IJPSR (2011), Vol. 2, Issue 9



INTERNATIONAL JOURNAL OF PHARMACEUTICAL SCIENCES AND RESEARCH (Review Article)



Received on 04 May, 2011; received in revised form 21 June, 2011; accepted 23 August, 2011

PRODUCTION OF REACTIVE OXYGEN SPECIES, ITS EFFECT, DRUGS AND PLANT EXTRACT USED AS AN ANTIOXIDANT, CHELATOR ON THALASSEMIC PATIENT: A REVIEW

Kuldeep K. Gupta*, Amit Mishra and Archana Tiwari

School of Biotechnology, Rajiv Gandhi Proudyogiki Vishwavidyalaya, Airport Bypass Road, Bhopal-462033 Madhya Pradesh, India

Keywords: Mutations, Transfusion, Reactive oxygen species, Antioxidant

Correspondence to Author:

Kuldeep Kumar Gupta

M. Tech., School Of Biotechnology, Rajiv Gandhi Proudyogiki Vishwavidyalaya, Airport Bypass Road, Bhopal-462033, Madhya Pradesh, India

ABSTRACT

 β - Thalassemia is an inherited genetic disorder which is caused by different kinds of mutations in the HBB gene in chromosome 11. Due to several types of mutation in β - gene, globin chains cannot synthesise completely and free α -globin is highly unstable and readily precipitates bound heme and iron. In β - thalassemia these precipitated iron, repeated blood transfusion and increased gastrointestinal iron absorption lead to iron overload in the body. The increased free iron in blood is responsible for the formation of Reactive Oxygen Species (ROS). If the production of ROS exceeds the capacity of enzymatic and non-enzymatic antioxidants systems to scavenge these species or if these protective systems are compromised, then oxidative stress occurs. This review summarizes the production of ROS, its effect and different drug and plant extract used as an antioxidant as well as chelating agent in thalassemic patient.

INTRODUCTION: β - Thalassemia is an autosomal recessive disorder characterized by microcytosis and hemolytic anemia, which is a result of the reduced synthesis of the β -globin chains of haemoglobin ¹. The disorder affects about 150 million people in the world ². β - Thalassemia is prevalent in Mediterranean countries, the Middle East, Central Asia, India, Southern China, and the Far East as well as countries along the north coast of Africa and in South America. The highest carrier frequency is reported in Cyprus (14%), Sardinia (10.3%), and Southeast Asia ³.

Although there are now more than 180 known β thalassemia mutations worldwide ⁴, a smaller collection of alleles accounts for the inactivation of most β -globin genes in each population or ethnic group. The genes involved in thalassemia control the production of a protein in red cells called haemoglobin. The synthesis of haemoglobin is controlled by two developmentally regulated multigene clusters: the alpha-like globin cluster on chromosome 16 and the beta-like globin cluster on chromosome 11. Human haemoglobin is a hetrotetramer protein, compose of two alpha and two beta subunits as shown in **Figure 1**. Each subunit contains a heme group, an iron containing compound that binds to oxygen.

In β -thalassemia, mutations in one or more of the β globins gene loci that result in reduced β -globin production. In addition to the direct effects of reduced β -globin synthesis, many of the symptoms of this disorder appear to be consequences of the resulting cytotoxic build up of free α -globin. Free α -globin is highly unstable and readily precipitates and release iron in reactive form ^{5, 6}. In addition to this, repeated blood transfusions and increased gastrointestinal iron absorption lead to iron overload in the body ⁷. Humans are unable to eliminate the iron, and the excess iron is deposited as hemosiderin and ferritin in the liver, spleen, endocrine organs and myocardium.



FIG. 1: HUMAN HAEMOGLOBIN

The accumulation of toxic quantities of iron causes tissue damage and leads to complications such as heart failure, endocrine abnormalities like diabetes, hypothyroidism, liver failure and ultimately early death ^{8, 9, and 10}. The deposited iron are responsible for the formation of reactive oxygen species such as superoxide anion (O_2^{-}) , hydroxyl radical (OH^{-}) , singlet oxygen and hydrogen peroxide (H_2O_2) . If the production of ROS exceeds, the capacity of enzymatic and non-enzymatic antioxidants systems to scavenge these species, or if these protective systems are compromised, then oxidative stress occurs 7, 11. This oxidative stress and a possible consequential accelerated apoptosis may contribute to shortened life of erythrocytes, primary or secondary span amenorrhoea, hypogonadism, osteoporosis and other endocrine disorders ¹².

Biomarkers of oxidative stress included plasma malondialdehyde (a marker of lipid per oxidation ¹³⁻¹⁶ and plasma protein carbonyls, a marker of oxidation to circulating proteins. Inflammatory biomarkers were cytokines (including interleukin-6) and high-sensitivity C-reactive protein (hsCRP), markers previously found useful in thalassaemia ^{14, 17, and 18}.

