



Received on 27 May, 2011; received in revised form 24 August, 2011; accepted 29 August, 2011

VERMICIDAL ACTIVITY OF *MILLETTIA PACHYCARPA* ON THE NEMATODE, *HETERAKIS GALLINARUM*

K. Lalchhandama

Department of Zoology, Pachhunga University College, Mizoram University, Aizawl 796001, Mizoram, India

ABSTRACT

Keywords:

Albendazole,
Heterakis gallinarum,
Medicinal Plant,
Millettia pachycarpa,
Nematode

Correspondence to Author:

K. Lalchhandama

Department of Zoology, Pachhunga
University College, Mizoram University,
Aizawl 796001, India

The stem and root barks, leaves and seeds of *Millettia pachycarpa* Benth. are used for various medicinal purposes in Chinese and Mizo traditional practices. The plant extract reportedly showed potent vermicide activity on cestodes. The ethanol extract of the root bark was tested against the poultry nematode, *Heterakis gallinarum* Schrank. *In vitro* treatment at 37±1°C revealed that a serial dose of the plant extract caused dose-dependent vermicide effect on the worm (significant efficacy at $P < 0.05$), except at the lowest dose (i.e. 5 mg/mL). A broad-spectrum drug albendazole also exhibited similar effect but with higher potency at all doses applied. The nematode treated with 80 mg/mL of the plant extract caused apparent destruction on the morphological structure, mostly of severe distortion in the cuticular organization throughout the body topography. Collapse of the lips and contraction of the body proper with prominent folding of the cuticle at the cephalic region were clearly discernible. At the posterior end were also extensive disintegration of the cuticle along with shrivelled cloacal mount and caudal papillae.

INTRODUCTION: Helminth parasites remain one of the major concerns in human health, particularly in developing countries where they inflict serious morbidity among children, and the principal cause of economic loss in livestock production. The problem is compounded by development of resistance in helminths to all kinds of antihelmintic drugs¹. Major efforts have been deployed in investigating medicinal plants to combat the pervasive helminth infections².

Millettia pachycarpa Benth. (Family: Fabaceae) is a leguminous perennial climbing tree native to south-east Asian region where it is used in various traditional practices. The root bark, seed and leaf are commonly used as a blood tonic, treatment of infertility, fish stupefying, anticancer and insecticidal agents^{3, 4}. A number of novel bioactive compounds is identified from the plant, of which barbigerone and isoflavones such as erysenegalensein E, isoerysenegalensein E, 6, 8-diprenylorobol, millewanins G and H, furowanin A

and B, and auriculasin were all demonstrated to have antiestrogenic activity⁵⁻⁷. Following the traditional usage of the Mizo tribes of north-east India, the extract of the root bark was demonstrated to have potent vermicide activity against the cestode, *Raillietina echinobothrida*^{8, 9}. This study is the first attempt to examine the antinematodal activity of the plant against a poultry cecal roundworm, *Heterakis gallinarum* Schrank.

MATERIALS AND METHODS:

Plant Material: The fresh roots of *M. pachycarpa* were collected from the nearby forest of Aizawl, Mizoram, India. Identification and authentication of the plant material were reported earlier⁸. The root barks were peeled off, thoroughly washed with deionized water, cut into small pieces, and dried in a hot air oven at 50°C. The dried parts were macerated to fine powder and then refluxed with ethanol (100 g/L) for 8 h at 60°C.

The solution obtained was filtered through Whatman filter paper (No. 1) and the solution was evaporated to complete dryness at 50°C. The ethanol extract was obtained as a deep brown powdered material, which was then refrigerated at 4°C until further use. 1 h prior to *in vitro* assay, a stock concentration of the extract, 80 mg/mL, was prepared by dissolving in 0.9% neutral phosphate-buffered saline (PBS, pH~ 7.1) supplemented with 1% dimethylsulfoxide (DMSO). Varying doses, viz. 5, 10, 20 and 40 mg/L, were then prepared by serial dilution of the stock solution using PBS with DMSO.

Chemicals and Drug: All chemicals used were of standard analytical grades, obtained either from Merck or S.D. Fine-Chemicals Limited, India, except where otherwise stated. Ethanol was supplied by Bengal Chemicals, Kolkata, India, and the reference drug albendazole (Zentel®) is a product of GlaxoSmithKline Pharmaceutical Limited, India.

