



Received on 24 October, 2013; received in revised form, 06 January, 2014; accepted, 13 February, 2014; published 01 March, 2014

PHYTOCHEMICAL STUDIES ON *DELONIX ELATA* BARK

Gopal Murugananthan*¹ and Mohan Shamanna²

Department of Pharmacognosy¹, Department of Pharmaceutical Chemistry², PES College of Pharmacy, Hanumanthnagar, Bangalore - 560 050, Karnataka, India

Keywords:

Phytochemical screening, Phenolics, Flavonoids, Caffeic acid, Apigenin

Correspondence to Author:

Gopal Murugananthan

Department of Pharmacognosy, PES
College of Pharmacy,
Hanumanthnagar, Bangalore - 560
050, Karnataka, India

E-mail: murugan13@gmail.com

ABSTRACT: *Delonix elata* bark was collected, authenticated, dried, coarsely powdered and extracted using 70% alcohol and subjected for preliminary phytochemical screening. Total phenolic and flavonoidal contents were determined using UV/ visible spectrophotometer. In addition the silica column chromatographic separation was carried out to separate the phytochemicals of *D. elata* bark extract. Preliminary phytochemical screening showed positive tests for saponins, flavonoids, tannins and carbohydrates in the alcoholic extract of *Delonix elata* bark. When the extract was analyzed quantitatively by UV/ visible spectrophotometer, 75 mg/gm of gallic acid equivalent total phenolics and 57.4 mg/gm of rutin equivalent flavonoids were observed. Two phenolic compounds, caffeic acid and apigenin, were isolated and characterized by spectral analysis from the alcoholic extract of the *Delonix elata* bark. Moreover, HPLC method was employed to analyze the isolated phytoconstituents i.e. caffeic acid and apigenin present in the fraction of alcoholic extract of *D. elata* bark which showed the similar R_f of separated constituents. The present study reveals the presence of flavonoids in *D. elata* bark as evidenced by the phytochemical screening, and identification of caffeic acid and apigenin.

INTRODUCTION: *Delonix elata* is commonly known as white gulmohur belonging to the family of *Fabaceae* and subfamily *Caesalpinoidae*¹. *Delonix elata* is not a classical Ayurvedic drug, but found included in Shodhala Nighantu under the Sanskrit name of ‘-Siddeshwara-’ during 12th century AD².

The medical usefulness of the tree is acknowledged by people living in the villages who take a decoction of the leaves and barks to get relief from rheumatic problems like pain and stiffness of the joints, especially affecting the knees^{3,4}.

It was observed that local people and Siddha practitioners in Tamil Nadu, India use the *Delonix elata* bark and leaves for treating inflammation and arthritic conditions. The benefits may be attributed to the chemical constituents like β -sitosterol, quercetin, lupelol, lysine, alanine, valine, tyrosine and rhamnose are which reported from *Delonix regia*. Quercetin 3-O-rhamnoglucoside and Quercetin-3O-galactoside are also reported³.

Extensive pharmacological studies on *Delonix elata* exhibited anti-inflammatory^{4, 5, 6, 7}, anti-arthritic^{4, 5}, immune modifying potentials⁵ and anti-oxidant activities⁸ were studied. Earlier studies in our investigations found promising anti-arthritic and immune modifying potentials in the alcoholic extract of *Delonix elata* bark^{4, 5}. Therefore, an attempt was made to carry out phytochemical studies on *D. elata* bark 70 % v/v alcoholic extract using advanced separation techniques.

	<p style="text-align: center;">DOI: 10.13040/IJPSR.0975-8232.5(3).982-88</p>
	<p style="text-align: center;">Article can be accessed online on: www.ijpsr.com</p>
<p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.5(3).982-88</p>	

MATERIALS AND METHODS:

Plant Material: The Barks of the plant *Delonix elata* were obtained from the outskirts of Tiruchengode, Namakkal Dt. Tamilnadu, India. It was identified and authenticated by Prof. K. P. Sreenath (Taxonomist) Bangalore University, Bangalore. A voucher specimen has been prepared and deposited at the Department of Pharmacognosy, PES College of Pharmacy, Bangalore. The collected bark was dried in shade, crushed to coarse-fine powder and used for the studies.

Chemicals: All the solvents and chemicals used for the studies are laboratory grade procured from SD fine chemicals. Gallic acid and Rutin were procured as gift samples from Himalaya drug company, Bangalore. Folin-Ciocalteu reagent was obtained from Merck (India).