Antioxidant are complex group of protein such as superoxide dismutase's, which convert superoxide to oxygen peroxide, catalase and glutathione peroxidase, which convert hydrogen peroxide to water, and nonenzymatic scavengers such as glutathione, peroxiredoxin, ascorbic acid and carotenoids. Malondialdehyde (MDA), a product of lipid peroxidation is generated in excess amounts in supporting the fact that large amount of membrane bound iron is present in thalassemic erythrocytes ^{19, 20}.

Chelation therapy reduces iron-related complications and thereby improves quality of life and overall survival ^{21, 22}. The poor oral bioavailability, short plasma half-life and severe side effects make available chelators suboptimal ^{22- 26}. Iron chelators mobilize tissue iron by forming soluble, stable complexes that are then excreted in the faeces and/or urine. It was reported that chelating agents are effective as secondary antioxidants because they reduce the redox potential, thereby stabilizing the oxidized form of the metal ion ²⁷.

Production of ROS in thalassemic Patient: Reactive oxygen species are partially reduced forms of atmospheric oxygen. They typically result from the excitation of O₂ to form singlet oxygen or from the transfer of one, two or three electrons to O₂ to form, respectively, a superoxide radical (O₂), hydrogen peroxide (H₂O₂) or hydroxyl radical (OH⁻). Free α globins is highly unstable and readily precipitates, and release iron in reactive form ^{5, 6}. In addition to this blood transfusions and increased repeated gastrointestinal iron absorption lead to iron overload in the body 7 . The deposited iron is responsible for the formation of reactive oxygen species which causes oxidative stress in thalassemic patient.

In mitochondria during oxidative phosphorylation for energy production O_2 molecules accept four electrons to form two molecules of water. The acceptance of a single electron by O_2 generates superoxide O_2^- . So mitochondria are a major source of superoxide. These produced superoxides undergo rapid dismutation both spontaneously and by a family of enzymes, super oxide dismuatase to form $H_2O_2^{-28, 29.30}$.



This H_2O_2 reacted with highly reactive Fe^{2+} ions which catalyze the Fenton's and Haber-Weiss reactions ³¹ these reactions takes place in the presence of hydrogen peroxide (H_2O_2) and superoxide anion radicals ($O_2^{\bullet-}$).

According to Fenton's;

Haem-Fe²⁺ + H₂O₂
$$\rightarrow$$
 Fe³⁺ + OH⁻ + [•]OH
Fe³⁺ + O₂^{•-} \rightarrow Fe²⁺ + O₂

In general, the O_2 carried by heme proteins is only bound directly to the ferrous iron with an oxidation state of two (Fe²⁺). In the presence of H₂O₂, ferrous iron and H₂O₂ are going to react to generate ferric iron (Fe³⁺) and highly reactive hydroxyl radicals ([•]OH). Addition of a reducing agent, such as ascorbate, leads to a cycle which increases the damage to biological molecules.

According to the Haber–Weiss reactions

$$O_2^{\bullet^-} + H_2O_2 \rightarrow \bullet OH + OH^- + O_2$$
$$\bullet OH + H_2O_2 \rightarrow O_2^{\bullet^-} + H_2O + H^+$$

The above reaction is catalysed by Fe^{3+} and is a possible source of [•]OH.

It was demonstrated that the release of Fe^{3+} from the porphyrin ring increased as a function of O_2 concentration as shown below

Heme-Fe²⁺ + O₂
$$\rightarrow$$
 Heme-Fe³⁺ + O₂^{•-}
Heme-Fe³⁺ \rightarrow Fe³⁺ + iron-free Heme

When Fe^{3+} is released, it could generate reactive oxygen species that would, in turn, damage cellular compartments ³². The accelerated ROS production also causes the expression of several specific metal-binding proteins (transferrin, cerruloplasmin, ferritin, lactoferitin, etc.) and subsequent progression of oxidative stress ³³.

MDA a Marker of oxidative stress: Malondialdehyde (MDA) is a three carbon dialdehyde which is widely produced in mammalian organisms as a side product of prostaglandin biosynthesis and an end product of polyunsaturated lipid peroxidation ³⁴⁻³⁶. MDA is generated in excess amounts in thalassemia, supporting the fact that large amounts of membrane bound iron are present in thalassemic erythrocytes. MDA is a bi functional reagent and has been reported to crosslink several cell constituents including membrane components ^{37, 38}.



Increased plasma MDA level was found in betathalassemia patients ^{39, 40}. Plasma MDA was studied as a marker of tissue injury and oxidative stress. In oxidative stress condition peroxidative damages to tissues and depletion of endogenous antioxidants may be expected ⁴¹. Peroxidative damage to lipids and protein is indicated by the increase of about two fold of the serum MDA, conjugated diene lipid hydro peroxides and protein carbonyls ⁴².

