



Received on 18 December, 2014; received in revised form, 18 February, 2015; accepted, 26 April, 2015; published 01 August, 2015

BIOLOGICAL INVESTIGATIONS OF THE LEAF EXTRACT OF *SPONDIAS PINNATA*

Sonia Kamal, Rumana Akhter, Nazma Akter Tithi, Md. Abdul Wadud, Syeda Najah Narjish, Mohammad Shahriar and Mohiuddin Ahmed Bhuiyan*

Phytochemistry Research Laboratory, Department of Pharmacy, University of Asia Pacific, Dhaka, Bangladesh

Keywords:

Spondias pinnata, Membrane stabilizing, Neuropharmacological activity, GI Motility, Acute Toxicity.

Correspondence to Author: Mohiuddin Ahmed Bhuiyan

Professor & Head, Department of Pharmacy, University of Asia Pacific
House no. 73, Road no. 5A, Dhanmondi,
Dhaka - 1209, Bangladesh

E-mail: mohiuddin@uap-bd.edu

ABSTRACT: The present study was done to evaluate in-vitro membrane stabilizing activity with human RBC, *in-vivo* neuropharmacological activity including open field and swimming test, gastrointestinal motility and acute toxicity of different leaf extracts of *Spondias pinnata* in Swiss albino mice following oral administration. *In-vivo* GI motility test was done by charcoal induced anti motility test, acute toxicity study was done by investigating mortality/morbidity status of test animal and *in-vitro* membrane stabilizing activity was done with human RBC in both hypotonic and heat induced method. In case of *in-vitro* membrane stabilizing activity for hypotonic solution induced haemolysis, the ethanol extract inhibited 76.92% haemolysis of RBCs as compared to 85.42% produced by acetyl salicylic acid and during heat induced condition different organic soluble materials of *S. pinnata* demonstrated 76.68%, 77.19% and 74.83% inhibition of haemolysis of RBCs respectively whereas ASA inhibited 78.04%. *In-vivo* neuropharmacological activity showed significant ($p < 0.05$) and ($p < 0.001$) results when methanol 200 mg/kg, ethanol 200 mg/kg, chloroform 100 & 200 mg/kg extracts were administered to evaluate the rate of movement with time in dose dependent manner when compared with corresponding value of control group. In case of forced swimming test methanol 200 mg/kg, ethanol 200 mg/kg, chloroform 100 & 200 mg/kg extracts showed significant ($p < 0.001$) and ($p < 0.05$) results corresponding value of control group. Both the dosage of methanol and ethanol extracts of 100 mg/kg and 200 mg/kg showed significant ($p < 0.05$) and ($p < 0.001$) results in gastrointestinal motility comparing with standard. None of the extracts showed any *in-vivo* acute toxicity effect on mice.

INTRODUCTION: *Spondias pinnata* is a highly ornamental tree, reaching a height of 12 to 18 m with upright growth. It is wholly or partly deciduous with occasionally small buttress. Its bark surface is smooth, with irregular cracks, greyish to pale reddish brown, exuding a clear, sticky sap with turpentine smell. The leaves are aromatic, acidic and astringent.

They are arranged spirally, leaflets are alternate to opposite. They have a sour taste and used for flavoring. Flowers are stalkless, white and bisexual. Fruit is yellow in color, fleshy, drupe with a finely flavored edible pulp; seed is hard, ridged and has a fibrous surface. It is being cultivated commercially in our country for both local and international market.

The bark is astringent and refrigerant; infusion of the bark is given in dysentery, diarrhoea and to prevent vomiting. Paste of the bark is used as an embrocation for both articular and muscular rheumatism. Decoction of the bark is given in gonorrhoea. Gum of the bark is demulcent. Roots are useful in regulating menstruation. The leaves

QUICK RESPONSE CODE 	DOI: 10.13040/IJPSR.0975-8232.6(8).3351-58
	Article can be accessed online on: www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.6(8).3351-58	

are appetizing and astringent. Fruit possesses antiscorbutic and astringent properties; used in bilious dyspepsia. The unripe fruit is good for rheumatism and sore throat. Ripe fruit is tonic, aphrodisiac and astringent to the bowels; cures burning sensation¹. As a part of our continuing studies on medicinal plants of Bangladesh the organic soluble materials of the leaf extracts of *Spondias pinnata* were evaluated for membrane stabilizing activity through hypotonic solution and heat induced haemolysis, *in-vivo* neuropharmacological activity through open field test and forced swimming test, gastrointestinal motility and acute toxicity for the first time²⁻⁶.