Preparation of Extract: The dried powdered bark (1000g) was subjected to continuous hot extraction using 70% v/v methanol in distilled water for 8 h. The extract was filtered, concentrated and the solvent was removed by rotary flash evaporator to get 98g extract. The extract was dried over a desiccator and used for the study.

Phytochemical analysis: The extract was subjected to preliminary qualitative tests to identify the various phytoconstituents i.e. saponins, flavonoids, tannins, alkaloids, glycosides, terpenoids, steroids and carbohydrates by standard procedure⁹.

Determination of total poly phenol content: Qualitative chemical tests revealed that the alcoholic extract of *D. elata* showed prominent results for phenolics and tannins. Hence, estimation of polyphenol content was performed. Total phenolic content of the extract was determined using Folin-Ciocalteu reagent method with Gallic acid as a standard. The alcoholic extract (1000 µg/ml) was taken in a 50 ml volumetric flask with 45 ml distilled water and 1 ml Folin-Ciocalteu reagent and the flask was shaken thoroughly. After 3 min, 3 ml solution 2% Na₂CO₃ was added and the mixture was allowed to stand for 2 h with intermittent shaking. Absorbance was measured at 760 nm using UV/ visible spectrophotometer.

The procedure was repeated for all the standard Gallic acid concentrations (20, 40, 60, 80 and 100 µg) and standard curve derived to obtain linear regression equation^{10,11}.

Total flavonoids assay: The total flavonoid content of *Delonix elata* bark extract was measured with Aluminum chloride assay. An aliquot of 0.2, 0.4, 0.6, 0.8 and 1.0 ml of standard Rutin and 2.5 ml of samples were separately transferred to a 10 ml volumetric flask and 4 ml of distilled water was added. All the flasks were added with 0.3 ml of 5% NaNO₂ and allowed to stand for 5 min. A 0.3 ml of AlCl₃ was added followed by 2 ml of 1 M NaOH and the total volume was made up to 10 ml with distilled water. The solution was mixed well and the absorbance was measured against a reagent blank at 510 nm with UV/ visible spectrophotometer¹¹.

Column Chromatography -1 (CC-1): The alcoholic extract (65 g) of *D. elata* bark loaded onto silica (60-120 mesh size) column prepared in chloroform. The fractions were eluted with chloroform- ethyl acetate (100: 0, 90: 10, 80: 20, 70: 30, 50: 50, 75: 25, 0: 100, v/v). All the elutes were analyzed by TLC method and pooled as master fraction (MF) which yielded seven fractions with yield of 140, 200, 220, 200, 340, 470 and 710 mg (MF 1 to MF7).

Later the silica column was eluted with ethyl acetate-methanol (90: 10, 80: 20, 70: 30, 60: 40, 40: 60, 20: 80 and 0: 100, v/v). The master fractions were yielded as 1.63, 2.26, 1.9, 2.7, 3.2, 2.93 and 5.8 g (MF 8 to MF14).

Based on their TLC profiles of CC-1 fractions which eluted with 70: 30, 60: 40, 40: 60 v/v of ethyl acetate-methanol were combined together (7.80 g) and subjected further for silica column chromatography-2 (CC-2) for purification. Similarly fractions obtained using 20: 80 and 0: 100, v/v ethyl acetate-methanol were combined together (8.73g) and further subjected for chromatographic purification i.e. column chromatography-3 (CC-3).

Column chromatography-2 (CC-2): The collective fraction (7.80g) was subjected for silica column chromatography.

Silica gel (100-200 mesh size) was used as adsorbent and the column was packed with ethyl acetate as initial solvent. Elution was carried out by increasing the polarity with methanol, which yielded a yellow colored compound (40 mg) with the elution ratio of 65:35, v/v ethyl acetate in methanol, named as DECP1.

Column chromatography-3 (CC-3): Similarly, the collective fraction (8.73g), as described above, was subjected using silica (100-200 mesh size) column chromatography with ethyl acetate as initial mobile phase. Elution was carried out by increasing the polarity with methanol, which yielded a yellow colored compound (35 mg) with the elution ratio of 10: 90, v/v of ethyl acetate in methanol, named as DECP2.

