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## **IN VITRO ANTIOXIDANT AND FREE RADICAL SCAVENGING POTENTIAL OF *HOLOSTEMMA ADA* - KODIEN K. SCHUM., AN IMPORTANT RARE MEDICINAL PLANT**

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### **ABSTRACT**

The present study was undertaken to evaluate the *in vitro* antioxidant activities of hexane, ethyl acetate and methanolic extracts of *Holostemma ada kodien* root tubers. The above extracts exhibited a dose dependent scavenging activity against DPPH radicals, Superoxide radicals, and Nitric oxide radicals. Further, the MeOH, EtOAC extracts showed relatively higher reducing power compare to that of BHT. TLC of the above extracts using the DPPH as a spraying reagent revealed yellow spots against purple background indicating the presence of potent antioxidant compounds.

**INTRODUCTION:** Hyperphysiological burden of free radicals causes imbalance in homeostatic phenomena between oxidants and antioxidants in the body. This imbalance leads to oxidative stress that is being suggested as the root cause of aging and various human polygenic diseases like atherosclerosis, stroke, diabetes, cancer, and neurodegenerative diseases such as Alzheimer's disease and Parkinsonism<sup>1</sup>.

Reactive oxygen species (ROS) and reactive nitrogen species (RNS) are various forms of activated oxygen and nitrogen, which include free radicals such as superoxide ions ( $O_2^{\bullet}$ ), hydroxyl ( $OH^{\bullet}$ ) and nitric oxide radicals ( $NO^{\bullet}$ ) as well as non-free radicals such as hydrogen peroxide ( $H_2O_2$ ) and nitrous acid ( $HNO_2$ )<sup>2,3,4</sup>. ROS can readily react with and oxidize most bio molecules, such as carbohydrates, proteins, lipids and DNA. The importance of ROS and free radicals has attracted an increasing attention over the past decade<sup>5,6,7</sup>. There is a crucial balance between the generation

of ROS and inactivation of ROS by the antioxidant system in organisms. Under pathological conditions, excessive production of ROS causes oxidative stress and macromolecular damage including protein oxidation and lipid peroxidation<sup>8,9</sup>. Malonaldehyde is inferred to be cytotoxic and it has been found at elevated levels in various diseases thought to be related to free radical damage<sup>10,11,12</sup>. Therefore, current research has been focused on the use of antioxidants, with reemphasis on naturally-derived antioxidants, which may inhibit ROS production and display protective effects.

Antioxidants can neutralize the ill effects of free radicals by scavenging or chain breaking (like vitamin A, C, beta carotene, etc.) or some other mechanism of action. These antioxidants must be constantly replenished since they are "used up" in the process of neutralizing free radicals<sup>13</sup>. Current research into free radicals has confirmed that foods rich in antioxidants

play an essential role in the prevention of cardiovascular diseases and cancers<sup>14</sup>. Many synthetic antioxidants such as butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), propyl gallate and tert-butyl hydroquinone have been shown to have some adverse effects including liver damage and carcinogenesis<sup>15, 16</sup>. Therefore, there is a growing interest on natural additives as potential antioxidants in reducing free radical-induced tissue injury. Numerous plant products have been screened and shown to have antioxidant activity due to the presence of antioxidant vitamins, flavonoids, and polyphenolic compounds.

Medicinal plants have been used traditionally to treat a variety of disease conditions that are now being explained by the oxidative stress theory. *Holostemma ada-kodien* is an important medicinal plant belonging to the family Asclepiadaceae, it was also called Jivanti, Arkapushpi, Kshira, Dodi, Suryavalli and is widely distributed in the Tropical rain forests in India<sup>17, 18</sup>. The plant is used for maintaining youthful vigor, strength and vitality<sup>19</sup>. The root tubers of the plant are used as tonic, ophthalmic, alterant, stimulant, aphrodisiac expectorant and gallactagogue<sup>20</sup>. The terpenoid sugars present in the root tubers of the plant responsible for the medicinal properties<sup>21</sup>. There is a huge demand for the root tubers of *Holostemma* and more than 150 tons of root tubers are required every year in the South Indian pharmacies.

## MATERIALS AND METHODS:

**Collection of the Plant Material:** Roots of *Holostemma ada-kodien* were collected from the Tirumala hill region and were shade dried, powdered and stored in desiccators and powder of root material was subjected to solvent extraction.