MATERIALS AND METHODS:

Collection and Processing of Plant Samples:

Fresh leaves of *Spondias pinnata* was collected from Dhaka, Bangladesh in July 2014 and a plant sample was submitted to the Bangladesh National Herbarium for identification (Accession number-39649). Leaf was sun dried for seven days. The dried leaf were then ground in coarse powder using high capacity grinding machine which was then stored in air-tight container with necessary markings for identification and kept in cool, dark and dry place for the investigation.

Extraction Procedure:

The powdered plant parts (22gm) were successively extracted in a soxhlet extractor at elevated temperature using 250 ml of distilled Methanol (40-60) °C which was followed by ethanol, and chloroform. After extraction all extracts kept in refrigerator 4°C for future investigation with their necessary markings for identification. Then kept in cool, dark and dry place for further investigation.

Blood sample:

Blood (n=6) was drawn from healthy human volunteers without a history of oral contraceptive or anticoagulant therapy and 5ml of blood was transferred to the previously weighed micro centrifuge tubes and was allowed to form clots.

Membrane stabilizing activity:

The membrane stabilization by hypotonic solution and heat-induced haemolysis method was used to assess anti-inflammatory activity of the plant

extracts by following standard protocol⁷⁻⁸. Since the erythrocyte membrane resembles to lysosomal membrane and as such, the effect of drugs on the stabilization of erythrocyte could be extrapolated to the stabilization of lysosomal membrane⁹. The membrane stabilizing activity of the extractives was assessed by using hypotonic solution-induced and heat-induced human erythrocyte haemolysis. To prepare the erythrocyte suspension, whole blood was obtained from healthy human volunteer and was taken in syringes (containing anticoagulant 3.1% Na-citrate). The blood was centrifuged and blood cells were washed three times with solution (154mM NaCl) in 10mM sodium phosphate buffer (pH 7.4) through centrifugation for 10min at 3000g¹⁰⁻¹¹.

Hypotonic solution-induced haemolysis:

The test sample consisted of stock erythrocyte (RBC) suspension (0.5mL) mixed with 5mL of hypotonic solution (50mM NaCl) in 10mM sodium phosphate buffered saline (pH 7.4) containing either the extract (1.0mg/mL) or acetyl salicylic acid (ASA) (0.1mg/mL). The control sample consisted of 0.5mL of RBCs mixed with hypotonic-buffered saline alone. The mixture was incubated for 10min at room temperature, centrifuged for 10min at 3000g and the absorbance of the supernatant was measured at 540nm. The percentage inhibition of either haemolysis or membrane stabilization was calculated using the following equation:

$$\% \text{ inhibition of hemolysis} = 100 \times (\text{OD}_1 - \text{OD}_2) / \text{OD}_1$$

where, OD₁= optical density of hypotonic-buffered saline solution alone (control)

OD₂= optical density of test sample in hypotonic solution

Heat-induced haemolysis:

Isotonic buffer containing aliquots (5ml) of the different extractives were put into two duplicate sets of centrifuge tubes. The vehicle, in the same amount, was added to another tube as control. Erythrocyte suspension was added to each tube and mixed gently by inversion. One pair of the tubes was incubated at 56°C for 30min in a water bath, while the other pair was maintained at (0-5) °C in an ice bath. The reaction mixture was centrifuged for 5min at 2500g and the absorbance of the

supernatant was measured at 560 nm. The percentage inhibition or acceleration of hemolysis in tests and was calculated according to the equation:

$$\% \text{ Inhibition of hemolysis} = 100 \times [1 - (\text{OD}_1 - \text{OD}_2) / (\text{OD}_3 - \text{OD}_1)]$$