HPLC analysis: Isolated compounds i.e. caffeic acid (DECP1) and apigenin (DECP2) and the silica column chromatography fractions i.e. CC-2 and CC-3 were analyzed on a Shimadzu HPLC system using C18 column using standard Apigenin and caffeic acid authentic markers. HPLC column was 5 micron with the dimension of 4.6 x 250 mm length. Mobile phase A was Buffer (consisting of 0.14g KH₂PO₄ with 0.5 ml H₃PO₄ in HPLC grade water QS to 1000 ml) and Mobile phase B was Acetonitrile. Gradient elution was performed with 95: 5, v/v to 20: 80 v/v of A: B. in 30 min, later 95: 5, v/v till 40 min. Samples in the volume of 20 µl was injected and analyzed at 360 nm wavelength. The chromatograms are shown in **Fig. 1-5**.

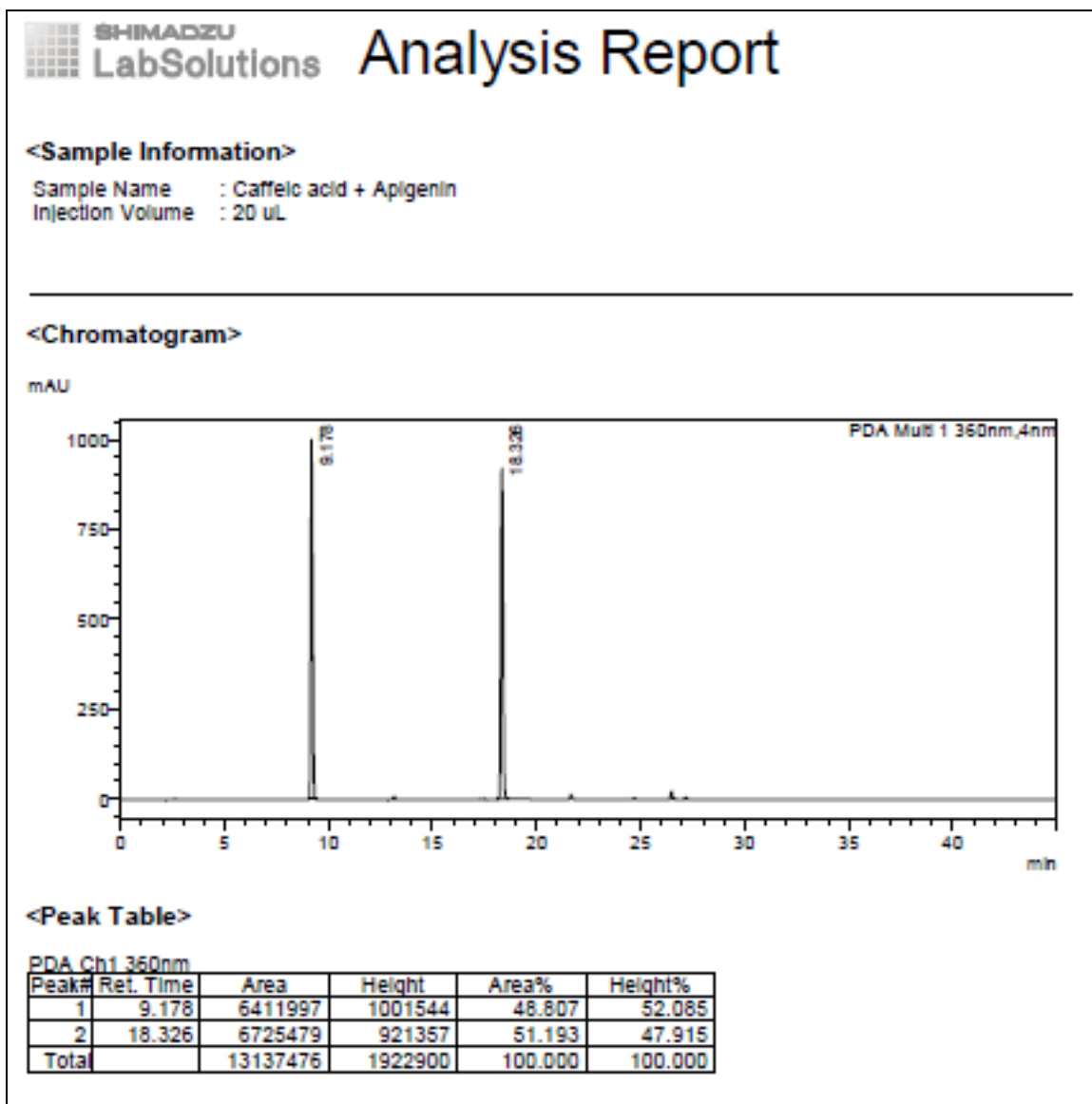


FIG. 1: HPLC CHROMATOGRAMS OF STANDARD APIGENIN AND CAFFIC ACID