Effect of oxidative stress on Enzymatic and Nonenzymatic antioxidant: Antioxidant capacity is an important determinant of tissue injury, especially in patients with increased oxidant stress ⁴³. Researchers were found significant differences in antioxidant capacity in terms of tocopherol level. Plasma α tocopherol was decreased in thalassemia ⁴⁴⁻⁴⁶. These low levels probably contribute to increased MDA levels. In addition to this all others antioxidant level markedly modified except albumin and glutathione. Concentrations of ascorbate and vitamin E (α tocopherol), the major lipid soluble antioxidant in human blood, decreased by 42%, other lipid soluble antioxidants such as vitamin A, and carotenoids such as carotene and lycopene, which are part of the unmeasured compounds in TEAC determination, were also markedly reduced ⁴². Serum levels of aspartate transaminase were inversely correlated with vitamin E, vitamin A, and lycopene, suggesting that liver damage may play a major role in the extent of depletion of these lipid soluble antioxidants. Levels of alanine aminotransferase were similarly inversely correlated with vitamin E, vitamin A, and lycopene ⁽⁴²⁾. Then, despite a mean decrease of 14% in the serum total antioxidant potential, a dramatic fall in the amount of ascorbate (44%) and lipid soluble antioxidants, vitamin E (42%), vitamin A (44%), *p* carotene(29%), and lycopene (67%) is observed in all patients ⁴².

Role of Antioxidant in thalassemic patient: The body's antioxidant system is an integrated one, in which some components may interact to spare or replace each other. However, the deficiency of individual antioxidants observed in thalassemia is such that no effective compensation could be brought about ⁽⁴²⁾. Antioxidant capacity is a result of the overall effect of water-soluble antioxidants, lipid-soluble antioxidants and antioxidant enzymes such as superoxide dismutase ³¹.

Dehydroascorbate cannot be regenerated to its reduced form, as its regenerating system involves erythrocyte glutathione, most of which in thalassemia patients can be oxidized. Moreover, as vitamin C is essential to maintain vitamin E status and function, depletion of vitamin C, in turn, contributes to further exacerbate the depletion of vitamin E. Although efficient antioxidants such as uric acid and bilirubin are high, they cannot compensate for lipid-soluble antioxidants, so that tissue lipid compartments are not suitably preserved⁴⁴.

Antioxidant activity depends on the kind of oxidative stress and on oxidative substrate. According to Packer *et al.*, 1995 47 , when we evaluate the antioxidant potential of a compound, criteria such as.

- a) Specificity of free radical scavenging,
- b) Interaction with other antioxidants,
- c) Metal-chelating activity,
- d) Effects on gene expression,
- e) Bioavailability,

- f) Location (in aqueous or membrane domains, or in both),
- g) Ability to repair oxidative damage ⁴⁸

LA/DHLA are considered ideal therapeutic antioxidant because they are naturally existing, low molecular weight compounds with very powerful antioxidant effective in both aqueous and lipid domains. Their effects include free radical quenching 49, metal chelation ⁵⁰ and regeneration of other antioxidant such as ascorbic acid, vitamin E and glutathione ⁵¹. The antioxidant role of vitamin E is attributed to its ability quenching highly reactive lipid peroxide in intermediate by donating hydrogen and this prevents extraction of hydrogen from PUFA. This assists in restricting self perpetuated lipid peroxidation chain reaction ^{52, 53}. Erythrocyte SOD scavenges superoxide radicals to form hydrogen peroxide and protects the cell membrane from its damage.

Increased Erythrocyte SOD activity may be due to blood transfusion and increase in the proportion of younger erythrocytes, as a compensatory mechanism after increased oxidative stress ⁵⁴. Since antioxidants seem to act co-operatively *in vivo*, the evaluation of TAOA in blood plasma could provide a more comprehensive assessment than the evaluation of individual antioxidants. TAOA is a parameter, summarizing the overall content and activity of the water-soluble antioxidants. The depletion of TAOA induced by oxidative stress in β - thalassemia major patients is probably eliminated by the release of stock organ antioxidants and the induction or activation of antioxidant enzymes⁵⁵.

Chelators: As the body has no effective means for removing iron, the only way to remove excess iron is to use iron binders (chelators), which allow iron excretion through the urine and/or stool. As a general rule, patients should start iron chelation treatment once they have had 10-20 transfusions or when ferritin levels rise above 1000ng/ml⁵⁶. Since till date there is no effective drug available in the market for the treatment of thalassemia, only chelation therapy is the way by which a patient can live long life. So different researcher used certain types of drugs and plant extracts for the inhibition of oxidative stress in thalassemic patient. These drug and plant extract work as chelator as well as antioxidant.