Where, OD_1 = optical density of unheated test sample

OD_2 = optical density of heated test sample

OD_3 = optical density of heated control sample

Experimental animals:

For the experiment Swiss albino mice of either sex, 4-5 weeks of age, weighing between 10-24 gm were collected from ICDDR, B, Dhaka. Animals were maintained under standard environmental conditions (temperature: $(27.0 \pm 1.0)^\circ\text{C}$, relative humidity: (55-65) % and 12 hour light/12 hour dark cycle) and free access to feed and water. The animals were acclimatized to laboratory condition for one week prior to experiments. All protocols for animal experiment were approved by the institutional animal ethical committee.

Neuropharmacological study:

To check the neuropharmacological effects or side-effects of drug, two types of experiment is carried out which are open field test and swimming test.

Open field test (OFT):

According to previous work with slight modification open field test was performed to monitor behavioral responses in mice that were placed in a novel and bright arena¹². Rodents tend to stay away from brightly illuminated areas. The experiment also assesses a range of anxiety-induced, locomotor activity and exploratory behaviors. The animals were divided into 8 groups of 5 mice each. The first group was given 10ml/kg of 1% Tween 80 orally and served as control. Group 2 was served 2mg of clonazepam per kg of body weight and it served as standard. Groups 3, 4 received methanol extract of the leaves of *Spondias pinnata* at 100 and 200mg/kg of body weight, and groups 5, 6 received ethanol extract of the leaves of *Spondias pinnata* at 100 and 200mg/kg of body weight. Group 7, 8 received chloroform extracts of the leaves of *Spondias pinnata* at 100 and 200mg/kg of body weight. The open field apparatus

is made of hardboard (60cm x 60cm; 40cm walls). Blue lines drawn on the floor divide the floor into thirty six squares 10cm x 10cm squares alternatively colored black and white and Central Square (10cm x 10cm) in the middle clearly marked. The number of squares visited by the animals was calculated for 2min at 0, 30, 60, 90 and 120min subsequent to oral administration of the experimental crude extracts¹².

Forced swimming test (FST):

According to Porsolt *et al.*, 1978 swimming test was performed with slight modification¹³. Animals were randomly divided into 8 groups with 5 mice on each group. Group 1 was given 10ml/kg of 1% Tween 80 which served as control and Group 2 was given 2mg of diazepam per kg of body weight which served as standard. Groups 3, 4 received methanol extracts of the leaf of *Spondias pinnata* at 100 and 200mg/kg of body weight. Groups 5, 6 received ethanol extracts of the leaf of *Spondias pinnata* at 100 and 200 mg/kg of body weight.

Group 7, 8 received chloroform extracts of *Spondias pinnata* at 100 and 200mg/kg of body weight. The forced swim test was carried out on mice individually forced to swim in an open acquire water tank apparatus (29cm x 19cm x 20cm), containing 9 cm of water at $25 \pm 1^\circ\text{C}$. The total duration of immobility during the 4-min test was scored as described. Each mouse was judged to be immobile when it ceased struggling and remained floating motionless in the water, making only those movements necessary to keep its head above water. The duration of immobility was recorded. Decrease in the duration of immobility during the FST was taken as a measure of antidepressant activity¹³.

Gastrointestinal motility determination:

Forty eight Swiss Albino mice, weighing between 10-20 g were selected and housed properly for 10 days before performing the experiment. On the test day, the animals were divided into eight groups of six mice each. They were weighed and deprived of food, with free access to water. Three hours after food deprivation, the animals in group 1 received orally by gavages 5 ml/kg body weight of 0.9% NaCl (normal saline) as control group, while those in group 2 received 5 mg/kg body weight of

butapen as standard group. The other six groups received their respective doses as shown in the table. After 90 min, 0.3 ml of an aqueous suspension of 5% charcoal in normal saline was administered to each animal orally by gavages (time 90 min). Sixty minutes later they had free access to food (time 150 min).

