identification of antimicrobial peptide⁵⁷.

Two cyclic thiopeptides containing thiazole and pyridine moieties, and several unusual amino acids, were obtained from a culture of *B. cereus* isolated from the marine sponge *Halichondria japonica*, which exhibited potent antibacterial activities against *Staphylococcus* and *Enterococcus* and were active against multiple drug resistant strains. These thiopeptides were only inactive against Gram-negative bacteria^{58, 59}.

The most promising antimicrobial substances appear to be 2-undecyl- 4-quinolone, DKPs, lipopeptides (surfactins, iturins and fengycins), manzamine-A, organo halogen (2,4,4'-trichloro- 2'-hydroxydiphenyl ether), phenazine, pyrone I, rifamycins and thio peptides. Among these substances, manzamine-A produced by an actinomycete appear to be great promise for the future.

Anti-Fungal Peptides: Two other peptides, discobahamin A and B, isolated from the Bahamian deep water marine sponge *Discodermia* sp., were evaluated as inhibitors of the growth of *C. albicans*⁶⁰. The depsipeptides halicylindramides A-C, D and E^{61, 62}, with antifungal and cytotoxic properties (against P388), were obtained from the Japanese marine sponge *Halichondria cylindrata*. Halicylindramides A-C are tetradecapeptides with an N-terminus blocked by a formyl group and the C-terminus lactonized with a threonine residue. Halicylindramide D is a tridecapeptide also with antifungal and cytotoxic properties, while halicylindramide E is a truncated linear peptide with a C-terminal amide.

Other antifungal cyclic peptides from sponges are the aciculitins A-C⁶³ and the theonegramide⁶⁴. The marine sponge *Theonella swinhoei* from Palau contains the bicyclic glycopeptide antifungal compound theopalauamide⁶⁵. Another example is the Haligramides-A and B two new cytotoxic from hexapeptides from the sponge *Haliclona nigra*⁶⁶.

Anti- HIV Peptides: HIV-inhibitory peptides from sponges constitute a recent discovery. HIV-inhibitory peptides from sponges constitute a recent discovery. The cyclic depsipeptides papuamides A-D⁶⁷ isolated from sponges of the genus *Theonella* contain a number of unusual amino acids and are also the first marine derived peptides reported to contain 3- hydroxyleucine and homoproline residues, and 2, 3- dihydroxy-2, 6, 8-trimethyldeca-(4Z, 6E)-dienoic acid moiety N-linked to a terminal glycine residue.

Papuamides A and B inhibited the infection of human T-lymphoblastoid cells by HIV-1 sub (RF) *in vitro* with a 50% effective concentration (EC50) of approximately 4 ng/ml. Another anti-HIV candidate from the sponge *Sidonops microspinosa* is the microspinosamide⁶⁸, a cyclic depsipeptide incorporating 13 amino acid residues and the first naturally occurring peptide containing a beta- hydroxy-*p*-bromophenylalanine residue. Microspinosamide inhibited the cytopathic effect of HIV-1 infection in an XTT-based *in vitro* assay.

A new sulfated cyclic depsipeptide, termed mutremdamide A, and six new highly *N*-methylated peptides, termed koshikamides C-H, were isolated from different deep-water specimens of *Theonella*

swinhoi and *Theonella cupola*. Mutremdamide A displays a rare 2-amino-3-(2-hydroxyphenyl)propanoic acid and a new *N*δ-carbamoyl-β-sulfated asparagine. Koshikamides C–E are linear undecapeptides, and koshikamides F–H are 17-residue depsipeptides containing a 10-residue macrolactone. Koshikamides F and G differ from B and H in part by the presence of the conjugated unit 2-(3-amino-5-oxopyrrolidin-2-ylidene)propanoic acid. Cyclic koshikamides F and H inhibited HIV-1 entry at low micromolar concentrations while their linear counterparts were inactive⁶⁹. The origin and role of bioactive peptides inside the sponges in many cases is unclear. Several of these substances possess a great potential for drug development, but none has given origin to a commercial medication so far.