**Preparation of Extracts:** 100g of fine powder was taken and soxhlated for 6-8hrs initially with hexane followed by ethyl acetate and methanol. The filtrate was distilled and concentrated under reduced pressure in the Buchi rotavapour R-200 and finally freeze dried to dense residues. The yields of the extracts were 4.2g (w/w), 3g (w/w) and 4.9g (w/w) for hexane, ethyl acetate and methanol respectively. The hexane extract was obtained in the form of oil. All the extracts were preserved in a refrigerator till further use. Preliminary

phytochemical analysis was carried out in all the 3 extracts by different methods of phytochemical analysis<sup>22</sup>.

**Antioxidant Activity:** The following methods were used to evaluate antioxidant activity of the above 3 extracts.

**DPPH Radical Scavenging Activity:** The hydrogen atom or electron donation abilities of the corresponding extracts were measured from the bleaching of the purple-colored methanol solution of 2, 2'-diphenyl-1-picrylhydrazyl (DPPH). This spectrophotometric assay uses the stable radical DPPH as a reagent<sup>23</sup>. 1ml of various concentrations of the extracts (25, 50, 75, 100 and 250µg/ml) in methanol were added to 4ml of 0.004% (w/v) methanol solution of DPPH. After a 30 min incubation period at room temperature, the absorbance was read against blank at 517nm. The ability to scavenge DPPH radical was calculated by the following equation;

$$\text{DPPH radical scavenging activity (\%)} = [(A \text{ control} - A \text{ sample}) / A \text{ blank}] \times 100;$$

Where A- control is the absorbance of the control reaction (containing all reagents except the test compound), A- sample is the absorbance of the test compound and A- blank is only Methanol. Tests were carried out in triplicate.

**Nitric Oxide Scavenging Activity:** Nitric oxide scavenging activity was measured by slightly modified methods<sup>24, 25</sup>. Nitric oxide radicals (NO<sup>•</sup>) were generated from sodium nitroprusside. 1ml of sodium nitroprusside (10mM), 1.5 ml of phosphate buffer saline (0.2M, pH7.4) was added to the different concentrations (25, 50, 75 and 100µg/ml) of the plant extracts and incubated for 150 min at 25°C. After incubation, 1ml of the reaction mixture was treated with 1ml of Griess reagent (1% sulfanilamide, 2% H<sub>3</sub>PO<sub>4</sub> and 0.1% naphthalenediamine dihydrochloride). The absorbance of the chromophore was measured at 546nm. Butylated hydroxy toluene (BHT) was used as a standard. The nitric oxide scavenging activity (%) was calculated by the following equation.

$$\text{Nitric oxide scavenging activity (\%)} = [(A \text{ control} - A \text{ sample}) / A \text{ blank}] \times 100;$$

Where, 'A- control' is the absorbance of the control reaction (containing all reagents except the test compound), 'A- sample' is the absorbance of the test compound and A- blank is only the phosphate buffer saline. Tests were carried out in triplicate.

**Superoxide Anion Scavenging Activity:** Measurement of superoxide anion scavenging activity of *Holostemma ada-kodien* was done based on the Nishimiki method<sup>26</sup>. About 1 ml of Nitroblue tetrazolium (NBT) solution (156µM NBT in 100mM Phosphate buffer, pH 7.4) and 0.1 ml of sample solution of *Holostemma ada-kodien* (100, 250, 500µg/ml) in methanol were mixed. The reaction was started by adding 100µl of phenazine methosulphate (PMS) solution (60µM PMS in 100mM Phosphate buffer, pH 7.4) to the mixture. The reaction mixture was incubated at 25°C for 5 min, and the absorbance of the reaction mixture was measured at 560 nm to study the superoxide anion scavenging activity. Ascorbic acid was used as a positive control.

**Reducing Power:** The reducing power was determined according to the method of Oyaizu<sup>27</sup>. Different concentrations of plant extracts (25, 50, 250 and 500µg/ml) prepared in methanol were mixed with phosphate buffer (2.5 ml, 0.2M, pH 6.6) and potassium ferricyanide [K<sub>3</sub>Fe(CN)<sub>6</sub>] (2.5ml, 1%). The mixture was incubated at 50°C for 20 min and 2.5ml of trichloro acetic acid (10%) was added to the mixture, which was then centrifuged at 3000 rpm for 10 min. The upper layer of the solution (2.5 ml) was mixed with distilled water (2.5 ml) and FeCl<sub>3</sub> (0.5 ml, 0.1%) and the absorbance was measured at 700nm. Increased absorbance of the reaction mixture indicated increased reducing power. Ascorbic acid was used as a standard.