The animals were observed at 5 min intervals until feces with charcoal were eliminated (maximum time of observation was 300 min). Charcoal was observed on the feces using normal light when it was easily visible, or using a microscope to help the identification of the black spots. The results were based on the time for the charcoal to be eliminated¹⁴.

Acute toxicity:

Acute toxicity describes the effects of a substance which result either from a single exposure or from multiple exposures in a short space of time (usually less than 24 hours). The acute toxicity study was conducted to find out LC₅₀ of the test samples. The test samples were administered orally to the test animals at different concentration (500 & 1000 mg/kg body weight).

After administration of the extract solutions mortality or sign of any toxicity was observed for one hour. Then the test animals were observed every 1 hour for next 5-6 hours. The animals were kept under observation for 1 week¹⁵⁻¹⁷.

RESULTS AND DISCUSSIONS:

Membrane stabilizing activity:

In the study of membrane stabilizing activity, the leaf extracts of *Spondias pinnata* at concentration of 1.0mg/mL were tested against the lysis of human erythrocyte membrane induced by hypotonic solution as well as heat, and compared with the standard acetyl salicylic acid (ASA) (Table 1). For hypotonic solution induced haemolysis, at a concentration of 1.0mg/ mL, the ethanol extract inhibited 76.92% haemolysis of RBCs as compared to 85.42% produced by acetyl salicylic acid (0.10mg/mL. The methanol and chloroform soluble extractives also revealed good inhibition of haemolysis of RBCs. On the other hand, during heat induced condition different organic soluble materials of *Spondias pinnata* demonstrated 76.68%, 77.19% and 74.83% inhibition of haemolysis of RBCs, respectively whereas ASA inhibited 78.04%.

To confirm the membrane stabilizing activity of *Spondias pinnata* of the above mentioned model, experiments were performed on the erythrocyte membrane. A possible explanation of the stabilizing activity of different extractives due to an increase in the surface area/volume ratio of the cells which could be brought about by an expansion of the membrane or shrinkage of the cell and an interaction with membrane proteins. The present investigation suggests that the membrane stabilizing activity of *Spondias pinnata* may be playing a significant role in its anti-inflammatory activity¹¹.

TABLE 1: EFFECT OF EXTRACTIVES OF *SPONDIAS PINNATA* ON HYPOTONIC SOLUTION AND HEAT INDUCED HAEMOLYSIS OF ERYTHROCYTE

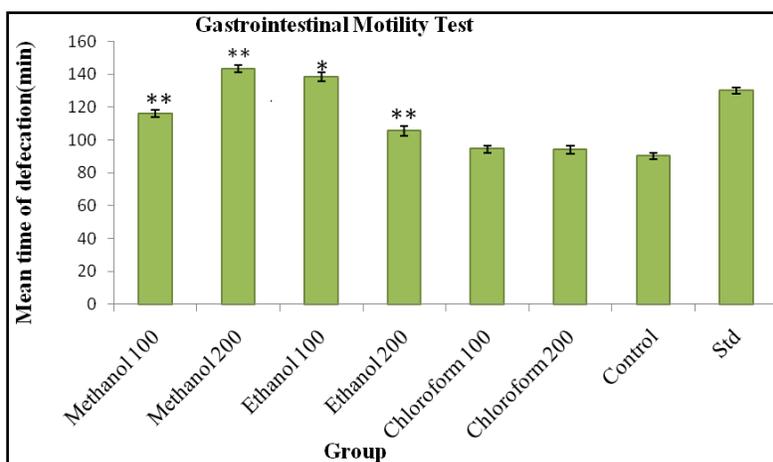
Samples	Concentration (mg/mL)	% Inhibition of Haemolysis	
		Hypotonic solution	Heat induced
Hypotonic solution(Control)	50mM	–	–
Acetyl salicylic acid (ASA)	0.1	85.42±0.025	78.04±0.033
Methanol extract	1	72.47±0.018	76.68±0.041
Ethanol extract	1	76.92±0.023	77.19±0.065
Chloroform extract	1	63.97±0.004	74.83±0.052

Values are mean ± SEM (n=6)

Gastrointestinal Motility Determination:

In-vivo gastrointestinal motility test was conducted on methanol, ethanol and chloroform extracts on the doses of 100mg/kg and 200mg/kg. The duration between charcoal administration and charcoal defecation is measured for gastrointestinal motility

determination. According to and Fig. 1, methanol extracts and ethanol extracts showed significant (p<0.05) and (p<0.001) increase in gastrointestinal motility comparing the standard drug butapen. Among all the test groups, the lowest defecation time was shown by chloroform extract 100mg/kg and chloroform extracts 200mg/kg.