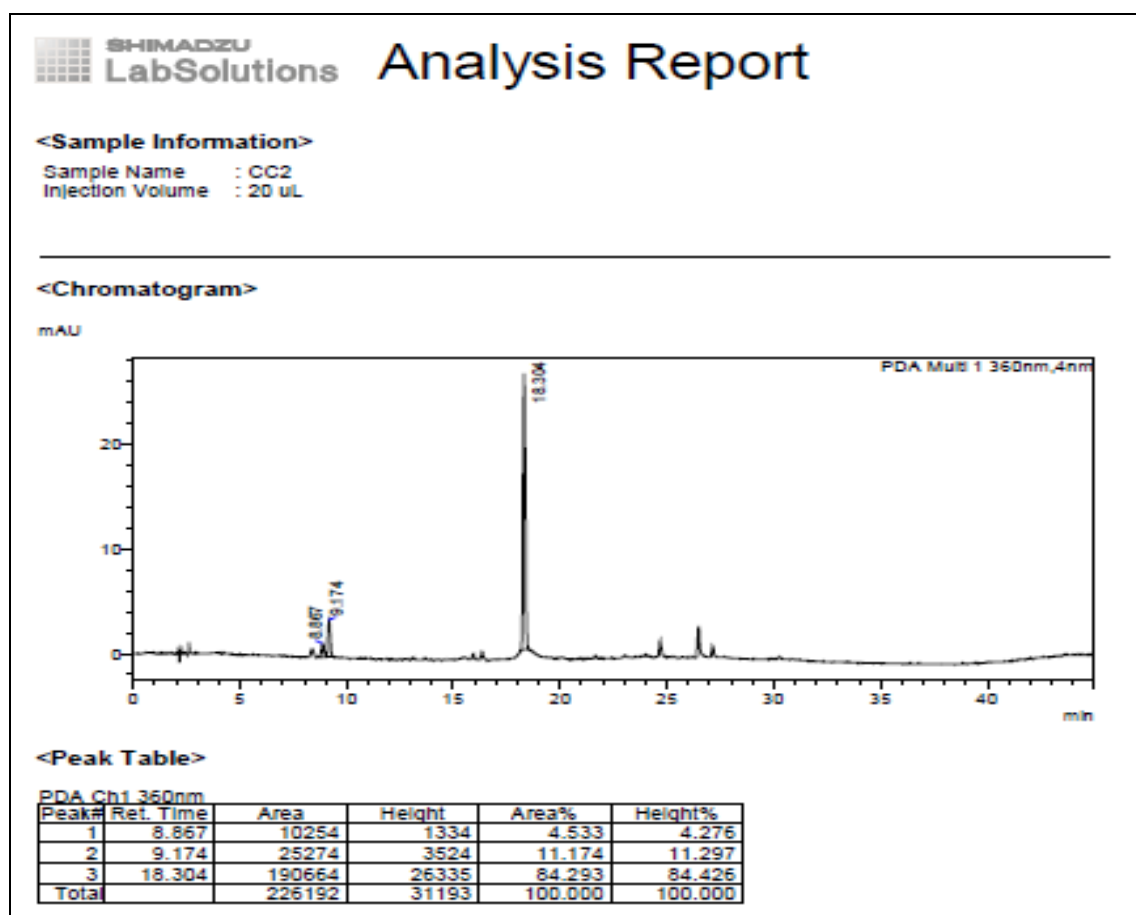


FIG. 2: HPLC CHROMATOGRAM OF CC2 FRACTION

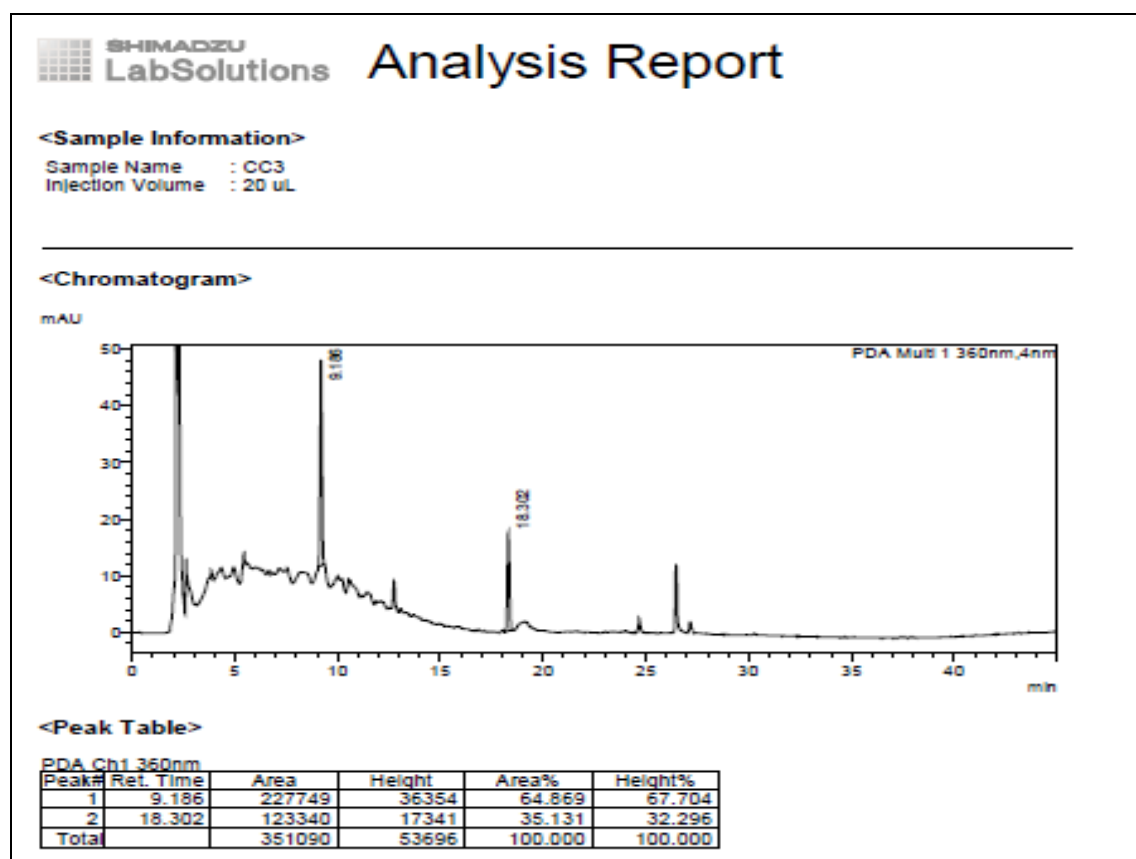


FIG. 3: HPLC CHROMATOGRAM FOR CC3 FRACTION

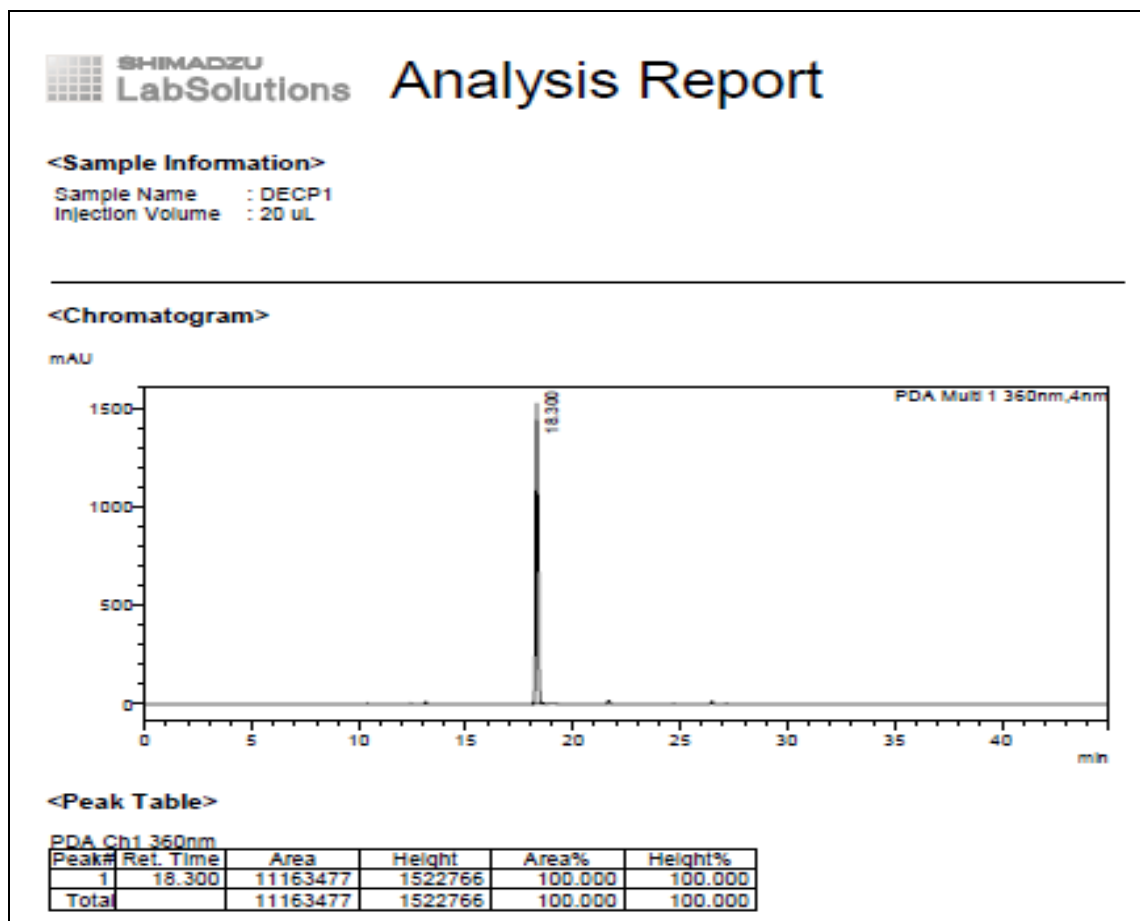


FIG. 4: HPLC CHROMATOGRAM FOR DECP1

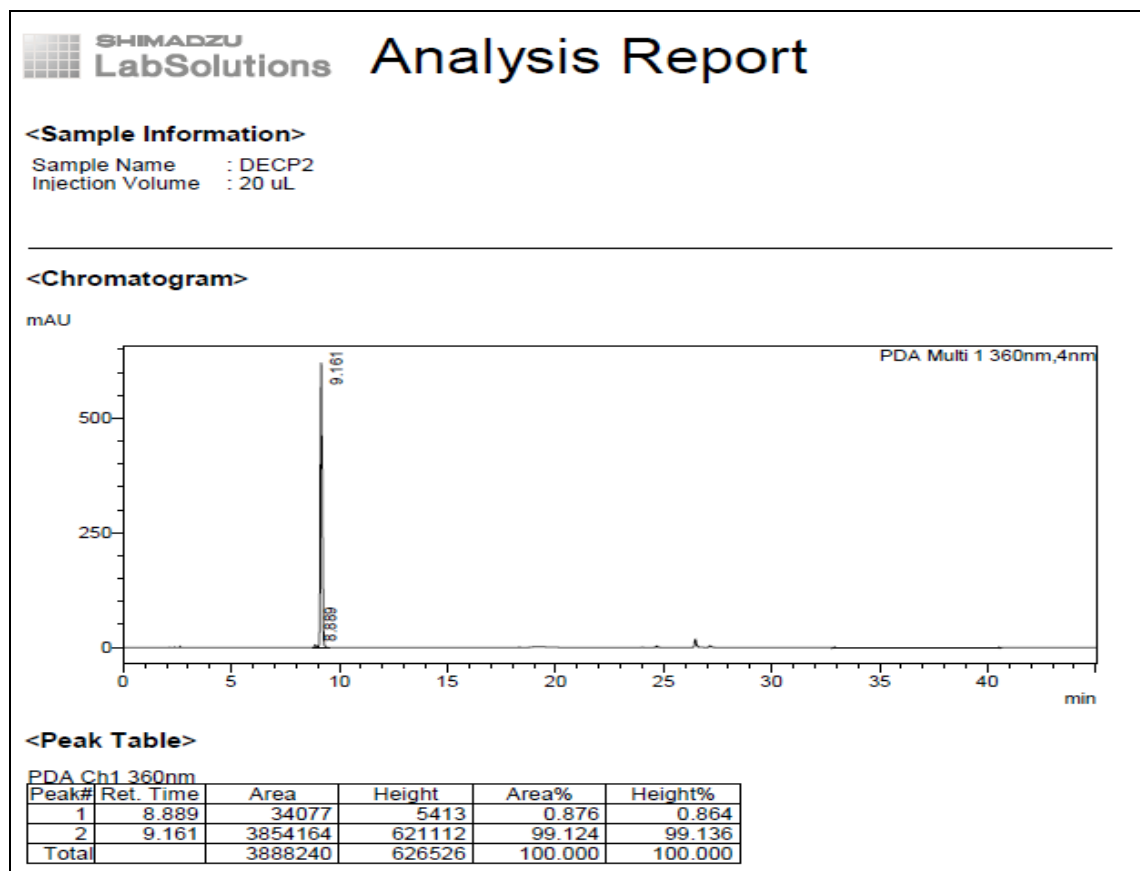


FIG. 5: HPLC CHROMATOGRAM OF DECP2

RESULTS AND DISCUSSION:

Phytochemical analysis: The alcoholic extract of *D. elata* showed the presence of flavonoids, tannins, terpenoids, steroids and carbohydrates in qualitative chemical analysis. UV/visible spectrophotometric determination revealed the presence of 75mg/gm polyphenol content equivalent to 100 g Gallic acid and the flavonoid content as 57.4 mg/gm equivalent to Rutin.