Six new depsipeptides belonging to two different structural classes, termed celebesides A–C and theopapuamides B–D, have been isolated from the marine sponge *Siliquariaspongia mirabilis*. Celebeside A neutralized HIV-1 in a single-round infectivity assay with an IC₅₀ value of 1.9±0.4 μg/mL while the nonphosphorylated analog celebeside C was inactive at concentrations as high as 50 μg/mL⁶⁹. HIV cyclodepsipeptide, homophymine A, was isolated from a New Caledonian collection of the marine sponge *Homophymia* sp. Homophymine A contains 11 amino acid residues and an amide-linked 3-hydroxy-2,4,6-trimethyloctanoic acid moiety. In a cell-based XTT assay, homophymine A exhibited cytoprotective activity against HIV-1 infection with a IC₅₀ of 75 nM⁷⁰.

Anti- Tumor Peptides: Koshikamide A2 (2) was isolated as a cytotoxic metabolite from a marine sponge of *Theonella* sp. Its structure was elucidated to be a linear undecapeptide by spectroscopic and chemical methods, together with enzymatic conversion to known koshikamide A1 (1). The new peptide moderately inhibited the growth of P388 murine leukemia cells⁷¹.

Koshikamide B (1) has been isolated from two separate collections of the marine sponge *Theonella* sp. as the major cytotoxic constituent. Koshikamide B is a 17-residue peptide lactone composed of six proteinogenic amino acids, two d-isomers of proteinogenic amino acids, seven *N*-methylated amino acids, and two unusual amino acid residues.

It exhibits cytotoxicity against P388 murine leukemia cells and the human colon tumor (HCT-116) cell line with an IC₅₀ value of 0.45 and 7.5 μg/mL, respectively⁷².

Nonribosomal cyclic peptide leucamide A was isolated from the sponge *Leucetta microraphis*, obtained from the Great Barrier Reef of Australia. The compound was found to inhibit the growth of three tumor cell lines (stomach carcinoma, liver carcinoma and liver carcinoma with mutated p53)³².

Keramamides B–D as well as Orbiculamide A, isolated from sponges of the genus *Theonella*⁷³ were cytotoxic against P388 murine leukemia cells (IC₅₀ = 4.7 ng/ml).

Pakellistatin 12 (1) is a new cancer cell growth inhibitory (P388 ED₅₀ 2.8 μg/mL) cyclodecapeptide that was isolated from a marine sponge *Phakellia* sp⁷⁴. Haliclona sp a marine sponge yielded Kendarimide A a novel peptide which reversed glycoprotein mediated multi drug resistance in tumor cells. HTI-286⁶¹, a synthetic analog of the tripeptide, hemiasterlin, originally isolated from the South African sponge *Hemiasterella minor*, depolymerizes microtubules and blocks cell growth. HTI-286 has also shown antitumor activity in human tumor xenograft murine models²¹.

Papuamides are representatives of a class of marine sponge derived cyclic depsipeptides, including callipeltin A, celebesides A and B, homophymine A, mirabamides, microspinosamide, neamphamide A and theopapuamides. They are thought to have cytoprotective activity against HIV-1 *in vitro* by inhibiting viral entry. Jasplakinolide, a representative member of marine sponge-derived cyclodepsipeptides that include arenastatin A, geodiamolides, homophymines, spongidepsin and theopapuamides, is a potent inducer of actin polymerization *in vitro*.

Although actin dynamics is essential for tumor metastasis, no actin targeting drugs have been used in clinical trials due to their severe cytotoxicity. Nonetheless, the actin cytoskeleton remains a potential target for anticancer drug development. These features imply the use of cyclodepsipeptides as molecular models in drug research⁷⁵.

CONCLUSION: Peptide compounds analysed here are obtained from very different marine organism of sponge exhibiting different chemicals and displaying the large variety of pharmacological effects of specific targets. The marine world has become an important source of therapeutic agents with novel mechanisms of action. A multidisciplinary and cooperative effort with the use of more sensitive and fast methods in the analysis of the structure of peptides, e.g. exact description of the molecular weight and the sequence, as well as in the pharmacological evaluation, will speed up drastically the discovery of novel active peptides from marine sources.

From another side, these compounds seem to be very useful and promising for biomedical research to clarify many normal and pathological mechanism of action in the human body as well as in the design of very specific and potent new pharmaceuticals for a wide variety of diseases. The discovery of the bio-regulatory role of different endogenous peptides in the marine sponges as well as the understanding of the molecular mechanisms of action as anti-bacterial, antifungal, anti-HIV and anti-tumor contributed to consider peptides also as promising lead drug candidates.