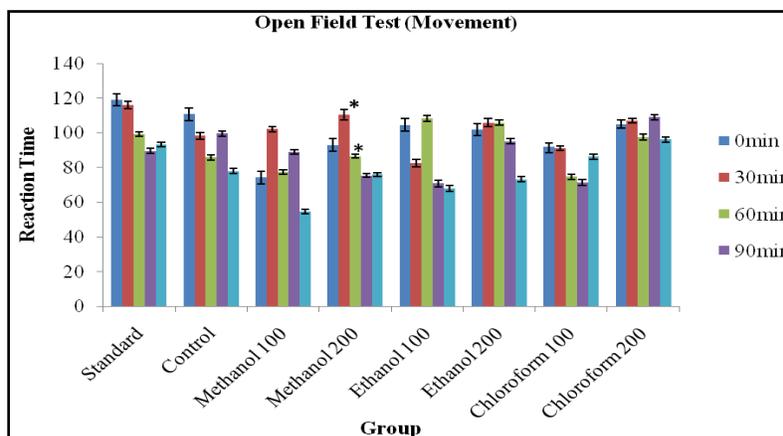
Values are mean ± SEM (n=6), * (p<0.05) and ** (p<0.001) significantly different when compared with the corresponding value of standard group, done by independent sample t-test

FIG. 1: TIME OF DEFECTION AT GASTROINTESTINAL MOTILITY ACTIVITY AFTER ADMINISTRATION OF DOSES OF CONTROL, STANDARD AND EXTRACTS

Neuropharmacology study:

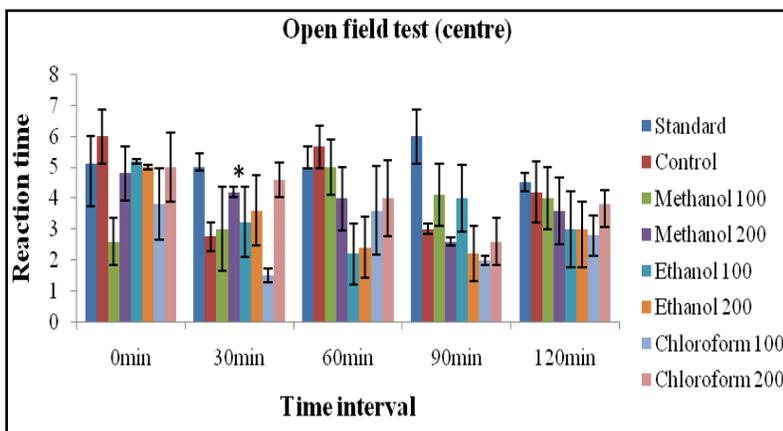
Open field test: This experiment was performed to assay general locomotor activity levels. After

investigation with leaf extracts of *Spondias pinnata* following data was observed (Table 2 and Fig.2, 3 and 4).



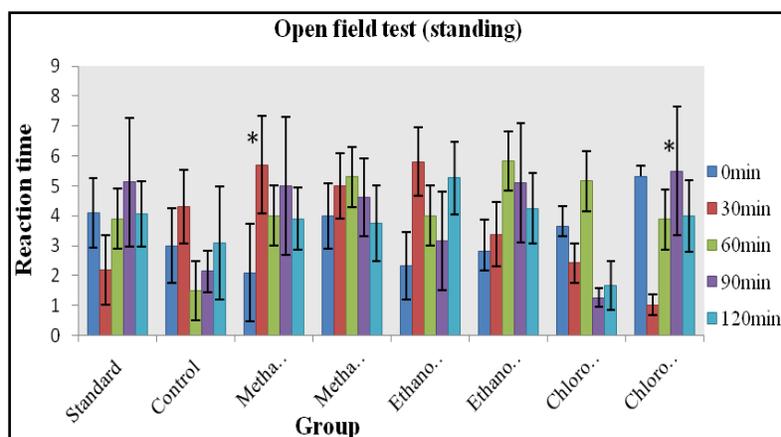
Values are mean ± SEM (n=6), * (p<0.05), significantly different when compared with the corresponding value of standard group, done by independent sample t-test