Drugs used as iron chelator in thalassemic patient: The first drug available for treatment of iron overload was *deferoxamine* (DFO), an exadentate iron chelator that is not orally absorbed and thus needs parenteral administration ^{56, 57}. The use of DFO decreases morbidity and mortality among those who are able to comply with regular prolonged infusions ⁵⁸. However, because of the side effects and the inconvenient parenteral administration, a consistent proportion of patients are non-compliant, limiting the usefulness of this chelator ⁵⁹.

According to Yasser Ali *et al.,* ⁶⁰ *deferiprone* (DFP) is an effective drug which can be used safely for iron chelation in thalassaemia major patient with iron overload. This drug is very effective in cardiac protection. The orphan drug DFP is an orally active iron chelator which has emerged from an extensive search for new treatment of iron overload ⁶¹. Retrospective and prospective studies have shown that DFP monotherapy is significantly more effective than DFO in decreasing myocardial siderosis in thalassemia major ⁶²⁻⁶⁴.

Researchers work on different drug like DFO and DFP and they used in combination and alone they found that, drug is to be most effective in combination rather than alone ⁶⁵⁻⁶⁹. Similar work has been conducted by Gharagozloo M *et al.*, but they used silymarin, a flavovonolignan complex isolated from silybum marianum and DFO. They found that significant improvement in liver alkaline phosphatase and glutathione levels of red blood cells was detected. This is the first report showing the beneficial effects of silymarin in thalassemia patients and suggests that silymarin in combination with DFO can be safely and effectively used in the treatment of iron-loaded patients. Silymarin has a strong antioxidant, hepatoprotective, and iron chelating activities.

Patrick B. Walter's *et al.*, research emphasized on MDA level of thalassemic patient. They treated the patient with deferaxasirox and deferaxamine and found that both drugs are equally effective in decreasing iron burden and MDA levels.

Plant extract used as chelation and anti oxidant: Plants are rich sources of natural antioxidants and the antioxidant effect of plant products is mainly attributed to phenolic compounds such as flavonoids, phenolic acids, tannins and phenolic diterpenes ⁷⁰. Among the various medicinal and culinary plants, some endemic species are of particular interest because they may be used for producing raw materials or preparations containing phytochemicals with significant antioxidant capacities and health benefits ⁷¹. Plant extracts of various plants are used for the treatment of various diseases, were highly regarded by the ancient civilizations. Even today, plant materials remain an important resource for combating illnesses ⁷².

It has been documented that flavonoids which contain hydroxyl functional group, show antioxidant activity and their effects on human nutrition and health are considerable. The mechanisms of action of flavonoids are through scavenging or chelating process ⁷³⁻⁷⁷. Phenolic compounds are a class of antioxidant compounds which act as free radical terminators ⁷⁵. There was a direct relation between chelatory activity and the content of active compounds, phenol and flavonoid. Plant extracts with high phenol and flavonoid contents showed good chelating of Fe^{++ 72} so such type of plant extract now used as chelator antioxidant for thalassemic patient. These are as follows-

According to Olabinri BM, Eniyansoro O.O. et al., ⁷⁸, the aqueous extract of Tetracarpidium conophorum (African Walnut) demonstrate dose dependent decrease in chelating ability. They found that the antioxidant activity of plant extract fully depend on their phenolic and flavonoid content. If the plant extract have high concentration of phenol and flavonoid then that plants have maximum antioxidant activity. Similar results were observed research conducted by Phalguni Srimani, Goutam Mandal et al., ⁷⁹ who demonstrated the antioxidant potential of ethanolic extract of leaves of Piper beetle Linn. (Pan). Although the extract of P. betle contained phenols, chevi, etol, allylpyrocatechol and their respective glycosides ⁸⁰ but the compound responsible for antioxidant activity is unknown. Effective antioxidant activity, reducing power activity Fe⁺⁺ chelating , nitric oxide and DPPH radical- scavenging property was exhibited by aerial plants extract of Leonurus cardiac subsp. Grammosciadium platycarpum, Onosma

demawendicum research conducted by Ebrahimzadeh , M. A. et al., 2010 81 .

According to Mohammad Ali Ebrahimzadeh *et al.*, 2008 ⁸², there was direct relation between chelatory activity and the content of active compounds phenol and flavonoid present in plant extracts. They showed that *Epilobium hirsutum, Melilotus arvensis, Feijoa sellowiana* showed similar result while corn milk with high phenol and flavonoid content showed very weak chelating activity, although, *Pistacia lentiscus* with low phenol and flavonoid content showed good chelating activity.

