



Received on 20 August, 2015; received in revised form, 12 October, 2015; accepted, 13 November, 2015; published 01 February, 2016

POMEGRANATE SEED OIL: A COMPREHENSIVE REVIEW ON ITS THERAPEUTIC EFFECTS

Mohammad Taher Boroushaki, Hamid Mollazadeh* and Amir Reza Afshari

Department of Pharmacology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran.

Keywords:

PSO, Punicic Acid,
Pharmacological Properties,
Antioxidant, Anti-Inflammatory

Correspondence to Author:

Hamid Mollazadeh

Department of Pharmacology,
Faculty of Medicine, Mashhad
University of Medical Sciences,
Azadi Square, Mashhad, Iran.

E-mail: mollazadehh901@mums.ac.ir

ABSTRACT: Pomegranate, *Punica granatum* L. (Punicaceae), as a medicinal and nutritional ancient fruit, has an outstanding medical history throughout the world. Each of compartments of pomegranate has interesting pharmacological activity. Juice, leaf, flower, and peels of pomegranate possess potent antioxidant properties, while juice, peel and oil are all weakly estrogenic activity. Pomegranate seeds have ethnomedical indication and high conjugated α -linolenic acids (CLn) contents. Pomegranate seed oil (PSO) with high amount of punicic acid (PA), a conjugated isomer of α -linolenic acid, has variety of pharmacological properties. The main properties are as follows; antioxidant, anti-inflammatory, nephroprotective, hepatoprotective, neuroprotective, anti-cancer, enhancing the immune system, enhancing carbohydrate metabolism and reducing insulin resistance. Based on some studies, effects of PSO on lipid profile are controversial and consistency of data is rare to find yet. Therefore, this review is aimed to highlight the PSO's composition and beneficial effects on human health and represent the mechanisms involved in its action.

INTRODUCTION: Natural products as an alternative source of medicinal compounds are interested in the researchers' viewpoint. Nowadays, the use and investigation about medicinal plants as curative and preventive agents against diseases are increasing because of their popularity, low adverse effects, easy to earn and safety. Some important drugs such as digoxine, quinine and vinca alkaloids are formulated from herbal medicines¹⁻³.


Plants produce chemical compounds with various effects as a part of their normal metabolic activities. About 120 active compounds currently extracted from the medicinal plants.

All aspects of traditional plants and their close mechanisms are not determined completely but many beneficial effects of them have been proven. Awareness about these characteristics of plants leads the researchers to discover the new dosage forms with best quality and minimum adverse effects^{2,4,5}.

In this review, we try to evaluate and collect the information about chemical compositions, mechanisms, efficacy, adverse effects, benefits and toxicity of pomegranate seed oil (PSO) as a vintage of pomegranate.

Pomegranate:

Pomegranate (*Punica granatum* L.), from family Punicaceae, has been traditionally used for thousands of years as a medicinal fruit **Fig.1**. Mediterranean regions (including Iran, India and Pakistan) have the highest rate of pomegranate cultivation in the world⁶. Based on excavations of the Early Bronze Age (3500–2000 BC), it is

QUICK RESPONSE CODE	DOI: 10.13040/IJPSR.0975-8232.7(2).430-42
	Article can be accessed online on: www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.7(2).430-42	

believed that the pomegranate was one of the first cultivated fruits for its beneficial properties. Pomegranate was held sacred by many of the world's major religions and nations. In the Holy Quran, pomegranate has been described as a paradise fruit and a symbol of insurgence and life everlasting in Christian art. In China, pomegranate is widely represented in art symbolizing fertility, posterity, abundance, numerous and virtuous offspring, and a blessed future. It was also a symbol of invincibility in battle by the Persians⁷⁻⁹. For over 4,000 years, human beings have cultivated pomegranate for its medicinal properties. Juice, seeds, leaves, flowers, bark and roots of pomegranate have various effects. Lowering fever, treating diabetes, anthelmintic, anti-diarrhea, blood tonic, stopping the bleeding, and healing ulcers are the most important traditional uses of pomegranate¹⁰⁻¹⁴.



FIG. 1: POMEGRANATE FRUIT

Chemical Composition and Pharmacological Properties:

The parts of the fruits of pomegranate have different amounts of chemical compounds including vitamins, polysaccharides, minerals, polyphenols, and carbohydrates that are the most abundant ingredients in pomegranate^{15, 16}. Important pharmacological components of pomegranate are summarized in **Table 1**. High antioxidant capacity of pomegranate besides its anti-invasive, anti-proliferative and pro-apoptotic features have been studied in many human and animal models^{15, 17-20}. Moreover, phytoestrogenic compounds isolated from seeds, juice and peels of the fruit have many hormonal activity^{21, 22}. Treatment of stomach-ache, inhibiting herpes and influenza viruses and suppressing the reproduction of cancer cells are the other pharmacological characteristics of pomegranate juice and seeds extracts. Antimicrobial, anti-inflammatory, protective effects against liver disease, cardiovascular protection, anti-diabetic and anti-obesity effects of pomegranate have been investigated and documented by many researchers^{11, 23-26}. The presence of different substances with various chemical structures reveals multiple therapeutic effects of pomegranate.

TABLE 1: MOST IMPORTANT PHARMACOLOGIC COMPOUNDS OF *PUNICA GRANATUM L.*^{10, 11, 27-29}

Chemical class	Compound(s) name	Part(s) of the plant
Simple sugars	Glucose, Fructose, Sucrose	Juice
Aliphatic organic acids	Citric acid, Malic acid, Tartaric acid	Juice
Hydroxybenzoic acids	Gallic acid, Ellagic acid, 3,3_-Di-O-methyl ellagic acid	Juice, Leaf, Flower, Seed
Flavan-3-ols	Flavan-3-ol, Catechin, Epicatechin	Juice, Leaf
Flavonol glycosides	Kaempferol 3-O-glycoside, Kaempferol 3-O-rhamnoglycoside	Leaf
Anthocyanidins	Delphinidin, Cyanidin	Leaf
Ellagitannins	Punicalin, Punicalagin, Corilagin	Leaf
Amino acids	Proline, Valine, Methionine	Juice
Conjugated fatty acids	Punicic acid	Seed
Non-conjugated fatty acids	Linoleic acid, Oleic acid, Palmitic acid	Seed
Sterols	Daucosterol, Campesterol	Seed
Sex steroids	17_-Estradiol, Testosterone	Seed
Phenyl aliphatic glycosides	Icariside D1	Seed

Pomegranate Seed Oil (PSO):

Mainly, 12–20% of total seed weight is made by PSO. Conjugated octadecatrienoic fatty acids are responsible for approximately 80% of seed contents and PSO is considered to be a rich source of those fatty acids; in particular, punicic acid (PA) (*cis*9,

*trans*11, *cis*13 acid) which is the main fatty acid among them. Other isomers of conjugated linolenic acids (CLnAs) are catalpic acid (C18:3-9*trans*,11*trans*,13*cis*) and α -eleostearic acid (C18:3-9*cis*, 11*trans*, 13*trans*) but the oil content of the seed and fatty acid composition are affected by

cultivation sites, harvesting time, fruit genotypes and climatic conditions^{15, 30, 31}. Total lipids in PSO comprise mainly of triglycerides (TG) that their composition is varied and the most important templates are CLnA-CLnA-P and CLnA-CLnA-CLnA³². Lipid profile of PSO is different due to its cultivar, environmental growth conditions and ripening stage but the common and important fatty acids found in PSO in various types of pomegranate have been listed in **Table 2**^{33, 34}.

Also, **Table 3** shows the different fatty acid composition of PSO in the Iranian strains of pomegranate³⁵. In addition to fatty acids, minor components of the oil including steroids, sterols, and, cerebroside (a key component of mammalian myelin sheaths), lignins, hydroxycinnamic acids, and the potent antioxidant lignin derivatives are found in PSO. PSO is abundant of phytosterols such as β -sitosterol, campesterol, stigmasterol and tocopherols such as α and γ -tocopherol.^{32, 36, 37}

TABLE 2: COMMON AND IMPORTANT FATTY ACIDS FOUND IN PSO IN VARIOUS TYPES OF POMEGRANATE. RESULTS ARE EXPRESSED AS AVERAGE (PERCENTAGE OF TOTAL FATTY ACIDS) \pm STANDARD DEVIATION^{33, 34}.