FIG. 2: GRAPHICAL REPRESENTATION OF EFFECT OF DIFFERENT EXTRACTS OF SPONDIAS PINNATA IN OPEN FIELD TEST (MOVEMENT)



Values are mean ± SEM (n=6), * (p<0.05), significantly different when compared with the corresponding value of standard group, done by independent sample t-test

FIG. 3: COMPARATIVE GRAPHICAL REPRESENTATION OF EFFECT OF DIFFERENT EXTRACTS OF SPONDIAS PINNATA IN OPEN FIELD (CENTRE)



Values are mean \pm SEM (n=6), * (p< 0.05), significantly different when compared with the corresponding value of standard group, done by independent sample t-test

FIG.4: GRAPHICAL REPRESENTATION OF EFFECT OF DIFFERENT EXTRACTS OF SPONDIAS PINNATA IN OPEN FIELD TEST (STANDING)

TABLE 2: EFFECT OF DIFFERENT EXTRACTS OF SPONDIAS PINNATA IN OPEN FIELD TEST (STOOL)

Group	Doses mg/kg	0 min	30 min	60 min	90 min	120 min
Control	10ml/kg	0.83 \pm 0.68	0 \pm 0	0 \pm 0	0.33 \pm 0.47	0.66 \pm 0.74
Standard	2mg/kg	0.66 \pm 0.74	0 \pm 0	0 \pm 0	0.66 \pm 0.74	0 \pm 0
Methanol extract	100	0.33 \pm 0.47*	0.66 \pm 0.74	0 \pm 0	0 \pm 0	0.83 \pm 0.68
	200	0.5 \pm 0.76	0 \pm 0	0.17 \pm 0.37	0.33 \pm 0.47	0 \pm 0
Ethanol extract	100	0.33 \pm 0.47	0.66 \pm 0.74	0 \pm 0	0 \pm 0	0.83 \pm 0.68
	200	0.17 \pm 0.37	0 \pm 0	0.17 \pm 0.37	0.5 \pm 0.76	0.33 \pm 0.47
Chloroform extract	100	0.83 \pm 0.68	0 \pm 0	0 \pm 0	0 \pm 0	0 \pm 0
	200	0.66 \pm 0.74	0 \pm 0	0 \pm 0	0.17 \pm 0.37	0.33 \pm 0.47

Values are mean \pm SEM (n=6), * (p< 0.05), significantly different when compared with the corresponding value of standard group, done by independent sample t-test

Table 2 and **Fig. 2, 3 & 4** represents the effect of different extracts of *Spondias pinnata* on various parameter of open field test. The most important step in evaluating drug action on CNS is to observe its effect on locomotors activity of the animal. The activity is a measure of the level of excitability of the CNS and this decrease may be loosely related to sedation resulting from depression of the central nervous system. The extract significantly decreased the locomotor activity as shown by the results of the open field. Chloroform extract decreased movement of rodents in a dose dependent manner but could not reach significance; whereas clonazepam decreased movement significantly. Clonazepam also decreased standing significantly. But extracts exhibited exert effect on standing and entrance in the center of the open field.

The effect of higher dose of methanol 200mg/kg and dose of chloroform 200mg/kg mg on defecation was like clonazepam. The results show that the methanol and chloroform extract has no ability to relieve stress and had an anxiolytic effect

on the rodents like clonazepam did. On the other hand, lower dose of methanol and both dose of chloroform decreased defecation which cannot support the previous results.