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REFERENCES:

- Faulkner DJ: Marine natural products. *Nat. Prod. Rep* 2000; 17: 7-55.
- Bergmann W and Feeney RJ: The isolation of a new thymine pentoside from sponges. *J. Am. Chem. Soc* 1950; 72: 2809-2810.
- Faulkner DJ. Marine natural products. *Nat. Prod. Rep.*2002; 19: 1-48.
- Sipkema D, Franssen MCR, Osinga R, Tramper J and Wijffels RH: Marine sponges as pharmacy. *Mar. Biotechnol* 2005; 7: 142-162.
- Joseph B and Nair VM: Marine Sponge Database (MSD): A Database on Marine Sponges in Kanyakumari District. *International Journal of Soft Computing and Bioinformatics* 2011; 2(2): 63- 68.
- Simmons TL, Andrianasolo E, McPhail K, Flatt P and Gerwick WH: Marine natural products as anticancer drugs. *Mol Cancer Ther* 2005; 4(2): 333-342.
- Firn RD and Jones CG: Natural products: a simple model to explain chemical diversity. *Nat Prod Rep* 2003; 20: 382-91.
- Sabdono A and Radjasa OK: Microbial symbionts in marine sponges: Marine natural product factory. *J. Coast. Dev* 2008; 11- 57-61.
- Muller WEG, Schroder HC, Wiens M, Perovic- Ottstadt S, Batel R and Muller IM: Traditional and Modern Biomedical Prospecting: Part II—the Benefits Approaches for a Sustainable Exploitation of Biodiversity (Secondary Metabolites and Biomaterials from Sponges), *Advance Access Pub* 2004; 1(2): 133-144.
- Thakur NL and Muller WEG: Biotechnological potential of marine sponges. *Current science* 2004; 86(11): 1506- 1512.
- Pawlik JR, McFall G and Zea Z: Does the odor from sponges of the genus *Ircinia* protect them from fish predators? *J. Chem. Ecol* 2002; 28: 1103- 1115.
- Becerro MA, Turon X and Uriz MJ: Multiple functions for secondary metabolites in encrusting marine invertebrates. *J. chem.. ecol* 1997; 23: 1527- 1547
- Belarbi EH, Gomez AC, Chisti Y, Camacho FG and Grima EM: Producing Drugs from Marine sponges. *Biotechnol. Adv* 2003; 21: 585- 598
- Sipkema D, Osinga R, Schatton W, Mendola D, Tramper J and Wijffels RH: Large-scale production of pharmaceuticals by marine sponges: sea, cell, or synthesis *Biotechnol. Bioeng* 2005; 90: 201-222.
- Donia M and Hamann M: Marine natural products and their potential applications as anti-infective agents. *Lancet Infect. Dis* 2003; 3: 338-48.
- Matsunaga S and Fusetani N: Nonribosomal Peptides from Marine Sponges. *Curr. Org. Chem* 2003; 7: 945- 966.
- Keyzers RA and Davies-Coleman MT: Anti-inflammatory metabolites from marine sponges. *Chem. Soc. Rev* 2005; 34: 355-365.
- Moore BS: Biosynthesis of Marine Natural Products: Macroorganisms (Part B). *Nat. Prod. Rep* 2006 23, 615 – 629.
- Piel J: "Bacterial symbionts: prospects for the sustainable production of invertebrate-derived pharmaceuticals". *Curr. Med. Chem* 2006; 13: 39-50.
- Mayer AMS and Gustafson KR: Marine pharmacology in 2000: Antitumor and Cytotoxic compounds. *International Journal of Cancer* 2003; 105: 291-299.
- Newman D and Cragg GM: Marine natural products and related compounds in clinical and advanced preclinical trial. *Journal of Natural Products* 2004; 67: 1216-1238.
- Cimino G and Ghiselin MT: *Marine Chemical Ecology* (eds McClintock, J. B. and Baker, B. J.), CRC Press, Boca Raton, 2001: 115-154.
- Thakur NL, Hentschel U, Krasko A, Anil AC, Müller WEG: Antibacterial activity of the sponge *Suberites domuncula* and its primmorphs: potential basis for chemical defense. *Aqua. Microbiol. Ecol* 2003; 31: 77-83.
- Bewley CA and Faulkner DJ: Lithistid sponges: Star performers or hosts to the stars. *Angew. Chem. Int. Ed.* 1998; 37: 2162-2178.
- Hentschel U, Hopke J, Horn M, Friedrich AB, Wagner M, Hacker J and Moore BS. Molecular Evidence for a Uniform Microbial Community in Sponges from Different Oceans. *Appl. Environ. Microbiol* 2002; 68: 4431- 4440.
- Piel J: "Metabolites from symbiotic bacteria" *Nat. Prod. Rep* 2004; 21: 519-538
- Holler U, Wright AD, Matthee GF, König GM, Draeger S, Aust HJ and Schulz B: Fungi from marine sponges: diversity, biological activity and secondary metabolites. *Mycol Res* 2000; 104:1354 – 65.