Fatty Acids	India Strain	China Strain	Turkey Strain
16:0	5.7 \pm 4.1	5.07 \pm 1.30	2.45 \pm 0.19
18:0	2.1 \pm 3.1	4.20 \pm 1.56	1.52 \pm 0.26
18:1 (ω -9)	9.0 \pm 5.6	7.86 \pm 2.25	4.19 \pm 0.61
18:2 (ω -6)	10.8 \pm 6.9	8.36 \pm 2.36	4.49 \pm 0.49
18:3 (9c11t13t)		10.70 \pm 4.44	6.41 \pm 0.27
18:3 (9t11t13t)		8.78 \pm 5.16	1.03 \pm 0.16
18:3 (9t11t13c)		15.24 \pm 6.17	3.48 \pm 0.34
18:3 (9c11t13c)	71.5 \pm 17.9	36.98 \pm 10.12	74.11 \pm 1.55

TABLE 3: FATTY ACID COMPOSITION OF PSO IN IRANIAN STRAINS OF POMEGRANATE³⁵.

Variety	Fatty acid (%)					
	18:3	18:2	18:1	18:0	16:0	Others
Red Seed Ardestani	79.43	6.95	6.62	2.09	3.16	1.72
Rizdavar's Dorpaye	79.22	6.87	6.30	2.50	3.36	1.73
Ashkzar's Sour Narak	82.40	5.22	5.71	1.99	2.95	1.70
Taft's Tokhm-e Mush	78.25	6.59	7.48	2.54	3.41	1.70
Chatrud's Sour Shahi	78.73	7.08	6.68	2.16	3.57	1.75

Punicic acid:

PA is a conjugated α -linolenic acid (CLnA) molecule, which is found in PSO and it may contribute to a lot of health benefits associated with pomegranate. PA contains a third double bond, is known as *cis*9, *trans*11, *cis*13, and is referred to as 18:3 fatty acids. It is an omega-5 long chain polyunsaturated fatty acid and a positional and geometric isomer of α -linolenic acid (LnA; C18:3-9c, 12c, 15c) **Fig. 2**³⁸⁻⁴⁰.

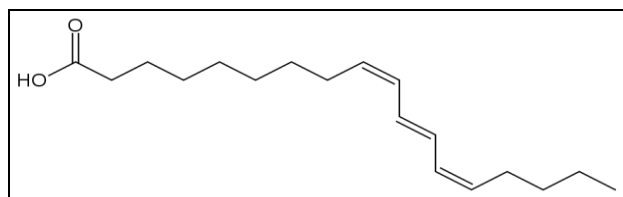


FIG.2: CHEMICAL STRUCTURE OF PUNICIC ACID

PA features *cis* and *trans*-type double bonds with parentage of 2:1. Numerous studies have shown that conjugated linoleic acids (CLAs) and α -linolenic acids (LnAs) have abundant health

benefits and PA has a very high structural similarity with them^{41, 42}. Anti-inflammatory, immunomodulatory, anti-cancer, anti-estrogen and beneficial effects on lipid profile are examples of beneficial effects of PA that revealed in many studies^{15, 43-45}.

Pharmacological actions of PSO:

PSO has been reported to have beneficial effects including cytotoxic and antitumor properties, body fat-reducing and lipid metabolism-normalizing effects, enhancing the immune system *in vivo*, Showing chemo preventive activity against hormone-related (prostate and breast) and colon cancers, reducing the accumulation of hepatic triglycerides and promoting epidermal tissue regeneration⁴⁶⁻⁵⁰. Antioxidant and anti-inflammatory activities are the main features of PSO which results from inhibition of lipid peroxidation and neutrophil-activation^{37, 39, 51}. Effects of PSO on body weight and serum lipid profile are inconsistent and many studies confirmed

weight reducing effects but the effects of PSO on reducing in serum triglyceride, phospholipid, cholesterol, and LDL-cholesterol levels are contradictory^{40, 52}. PSO reduces body weight, leptin and insulin levels, enhances glucose tolerance, improves peripheral insulin sensitivity, increases carbohydrate oxidative capacity, and inhibits the progression of type2 diabetes^{31, 53}.

PSO is opulent of CLnAs which consists of octadecatrienoic fatty acid isomers with three conjugated double bonds (C18:3). Beneficial health effects of PSO are due to the high level of its CLnAs^{30, 54}. The exact mechanisms of action of CLnAs are not completely understood but probable and definite mechanisms are summarized in **Table 4**.

TABLE 4: CLINICAL EFFECTS AND MECHANISMS OF CLnAs

Effect(s)	Mechanism(s)	Reference(s)
Antitumor and anticancer	Induce apoptosis through lipid peroxidation and protein kinase C pathway,	55
	Act as selective estrogen receptor modulators and inhibit estrogen receptors α and β	22
Anti-diabetic	Improve insulin sensitivity,	31
	Suppress NF- κ B and TNF- α activation and up regulate PPAR α and γ -responsive genes	53
Reductions in hepatic and plasma TG levels	Increase the levels of PPAR- γ and α mRNA,	33
	Suppress the delta-9 desaturation	56
Antioxidant	Enhance the levels of the antioxidant enzymes,	42, 57
	Reduce lipid peroxidation, oxidative stress	
Anti-inflammatory	Free radical addition to one of the conjugated double bonds of CLnAs	39, 58
	Inhibit TNF α -induced priming of ROS production and MPO,	
	Up regulate colonic PPAR- δ expression,	
	Increase the levels of IL-17 and IFN- γ ,	
	Modulates mucosal immune responses,	
	Reducing the expression of TNF- α and IL-6	

NF- κ B: nuclear factor kappa-light-chain-enhancer of activated B cells, **TNF- α** : Tumor necrosis factor- α , **PPAR**: Peroxisome proliferator-activated receptor, **CLnAs**: conjugated linolenic acid, **ROS**: Reactive oxygen species, **MPO**: Myeloperoxidase, **IL**: Interleukin, **IFN**: Interferon

Pharmacological studies of PSO:

Anti-inflammatory activity:

Inhibition of cyclooxygenase (COX) and lipoxygenase (LOX) by PSO have been shown in Schubert study⁵⁹. The results of Jiang *et al.* study showed that, inflammation and its important mediators such as prostaglandin E₂ (PGE₂) have a key role in progression of diseases such as cancer and vascular heart disease. γ -tocopherol in PSO can inhibit PGE₂ formation. They showed that, PGE₂ synthesis in both IL-1 β -treated A549 human epithelial cell and lipopolysaccharide (LPS)-stimulated RAW264.7 macrophages was reduced with γ -tocopherol with IC₅₀ of 4 and 7.5 μ M, respectively. γ -tocopherol and its metabolite (γ t, 2,7,8-trimethyl - 2 - (beta-carboxyethyl)-6-hydroxy chroman) can inhibit PGE₂ formation after the explosion for 1 hr to COX-2- preinduced cells followed by addition of arachidonic acid. Also, suppression of inducible nitric oxide synthase expression was reduced by γ -tocopherol⁶⁰.

Enhancing B-cell function in vivo by PA was shown in Yamasaki *et al.* study. Diets containing 0%, 0.12%, or 1.2% of PSO was treated to mice for 3 weeks. After this period amounts of immunoglobulin G and M were larger than control group in splenocytes isolated from mice⁴³. Ellagic acid is another polyphenolic compound with potent antioxidant and anti-inflammatory activity. Ellagic acid inhibited proliferation and migration of platelet-derived growth factor- BBand it can inhibit IL-1 β - and tumor necrosis factor- α (TNF- α)-induced activation of activator protein-1 (AP1) and mitogen-activated protein kinases (MAPK).

In addition, ellagic acid inhibited proliferation and migration of monocyte chemo attractant protein-1 production which have a key role in inflammation⁶¹. Gallic acid is a polyphenolic compound with anti-inflammatory properties which is found in PSO. Madlener *et al.* showed that gallic acid can inhibit COX-1 and COX-2 in humanHL-60promyelocytic leukemia cells with IC₅₀ values of 3.5 and 4.4nM⁶². About 80% w/w of hydrophilic

fraction of PSO is PA. Costantini *et al.* evaluated the anti-inflammatory effect of the aqueous methanolic extract of PSO on some human cell lines.