Forced swimming test:

Forced Swimming test (FST) was performed to evaluate the effect of anti-depressant effect of leaf extracts of *Spondias pinnata* on mice. After investigation of leaf extracts of *Spondias pinnata* following data was observed (**Table 3**). Where methanol extract 100 mg/kg b.w dose, ethanol 200 mg/kg b.w and both doses (100 & 200 mg/kg b.w) of chloroform extract shows significant values compared to standard.

Ethanol and chloroform extract showed decreased immobility with dose dependently like the standard diazepam which indicates its anti-depressant effect. This plant part has a great sensitivity with all the antidepressant classes and all the mechanisms of action of treatments could be determined, but clinical correlations should be considered very

carefully. When rodents are forced to swim in a confined place, they tend to become immobile after vigorous activity (struggling). This stressful inescapable situation can be evaluated by assessing different behavioral strategies and immobility

during the test could be an efficient adaptive response to the stress. The development of immobility when the rodents are placed in an inescapable container of water reflects the cessation of persistent escape directed behavior¹⁸.

TABLE 3: EFFECT OF DIFFERENT EXTRACT OF SPONDIAS PINNATA IN SWIMMING TEST

Group	Dose	Time of immobility (sec)
Control	1ml/kg	117.4±5.05
Standard	5mg/kg	68.2 ±6.74
Methanol extract	100mg/kg	115.2±6.33**
	200mg/kg	109.6±6.50
Ethanol extract	100mg/kg	105±7.23
	200mg/kg	94.6±6.16*
Chloroform extract	100mg/kg	109.8±6.85**
	200mg/kg	86.2±6.18*

Values are mean ± S.E.M (n=6), * (p< 0.05) and ** (p< 0.001), significantly different when compared with the corresponding value of standard group, done by independent sample t-test.

Acute toxicity: In the time of investigation of acute toxicity none of the extracts ranging from 500 &

1000mg/kg body weight showed any sign of toxicity in the period of one week (**Table 4**).

TABLE 4: ACUTE TOXICITY TEST OF DIFFERENT EXTRACTS OF SPONDIAS PINNATA ON MICE

Group	Dose	No. of Deaths/sign of Toxicity
Control		None
Methanol extract	500mg/kg	None
	1000mg/kg	None
Ethanol extract	500mg/kg	None
	1000mg/kg	None
Chloroform extract	500mg/kg	None
	1000mg/kg	None

CONCLUSION: So far some chemical and biological investigations have been carried out on *Spondias pinnata* plant in this study mainly focusing on the leaf of the plant but those are considered to be preliminary. As a result more sophisticated research is necessary to reach concrete conclusion about the finding of the present study. On the basis of above result and available reports, all three leaf extracts of *Spondias pinnata* had potent membrane stabilizing activity. This study also exhibited little gastrointestinal motility activity and neuropharmacological activity. But all three leaf extracts of *Spondias pinnata* did not have any effect on the acute toxicity test.

Therefore, further chemical and pharmacological studies with extract of *Spondias pinnata* for *in vivo* antioxidant, analgesic activity, anti-pyretic activity, hypoglycemic effect, anti-cancer activity, isolating new bioactive compounds and evolution of their extracts mode of action and chronic toxicity profile

might be the next steps to be followed to eventually find new led compounds. The plant can be further screened against various diseases in order to find out its unexplored efficacy and can be a potential source of chemically interesting and biologically important drug candidates.

ACKNOWLEDGEMENT: We thank Office of the Institute of Energy, Environment Research and Development (IEERD), University of Asia Pacific (UAP), Dhaka, Bangladesh for their assistance.

REFERENCES:

1. Islam SM, Ahmed KT, Manik MK, Wahid MA and Kamal CS: A comparative study of the antioxidant, antimicrobial, cytotoxic and thrombolytic potential of the fruits and leaves of *Spondias dulcis*. Asian Pacific Journal of Tropical Biomedicine 2013; 3(9): 682-691.
2. Alam F, Shahriar M and Bhuiyan MA: *In-vivo* pharmacological investigation of bark extracts of *carissa carandas*. International Journal of Pharmacy and Pharmaceutical Sciences 2014; 6(6): 180-185.