CONCLUSION: β-thalassemia is a genetic disorder, symptoms of which appear only in thalassemia major patient. Such type of patient needs regular blood transfusion for survival, resulting in the increase of blood iron concentration which causes oxidative stress leading to the development of other abnormality in the body. Till date, the technique available for the treatment of this genetic disorder are bone marrow transplantation and stem cell therapy but these treatment methods are very expensive and hence people depends on regular blood transfusions throughout their life. The only way to increase the survival rate of such patients is by inhibiting the effect of oxidative stress. So researcher should explore such drugs which may be derive from plant extracts; which can minimize the effect of oxidative stress by removing free radical or remove excess iron from thalassemic patients.

REFERENCE:

- Weatherall DJ, Clegg JB: The Thalassemia Syndromes. 3rd ed. Oxford: Blackwell Scientific Publications 1981; 299-300.
- WHO: Community Control of Hereditary Anemias: Memorandum from a WHO Meeting. Bulletin WHO 1983; 61-63.
- Flint J, Harding RM, Boyce AJ, Clegg JB: The population genetics of the hemoglobinopathies. Bailliere's Clinical Hematology 1998; 11:1-50.
- 4. Baysal E, Carver MFH: The β and δ -Thalassemia Repository (8thed.). Hemoglobin 1995; 19:213.
- 5. Brunori M, G. Falcioni, Fioretti E, Giardina B, Rotilio G: Formation of superoxide in the autoxidation of the isolated α and β chains of human hemoglobin and its involvement in hemichrome precipitation, European Journal of Biochemistry 1975;53: 99–104.
- 6. Scott MD, van den berg JJ, Repka T, Rouyer-Fessard P, Hebbel RP, Beuzard Y, Lubin BH: Effect of excess α -hemoglobin chains on cellular and membrane oxidation in model β -thalassemic

erythrocytes. Journal of Clinical Investigation 1993; 91: 1706–1712.

- 7. Milena R, Branka Z, Biljana S et al.: Thalassemia syndrome in Serbia. Heamoglobin 2010; 34(5):477-485.
- 8. Taher A, Isma'eel H, Cappellini MD: Thalassemia intermedia: revisited. Blood Cells Mol. Dis. 2006; 37: 12-20.
- 9. Rund D, Rachmilewitz E: Beta-thalassemia, England Journal of Medicine 2005; 353: 1135-1146.
- 10. Loukopoulos D: Combined therapy with deferiprone and desferrioxamine in thalassemia major. Hematology Journal 2005; 90: 1305-1305.
- 11. Repka T, Shalev O, Reddy R, Yuan J, Abrahamov A, Rachmilewitz A, Low PS, Hebbel RP: Non random association of free iron with membranes of sickle and beta-thalassemic erythrocytes. Blood 1993; 82:3204–3210.
- 12. Bronspiegel-Weintrob N, Olivieri NF, Tyler B, Andrews DF, Freedman MH, Holland FJ: Effect of age at the start of iron chelation therapy on gonadal function in beta-thalassemia major. New England Journal of Medicine 1990; 323:713–719.
- 13. Cighetti G, Duca L, Bortone L, Sala S, Nava I, Fiorelli G: Oxidative status and malondialdehyde in beta thalassemia patients. European Journal of Clinical Investigation 2002; 32:55-60.
- 14. Walter PB, Fung E, Killilea DW, Jiang Q, Hudes M, Madden J: Oxidative stress and inflammation in iron-overloaded Patients with beta thalassemia or sickle cell disease. British Journal of Haematology 2006; 135:254-63.
- 15. Kadiiska MB, Gladen BC, Baird DD, Germolec D, Graham LB, Parker CE, *et al.*: Biomarkers of oxidative stress study II: Are oxidation products of lipids, proteins, and DNA markers of CCl4 poisoning. Free Radical Biology and Medicine 2005; 38:698-710.
- Kadiiska MB, Gladen BC, Baird DD, Graham LB, Parker CE, Ames BN et al., : Biomarkers of oxidative stress study: III. Effects of the nonsteroidal anti-inflammatory agents indomethacin and meclofenamic acid on measurements of oxidative products of lipids in CCl4 poisoning. Free Radical Biology and Medicine 2005; 15:25-40.
- 17. Tong PC, Ng MC, Ho CS, So WY, Li JK, and Lam CW: C-reactive protein and insulin resistance in subjects with thalassemia minor and a family history of diabetes. Diabet Care 2002; 25:1480-1.
- Archararit N, Chuncharunee S, Pornvoranunt A, Atamasirikul K, Rachakom B, Atichartakarn V: Serum C-reactive protein level in post splenectomized thalassemic patients. Journal of the Medical Association of Thailand 2000; 83:S63-9.
- Das N, Chowdhury TD, Chattopadhyay A, Datta: Attenuation of oxidation stress induced changes in thalassemic erythrocytes by Vitamin E. Polish Journal of Pharmacology 2004; 56:85-96.
- 20. Tesoriere L, Pintaudi AM, Calabrese A, MaggioA, Freisleben HJ, et al.: Oxidative stress and Antioxidant status in beta thalassemia major: Iron overload and depletion of lipid - soluble antioxidants. Blood 1996; 88: 3608-14.
- 21. Shinar E, Rachmilewitz EA: Oxidation denaturation of red blood cells in thalassemia. Semin Hematol 1990; 27: 70-91.
- 22. Hebbel RP, Leung A, Mohandas N: Oxidation-induced changes in micro heological properties of the red cell membrane. Blood 1990; 76:1015-1022.
- Grinberg LN, Rachmilewitz EA, Kitrossky N, Chevion M: Hydroxyl radical generation in
 ß-thalassemia red blood cells. Free Radical Biology and Medicine 1995; 18: 611-615.
- Kukongviriyapan V, Somparn N, Senggunprai L, Prawan A, Kukongviriyapan U, Jetsrisuparb A : Endothelial Dysfunction and Oxidant Status in Pediatric Patients with Hemoglobin EbetaThalassemia. Pediatric Cardiology 2008:29: 130-135.