They showed that the levels of vascular endothelial growth factor and nine pro-inflammatory cytokines was reduced after treating with PSO, dose-dependently⁶³. TNF- α is a proinflammatory cytokine that rises in many inflammatory conditions. The expression of NF- κ B and AP-1 has been increased in the PSO's effect. The effects of PSO and PA on serum TNF- α concentration is controversial. PSO administration (800mg PSO/day for 4 weeks) to hyperlipidemic patients had no effect on serum TNF- α concentration but in Hontecillas *et al.* study, PA can decrease TNF- α plasma concentration in mice. Changes in serum triglyceride, HDL-C and TNF- α are related to each other. TNF- α promotes lipolysis and can activate the endothelium and cause vascular damage. TNF- α and insulin resistance are related to each other.

It is probable that anti-TNF- α effect of PSO was liable for its anti-diabetic and triglyceride lowering effect⁶⁴. Decrease in TNF- α and IL-6 was reported by Saha and Ghosh after treatment of diabetic rats for four weeks with CLnAs (α -ESA and PA at 0.5 % total lipids)⁶⁵. PSO decreased intestinal inflammation in rats with necrotizing enterocolitis (NEC). In this model of inflammation, rats fed with 1.5% of PSO orally and then were exposed to asphyxia/cold stress to induce NEC. Decrease in incidence of NEC from 61% in control group to 28% in therapeutic group was seen. Also, PSO normalized the serum levels of proinflammatory cytokines such as IL-6, IL-8, IL-12, IL-23 and TNF- α . Enterocyte proliferation was reduced after administration of PSO⁶⁶.

All of above studies confirmed anti-inflammatory properties of PSO. In many studies anti-inflammatory effects of PSO were investigated with its antioxidant and anticancer effects, but in the present paper we mentioned the PSO's beneficial effects separately.

Antioxidant activity: Oxidation dependent mechanisms are very important ways to induce

diseases such as many types of cancers and inflammatory diseases⁶⁷. Reduction-oxidation state may activate and deactivate certain genes that promotes many secondary steps in disease conditions⁶⁸. NF- κ B and AP-1 are two examples of this one. Reactive oxygen species (ROS) act as 'signal transduction messengers to promote the activity of the cytokines'⁶⁹. Genes involved in carcinogenesis, atherosclerotic mechanisms, diabetic changes and HIV replication are near the receptor sites of these cytokines⁵⁹. Thus, antioxidant effects of PSO are effective in anti-inflammatory and cardioprotective activities. Lipid peroxidation and ROS production are involved in many organs toxicity due to various factors such as xenobiotics, drugs, environmental pollutants and etc.⁷⁰. Nephrotoxicity, hepatotoxicity, cardiotoxicity, and neurotoxicity are the main examples of interference of oxidative stress and will be discussed later.

In this section, we have an overview the antioxidant action of PSO. Mukherjee *et al.* showed that PA at concentration of 0.6% has maximum antioxidant activity. Peroxidation of polyunsaturated fatty acids in lipids and formation of free radicals showed a reduction. Alternatively, conjugated double bonds of PA and other CLnAs in PSO entrapped free radicals⁷¹. Membrane lipid peroxidation was decreased by diet containing 0.25 % CLnAs compare to diet without CLnAs⁷². Lowered oxidative stress and enhanced antioxidant enzyme serum levels were observed in Saha and Ghosh study. They treated streptozotocin-induced diabetic rats with α -ESA or PA with 0.5% total lipids. After the procedure, the levels of superoxide dismutase (SOD), glutathione peroxidase (GPx) and catalase (CAT) were higher in treated group compared to control group⁴². This result was confirmed by other study performed by them in sodium arsenite-induced oxidative stress in rat models⁴².

Antioxidant activity of PSO is contributed to tocopherols and polyphenolic compound contents⁷³. Oxidative stress reducing agents in PSO have metallic chelating potential, singlet oxygen quenchers and hydrogen donors⁷⁴. Antioxidant effect of PSO will be discussed later.

Effects on insulin resistance:

TNF- α is one of the most important pro-inflammatory cytokines which plays a pivotal role in obesity, inflammation and insulin resistance. Higher serum levels of TNF- α are accompanied with insulin resistance. Anusree *et al.* showed that TNF- α (10ng/mL for 24 hr) decreased 1.7 fold in insulin stimulated glucose uptake with respect to control group in 3T3-L1 adipocytes, but PA with doses of 5, 10, 30 μ M restored the glucose uptake capacity significantly and dose- dependently. PA at dose of 30 μ M is comparable to rosiglitazone (100nM) in insulin stimulated glucose uptake. The similar condition was observed in production of ROS with the effect of TNF- α and its restoration with the effect of PA. TNF- α treatment caused significant alterations in mitochondrial transmembrane potential, reduction in ATP production, O₂ consumption, overall increase in cellular ROS generation and a decrease in aconitase activity. The use of PA restored these states to the baseline⁷⁵.

Peroxisome proliferator activated receptor γ (PPAR- γ) agonists increase mitochondrial biogenesis and normalize the fission fusion ratio in this organelle that be altered with inflammation and high levels of TNF- α ⁷⁶. Agonistic effects of PA on PPAR- γ , lowering oxidative stress and serum TNF- α levels and positive effects of PA on mitochondrial functions are the main anti-diabetic mechanisms of PSO⁷⁵.

Effects of PSO on obesity and insulin resistance induced by rich fat diet in mice showed that, rich fat diet with 1% PSO after 12 weeks induced higher peripheral insulin sensitivity in treated group compared to control group (164 \pm 52% vs. 92 \pm 24% respectively), but liver insulin sensitivity showed no significant difference between two groups. In this study, obesity and fat deposition induced by this regimen was ameliorated by PSO, significantly³¹. A similar study was performed by Miranda *et al.* That used diet with 0.5% PA in rats. The results of this study showed no changes in adipose tissue weights and insulin resistance, but the glycemic value in the PA group had decreased, significantly⁷⁷. Improvement in insulin sensitivity is associated with reducing the risk of developing type 2 diabetes. Insulin sensitivity was improved with 61.79mg/day

PSO in mice which was treated with rich fat regimen compared to control group. In addition, final body weight, body weight accumulation, serum adiponectin, serum leptin, and serum insulin were reduced in treated group⁷⁸.

The use of PSO (200 and 400mg/day for 28 days) in streptozocin-nicotinamide induced diabetic rats showed that serum insulin levels increased but the serum glucose level had no change. Up-regulation of PPAR- γ responsive genes by PA may be a mechanism of PSO-induced increase in serum insulin⁷⁹. Anti-diabetic and insulin sensitizing effects of α -linolenic acid were also reported in other studies^{53, 78-81}.

Nephroprotective activity:

Investigating nephroprotective effects of PSO was carried out mainly by Boroushaki *et al.* In five separated studies, four nephrotoxic agents, mercuric chloride⁸², diazinon⁸³, gentamicin⁸⁴, and hexachlorobutadien⁸⁵ and cis-platin (under publication) were used and protective effect of PSO was shown. Elevated serum levels of urea and creatinine and urinary levels of glucose and protein as well as histopathological changes in kidney, such as severe tubular necrosis and atrophy, represented acute nephrotoxicity by use of these toxic agents. Inducing oxidative stress by these agents was shown in above mentioned studies by elevated kidney malondialdehyde content (MDA, as a biomarker of oxidative stress) and decreased total thiol content in the kidney.

The use of PSO in these models of nephrotoxicity was accompanied by its protective effects and PSO preserved renal functions and decreased histopathological changes in kidney. Restoration of serum urea and creatinine and decrease in urine glucose and protein were established after PSO treatment. Increasing in total thiol content was seen but was not dose dependent in all studies. Kidney was vulnerable to oxidative stress and each factor that induces this state can be a toxic agent on this organ. PSO due to its high content of polyphenolic compounds is a potent antioxidant. Moreover, hydroxybenzoic acid derivatives and sterols in PSO have an antioxidant effect and decrease lipid peroxidation.