Column Chromatography: During CC-2 silica column chromatographic separation, a yellow colored compound (40 mg) was obtained with the elution ratio of 65:35, v/v ethyl acetate-methanol. After spectral characterization, it was identified as caffeic acid. Similarly, during CC-3 Silica column chromatographic separation, a dull yellowish white colored compound (35 mg) was isolated with ethyl acetate-methanol (10: 90, v/v). After spectral characterization, it was identified as Apigenin, a flavonoid.

HPLC Analysis: HPLC studies showed that standard caffeic acid and apigenin were eluted at retention time (Rt) of 18.32 min and 9.17 min respectively. The isolated compounds i.e. DECP 1 (caffeic acid) showed Rt of 18.30 while DECP2 (apigenin) eluted out at Rt 9.16. The CC-2 loaded fraction exhibited prominent major peaks at Rt 18.30 with some other minor impurities. Similarly CC-3 loaded fraction showed peak at both Rt 9.16 and 18.30 along with some impurities (Figure 2-6). The present work, therefore, suggests that the caffeic acid and apigenin are present in sufficient concentrations to be determined by HPLC method described above.

Spectral data of DECP 1: $^1\text{H-NMR}$ ($\text{DMSO-}d_6$): δ in ppm as 6.1 (1H, d, H-2'), 6.7 (1H, d, H6), 6.9 (1H, d, H-1'), 7.0 (1H, s, H2), 7.4 (1H, d, H5) (aromatic protons); 9.5 (s) and 9.1 (s) (phenolic protons) and 12 (s) (carboxylic acid proton).

EI-MS: $m/z = 179$ (corresponds to molecular formula $\text{C}_9\text{H}_8\text{O}_4$ of the compound caffeic acid).