Recovery and Treatments of Nematodes: Native bred live chickens (*Gallus domesticus* Linn.) were obtained from the local abattoir in Aizawl, Mizoram, India. They were sacrificed and upon immediate necropsy, *H. gallinarum* were recovered from the cecum. Guidelines of the institutional Animal Care and Use Committee were observed. The worms were washed gently with and collected in PBS and then maintained at $37 \pm 1^\circ\text{C}$ in a digital glass-chambered incubator. The fresh worms were directly introduced to the serial doses (5, 10, 20, 40 and 80 mg/mL) of the plant extract. Similar treatment with corresponding doses was performed for albendazole. The control experiment consisted of nematodes maintained in a medium containing only PBS with 1% DMSO. Each medium was made in 5 replicates. Death was substantiated when complete immobility was noted upon dipping the parasites in tepid PBS ($\sim 45^\circ\text{C}$) that induced movement in sentient worms.

Scanning Electron Microscopy: A set of nematodes from each of the plant extract-treated and control media were washed in PBS, and then fixed in 4% formaldehyde at 4°C for 24 h. After post fixation in 1% buffered osmium tetroxide for 1 h, the worms were dehydrated through ascending concentration of acetone. Following the standardized method of Roy and Tandon for helminth parasites⁹, the specimens

were treated with tetramethylsilane for 10 min and then allowed to dry at room temperature ($25 \pm 2^\circ\text{C}$). The specimens were placed on metal stubs and sputter-coated with gold in a fine-coat ion sputter, JFC-1100 (JEOL), and finally observed under scanning electron microscope (JSM-5360) at an electron accelerating voltage of 20 kV.

Data Analysis: Data were presented as means plus or minus the standard deviation (SD) of the mean. Comparison of the mean values of the experimental treatments against those of the control group was made using unpaired Student's *t*-test, and the level of probability considered significant when $P < 0.05$.

RESULTS: *H. gallinarum* in the control experiment survived for 97.73 ± 0.90 h in a medium composed only of PBS with 1% DMSO. The nematodes thrived relentlessly, but once their movement ceased, death ensued abruptly. **Table 1** presents the response in physical activity of the nematodes after treatment with a broad-spectrum antiparasitic drug, albendazole, and the extracts of *M. pachycarpa* at the doses of 5, 10, 20, 40 and 80 mg/mL, respectively. Albendazole was found to be a highly effective nematocide exerting profound dose-dependent activity at all doses tested. The plant extract also indicated concentration-dependent efficacy on the nematode. However, at the lowest concentration, i.e. 0.5 mg/mL, the nematodes did not show any significant mortality with respect to the control, although average death was observed earlier than those in control group. **Figure 1** shows the comparative efficacy of the drug and the plant extract, depicting dose-dependent activity for both cases.

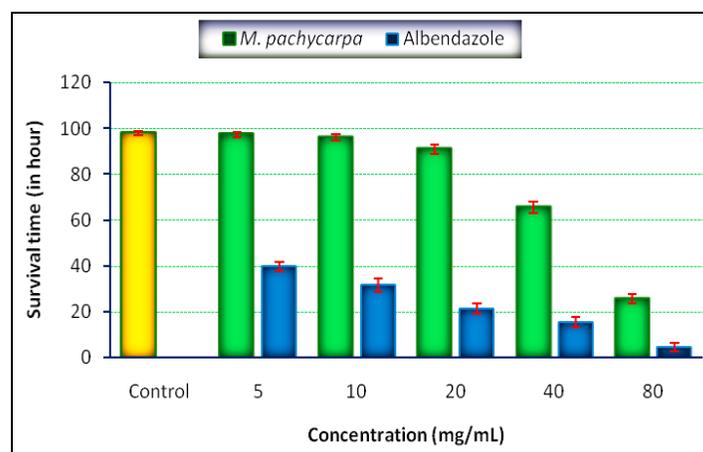


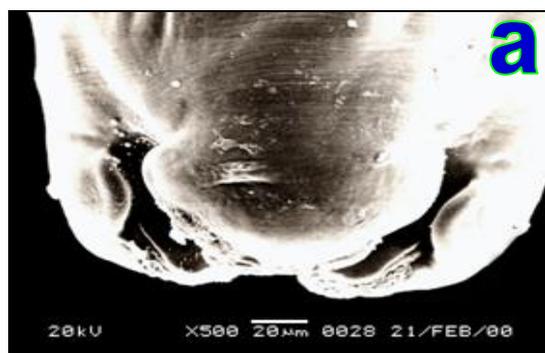
FIG. 1: COMPARISON OF THE EFFICACY OF ALBENDAZOLE AND THE EXTRACT OF *M. PACHYCARPA* ROOT BARK ON THE SURVIVAL OF *H. GALLINARUM*