28. Tresa RAT, Devanand PK, Ponnappakkam A and LokaBharathi: Marine Drugs from Sponge-Microbe Association—A Review *Mar. Drugs* 2010; 8: 1417-1468.
29. Bewley CA, He H, Williams DH and Faulkner DJ: Anti-fungal cyclic peptides from sponges *Microscleroderma* sp. *J. Am. Chem. Soc* 1996; 118: 4314-4321.
30. Lee YM, Dang HT, Hong J, Lee CO, Bae KS, Kim DK and Jung JH: A Cytotoxic Lipopeptide from the Sponge-Derived Fungus *Aspergillus versicolor*. *Bull. Korean Chem. Soc* 2010; 31: 205–208.
31. Proksch P, Edrada RA and Ebel R: Drugs from the sea—current status and microbiological implications. *Appl Microbiol Biotechnol* 2002; 59:125– 34.
32. Kamke J, Taylor MW and Schmitt S: Activity profiles for marine sponge-associated bacteria obtained by 16S rRNA vs 16S rRNA gene comparisons. *ISME J* 2010; 4: 498–508.
33. Richelle-Maurer E, Gomez R, Braekman JC, van de Vyver G, van Soest RWM and Devijver C. Primary cultures from the marine sponge *Xestospongia muta* (Petrosiidae, Haplosclerida). *J. Biotechnol* 2003; 100:169–76.
34. Pettit GR, Knight JC, Collins JC, Herald DL and Young VG: Antineoplastic agent 430 isolation and structure of cribostatins 3, 4 and 5 from the Republic of Maldives *Cribrochalina* sp. *J. Natl. Prod* 2000; 63: 793–8.
35. Bringmann G and Lang G: Full absolute stereostructures of natural products directly from crude extracts: the HPLC-MS/MS-NMR-CD 'triad'. WEG Müller, editor. *Sponges (Porifera)*. Marine Molecular Biotechnology Berlin: Springer. 2003; 89–116.
36. Oclarit JM, Okada H, Ohta S, Kaminura K, Yamaoka Y, Iizuka T, Mivashiro S and Ikegami S: Anti-bacillus substance in the marine sponge, *Hyatella* species, produced by an associated *Vibrio* species bacterium. *Microbios* 1994; 78: 7- 16.
37. Blunt JW, Copp BR, Munro MHG, Northcote PT and Prinsep MR: Marine natural products. *Nat Prod Rep* 2009; 26: 170–244.
38. Speitling M, Smetanina OF, Kuznetsova TA and Laatsch H: Bromoalterochromides A and A, Unprecedented chromopeptides from a marine *Pseudoalteromonas maricaloris* strain KMM 636T. *J. Antibiot* 2007; 60: 36–42.
39. König GM, Kehraus S, Seibert SF, Abdel-Lateff A and Müller D: Natural products from marine organisms and their associated microbes. *ChemBioChem* 2005; 7: 229–238.
40. Herken H and Hucho F: Purification of a New Dimeric Protein from *Cliona vastifica*. *Handbook of Experimental Pharmacology: Selective Neurotoxicity*. Springer-Verlag, Berlin 1992.
41. Matsunaga S, Fusetani N and Konosu S: Bioactive marine metabolites, IV. Isolation and the amino acid composition of discodermin A, an antimicrobial peptide, from the marine sponge *Discodermia kiiensis*. *J. Nat. Prod* 1985; 48:236-241.
42. Bultel-Ponce V, Berge J, Debitus C, Nicolas J and Guyot M: Metabolites from the sponge associated bacterium *Pseudomonas* species. *Mar. Biotechnol* 1999; 1: 384–390.
43. Capon RJ, Ford J, Lacey E, Gill JH, Heiland K and Friedel T: Phoriospongins A and B: Two new nematocidal depsipeptides from the Australian marine sponges *Phoriospongia* sp. and *Callyspongia bilamellata*. *J. Nat. Prod* 2002; 65: 358–363.
44. Hamada T, Matsunaga S, Fujiwara M, Fujita K, Hirota H, Schmucki, Gontert P and Fusetani N: Solution Structure of Polytheonamide B, a Highly Cytotoxic Nonribosomal Polypeptide from Marine Sponge. *J. Am. Chem. Soc* 2010; 132 (37): 12941–12945.
45. Laird DW, LaBarbera DV, Feng X, Bugni TS, Harper MK and Ireland CM: Halogenated Cyclic Peptides Isolated from the Sponge *Corticium* sp. *J. Nat. Prod* 2007; 70 (5): 741–746