- Filburn CR, Kettenacker R, Griffin DW: Bioavailability of a silybin-phosphatidylcholine complex in dogs. Journal of Veterinary Pharmacology and Therapeutics 2007; 30: 132-138.
- 26. Rachmilewitz EA, Shifter A, Kahane I: Vitamin E deficiency in ß thalassemia major: Changes in haematological and biochemical parameters after a therapeutic trial with alpha tocopherol. American Journal of Clinical Nutrition 1979; 32: 1850-1858.
- Ebrahimzadeh MA, Nabavi SF, Nabavi SM, Eslami B: Antihypoxic and antioxidant activity of *Hibiscus esculentus* seeds. Grasas Aceites, 2010 b: 61(1): 30-36.
- 28. Beckman KB and Ames BN: The free radical theory of aging matures. Physiol Rev 1998; 78: 547–581.
- 29. Droge W: Free radicals in the physiological control of cell function. Physiological Review 2002; 82: 47–95.
- 30. Finkel T: Oxidant signals and oxidative stress. Current Opinion in Cell Biology 2003; 15: 247–254.
- 31. Pavlova LE, Savov VM, Petkov HG, Charova IP: Oxidative stress in Patient with β thalassemia major. Sec. Biol. Med. Sci.2007; 145-154.
- Anastassopoulou J, Anifantakis B, Anifantakis ZA, Dovas A, Theophanides T: The role of free radical reactions with haemoglobin and thalassaemia. Journal of Inorganic Biochemistry 2000; 79:327-329.
- Vladimirov JA., Azizova OA., Deev AI, Kozlov AV., Ossipov AN, Roshtupkin DI. : Free radicals in living systems, in Science and technique reviews, Biophysics series, VINITI, Moscow 1991; 29: 1–252.
- Bernheim F, Bernheim MLC, Wilbur KM: The reaction between thiobarbituric acid and the oxidation products of certain lipids. The Journal of Biological Chemistry1948; 174: 257-264.
- Diczfalusy U, Falardeau P, Hamrnarstrim S: Conversion of prostaglandin endoperoxides to CI Thydroxyacids catalyzed by human platelet thromboxane synthase, FEBS Letters1997; 84: 271-274.
- Hamberg M, Samuelsson B: Oxygenation of unsaturated fatty acids by the vesicular gland of sheep. The Journal of Biological Chemistry 1967; 242: 5344-5354.
- Cighetti G, Duca L, Bortone L, Sala S, Nava I, Fiorelli G et al.: Oxidative status and malondialdehyde in beta thalassaemia patients. European Journal of Clinical Investigation 2002; 32 Suppl 1: 55-60.
- Nielsen F, Mikkelsen BB, Nielsen JB, Andersen HR, Grandjean P. Plasma malondialdehyde as biomarker foroxidative stress: reference interval and effects of life-style factors. Clinical Chemistry 1997, 43: 1209-1214.
- Aldouri MA, Hoffbrand AV, Flynn DM, Ward SE, Agnew JE, Hilson AJW: High incidence of cardiomyopathy in betathalassemiapatients receiving regular transfusions and iron chelation: Reversalby intensified chelation. Acta Haematol 1990; 841: 13.
- Zannos-Mariolea L, Tzortzatou F, Dendaki-Svolaki K, Katerellos C, Kavallari M, Matsaniotis N: Serum vitamin E levels with betathalassemia major: preliminary report. British Journal of Haematology 2008; 26:193-199.
- Halliwell B, Gutteridge JMC: Role of free radicals and catalytic metal ions in human disease: An overview, in Packer L, GlazerAN (eds): Methods in Enzymology, San Diego, CA, Academic, 1990; 186: 1.
- Livrea MA, Tesoriere L, Pintaudi AM, Calabrese A, Maggio A et al., Oxidative stress and antioxidant status in β thalassaemia major. The American society of Haematology1996; 0006-4971/96/8809-001.
- 43. Patric WB, Ellen FB, Killilea WD, Jiang Q: Oxidative Stress and Inflamation in Iron Overload patients with β -Thalassemia or

sickle cell disease. British Journal of Haematology 2006; 135(2):254-263.