**TLC-DPPH Antioxidant Screening:** This method is generally used for the screening of potential antioxidant activity in crude extracts. It involves the chromatographic separation of the crude plant extract, after which the developed chromatogram is sprayed with a colored radical solution and the presence of antioxidant compounds indicated by the disappearance of radical color. 10µl of each extract was loaded as a 1cm band on the origin of TLC (Merck, Silica gel 60 F254 plates). Plates were developed using different proportions of hexane, ethyl acetate and methanol solvents. Plates loaded with Hexane extract were run with Hexane: Ethyl acetate (1:1); Ethyl

acetate extract loaded plates were run with Ethyl acetate: Methanol (8:2) and Methanolic extract loaded plates were run with 100 % methanol. To detect antioxidant activity, chromatograms were sprayed with 0.2 % (w/v) DPPH in methanol<sup>28</sup>. The presence of antioxidant compounds were detected as yellow spots against a purple background on TLC plates.

**Statistical analysis:** All values are expressed as mean ± S.E. The data was statistically analyzed with Students 't' test.

## RESULTS:

**DPPH Radical Scavenging Activity:** Fig. 1 shows the DPPH radical scavenging activity of Hexane, Ethyl acetate and Methanolic extracts of *Holostemma ada-kodien*. These extracts exhibited the significant dose dependent DPPH radical scavenging activity, with a 50% inhibition (IC<sub>50</sub>) at a concentration of 265.95, 151.51 and 68.18µg/ml respectively. The IC<sub>50</sub> value of the hexane and ethyl acetate extract was found to be lesser than the standards, Ascorbic acid (IC<sub>50</sub> 26.59 µg/ml), BHT (IC<sub>50</sub> 73.52.59µg/ml). Methanolic extract showed good scavenging activity compared to that of BHT.

Ascorbic acid > Methanolic extract > BHT > Ethyl acetate extract > Hexane extract

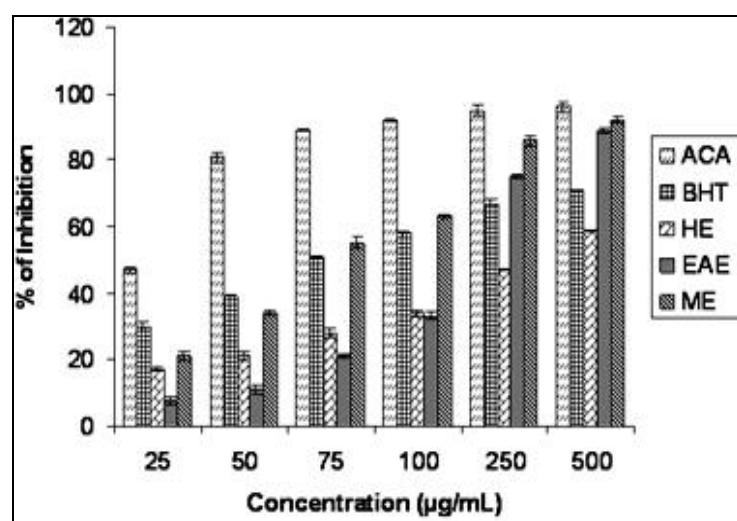
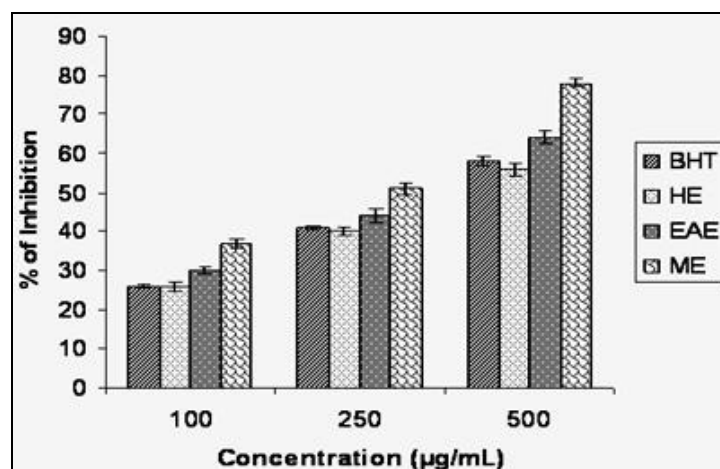