The possible nephroprotective mechanisms of PSO mentioned in these articles includes removing peroxy radicals and prevents Cu^{2+} - induced lipid peroxidation, inhibition of TNF- α - induced priming of ROS production and myeloperoxidase, hydroperoxide formation and biohydrogenation of CLnAs and chelating transition metals⁸⁶.

Effects on memory:

As mentioned, polyphenols have been found to possess antioxidant properties and recent studies indicated that seed oil extract of pomegranate has the highest concentration of polyphenols. Sarkaki *et al.* in their *in vivo* study demonstrated that, administration of PSO in permanent cerebral ischemia causes a remarkable improvement on memory with criterion condition responses (CCRs) in Y-maze and step-through latency (STL) in two-way shuttle box. The results showed that, both active and passive avoidance memories were meaningfully impaired in rats after cerebral hypoxia-ischemia (CHI) ($P < 0.001$) and PSO treatment significantly ameliorated passive and active memory impairments with bilateral common carotid arteries occlusion (2CCAO) ($P < 0.05$, $P < 0.01$, and $P < 0.001$)⁸⁷. Gabizon *et al.* Reported that administration of large concentrations of PSO may postpone the manifestation of disease in young transgenic mice (Tgs), in addition, lower doses of Nano-PSO significantly delayed disease onset in asymptomatic TgMHu2ME199K mice and postponed disease aggravation in already sick mice. Therefore, PSO formulations may be impressive on neurodegenerative diseases⁸⁸.

Anti- cancer effects:

PSO has a potent effect on tumor cells. Hora *et al.* investigated the chemo preventive effect of PSO on skin tumor development in CD1 mice. They concluded that, PSO (5%) significantly decreased tumor incidence ($P < 0.05$), multiplicity, and TPA (12-O tetradecanoylphorbol 13-acetate) - induced ODC (ornithine decarboxylase) activity during 20 weeks of promotion. The mechanism for this effect can be inhibition of prostaglandin biosynthesis (COX-1, COX-2, and LOX) by punicic acid. In this study, in addition, topical application of 5% pomegranate seed oil remarkably inhibited ($P < 0.05$) the TPA-induced epidermal ODC activity. Overall, PSO appears to be a good natural product

with a potential chemopreventive effect against skin cancer⁸⁹.

γ -Tocopherol is the most important constituents of PSO that is responsible for anti-cancer activity. Jiang *et al.* investigated that, γ - tocopherol, inhibits proliferation of prostate cancer cells but appears to have no effect on the growth of a normal prostate epithelial cell. Their proposed mechanism for this effect was inhibition of sphingolipid synthesis *de novo*⁹⁰. In another study performed by Jiang *et al.* γ -tocopherol inhibited cyclooxygenase activity in macrophages and epithelial cells. This mechanism is beneficial for human tumor tissues, including human colon cancer, have been reported to contain enhanced COX-2 expression and PGE_2 , because PGE_2 has been shown to promote proliferation in certain cancer cells⁶⁰. Another similar study showed a significant role for induction of cell death for all cancer cell lines such as colon cancer, prostate carcinoma cells, and osteosarcoma. According to this study, γ -tocopherol has chemopreventive properties by mechanisms such as reduction levels of C- reactive protein, inhibition of neoplastic transformation, suppression of ras p-21, inhibition of COX-2 activity, down-regulation of cyclins, and up-regulation of PPAR - γ ⁹¹.

As mentioned, PA as an important component of PSO has various anti- cancer effects. In a research performed by Lansky *et al.* PA was tested as a potential inhibitor of *in vitro* invasion of human PC-3 prostate cancer cells in an assay employing MatrigelTM artificial membranes. Results showed a considerable inhibition of PC-3 prostate cancer cell invasion¹⁵. In another study PSO exhibited an enhancing B-cell function *in vivo* that can be effective in some cancers⁴³. In a study PSO via lipid peroxidation mechanism showed a cytotoxic effect against leukemia cells⁹².

It is known that dietary phenolic compounds can elicit vital cellular responses such as cytotoxicity, cell cycle arrest and apoptosis by activating a cascade of molecular events. Hydroxybenzoic acids, such as ellagic acid and gallic acid are other important components of PSO. In LI *et al.* study, ellagic acid had shown an anti-cancer effect with flow cytometric assay, polymerase chain reaction (PCR) and determination of caspase-3 activity

methods. Ellagic acid significantly induced p53/p21 expression, G1 arrest and apoptosis in bladder cancer cells⁹³. In another study, gallic acid showed a strong dose and time-dependent growth inhibition and apoptotic death of human DU-145 prostate cancer cells⁹⁴. In a study provided by Madlener *et al.* Gallic acid induced a dose-dependent apoptosis in HL-60 cells and attenuated G0/G1 to the S phase, and COX in HL-60 leukemia cells⁶².

Other constituent of PSO, ursolic acid, with mechanisms such as apoptosis in MCF-7 via p-53 up regulation⁹⁵, apoptosis in endometrial cancer cells via caspase-3 pathway⁹⁶, and apoptosis in melanoma cells via the intrinsic cell death pathway and caspase 3 activation⁹⁷ showed anti-cancer activity.

Furthermore, sterols (daucosterol, campesterol, stigmasterol, beta-sitosterol) have shown an anti-cancer effect via; Inhibition of pro-inflammatory cytokines⁹⁸, PC-3 apoptosis and cell cycle arrest via ROS changes and prostaglandin release^{99, 100}, and Reverts impaired glutathione/oxidized glutathione ratio via estrogen/phosphatidylinositol 3-kinase pathway¹⁰¹.

Effects on lipid profile:

Conjugated fatty acids are polyunsaturated fatty acids in which at least one pair of double bonds are separated by only one single bond, as in conjugated linoleic acid. These fatty acids have useful biological effects. Arao *et al.* investigated the effects of PSO rich in PA (9cis, 11trans, 13cis-conjugated linolenic acid; 9c, 11t, 13c-CLNA in obese, hyperlipidemic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. Results showed that, feeding with 1% pomegranate did not affect serum lipid levels and abdominal white adipose tissue weights.

However, 2 weeks feeding of the diet supplemented with 5% PSO showed a significant reduction of omental white adipose tissue weight. Also, the accumulated hepatic triacylglycerol was significantly decreased by PA regimen. In addition, the activity of hepatic enzymes was not altered by PA regimen. Also, PA contained diet could decrease level of monounsaturated fatty acid

(MUFA) in OLETF rats. Furthermore, this in vivo study demonstrated that PA can inhibit the delta-9 desaturation which leads to reduction in hepatic triacylglycerol (TAG) accumulation¹⁰². In contrast, yang *et al.* reported that PSO does not alter serum cholesterol concentration¹⁰³. Vroegrijk *et al.* in their study demonstrated that PSO regimen ameliorates rich-fat diet induced obesity and causes a reduction in body fat mass independent of changes in food intake or energy expenditure³¹. Anti-atherogenic effects of PA, as a type of conjugated fatty acid and the main constituent of PSO, has been investigated by Mirmiran *et al.* This double-blind placebo-controlled clinical trial with 400 mg PSO exhibited the mean concentration of TAG and the TAG: HDL cholesterol (HDL-C) ratio, were significantly decreased after 4 weeks. Also, the results showed a diminution of cholesterol HDL-C ratio while, the serum cholesterol, LDL cholesterol remained unchanged. Therefore, this study could demonstrate a beneficial effect of lipid profile including TAG and TAG:HDL-C ratio⁵². Mc Farlin *et al.* demonstrated the regimen with PSO (approximately 61mg/d) during 14 weeks can reduce the body weight with the mechanism mediated by a leptin/adiponectin pathway.

This study showed that weight gain was associated with an increase in biomarkers of cholesterol profile, adipose tissue accumulation (P<0.05). PSO resulted in a decrease in total weight gain, leptin and insulin, and an increase in plasma adiponectin concentration (P<0.05)⁷⁸. Miranda *et al.* demonstrated that, dietary supplementation of 0.5% PA did not cause to decreased fat accumulation in adipose tissue, liver, or skeletal muscle and there was no significant difference between the experimental groups in serum TGs, HDL cholesterol, or non-HDL cholesterol. An interesting result from this research was hypoplasia induced in the liver due to the anti-proliferative effect on hepatocytes increased transaminase levels. Also, this issue should be considered before proposing PA as a functional ingredient⁷⁷. Another study performed by Yamasaki *et al.* with diets containing 0%, 0.12%, or 1.2% PSO for 3 weeks, showed a meaningful increase in serum triacylglycerol and phospholipid levels but not in total cholesterol in the PSO treated groups⁴³.