- 44. Rachmilewitz EA, Shohet SB, Lubin BH. Lipid membrane peroxidation in β -thalassemia major. Blood 1976; 47:495–505.
- 45. De Luca C, Filosa A, Grandinetti M, Maggio F, Lamba M, Passi S. Blood antioxidant status and urinary levels of catecholamine metabolites in β -thalassemia. Free Radical Research 1999; 30:453–462.
- Kassab-Chekir A, Laradi S, Ferchichi S, Haj Khelil A, Feki M, Amri F, Selmi H, Bejaoui M, Miled A. Oxidant, antioxidant status and metabolic data in patients with beta-thalassemia. Clinica Chimica Acta 2003; 338:79–86.
- Packer L, Witt EH, Tritschler HJ. Alpha-lipoic acid as a biological antioxidant, Free Radical Biology and Medicine, 1995; 19:227– 250.
- 48. Navari-izzo, Flavia, Quartacci MF, Sgherri C. Lipoic acid: a unique antioxidant in the detoxification of activated oxygen species. Plant Physiology and Biochemistry 2002; 40:463–470.
- Matsugo S, Yan LJ, Han D, Trischler HJ, Packer L. Elucidation of activity of alpha-lipoic acid toward hydroxyl radical. Biochemical and Biophysical Research Communication 1995; 208:161–167.
- Ou P, Tritschler HJ, Wolff SP. Thioctic (lipoic) acid: a therapeutic metal-chelating antioxidant? Biochemical Pharmacology 1995; 50: 123–126.
- Guo Q, Packer L. Ascorbate-dependent recycling of the vitamin E homologue Trolox by dihydrolipoate and glutathione in murine skin homogenates. Free Radical Biology & Medicine 2000; 29:368–374.
- Das N, Chowdhury TD, Chattopadhyay A, Datta: Attenuation of oxidation stress - induced changes in thalassemic erythrocytes by Vitamin E. Polish Journal of Pharmocology 2004; 56:85-96.
- 53. Baumgartener TG. Vitamins In: Van Way C.W. ed. Nutrition secrets. Philadelphia, Hanley and Belfus, 1999: 13-20.
- 54. Simsek F, Ozturk G, Kemahli S, Erbas D, Hasanoglu A: Oxidant and antioxidant status in beta thalassemia major patients. Ankara University Tip Fakultesi Mecmuasi 2005; 58: 34-8.
- Dubinina EE, Babenko GA, Shcherbak IG: Molecular heterogeneity of plasma superoxide dismutase. Free Radical Biology & Medicine 1992; 13(1): 1–7.
- 56. Thalassemia International Federation: Guidelines for the clinical management of thalassemia 2nd edition. 2008 [http://www.thalassemia.org.cy].
- Borgna-Pignatti C, Galanello R: Thalassemias and related disorders: Quantitative disorders of hemoglobin synthesis. In Wintrobe's Clinical Haematology Volume 42. 11th edition. Lippincott Williams & Wilkins. Philadelphia; 2004:1319-1365.
- 58. Gabutti V, Piga A: Results of long-term iron-chelating therapy. Acta Haematol 1996; 95:26-36.
- 59. Cunningham MJ, Macklin EA, Neufeld EJ, Cohen AR: Thalassemia Clinical Research Network. Complications of betathalassemia major in North America 2004.
- 60. Yasser W, Azza AS, Shabina D. Agranulocyosis in beta Thalassemia major patient treated with Oral Iron Chelating Agent. Oman medical journal 2008; 23:275-277.
- 61. Galanello R: Deferiprone in the treatment of transfusiondependent thalassemia: a review and perspective. Therapeutics and Clinical Risk Management 2003; 3:795-805.
- Anderson LJ, Wonke B, Prescott E, Holden S, Walker JM, Pennell DJ: Effects of oral deferiprone and subcutaneous desferrioxamine on myocardial iron concentrations and ventricular function in beta-thalassaemia. Lancet 2002; 360:516-520.