FIG. 1: SCAVENGING EFFECT OF *HOLOSTEMMA ADA-KODIEN* DIFFERENT EXTRACTS, STANDARDS ASCORBIC ACID AND BHT ON 2, 2'-DIPHENYL-1-PICRYL HYDRAZYL (DPPH) RADICAL. Results are mean ± S.E of three parallel measurements

**Nitric Oxide Radical Scavenging Assay:** The scavenging of nitric oxide by Hexane, Ethyl acetate and Methanolic extracts of *Holostemma ada-kodien* increased in a dose-dependent manner as illustrated in **Fig. 2**. The IC<sub>50</sub> values for the above test extracts are 312µg/ml, 284.09µg/ml, 245.098µg/ml respectively. BHT was used as the positive control shown 304µg/ml as IC<sub>50</sub> value. The IC<sub>50</sub> value of Hexane extract found to be lesser than the standard, where as IC<sub>50</sub> values of Ethyl acetate and Methanolic extracts were found to be higher compared to the BHT. IC<sub>50</sub> values of the three extracts are in the following order;

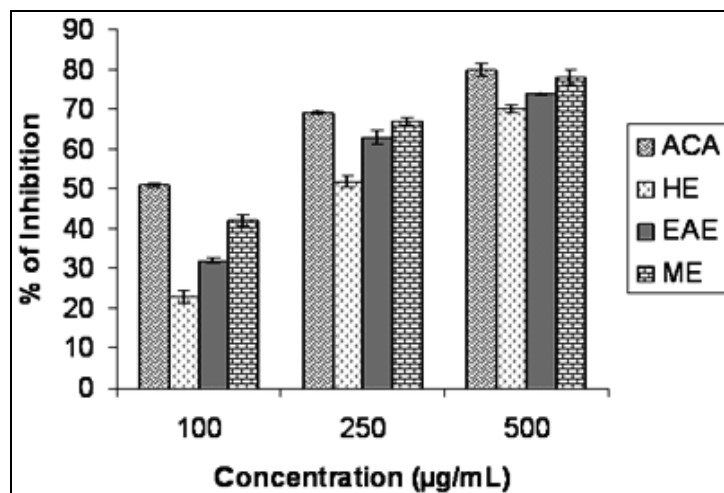
Methanolic extract > Ethyl acetate extract > BHT > Hexane extract



**FIG. 2: SCAVENGING EFFECT OF *HOLOSTEMMA ADA-KODIEN* DIFFERENT EXTRACTS AND STANDARD BHT ON NITRIC OXIDE RADICAL.** Results are mean ± S.E of three parallel measurements.

**Superoxide Anion Scavenging Activity:** The superoxide anion derived from dissolved oxygen by phenazine methosulphate/NADH coupling reaction reduces Nitroblue tetrazolium. The decrease the absorbance at 560 nm with the plant extract thus indicates the consumption of superoxide anion in the reaction mixture. As shown in **Fig. 3**, for the plant extracts Hexane, Ethyl acetate and Methanol, IC<sub>50</sub> values are 240.38µg/ml, 198.4, 119.04µg/ml respectively. IC<sub>50</sub> value of Ascorbic acid (98.03µg/ml) was comparatively good over the above test extracts. Among the three tested extracts, Methanolic extract showed high scavenging compared with Ethyl acetate and hexane extract respectively. Scavenging activity of the extracts is in the following order;

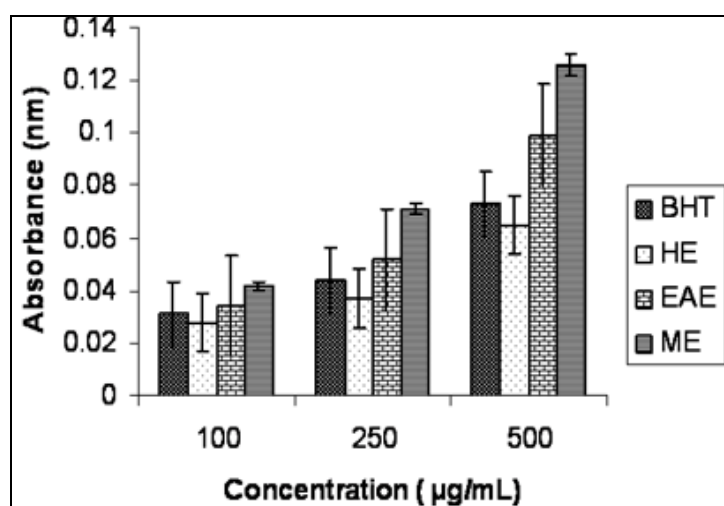
Ascorbic acid > Methanol extract > Ethyl acetate extract > Hexane extract.