FTIR (KBr): 3427 cm^{-1} (-OH of carboxylic acid); 3234 cm^{-1} (Phenolic -OH); 3022 cm^{-1} (aromatic -C-H stretch); 1647 cm^{-1} (carbonyl peak of carboxylic acid); 1282 cm^{-1} (vinyl stretching) and 1610 cm^{-1} (aromatic -C=C-).

Spectral data of DECP2: $^1\text{H-NMR}$ ($\text{DMSO-}d_6$): δ values in ppm as 6.1 (1H, d, H3), 6.4 (1H, d, H8) 6.7 (1H, s, H6), 6.9 (2H, m, H2' and H6'), 7.9 (2H, m, H3' and H5') (corresponding to aromatic protons); 10.3 and 10.8 (phenolic protons at 5th and 7th C-atom) and 13 (phenolic proton at C-4' position).

EI-MS: a molecular ion peak at $m/z = 271$ which corresponds to molecular formula $\text{C}_{15}\text{H}_{10}\text{O}_5$ of apigenin.

FTIR (KBr): 3280 cm^{-1} (phenolic -OH group); 3016 cm^{-1} (aromatic C-H stretch); 1656 cm^{-1} (carbonyl group); 1606 cm^{-1} (aromatic C=C peak) and 1242 cm^{-1} (-C-O-C stretching of the flavone ring).