46. Lemmens-Gruber R, Kamyar MR and Dornetshuber R: Cyclodepsipeptides - Potential drugs and lead compounds in the drug development process. *Curr. Med. Chem* 2009; 16: 1122–1137.
47. Sera Y, Adachi K, Fujii K and Shizuri Y: Isolation of Haliclonomides: New peptides as antifouling substances from a marine sponge species, *Haliclona*. *Marine Biotechnology*. 2002; 3:441-446.
48. Mitova M, Popov S and De Rosa S: Cyclic Peptides from a *Ruegeria* Strain of Bacteria Associated with the Sponge *Suberites domuncula*. *J. Nat. Prod* 2004; 67: 1178-1181.
49. Morel J, Drobecq H, Sautiere P, Tartar A, Mironneau J, Qar J, Lavie J and Hugues M: Sponge, which Specifically Blocks a Non-L-Type Calcium Channel in Mouse Duodenal Myocytes. *Molecular Pharmacology* 1997; 51:1042–1052.
50. Schmidt EW and Faulkner DJ: Microsclerodermins C - E, antifungal cyclic peptides from the lithistid marine sponges *Theonella* sp. and *Microscleroderma* sp. *Tetrahedron* 1998; 54: 3043-3056.
51. Fusani N, Warabi K, Nogata Y, Nakao Y, Matsunaga S and Soest RM: Koshikamide A1, a new cytotoxic linear peptide isolated from a marine sponge, *Theonella* sp. *Tetrahedron Letters* 1999; 40(25): 4687-4690.
52. Vente J, Vera B, Rodríguez AD, Rodríguez-Escudero I and Raptis RG: Euryjanicin A: a new cycloheptapeptide from the Caribbean marine sponge *Prosuberites laughlini* *Tetrahedron Letters* 2009; 50 (32): 4571-4574.
53. Zhang H, Yi Y, Yang G, Hu M, Cao G, Yang F and Lin H: Proline-Containing Cyclopeptides from the Marine Sponge *Phakellia fusca*. *J. Nat. Prod* 2010; 73 (4): 650–655.
54. Mohammed R, Peng J, Kelly M, and Hamann MT: Cyclic Heptapeptides from the Jamaican Sponge *Stylissa caribica*. *J. Nat. Prod* 2006; 69 (12): 1739–1744.
55. Cychon C and Kock M: Stylissamides E and F, Cyclic Heptapeptides from the Caribbean Sponge *Stylissa caribica*. *J. Nat. Prod* 2010; 73 (4): 738–742.
56. Hsieh FC, Lin TC, Meng M and Kao SS: Comparing Methods for Identifying *Bacillus* Strains Capable of Producing the Antifungal Lipopeptide Iturin A. *Curr. Microbiol* 2008; 56: 1-5.
57. Duraikannu K, Damodar Edupalli, Rameshkumar G and Ravichandran S: Antimicrobial Peptide from Marine Sponge *Clathria indica*, (Dendy,1889). *American-Eurasian Journal of Scientific Research* 2009; 4 (1): 47-53.
58. Nagai K, Kamigiri K, Arao N, Suzumura K, Kawano Y, Yamaoka M, Zhang H, Watanabe M and Suzuki K: YM-266183 and YM-266184, novel thiopeptide antibiotics produced by *Bacillus cereus* isolated from a marine sponge. I. Taxonomy, fermentation, isolation, physico-chemical properties and biological properties. *J. Antibiot* 2003; 56: 123-128.
59. Suzumura K, Yokoi T, Funatsu M, Nagai K, Tanaka K, Zhang H and Suzuki K: YM-266183 and YM-266184, Novel Thiopeptide Antibiotics Produced by *Bacillus cereus* Isolated from a Marine Sponge. II. Structure Elucidation. *J. Antibiot* 2003; 56: 129-134.
60. Gunasekara SP, Pimponi SA and Mc Carthy PJ: Isolated from the Bahamian deep water marine sponge *Discodermia* sp. *J. Nat. Prod. Lloydia* 1994; 57:79.
61. Li H, Matsunaga S and Fusetani N: Halicylindramides A-C, antifungal and cytotoxic depsipeptides from the marine sponge *Halichondria cylindrata*. *J. Med. Chem* 1995; 38: 338-343.
62. Li H, Matsunaga S and Fusetani N: Halicylindramides D and E, Antifungal Peptides from the Marine Sponge *Halichondria cylindrata*. *J. Nat. Prod* 1996; 59 (2):163–166
63. Bewley CA and Faulkner DJ: Anti-fungal cyclic peptides from sponges. *J. Org. Chem* 1994; 59: 4849-4852.