- 63. Piga A, Gaglioti C, Fogliacco E, Tricta F: Comparative effects of deferiprone and deferoxamine on survival and cardiac disease in patients with thalassemia major: a retrospective analysis. Haematologica 2003; 88:489-496.
- 64. Pennell DJ, Berdoukas V, Karagiorga M, Ladis V, Piga A, Aessopos A, Gotsis ED, Tanner MA, Smith GC, Westwood MA, Wonke B, Galanello R: Randomized controlled trial of deferiprone or deferoxamine in β- thalassemia major patients with asymptomatic myocardial siderosis. Blood 2006; 107:3738-3744.
- 65. Wonke B, Wright C, Hoffbrand AV: Combined therapy with deferiprone and desferrioxamine. British Journal of Haematology 1998; 103:361-364.
- Alymara V, Bourantas D, Chaidos A, Bouranta P, Gouva M, Vassou A,Tzouvara E, Bourantas KL: Effectiveness and safety of combined iron chelation therapy with deferoxamine and deferiprone. Hematol Journal 2004; 5:475-479.
- Farmaki K, Anagnostopoulos G, Platis O, Gotsis E, Toulas P: Combined chelation therapy in patients with thalassemia major: A fast and effective method of reducing ferritin levels and cardiological complications. Hematol Journal 2002; 3(Suppl 1):79.
- 68. Kattamis A, Kassou C, Ladis V, Berdoussi H, Papasotiriou I, Kattamis C: Safety and efficacy of combining deferiprone and deferoxamine in iron chelation therapy in patients with thalassemia . Blood 2002; 100:120a.
- 69. Origa R, Bina P, Agus A, Crobu G, Defraia E, Dessi C, Leoni GB, Muroni PP,Galanello R: Combined therapy with deferiprone and desferrioxamine in thalassemia major. Haematologica 2005; 90:1309-1314.
- 70. Pietta PG; Flavonoids as antioxidants. Journal of Natural Product 2000; 63: 1035-1042.
- Exarchou V, Nenadis N, Tsimidou M, Gerothanassis IP, Troganis A,Boskou D : Antioxidant activities and phenolic composition ofextracts from Greek oregano, Greek sage, and summer savory. Journal of Agriculture and Food Chemistry 2002; 50: 5294-5299.
- 72. Ebrahimzadeh M A, Pourmorad F, Bekhradnia AR: Iron chelating activity, phenol and flavonoid content of some

medicinal plants from Iran. African journal of Biotechnology 2008d; 7(18):3188-3192.

- 73. Kessler M, Ubeaud G, Jung L: Anti- and pro-oxidant activity ofrutin and quercetin derivatives. Journal of Pharmacy and Pharmacology 2003; 55: 131-142.
- 74. Cook NC, Samman S: Flavonoids- chemistry, metabolism, cardioprotective effects, and dietary sources. Nutritional Biochemistry 1996; 7: 66-76.
- 75. Shahidi F, Wanasundara PKJPD: Phenolic antioxidant. Nutrition & Food Science 1992; 32: 67-103.
- Das NP, Pereira TA: Effects of flavonoids on thermalautooxidation of Palm oil: structure- activity relationship. Journal of the American Oil Chemists Society 1990; 67: 255- 258.
- 77. Younes M: Inhibitory action of some flavonoids on enhancedspontaneous lipid peroxidation following glutathione depletion. Plant Med 1981; 43: 240-245.
- 78. Olabinri BM, Eniyansoro OO, CO Okoronkwo CO: Evaluation of chelating ability of aqueous extract of *Tetracarpidium conophorum* in vitro. International journal of applied research in natural products 2010; 3(3):13-18.
- 79. Srimani P, Mandal G, Ganguly S. Saha P. et al.,: Antioxidant effect of ethanolic extract of *Piper betle* on erythrocytes from patients with HbE- β-thalassemia. Indian journal of Biochemistry& Biophysics 2009; 46: 241-246.
- D, Saha P, Gamre S, Bhattacharjee S, Hariharan C, ganguly S, Sen R, Mandal G: Anti-inflammatory effect of allyl pyrocatechol in LPS-induced macrophages is mediated by suppression of iNOS and COX-2 via the NF-κB pathway: International immunopharmacology 2008;8: 1264- !271.
- Ebrahimzadeh MA, Nabavi SF, Fazelian M, Eslami B: *Invitro* antioxidant and free radical scavenging activity of *Leonurs platycarpum subsps. Persicus, Gammosiadiump platycarpum* and *Onosma demawendicum;* African Journal of Biotechnology 2010; 9 (51): 8865-8871.
- Ebrahimzadeh MA, Poumorad F, Bekhradnia AR: Iron chelating activity, phenol and flavonoid content of some medicinal plants from Iran. African Journal of Biotechnology 2008; 7(18):3188-3192.