Other beneficial effects of PSO: As mentioned, PSO has many pharmacological effects, other beneficial effects of PSO is summarized in the **Table 5**.

TABLE 5: OTHER BENEFICIAL EFFECTS OF PSO.

Pharmacological effect	Mechanism(s) and clinical evidence(s)	Reference(s)
Anti- osteoporosis	Down-regulation of expression RANK-RANKL downstream signaling targets and osteoclast differentiation markers in osteoclast-like cells, increasing alkaline phosphatase activity, mineralization of matrix and transcriptional levels of major osteoblast lineage markers involving the Wnt/ β -catenin signaling pathways	104
Anti- pancreatitis	Anti- inflammatory and antioxidant mechanisms of PSO caused reduction in amylase and lipase activity in serum, pancreatic MPO activity, edema, leukocyte infiltration and vacuolization.	105
Hepatoprotective	Decrease in MDA, DNA fragmentation, caspase- 3 and GSR activities, elevation in levels of GSH, SOD, GST and t-GPx activities. Consequently reduction in oxidative stress and apoptosis.	106
Improving in insulin secretion	increasing serum insulin and glutathione peroxidase activity	79
Anti-atherogenic	Decrease in TAG and the TAG:HDL cholesterol (HDL-C) ratio,	52
Neuroprotective	Neutralize ROS or enhance the expression of antioxidant gene, decrease in lactate/pyruvate ratio, extracellular nitric oxide, and lactase dehydrogenase generation	107
Anti-menopausal symptoms	Reduction in lipid oxidation and neuronal loss	108
	Reduction in the number of hot flashes per day, Reduction in sum score of the Menopause Rating Scale II parameters in the treated group after 12 weeks but not significant in comparison to control group	109
Cosmetic	Stimulate keratinocyte proliferation, mild thickening of epidermis, stimulating keratinocyte proliferation, stimulating type I procollagen synthesis, inhibiting matrix metalloproteinase-1	110

RANK: Receptor Activator of Nuclear Factor κ B, **RANKL:** Receptor Activator of Nuclear Factor κ B ligand, **MPO:** Myeloperoxidase, **MDA:** Malondialdehyde, **GSR:** Glutathione reductase, **GSH:** Glutathione, **SOD:** Superoxide dismutase, **t-GPx:** Glutathione peroxidase, **TAG:** triacylglyceride, **HDL:** High-density lipoprotein, **ROS:** Reactive oxygen species

Toxicity:

In our study on PSO toxicity, only one study had been found. In this study, *in vitro* toxicity was done with Ames and Chromosomal aberration test and *in vivo* toxicity was done with acute and chronic (28 days consumption) test. No mutagenicity of PSO was observed with Ames test and Chromosome aberration test with 5000 and 333 μ g/ml, respectively. 2000mg/kg PSO did not show any significant side effects in rat. Use of 0, 10,000, 50,000 and 150,000ppm of PSO per day in a 28 days period did not show any adverse effects but in 150,000ppm, that was much higher than routine anti-diabetic and anti-inflammatory dose, increasing in hepatic enzymes and weight was deduced and in this study this dose was considered as no observable adverse effect level (NOAEL) ¹¹¹.

CONCLUSION: The use of pomegranate has been increased due to its reported health benefits. Pomegranate and its derivatives, especially PSO, are rich source of several chemical compounds with potential physiological activities. The

information presented in this review article has shown some pharmacological and toxicological mechanisms and properties of PSO. It has been demonstrated that monotherapy or supplementation therapy with PSO may have protective effects against several diseases, including cancers, diabetes, cardiovascular disease, inflammation, neurotoxicity, mouth and skin disorders, pancreatitis, and osteoporosis. These effects act possibly due to highly antioxidant activity of PSO. Therefore, it is recommended that, PSO can be a very beneficial medicine for treatment and protection of certain diseases and disorders, although probably underlying mechanisms of this protection need further explorations.

ACKNOWLEDGEMENTS: The authors are grateful to the assistance of Dr. Sima Khosravi and all scientists in Department of Pharmacology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran, who helped us to create this review with their honest cooperation.

REFERENCES:

1. Kashani HH, Hoseini ES, Nikzad H and Aarabi MH: Pharmacological properties of medicinal herbs by focus on secondary metabolites. Life science journal-acta zhengzhou university overseas edition 2012; 9:509-520.
2. Goldman P: Herbal medicines today and the roots of modern pharmacology. Annals of internal medicine 2001; 135:594-600.
3. Dirin MM, Mousavi S, Afshari AR, Tabrizian K and Ashrafi MH: Potential drug-drug interactions in prescriptions dispensed in community and hospital pharmacies in East of Iran. Journal of research in pharmacy practice 2014; 3:104.
4. Roodsari MR, Zamanian-Azodi M and Salimpour F: Herbal remedies and medicine; introducing some Iranian plants. Journal of Paramedical Sciences 2013; 4.
5. Chan K, Zhang H and Lin ZX: An overview on adverse drug reactions to traditional Chinese medicines. British journal of clinical pharmacology 2015.
6. Modaresi J, Nasri MF, Rashidi L, Dayani O and Kebreab E: Short communication: Effects of supplementation with pomegranate seed pulp on concentrations of conjugated linoleic acid and punonic acid in goat milk. Journal of dairy science 2011; 94:4075-4080.
7. Langley P: Why a pomegranate? BMJ: British Medical Journal 2000; 321:1153.
8. Albanese CL: 1994 Presidential Address: Refusing the Wild Pomegranate Seed: America, Religious History, and the Life of the Academy. Journal of the American Academy of Religion 1995:205-229.
9. Heber D, Schulman RN and Seeram NP: Pomegranates: ancient roots to modern medicine: CRC press; 2006.
10. Lansky EP and Newman RA: Punica granatum (pomegranate) and its potential for prevention and treatment of inflammation and cancer. Journal of ethnopharmacology 2007; 109:177-206.
11. Miguel MG, Neves MA and Antunes MD: Pomegranate (Punica granatum L.): A medicinal plant with myriad biological properties-A short review. Journal of Medicinal Plants Research 2010; 4:2836-2847.
12. Kapoor R, Ronnenberg A, Puleo E, Chatterton Jr RT, Dorgan JF, Seeram NP and Sturgeon SR: Effects of Pomegranate Juice on Hormonal Biomarkers of Breast Cancer Risk. Nutrition and cancer 2015:1-7.
13. Wu P-T, Fitschen PJ, Kistler BM, Jeong JH, Chung HR, Aviram M, Phillips SA, Fernhall B and Wilund KR: Effects of Pomegranate Extract Supplementation on Cardiovascular Risk Factors and Physical Function in Hemodialysis Patients. Journal of medicinal food 2015.
14. Fan L: Antimicrobial Effect of Pomegranate Juice on *Listeria innocua* and *Escherichia coli* in Different Culture Systems. 2015 Annual Meeting (July 25-28, 2015); 2015: Iafp.
15. Lansky EP, Harrison G, Fromm P and Jiang WG: Pomegranate (Punica granatum) pure chemicals show possible synergistic inhibition of human PC-3 prostate cancer cell invasion across Matrigel™. Investigational new drugs 2005; 23:121-122.
16. Arafat S, Basuny AM and Elsayy HA: Utilization from leaves of olive and pomegranate as a source of bioactive components. Journal of Advances in Biological and Basic Research 2015; 1:28-34.
17. Syed DN, Afaq F and Mukhtar H: Pomegranate derived products for cancer chemoprevention. Seminars in cancer biology; 2007: Elsevier.
18. Pantuck AJ, Leppert JT, Zomorodian N, Aronson W, Hong J, Barnard RJ, Seeram N, Liker H, Wang H and Elashoff R: Phase II study of pomegranate juice for men with rising prostate-specific antigen following surgery or radiation for prostate cancer. Clinical Cancer Research 2006; 12:4018-4026.
19. Kim ND, Mehta R, Yu W, Neeman I, Livney T, Amichay A, Poirier D, Nicholls P, Kirby A and Jiang W: Chemopreventive and adjuvant therapeutic potential of pomegranate (Punica granatum) for human breast cancer. Breast cancer research and treatment 2002; 71:203-217.
20. Les F, Prieto JM, Arbonés-Mainar JM, Valero MS and López V: Bioactive properties of commercialised pomegranate (Punica granatum) juice: antioxidant, antiproliferative and enzyme inhibiting activities. Food & Function 2015.
21. Johanningsmeier SD and Harris GK: Pomegranate as a functional food and nutraceutical source. Annual review of food science and technology 2011; 2:181-201.
22. Tran HNA, Bae S-Y, Song B-H, Lee B-H, Bae Y-S, Kim Y-H, Lansky EP and Newman RA: Pomegranate (Punica granatum) seed linolenic acid isomers: concentration-dependent modulation of estrogen receptor activity. Endocrine research 2010; 35:1-16.
23. Medjakovic S and Jungbauer A: Pomegranate: a fruit that ameliorates metabolic syndrome. Food & function 2013; 4:19-39.
24. Shiraiishi T, Abe M and Miyagawa T: Cheese foods containing conjugated polyunsaturated fatty acid glycerides. Japanese Patent: JP 2002; 2002176913.
25. Lei F, Zhang X, Wang W, Xing D, Xie W, Su H and Du L: Evidence of anti-obesity effects of the pomegranate leaf extract in high-fat diet induced obese mice. International Journal of Obesity 2007; 31:1023-1029.
26. Ahmed MM, Samir E-SA, El-Shehawi AM and Alkafafy ME: Anti-obesity effects of taif and egyptian pomegranates: Molecular study. Bioscience, biotechnology, and biochemistry 2015; 79:598-609.
27. Ismail T, Sestili P and Akhtar S: Pomegranate peel and fruit extracts: a review of potential anti-inflammatory and anti-infective effects. Journal of Ethnopharmacology 2012; 143:397-405.
28. Shukla M, Gupta K, Rasheed Z, Khan KA and Haqqi TM: Bioavailable constituents/metabolites of pomegranate (Punica granatum L) preferentially inhibit COX2 activity ex vivo and IL-1beta-induced PGE2 production in human chondrocytes in vitro. Journal of inflammation 2008; 5:9.
29. Arun N and Singh D: Punica granatum: a review on pharmacological and therapeutic properties. International Journal Of Pharmaceutical Sciences And Research 2012; 3:1240-1245.
30. Kýralan M, Gölükcü M and Tokgöz H: Oil and conjugated linolenic acid contents of seeds from important pomegranate cultivars (Punica granatum L.) grown in Turkey. Journal of the American Oil Chemists' Society 2009; 86:985-990.
31. Vroegrijk IO, van Diepen JA, van den Berg S, Westbroek I, Keizer H, Gambelli L, Hontecillas R, Bassaganya-Riera J, Zondag GC and Romijn JA: Pomegranate seed oil, a rich source of punonic acid, prevents diet-induced obesity and insulin resistance in mice. Food and Chemical Toxicology 2011; 49:1426-1430.
32. Elfalleh W, Ying M, Nasri N, Sheng-Hua H, Guasmi F and Ferchichi A: Fatty acids from Tunisian and Chinese pomegranate (Punica granatum L.) seeds. International journal of food sciences and nutrition 2011; 62:200-206.