TABLE 1: EFFECTS OF ALBENDAZOLE AND THE EXTRACT OF *M. PACHYCARPA* ROOT BARK ON THE SURVIVAL OF *H. GALLINARUM*

Incubation medium	Dose (mg/mL)	Time (h) taken for death	df	t value	Probability level
Control	0	97.73 ± 0.90	--	--	--
	5	97.28 ± 1.13	8	0.69	$P > 0.05$
<i>M. pachycarpa</i> extract	10	96.10 ± 1.31	8	2.29	$P < 0.05$
	20	90.84 ± 1.89	8	7.36	$P < 0.05$
	40	56.59 ± 1.91	8	27.21	$P < 0.05$
	80	07.25 ± 1.10	8	76.39	$P < 0.05$
	5	39.71 ± 2.03	8	58.36	$P < 0.05$
Albendazole	10	31.55 ± 3.03	8	46.76	$P < 0.05$
	20	21.33 ± 2.30	8	69.13	$P < 0.05$
	40	15.41 ± 2.20	8	77.54	$P < 0.05$
	80	04.52 ± 1.84	8	101.65	$P < 0.05$

Values are expressed as mean ± SD ($n = 5$); P value significant at < 0.05 in comparison with control group; level of confidence at 95%.

Scanning electron microscopy of the untreated nematode revealed prominent triangular mouth at the extreme anterior end, which is surrounded by three conspicuous lips having mouth cuticle. Sensory organs named labial papillae are situated on each lip (**Figure 2a**). The posterior end is structurally elaborate and complex in males than in females. The male tail is characteristically curved ventrally and studded with numerous sensory caudal papillae, (**Figure 2b**).



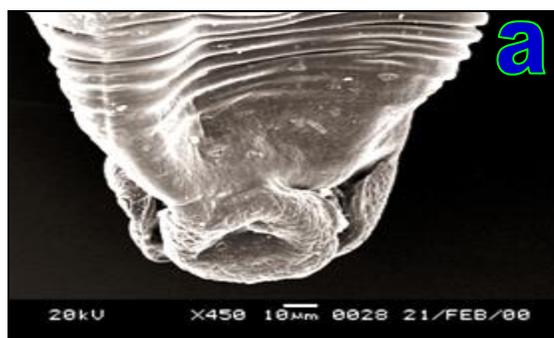
a) Anterior end showing three denticulate lips surrounding a central mouth; the cuticle with distinct transverse striations on the body proper



b) Tail end of male nematode showing a prominent cloaca and surrounding bulb-like papillae

FIG. 2: SCANNING ELECTRON MICROGRAPHS OF NORMAL *HETERAKIS GALLINARUM*

The nematode treated with 80 mg/mL of the plant was selected as it indicated the most elaborate structural alterations. At the anterior end, all the three lips showed signs of loosening and collapse, with conspicuous aberrant wrinkles all over the surface (**Figure 3a**). Cuticular contraction extended throughout the body, and the tail region of the male showed collapse of the cloacal mount due to shrinkage of the cuticle and disappearance of the papillae (**Figure 3b**).



a) Anterior end with severe shrinkage, lips collapsed and depressed at the centre, and the cuticle wrinkled with mutilations



b) Tail end indicating extensive contraction and collapse of the cloacal mount, with disappearance of the papillae

FIG. 3: SCANNING ELECTRON MICROGRAPHS OF *HETERAKIS GALLINARUM* TREATED WITH 80 MG/ML OF THE EXTRACT OF *M. PACHYCARPA* ROOT BARK

DISCUSSION: A considerable number of plants such as *Acacia polyacantha*, *Albizia lebbek*, *Allium sativum*, *Anogeissus leiocarpus*, *Azima tetracantha*, *Bridelia micrantha*, *Butea monosperma*, *Caesalpinia crista*, *Capparis decidua*, *Carica papaya*, *Cassia sieberiana*, *Centrathecum anthelminticum*, *Cleome icosandra*, *Commiphora mukul*, *Cucurbita mexicana*, *Mimosup elengi*, *Neurolaena lobata*, *Nigella sativa*, *Ocimum sanctum*, *Piliostigma thonningii*, *Piper longum*, *Punica granatum*, *Pterospermum acerifolium*, *Rhynchosia minima*, *Strobilanthes discolor*, *Thespesia lampas*, *Trachuspermum ammi*, *Trifolium repens*, *Xylopia aethiopica* and *Zingiber officinale* have been documented to exhibit significant activity on a variety of nematodes¹¹⁻¹⁵.