3. Ahmed T, Akter R, Sharif S, Shahriar M and Bhuiyan MA: *In vitro* antioxidant activities and *In vivo* antinociceptive and neuropharmacological activities of *Mimosa pudica*. International Journal of Pharmacy 2014; 4(2): 70-78.
4. Shahriar M, Ahmed T, Sharif S, Akter R, Haque MA and Bhuiyan MA: Phytochemical screenings, membrane stabilizing activity, thrombolytic activity and cytotoxic properties of leaf extracts of *Mimosa pudica*. International Journal of Pharmacy 2014; 4(2): 155-158.
5. Akther R, Haque MA, Bhuiyan MA and Shahriar M: *In vivo* pharmacological investigation of leaf extract of *Polygonum hydropiper* (L.). Dhaka University Journal of Pharmaceutical Sciences 2013; 12(2): 165-169.
6. Uddin MMN, Basak A, Amin MR and Shahriar M: Pharmacological Investigations on Flowers of *Ixora coccinea*. International Journal of Pharmacognosy and Phytochemistry 2014; 29(1): 1208-1213.
7. Shinde U, Phadke A, Nair A, Mungantiwar A, Dikshit V and Saraf M: Membrane stabilizing activity-a possible mechanism of action for the anti-inflammatory activity of *Cedrus deodara* wood oil. Fitoterapia 1999; 251-257.
8. Sikder M, Rahman M, Kaiser M, Rahman MS, Hasan C and Rashid M: In vitro antioxidant, reducing power, free radical scavenging and membrane stabilizing activities Maisuthisakul P, Suttajit M and Pongsawatmanit R: Assessment of phenolic content and free radical-scavenging capacity of some Thai indigenous plants. Food Chemistry 2007; 100(4):1409-1418.
9. Omale J and Okafor P: Comparative antioxidant capacity, membrane stabilization, polyphenols composition and cytotoxicity of the leaf and stem of *Cissus multistriata*. African Journal of Biotechnology 2008; 7:3129-33.
10. Ahmed M, Shikha H, Sadhu S, Rahman M and Datta B: Analgesic, diuretic, and anti-inflammatory principle from *Scoparia dulcis*. Pharmazie 2001; 56(8):657-660.
11. Shahriar M, Alam F and Uddin MMN: Membrane stabilizing and thrombolytic activity of *Withania somnifera* root. American Journal of Phytomedicine and Clinical Therapeutics 2014; 2(2):252-256.
12. Gupta B, Dandiya P and Gupta M: A psychopharmacological analysis of behaviour in rats. Japanese Journal of Pharmacology 1971; 21(3):293-8.
13. Porsolt R, Bertin A and Jalfre M: Behavioral despair in mice: a primary screening test for antidepressants. Archives Internationales De Pharmacodynamie Et De Therapie Journal 1978; 229(2):327-36.
14. Marona H and Lucchesi M: Protocol to refine intestinal motility test in mice. Laboratory Animals 2004; 38(3):257-60.
15. OECD. Guidance Document on Acute Oral Toxicity Testing. Environmental Health and Safety Monograph Series on Testing and Assessment N.24. Paris; 2001.
16. OECD. OECD Guidelines for Acute Toxicity of Chemicals. Organization for Economic Co-operation and Development. No. 420, 423, 425. Paris; 2001.
17. Walum E: Acute oral toxicity. Environmental health perspectives 1998; 497-503
18. Lucki I: The forced swimming test as a model for core and component behavioral effects of anti-depressant drugs. Behavioural Pharmacology 1997; 8(6-7):523-32.

How to cite this article:

Kamal S, Akhter R, Tithi NA, Md. Wadud A, Narjish SN, Md. Shahriar and Bhuiyan MA: Biological Investigations of the Leaf Extract of *Spondias Pinnata*. Int J Pharm Sci Res 2015; 6(8): 3351-58. doi: 10.13040/IJPSR.0975-8232.6(8).3351-58.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)