33. Carvalho Filho JM: Pomegranate Seed Oil (*Punica Granatum L.*): A Source of Punicic Acid (Conjugated α -Linolenic Acid). *Journal of Human Nutrition & Food Science* 2014.
34. Pande G. Antioxidant capacity and lipid characterization of Georgia-grown underutilized fruit crops: University of Georgia; 2009.
35. Habibnia M, Ghavami M, Ansaripour M and Vosough S: Chemical evaluation of oils extracted from five different varieties of Iranian pomegranate seeds. *Journal of Food Biosciences and Technology* 2012; 2:35-40.
36. Tsuyuki H, Ito S and Nakatsukasa Y: Lipids in pomegranate seeds. *Nihon Daigaku No-Juigakubu Gakujutsu Kenkyu Hokoku* 1981; 38:141-148.
37. Wang R-F, Xie W-D, Zhang Z, Xing D-M, Ding Y, Wang W, Ma C and Du L-J: Bioactive Compounds from the Seeds of *Punica granatum* (Pomegranate). *Journal of Natural Products* 2004; 67:2096-2098.
38. Carvalho EBTd, Melo ILPd and Mancini-Filho J: Chemical and physiological aspects of isomers of conjugated fatty acids. *Food Science and Technology (Campinas)* 2010; 30:295-307.
39. Yuan G, Sinclair AJ, Xu C and Li D: Research Article Incorporation and metabolism of punicic acid in healthy young humans. *Mol Nutr Food Res* 2009; 53:1336-1342.
40. Yuan GF, Wahlqvist ML, Yuan JQ, Wang QM and Li D: Effect of punicic acid naturally occurring in food on lipid peroxidation in healthy young humans. *Journal of the Science of Food and Agriculture* 2009; 89:2331-2335.
41. Grossmann ME, Mizuno NK, Schuster T and Cleary MP: Punicic acid is an ω -5 fatty acid capable of inhibiting breast cancer proliferation. *International journal of oncology* 2010; 36:421-426.
42. Saha S and Ghosh M: Comparative study of antioxidant activity of α -eleostearic acid and punicic acid against oxidative stress generated by sodium arsenite. *Food and Chemical Toxicology* 2009; 47:2551-2556.
43. Yamasaki M, Kitagawa T, Koyanagi N, Chujo H, Maeda H, Kohno-Murase J, Imamura J, Tachibana H and Yamada K: Dietary effect of pomegranate seed oil on immune function and lipid metabolism in mice. *Nutrition* 2006; 22:54-59.
44. Morikawa K, Nonaka M, Narahara M, Torii I, Kawaguchi K, Yoshikawa T, Kumazawa Y and Morikawa S: Inhibitory effect of quercetin on carrageenan-induced inflammation in rats. *Life Sciences* 2003; 74:709-721.
45. Bassaganya-Riera J. Method of using punicic acid to enhance immune response and prevent metabolic disorders. Google Patents; 2014.
46. Caligiani A, Bonzanini F, Palla G, Cirlini M and Bruni R: Characterization of a potential nutraceutical ingredient: pomegranate (*Punica granatum L.*) seed oil unsaponifiable fraction. *Plant foods for human nutrition* 2010; 65:277-283.
47. Sassano G, Sanderson P, Franx J, Groot P, van Straalen J and Bassaganya-Riera J: Analysis of pomegranate seed oil for the presence of jacaric acid. *Journal of the Science of Food and Agriculture* 2009; 89:1046-1052.
48. Koba K, Imamura J, Akashoshi A, Kohno-Murase J, Nishizono S, Iwabuchi M, Tanaka K and Sugano M: Genetically modified rapeseed oil containing cis-9, trans-11, cis-13-octadecatrienoic acid affects body fat mass and lipid metabolism in mice. *Journal of agricultural and food chemistry* 2007; 55:3741-3748.
49. Račková L, Ergin V, Burcu Bali E, Kuniaková M and Karasu Ç: Pomegranate Seed Oil Modulates Functions and Survival of BV-2 Microglial Cells in vitro. *International journal for vitamin and nutrition research Internationale Zeitschrift für Vitamin-und Ernährungsforschung Journal international de vitaminologie et de nutrition* 2014; 84:295.
50. Topkafa M, Kara H and Sherazi STH: Evaluation of the Triglyceride Composition of Pomegranate Seed Oil by RP-HPLC Followed by GC-MS. *Journal of the American Oil Chemists' Society* 2015; 92:791-800.
51. Boussetta T, Raad H, Lettéron P, Gougerot-Pocidallo M-A, Marie J-C, Driss F and El-Benna J: Punicic acid a conjugated linolenic acid inhibits TNF α -induced neutrophil hyperactivation and protects from experimental colon inflammation in rats. *PloS one* 2009; 4:e6458.
52. Mirmiran P, Fazeli MR, Asghari G, Shafiee A and Azizi F: Effect of pomegranate seed oil on hyperlipidaemic subjects: a double-blind placebo-controlled clinical trial. *British journal of nutrition* 2010; 104:402-406.
53. Hontecillas R, O'Shea M, Einerhand A, Diguardo M and Bassaganya-Riera J: Activation of PPAR γ and α by punicic acid ameliorates glucose tolerance and suppresses obesity-related inflammation. *Journal of the American College of Nutrition* 2009; 28:184-195.
54. Nagao K and Yanagita T: Conjugated fatty acids in food and their health benefits. *Journal of bioscience and bioengineering* 2005; 100:152-157.
55. Igarashi M and Miyazawa T: Newly recognized cytotoxic effect of conjugated trienoic fatty acids on cultured human tumor cells. *Cancer letters* 2000; 148:173-179.
56. Arao K, Yotsumoto H, Han S-Y, Nagao K and Yanagita T: The 9 cis, 11 trans, 13 cis isomer of conjugated linolenic acid reduces apolipoprotein B100 secretion and triacylglycerol synthesis in HepG2 cells. *Bioscience, biotechnology, and biochemistry* 2004; 68:2643-2645.
57. Saha SS and Ghosh M: Antioxidant effect of vegetable oils containing conjugated linolenic acid isomers against induced tissue lipid peroxidation and inflammation in rat model. *Chemico-biological interactions* 2011; 190:109-120.
58. Bassaganya-Riera J, Diguardo M, Climent M, Vives C, de Horna A, Sanchez S, Orpi M, Duran E, Einerhand AW and O'Shea M: Punicic acid modulates mucosal immune responses and prevents gut inflammation through PPAR $\{\gamma\}$ and $\{\delta\}$ -dependent mechanisms. *The FASEB Journal* 2010; 24:926-926.
59. Schubert SY, Lansky EP and Neeman I: Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids. *Journal of ethnopharmacology* 1999; 66:11-17.
60. Jiang Q, Elson-Schwab I, Courtemanche C and Ames BN: γ -Tocopherol and its major metabolite, in contrast to α -tocopherol, inhibit cyclooxygenase activity in macrophages and epithelial cells. *Proceedings of the National Academy of Sciences* 2000; 97:11494-11499.
61. Masamune A, Satoh M, Kikuta K, Suzuki N, Satoh K and Shimosegawa T: Ellagic acid blocks activation of pancreatic stellate cells. *Biochemical pharmacology* 2005; 70:869-878.
62. Madlener S, Illmer C, Horvath Z, Saiko P, Losert A, Herbacek I, Grusch M, Elford HL, Krupitza G and Bernhaus A: Gallic acid inhibits ribonucleotide reductase and cyclooxygenases in human HL-60 promyelocytic leukemia cells. *Cancer letters* 2007; 245:156-162.
63. Costantini S, Rusolo F, De Vito V, Moccia S, Picariello G, Capone F, Guerriero E, Castello G and Volpe MG: Potential anti-inflammatory effects of the hydrophilic fraction of pomegranate (*Punica granatum L.*) seed oil on breast cancer cell lines. *Molecules* 2014; 19:8644-8660.