However, further studies on the actual antihelmintic effects on the worms are scanty. The cuticle of nematodes is a complex proteinaceous layer that is metabolically active and specialized to perform selective absorption of nutrients, secretion of glycoproteins for immuno-protection, osmoregulation and (insofar as it supports sense organs) sensory reception. Consequently, passive diffusion through the cuticle is the principal mechanism by which antihelmintic compounds enter the nematode body¹⁶. Apparently, it has been firmly documented that one of the hallmark effects of any antihelmintic is the direct destruction of the worm's surface^{17, 18}. Indeed a number of antihelmintic agents have been shown to act on nematodes by causing structural alterations most notably on the cuticle.

Albendazole reportedly caused severe damage on the cuticle of *Trichinella spiralis* upon *in vivo* treatment¹⁹. Surface distortion and loss of regular cuticular annulations were also observed on *Brugia malayi*¹⁸. Adult *Wuchereria bancrofti* subjected to albendazole and diethylcarbamazine combination therapy exhibited swollen cuticle, formation of spherical and spike-like projections at the anterior region, and leaf-like expansion on the general cuticle²⁰. *Angiostrongylus contonensis* also developed severe shrinkage and formation of rounded leaf-like expansions on the cuticle throughout the body after *in vivo* treatment with imidacloprid and moxidectin combination¹⁷. Cysteine proteinases isolated from different fruits were shown to cause wrinkles and folds of the cuticle,

often followed by blistering and gradual digestion of the cuticle on different nematodes²¹⁻²³.

Extensive deformity on the cuticle, lips, body striations and musculature were observed on *Ascaridia galli* after treatment with the extract of *Acacia oxyphylla* stem bark²⁴. Tribendimidine also caused severe disruption of the cuticle and intestinal epithelium in *Necator americanus*²⁶. Cyclosporin A caused massive disorganization of the cuticle in *Trichinella spiralis*²⁷.

Morphological alterations due to *M. pachycarpa* on *H. gallinarum* in the present study are most probably a common primary antihelmintic action whereby cuticular organization is the prime target. However, the active principle of the plant and its mode of action at physiological level remain uncertain.

REFERENCES:

1. Woods DJ and Knauer CS: Discovery of veterinary antiparasitic agents in the 21st century: A view from industry. *Int. J. Parasitol.* 2010; 40:1177-1181.
2. Murthy PK, Joseph SK and Murthy PS: Plant products in the treatment and control of filariasis and other helminth infections and assay systems for antifilarial/antihelmintic activity. *Planta Med.* 2011; 77:647-661.
3. Perry LM and Metzger J: Medicinal Plants of East and Southeast Asia: Attributed Properties and Uses. MIT Press, Massachusetts, USA, 1980; pp. 219-220.
4. Lalchhandama K: Pharmacology of Some Traditional Antihelmintic Plants: Biochemical and Microscopic Studies. Lambert Academic Publishing, Saarbrücken, Germany, 2010; pp. 145.
5. Ito C, Itoigawa M, Kumagaya M, Okamoto Y, Ueda K, Nishihara T, Kojima N and Furukawa H: Isoflavonoids with antiestrogenic activity from *Millettia pachycarpa*. *J. Nat. Prod.* 2006; 69:138-141.
6. Okamoto Y, Suzuki A, Ueda K, Ito C, Itoigawa M, Furukawa H, Nishihara T and Kojima N: Anti-estrogenic activity of prenylated isoflavonoids from *Millettia pachycarpa*: implications for pharmacophores and unique mechanisms. *J. Health Sci.* 2006; 52:186-191.
7. Ye H, Zhong S, Li Y, Tang M, Peng A, Hu J, Shi J, He S, Wu W and Chen L: Enrichment and isolation of barbigerone from *Millettia pachycarpa* Benth. using high-speed counter-current chromatography and preparative HPLC. *J. Sep. Sci.* 2010; 33:1010-1017.
8. Roy B, Lalchhandama K and Dutta BK: Scanning electron microscopic observations on the *in vitro* antihelmintic effects of *Millettia pachycarpa* on *Raillietina echinobothrida*. *Pharmacogn. Mag.* 2008; 4:20-26.
9. Lalchhandama K, Roy B and Dutta BK: Effect of *Millettia pachycarpa* on the trace metals and tegumental enzymes of *Raillietina echinobothrida*. *Pharmacogn. Mag.* 2008; 4:254-261.
10. Roy B and Tandon V: Usefulness of tetramethylsilane in the preparation of helminth parasites for scanning electron microscopy. *Riv. Parasitol.* 1991; 8:207-215.