64. Bewley CA, Debitus C and Faulkner DJ: Microsclerodermins A and B. Antifungal Cyclic Peptides from the Lithistid Sponge *Microscleroderma* sp. J. Am. Chem. Soc 1994; 116 (17): 7631-7636.
65. Fusetani N, Warabi K, Nogata Y, Nakao Y, Matsunaga S and van Soest RRM: Koshikamide A₁, a New Cytotoxic Linear Peptide Isolated from a Marine Sponge, *Theonella* sp. Tetrahedron Letters 1999; 40(25): 4687-4690.
66. Rashid MA, Gustafson KR, Cartner LK, Shigematsu N, Pannell LK and Boyd MR: Anti- HIV candidate from the sponge *Sidenops microspinosa*. J. Nat. Prod 2001; 64: 117-121.
67. Ford PW, Gustafson KR, McKee TC, Shigematsu N, Maurizi LK, Pannell LK, Williams DE, De-Silva ED, Lassota P, Allen TM, Van-Soest R, Andersen RJ and Boyd MR: Papuamides A–D, HIV-Inhibitory and Cytotoxic Depsipeptides from the Sponges *Theonella mirabilis* and *Theonella swinhoei* Collected in Papua New Guinea. J. Am. Chem. Soc 1999; 121: 5899-5909
68. Rashid MA, Gustafson KR, Boswell J and Boyd MR: Isolation of new cytotoxic hexapeptides from sponge *Haliclona nigra*. J. Nat. Prod 2000; 63: 956.
69. Plaza A, Bifulco G, Masullo M, Lloyd JR, Keffer JL, Colin PL, Hooper JNA, Bell LJ and Bewley CA: Mutremdamide A and Koshikamides C–H, Peptide Inhibitors of HIV-1 Entry from Different *Theonella* Species. J. Org. Chem 2010; 75 (13): 4344–4355.
70. Zampella A, Sepe V, Luciano P, Bellotta F, Monti MC, D’Auria M V, Jepsen T, Petek S, Adeline MT, Laprevote O, Aubertin AM, Debitus C, Poupat C and Ahond A: Homophymine A, an Anti-HIV Cyclodepsipeptide from the Sponge *Homophymia* sp. J. Org. Chem 2008; 73 (14): 5319–5327.
71. Araki T, Matsunaga S and Fusetani N: Koshikamide A₂, a Cytotoxic Linear Undecapeptide Isolated from a Marine Sponge of *Theonella* sp. Bioscience, Biotechnology and Biochemistry 2005; 69(7): 1318- 1322.
72. Araki T, Matsunaga S, Nakao Y, Furihata K, West L, Faulkner D J and Fusetani N: Koshikamide B, a Cytotoxic Peptide Lactone from a Marine Sponge *Theonella* sp. J. Org. Chem 2008; 73 (20): 7889–7894.
73. Kobayashi J, Itagaki F, Shigemori H, Ishibashi M, Takahashi K, Ogura M, Nagasawa S, Nakamura T and Hirota H: Keramamides B .apprx. D, novel peptides from the Okinawan marine sponge *Theonella* sp. J. Am. Chem. Soc; 1999; 113 (20): 7812–7813.
74. Ali L, Musharraf SG and Shaheen F: Solid-Phase Total Synthesis of Cyclic Decapeptide Phakellistatin 12. J. Nat. Prod 2008; 71 (6): 1059–1062.
75. Andavan GSB and Lemmens-Gruber R: Cyclodepsipeptides from Marine Sponges: Natural Agents for Drug Research. Mar. Drugs 2010; 8: 810-834.

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