64. Asghari G, Sheikholeslami S, Mirmiran P, Chary A, Hedayati M, Shafiee A and Azizi F: Effect of pomegranate seed oil on serum TNF- α level in dyslipidemic patients. *International journal of food sciences and nutrition* 2012; 63:368-371.
65. Saha SS and Ghosh M: Antioxidant and anti-inflammatory effect of conjugated linolenic acid isomers against streptozotocin-induced diabetes. *British Journal of Nutrition* 2012; 108:974-983.
66. Coursodon-Boydiddle CF, Snarrenberg CL, Adkins-Rieck CK, Bassaganya-Riera J, Hontecillas R, Lawrence P, Brenna JT, Jouni ZE and Dvorak B: Pomegranate seed oil reduces intestinal damage in a rat model of necrotizing enterocolitis. *American Journal of Physiology-Gastrointestinal and Liver Physiology* 2012; 303:G744-G751.
67. Reuter S, Gupta SC, Chaturvedi MM and Aggarwal BB: Oxidative stress, inflammation, and cancer: how are they linked? *Free Radical Biology and Medicine* 2010; 49:1603-1616.
68. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M and Telser J: Free radicals and antioxidants in normal physiological functions and human disease. *The international journal of biochemistry & cell biology* 2007; 39:44-84.
69. Gloire G, Legrand-Poels S and Piette J: NF- κ B activation by reactive oxygen species: fifteen years later. *Biochemical pharmacology* 2006; 72:1493-1505.
70. Stohs S and Bagchi D: Oxidative mechanisms in the toxicity of metal ions. *Free radical biology and medicine* 1995; 18:321-336.
71. Mukherjee C, Bhattacharyya S, Ghosh S and Bhattacharyya DK: Dietary Effects of Punicic Acid on the Composition and Peroxidation of Rat Plasma Lipid. *Journal of Oleo Science* 2002; 51:513-522.
72. Dhar P, Bhattacharyya D, Bhattacharyya D and Ghosh S: Dietary comparison of conjugated linolenic acid (9 cis, 11 trans, 13 trans) and α -tocopherol effects on blood lipids and lipid peroxidation in alloxan-induced diabetes mellitus in rats. *Lipids* 2006; 41:49-54.
73. Elfalleh W, Tlili N, Nasri N, Yahia Y, Hannachi H, Chaira N, Ying M and Ferchichi A: Antioxidant capacities of phenolic compounds and tocopherols from Tunisian pomegranate (*Punica granatum*) fruits. *Journal of Food Science* 2011; 76:C707-C713.
74. Seeram NP, Adams LS, Henning SM, Niu Y, Zhang Y, Nair MG and Heber D: In vitro antiproliferative, apoptotic and antioxidant activities of punicalagin, ellagic acid and a total pomegranate tannin extract are enhanced in combination with other polyphenols as found in pomegranate juice. *The Journal of nutritional biochemistry* 2005; 16:360-367.
75. Anusree S, Nisha V, Priyanka A and Raghu K: Insulin resistance by TNF- α is associated with mitochondrial dysfunction in 3T3-L1 adipocytes and is ameliorated by punicalagin, a PPAR γ agonist. *Molecular and cellular endocrinology* 2015; 413:120-128.
76. Zolezzi JM, Silva-Alvarez C, Ordenes D, Godoy JA, Carvajal FJ, Santos MJ and Inestrosa NC: Peroxisome proliferator-activated receptor (PPAR) γ and PPAR α agonists modulate mitochondrial fusion-fission dynamics: relevance to reactive oxygen species (ROS)-related neurodegenerative disorders? 2013.
77. Miranda J, Aguirre L, Fernández-Quintela A, Macarulla MT, Martínez-Castaño MG, Ayo J, Bilbao E and Portillo MP: Effects of pomegranate seed oil on glucose and lipid metabolism-related organs in rats fed an obesogenic diet. *Journal of agricultural and food chemistry* 2013; 61:5089-5096.
78. McFarlin BK, Strohacker KA and Kueht ML: Pomegranate seed oil consumption during a period of high-fat feeding reduces weight gain and reduces type 2 diabetes risk in CD-1 mice. *British Journal of Nutrition* 2009; 102:54-59.
79. Nekooeian AA, Eftekhari MH, Adibi S and Rajaeifard A: Effects of Pomegranate Seed Oil on Insulin Release in Rats with Type 2 Diabetes. *Iranian Journal of Medical Sciences* 2014; 39:130-135.
80. Bhandari PR: Pomegranate (*Punica granatum* L). Ancient seeds for modern cure? Review of potential therapeutic applications. *International Journal of Nutrition, Pharmacology, Neurological Diseases* 2012; 2:171.
81. Banihani S, Swedan S and Alguraan Z: Pomegranate and type 2 diabetes. *Nutrition research* 2013; 33:341-348.
82. Boroushaki MT, Mollazadeh H, Rajabian A, Dolati K, Hoseini A, Paseban M and Farzadnia M: Protective effect of pomegranate seed oil against mercuric chloride-induced nephrotoxicity in rat. *Ren Fail* 2014; 36:1581-1586.
83. Boroushaki MT, Arshadi D, Jalili-Rasti H, Asadpour E and Hosseini A: Protective effect of pomegranate seed oil against acute toxicity of diazinon in rat kidney. *Iranian journal of pharmaceutical research: IJPR* 2013; 12:821.
84. Asadpour E, Boroushaki MT and Sadeghnia H: Protective effect of pomegranate seed oil against gentamicin induced nephrotoxicity in rat. *Toxicology Letters* 2010; 196:S232.
85. Boroushaki MT, Sadeghnia HR, Banihasan M and Yavari S: Protective effect of pomegranate seed oil on hexachlorobutadiene-induced nephrotoxicity in rat kidneys. *Renal failure* 2010; 32:612-617.
86. Boroushaki MT, Mollazadeh H, Rajabian A, Dolati K, Hoseini A, Paseban M and Farzadnia M: Protective effect of pomegranate seed oil against mercuric chloride-induced nephrotoxicity in rat. *Renal failure* 2014; 36:1581-1586.
87. Sarkaki A and Rezaei M: Improving active and passive avoidance memories deficits due to permanent cerebral ischemia by pomegranate seed extract in female rats. *The Malaysian journal of medical sciences: MJMS* 2013; 20:25.
88. Gabizon R, Mizrahi M, Friedman-Levy Y, Larush L, Frid K, Binyamin O, Feinstein N, Dori D, Ovadia H and Ben-Hur T: Novel Pomegranate Oil Nano-Emulsions for the Prevention and Treatment of Neurodegenerative Diseases: The Case of Genetic CJD (P5. 236). *Neurology* 2014; 82:P5. 236-P235. 236.
89. Hora JJ, Maydew ER, Lansky EP and Dwivedi C: Chemopreventive effects of pomegranate seed oil on skin tumor development in CD1 mice. *Journal of medicinal food* 2003; 6:157-161.
90. Jiang Q, Wong J, Fyrst H, Saba JD and Ames BN: γ -Tocopherol or combinations of vitamin E forms induce cell death in human prostate cancer cells by interrupting sphingolipid synthesis. *Proceedings of the National Academy of Sciences of the United States of America* 2004; 101:17825-17830.
91. Campbell SE, Stone WL, Lee S, Whaley S, Yang H, Qui M, Goforth P, Sherman D, McHaffie D and Krishnan K: Comparative effects of RRR- α - and RRR- γ -tocopherol on proliferation and apoptosis in human colon cancer cell lines. *BMC cancer* 2006; 6:13.
92. Suzuki R, Noguchi R, Ota T, Abe M, Miyashita K and Kawada T: Cytotoxic effect of conjugated trienoic fatty acids on mouse tumor and human monocytic leukemia cells. *Lipids* 2001; 36:477-482.