**FIG. 3: SCAVENGING EFFECT OF *HOLOSTEMMA ADA-KODIEN* DIFFERENT EXTRACTS AND STANDARD ASCORBOIC ACID ON SUPEROXIDE RADICAL.** Results are mean ± S.E of three parallel measurements

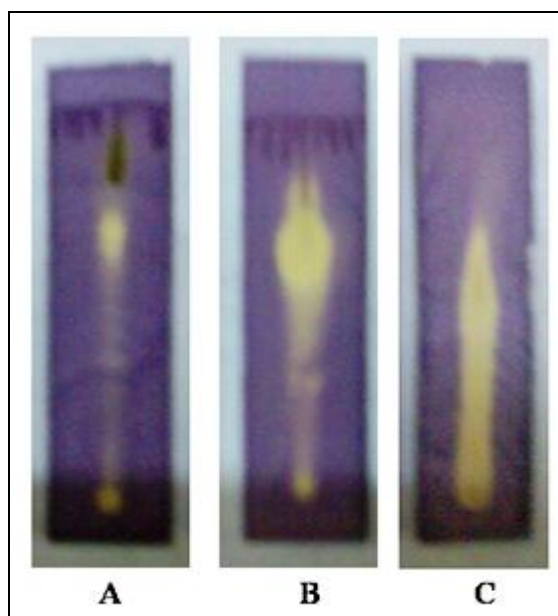
**Reducing Power:** The reducing power of Hexane, Ethyl acetate and Methanolic extracts of *Holostemma ada-kodien* was observed to be very potent. The reducing power of the extracts increased with increasing concentrations from 100, 250, 500µg/ml and the values showed 0.028, 0.037 & 0.065 with Hexane extract, 0.034, 0.052 & 0.099 with Ethyl acetate extract, 0.042, 0.071 & 0.126 with Methanolic extract, and (**Fig. 4**) 0.031, 0.044 & 0.073 with Butylated hydroxyl toluene respectively. The order of reducing ability of the extracts and the standard are as follows.

Methanolic extract > Ethyl acetate extract > BHT > Hexane



**FIG. 4: THE REDUCTIVE ABILITY OF *HOLOSTEMMA ADA-KODIEN* DIFFERENT EXTRACTS AND STANDARD BHT.** Results are mean ± S.E. of three parallel measurements.

**TLC DPPH Assay:** The TLC DPPH assay of three tested extracts showed potent antioxidant activity (**Fig. 5**). Among the three, Methanol extracts had shown highest activity followed by Ethyl acetate and Hexane extract.



**FIG. 5: TLC DPPH CHROMATOGRAMS OF *HOLOSTEMMA ADAKODIEN* DIFFERENT EXTRACTS. (A) HEXANE EXTRACT (B) ETHYL ACETATE EXTRACT (C) METHANOLIC EXTRACT**

**DISCUSSION:** The antioxidant activity was directly correlated with phenolic content. This close correlation between antioxidant activity and phenolic content of extracts obtained from various natural sources has been demonstrated by many workers<sup>29, 30</sup>. It was reported that the solvent used in extraction may also be important in the antioxidant activity of the extract, depending on the phenolic content. DPPH radical scavenging activity was known to correlate well with the inhibitory capacity of lipid peroxide of a test compound<sup>31</sup>.

Sodium nitroprusside in aqueous solution at physiological pH spontaneously generates nitric oxide which interacts with oxygen to produce nitrite ions that can be estimated using Griess reagent. Scavengers of nitric oxide compete with oxygen, leading to reduced production of nitrite ions. In addition to reactive oxygen species, nitric oxide was also implicated in inflammation, cancer and other pathological conditions. The nitric oxide scavenging activity of Hexane, Ethyl acetate and Methanolic extracts increased in a dose dependent manner as illustrated in **Fig. 2**.