11. Stepek G, Behnke JM, Buttle DJ and Duce IR: Natural plant cysteine proteinases as antihelminthic? Trends Parasitol. 2004; 20:322-327.
12. Iqbal Z, Jabbar A, Akhtar MS, Muhammad G and Lateef M: Possible role of ethnoveterinary medicine in poverty reduction in Pakistan: use of botanical anthelmintics as an example. J. Agri. Soc. Sci. 2005; 1:187-195.
13. Mali RG and Mehta AA: A review on antihelminthic plants. Nat. Prod. Rad. 2008; 7:466-475.
14. Waterman C, Smith RA, Pontiggia L and DerMarderosian A: Antihelminthic screening of Sub-Saharan African plants used in traditional medicine. J. Ethnopharmacol. 2010; 127:755-759.
15. Mahesh B and Ramling P: Indian medicinal plants as natural antihelminthic agents: an overview. J. Pharmaceut. 2010; 1:11-18.
16. Alvarez LI, Mottier ML and Lanusse CE: Drug transfer into target helminth parasites. Trends Parasitol. 2007; 23:97-104.
17. Schmahl G, Mehlhorn H, Harder A, Klimpel S and Krieger KJ: Efficacy of a combination of imidacloprid plus moxidectin against larval and adult stages of nematodes (*Trichuris muris*, *Angiostrongylus cantonensis*) in rodents. Parasitol. Res. 2007; 101:S85-S92.
18. Tippawangkosol P, Choochote W, Na-Bangchang K, Jitpakdi A, Pitasawat B and Riyong D: The *in vitro* effect of albendazole, ivermectin, diethylcarbamazine, and their combinations against infective third-stage larvae of nocturnally subperiodic *Brugia malayi* (Narathiwat strain): Scanning electron microscopy. J. Vector Ecol. 2004; 29:101-108.
19. Hrkova G, Velebný S and Horak J: A morphological study of the effects of liposomized albendazole on the muscle phase of *Trichinella spiralis* in mice. J. Helminthol. 1993; 67:24-30.
20. Oliveira-Menezes A, Lins R, Norões J, Dreyer G and Lanfredi RM: Comparative analysis of a chemotherapy effect on the cuticular surface of *Wuchereria bancrofti* adult worms *in vivo*. Parasitol. Res. 2007; 101:1311-1317.
21. Stepek G, Buttle DJ, Duce IR, Lowe AE and Behnke JM: Assessment of the antihelminthic effect of natural plant cysteine proteinases against the gastrointestinal nematode, *Heligmosomoides polygyrus*, *in vitro*. Parasitology 2005; 130:1-9.
22. Stepek G, Lowe AE, Buttle DJ, Duce IR and Behnke JM: *In vitro* and *in vivo* antihelminthic efficacy of plant cysteine proteinases against the rodent gastrointestinal nematode, *Trichuris muris*. Parasitology 2006; 132:681-689.
23. Stepek G, Lowe AE, Buttle DJ, Duce IR and Behnke JM: Anthelmintic action of plant cysteine proteinases against the rodent stomach nematode, *Protospirura muricola*, *in vitro* and *in vivo*. Parasitology 2007; 134:103-112.
24. Lalchandama K: Nematocidal effects of piperazine and the extract of *Acacia oxyphylla* stem bark on the poultry nematode, *Ascaridia galli*. Pharmacologyonline 2008; 3:864-869.
25. Lalchandama K, Roy B and Dutta BK: Antihelminthic activity of *Acacia oxyphylla* stems bark against *Ascaridia galli*. Pharmaceutical Biology 2009; 47:578-583.
26. Xiao SH, Ren HN, Da-i ZQ, Yang YQ and Zhang CW: Light and EM observations on effects of tribendimidin on cuticle of *Necator americanus* and small intestinal mucosa of infected golden hamsters. Acta Pharmacol. Sin. 1989; 10:90-92.
27. Boulous LM, Abu-Samra LM and el-Azzouni MZ: Cyclosporin A in experimental trichinosis scanning electron microscopic study. J. Egypt. Soc. Parasitol. 1992; 22:767-773.