93. Li T-M, Chen G-W, Su C-C, Lin J-G, Yeh C-C, Cheng K-C and Chung J-G: Ellagic acid induced p53/p21 expression, G1 arrest and apoptosis in human bladder cancer T24 cells. *Anticancer Research* 2005; 25:971-979.
94. Veluri R, Singh RP, Liu Z, Thompson JA, Agarwal R and Agarwal C: Fractionation of grape seed extract and identification of gallic acid as one of the major active constituents causing growth inhibition and apoptotic death of DU145 human prostate carcinoma cells. *Carcinogenesis* 2006; 27:1445-1453.
95. Wang X, Zhang F, Yang L, Mei Y, Long H, Zhang X, Zhang J and Su X: Ursolic acid inhibits proliferation and induces apoptosis of cancer cells in vitro and in vivo. *BioMed Research International* 2011; 2011.
96. Achiwa Y, Hasegawa K, Komiya T and Udagawa Y: Ursolic acid induces Bax-dependent apoptosis through the caspase-3 pathway in endometrial cancer SNG-II cells. *Oncology reports* 2005; 13:51-57.
97. Harmand PO, Duval R, Delage C and Simon A: Ursolic acid induces apoptosis through mitochondrial intrinsic pathway and caspase-3 activation in M4Beu melanoma cells. *International Journal of Cancer* 2005; 114:1-11.
98. Nashed B, Yeganeh B, HayGlass KT and Moghadasian MH: Antiatherogenic effects of dietary plant sterols are associated with inhibition of proinflammatory cytokine production in Apo E-KO mice. *The Journal of nutrition* 2005; 135:2438-2444.
99. Awad AB and Fink CS: Phytosterols as anticancer dietary components: evidence and mechanism of action. *The Journal of nutrition* 2000; 130:2127-2130.
100. Awad AB, Burr AT and Fink CS: Effect of resveratrol and β -sitosterol in combination on reactive oxygen species and prostaglandin release by PC-3 cells. *Prostaglandins, leukotrienes and essential fatty acids* 2005; 72:219-226.
101. Vivancos M and Moreno JJ: β -Sitosterol modulates antioxidant enzyme response in RAW 264.7 macrophages. *Free Radical Biology and Medicine* 2005; 39:91-97.
102. Arao K, Wang Y-M, Inoue N, Hirata J, Cha J-Y, Nagao K and Yanagita T: Dietary effect of pomegranate seed oil rich in 9cis, 11trans, 13cis conjugated linolenic acid on lipid metabolism in obese, hyperlipidemic OLETF rats. *Lipids Health Dis* 2004; 3:1-7.
103. Yang L, Leung KY, Cao Y, Huang Y, Ratnayake W and Chen Z-Y: α -Linolenic acid but not conjugated linolenic acid is hypocholesterolaemic in hamsters. *British journal of nutrition* 2005; 93:433-438.
104. Spilmont M, Léotoing L, Davicco M-J, Lebecque P, Mercier S, Miot-Noirault E, Pilet P, Rios L, Wittrant Y and Coxam V: Pomegranate seed oil prevents bone loss in a mice model of osteoporosis, through osteoblastic stimulation, osteoclastic inhibition and decreased inflammatory status. *The Journal of nutritional biochemistry* 2013; 24:1840-1848.
105. Minaiyan M, Zolfaghari B, Taheri D and Gomarian M: Preventive Effect of Three Pomegranate (Punica granatum L.) Seeds Fractions on Cerulein-Induced Acute Pancreatitis in Mice. *International Journal of Preventive Medicine* 2014; 5:394-404.
106. Shaban NZ, El-Kersh MA, El-Rashidy FH and Habashy NH: Protective role of Punica granatum (pomegranate) peel and seed oil extracts on diethylnitrosamine and phenobarbital-induced hepatic injury in male rats. *Food chemistry* 2013; 141:1587-1596.
107. Al-Sabahi BN, Fatope MO, Essa MM, Subash S, Al-Busafi SN, Al-Kusaibi FS and Manivasagam T: Pomegranate seed oil: Effect on 3-nitropropionic acid-induced neurotoxicity in PC12 cells and elucidation of unsaturated fatty acids composition. *Nutritional neuroscience* 2014.
108. Mizrahi M, Friedman-Levi Y, Larush L, Frid K, Binyamin O, Dori D, Fainstein N, Ovadia H, Ben-Hur T and Magdassi S: Pomegranate seed oil nanoemulsions for the prevention and treatment of neurodegenerative diseases: the case of genetic CJD. *Nanomedicine: Nanotechnology, Biology and Medicine* 2014; 10:1353-1363.
109. Auerbach L, Rakus J, Bauer C, Gerner C, Ullmann R, Wimmer H and Huber J: Pomegranate seed oil in women with menopausal symptoms: a prospective randomized, placebo-controlled, double-blinded trial. *Menopause* 2012; 19:426-432.
110. Aslam MN, Lansky EP and Varani J: Pomegranate as a cosmeceutical source: pomegranate fractions promote proliferation and procollagen synthesis and inhibit matrix metalloproteinase-1 production in human skin cells. *Journal of ethnopharmacology* 2006; 103:311-318.
111. Meerts I, Verspeek-Rip C, Buskens C, Keizer H, Bassaganya-Riera J, Jouni Z, Van Huygevoort A, Van Otterdijk F and Van de Waart E: Toxicological evaluation of pomegranate seed oil. *Food and Chemical Toxicology* 2009; 47:1085-1092.

How to cite this article:

Md. Boroushaki T, Mollazadeh H and Afshari AR: Pomegranate Seed Oil: A Comprehensive Review on Its Therapeutic Effects. *Int J Pharm Sci Res* 2016; 7(2): 430-42. doi: 10.13040/IJPSR.0975-8232.7(2).430-42.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)