Superoxide anion was a free radical created from the normal process of energy generation in the human body. Superoxide radical was known to be very harmful to cellular components as a processor of more reactive oxygen species. Although superoxide anion was a weak oxidant, it gives rise to generation of powerful and dangerous hydroxyl radicals as well as singlet oxygen, both of which contribute to the oxidative stress<sup>32</sup>. The superoxide anion derived from dissolved oxygen by phenazine methosulphate/NADH coupling reaction reduces nitroblue tetrazolium. The decrease in the absorbance at 560nm with the plant extracts indicates the consumption of superoxide anion in the reaction mixture.

In reducing power,  $Fe^{+++}$  reduction was often used as an indicator of electron donating activity, which was an important mechanism of phenolic antioxidant action and can be strongly correlated with other antioxidant properties<sup>33</sup>. Our results were in accordance with other investigators who have also reported that antioxidant properties were concomitant with development of reducing power<sup>34, 35, 36</sup>.

The TLC-DPPH chromatograms (Fig. 5) reveals the presence of antioxidant compounds in all 3 extracts, which were detected as yellow spots against a purple background on TLC plate sprayed with 0.2% DPPH in methanol. From the chromatograms it was clear that methanolic extract having more potent antioxidant compounds than hexane and ethyl acetate extracts.

The present study has shown that Hexane, Ethyl acetate and Methanolic extracts of the *Holostemma adakodien* root tuberous have exhibited very potent antioxidant, free radical scavenging activities.

## REFERENCES:

1. Tiwari AK: Antioxidant: New generation therapeutics base for treatment of polygenic disorders. *Current science* 2004; 86: 1092-1102.
2. Halliwell B: How to characterize an antioxidant: an update. *Biochem. Soc. Symp* 1995; 61: 73-101.
3. Aruoma OI and Cuppett SL: Antioxidant methodology *in vivo* and *in vitro* concepts, AOCS press, Il-lionis. 1997: 1-22.
4. Squadrito GL and Pryor WA: Oxidative chemistry of nitric oxide: the roles of superoxide, peroxynitrite and carbon dioxide. *Free Rad. Biol. Med* 1998; 25: 392-403.
5. Halliwell B and Gutteridge JMC: *Free Radical in Biology and Medicine*. Clarendon Press, Oxford 1989: 23-30.

6. Gulcin I, Buyukokuroglu ME, Oktay M and Kufrevioglu OI: On the *in vitro* antioxidant properties of melatonin. *J. Pineal. Res* 2002; 33: 167-171.
7. Gulcin I, Oktay M, Kufrevioglu OI and Aslan A: Determination of antioxidant activity of lichen *Cetraria islandica* (L). *Ach. J. Ethnopharmacol* 2002; 79: 325-329.
8. Duh PD, Tu YY and Yen GC: Antioxidant activity of water extract of harnng jjur (*Chrysanthemum morifolium Ramat*). *Lebens. Wiss. Technol* 1999; 32: 269-277.
9. Buyukokuroglu ME, Taysi S, Polat F and Gocer F: Mechanism of the beneficial effects of dantrolene sodium on ethanol-induced acute gastric mucosal injury in rats. *Pharmacol. Res* 2002; 45: 421-425.
10. Raquel M and Laura B: Chromatographic and electrophoretic methods for the analysis of biomarkers of oxidative damage to macromolecules (DNA, lipids, and proteins). *J. Sep. Sci* 2007; 30: 175-91.
11. Grotto D, Santa MLD, Boeira S, Valentini J, Charao MF, Moroa AM, Nascimento PC, Pomblumc VJ and Garcia SC: Rapid quantification of malondialdehyde in plasma by high performance liquid chromatography-visible detection. *J. Phr. Biomed. Analysis* 2007; 43: 619-24.
12. Jurgen P, Ingolf M and Christoph HG: Measurement of free and bound malondialdehyde in plasma by high-performance liquid chromatography as the 2, 4-dinitrophenylhydrazine derivative. *J. Chromatography* 2000; B. 742: 315-25.
13. Kumar V and Sharma SK: Antioxidant studies on some plants: a review. *Hamdard Medicus* 2006; 49: 25-36.
14. Kris-Etherton PM, Hecker KD, Bonanome A, Coval SM, Binkoski AE and Hilpert KF: Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. *Am. J. Med* 2000; 113: 715-88.
15. Grice HC: Safety evaluation of butylated hydroxytoluene (BHT) in the liver, lung and gastrointestinal tract. *Food Chem. Toxicol* 1986; 24: 1127-1130.
16. Sherwin ER: Antioxidants. In R. Branen, eds. *Food Additives*. Marcel Dekker, Inc., New York. 1990: 139-193.
17. Kolammal M: Pharmacognosy of Ayurvedic drugs, Department of Pharmacognosy, University of Kerala, Trivandrum. 1979: 21.
18. Sivarajan VV and Balachandran I: *Ayurvedic Drugs and Their Plant Sources*. Oxford & IBM Publ. Co. Pvt. Ltd, New Delhi 1994: 195.
19. Gupta RC: Botanical identity of Jivanti the ayurvedic rejuvenant par excellence. *Appl. Bot. Abstr* 1997; 17: 49-63.
20. Warriar PK, Nambiar VPK and Ramankutti C: *Indian Medicinal Plants: A Compendium of 500 Species*, Vol. 3. Orient Longman. 1995: 167-171.
21. Ramiah N, Nair GA and Prasad NBR: Chemical components of *Holostemma annulare* K. Schum. *J. Sci. Res. Pl. Med* 1981; 2: 76-78.
22. Harborne JB: *Phytochemical Methods: A Guide to Modern Techniques of Plant analysis*, 3<sup>rd</sup> ed. Springer (India) Pvt. Ltd, New Delhi, 1998.
23. Burits M and F Bucar: Antioxidant activity of *Nigella sativa* essential oil. *Phytother Res* 2000; 14: 323-328.
24. Green LC, Wagner DA, Glogowski Skipper PL, Wishnok JS and Tannenbaum SR: Analysis of nitrate, nitrite, and [15N] nitrate in biological fluids. *Anal Biochem* 1982; 126: 131-138.
25. Marcocci L, Maguire JJ, Droy-Lefaix MT and Packer L: The nitric oxide-scavenging properties of *Ginkgo biloba* extract. *Biochem Biophys Res Commun* 1994; 201: 748-55.
26. Nishimiki M, Appaji N and Yagi K: The occurrence of superoxide anion in the reaction of reduced phenazine methosulphate and molecular oxygen. *Biochem. Biophys. Res. Commun.* 1972; 46: 849.
27. Oyaizu M: Studies on products of browning reactions: antioxidant activities of products of browning reaction prepared from glucose amine. *Jap. J. Nutr* 1986; 44: 307-315.
28. Deby C and Margotteaux G: Relationship between essential fatty acids and tissue antioxidant levels in mice. *CR Seances Society Soc Biol Fil* 1970; 165: 2675-2681.
29. Liu X, Dong M, Chen X, Jiang M, Lv X, and Yan G: Antioxidant activity and phenolics of an endophytic *Xylaria* sp. From *Ginkgo biloba*. *Food Chem* 2007; 105: 548-554.
30. Verzelloni E, Tagliacuzzi D and Conte A: Relationship between the antioxidant properties and the phenolic and flavonoid content in traditional balsam vinegar. *Food Chem* 2007; 105: 564-571.
31. Rekkas E and Kourounakis PN: Effect of hydroxyethyl rutosides and related compounds on lipid peroxidation and free radical scavenging activity. Some structural aspects. *J. Pharm. Pharmacol* 1991; 43: 486-91.
32. Mayer AS and Isaksen A: Application of enzymes as food antioxidants. *Trends Food Sci. Technol* 1995; 6: 300-304.
33. Dorman HJD, Peltoketo A, Hiltunen R and Tikkanen MJ: Characterization of the antioxidant properties of de-odorized aqueous extracts from selected Lamiaceae herbs. *Food Chem* 2003; 83: 255-262.
34. Chung YC, Chen SJ, Hsu CK, Chang CT and Chou ST: Studies on the antioxidative activity of *Graptopetalum paraguayense* E. Walther. *Food Chem* 2005; 91: 419-424.
35. Duh PD and Yen GC: Antioxidative activity of three herbal water extracts. *Food Chem* 1997; 60: 639-645.
36. Kanatt SR, Chander R and Sharma A: Antioxidant potential of mint (*Mentha spicata* L.) in radiation-processed lamb meat. *Food Chem* 2

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