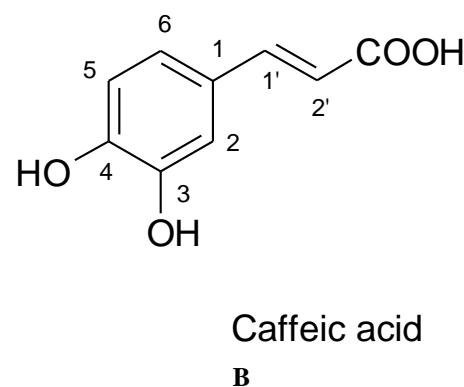
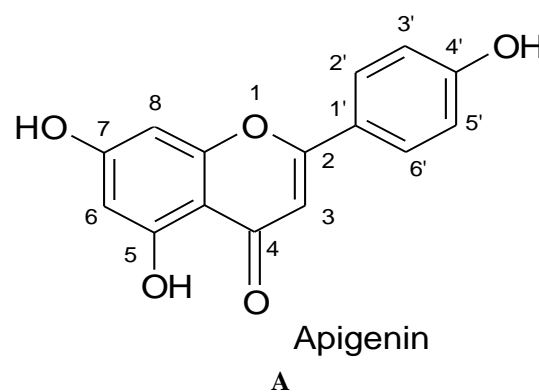


FIGURE 1: CHEMICAL STRUCTURES OF A. APIGENIN AND B. CAFFEIC ACID.

CONCLUSION: An equivalent 75 mg/gm of the total polyphenol and 57.4 mg/gm of the flavonoid content was found in the alcoholic extract of the *D. elata* bark extract. Apigenin and caffeic acid were isolated and characterized for the first time from *D. elata* bark. These constituents are found in *Delonix elata* bark where comparable to other plant-based sources.

Our work hence, confirms and quantitates the reliable sources of Apigenin and Caffeic acids from *Delonix elata* bark which will be helpful for future studies in the extraction of these pharmacologically active compounds.

ACKNOWLEDGEMENT: Authors are thankful to Management, PES Institutions, Bangalore, India for providing facilities to carry out the research work. We are grateful to Natural Remedies Pvt. Ltd., Bangalore and Interdisciplinary School of Indian System of Medicine (ISISM), S.R.M. University, Kattankulathur for providing spectral analysis.

REFERENCES:

1. Ghada Abd El- MH, Invitro studies on *Delonix elata* L. an endangered medicinal plant. World Applied Sci J, 2011; 14(5): 679-686.
2. Wijayasiriwardena C, Sharma PP, Chauhan MG Pillai APG, Pharmacognostical investigation of *Delonix elata* L, from folklore practice. Ayurveda 2009; 30(1): 68-72.
3. Kirtikar, K.R., Basu, B.D., 1956. Indian Medicinal Plants, vol. II, Second ed. Lalit Mohan Basu, Allahabad, India, p. 852
4. Murugananthan G, Mohan S, Anti- Inflammatory and Anti-arthritic activities of *Delonix elata* bark Extracts. Int J of Res in Ayur and Pharm, 2011; 2(6): 1819-1821.
5. Murugananthan G, Mohan S, Anti- arthritic and immune modifying potential of *Delonix elata* Bark extracts. Res J Pharma Biol and Chem Sci, 2013; 4(2):1642-1648.
6. Manimekalai K, Kartik JS, Harsha MS, Evaluation of the effect of the ethanolic extract of *Delonix elata* on acute inflammation in rats. J Natural Pharm, 2011; 2(3):149-153.
7. Krishan Rao RV, Ganapathy P, Mallikarjuna Rao, Ganga Rao B, Anti-inflammatory activity of the leaves and barks of *Delonix elata*. Ancient Sci of Life. 1997; 17(2): 01-03.
8. Ghada Abd El- MH, In-vitro studies on *Delonix elata* L. – An Endangered Medicinal Plant. World App Sci J, 2011; 14(5): 679-686.
9. Kokate C, K. Plant Constituents. In: Practical Pharmacognosy. 1st ed. Delhi: Vallabh Prakashan; 1986, 111-5.
10. Chinedu PA, Ijeoma E, Olusola A, Ayobami OA, Polyphenolic content and antioxidant activity of Hibiscus subdariffa Calyx. Res J of Med Plants, 2011; 5(5): 557-566.
11. Atanassova M, Georgieva S, Ivancheva K. Total phenolic and total flavonoid contents, antioxidant capacity and biological contaminants in medicinal herbs, J of the University of Che Tech and Metallur. 2011; 46(1):81-88.
12. Vishal J, Murugananthan G, Deepak M, Viswanatha GL, Manohar D, Isolation and Standardization of Various Phytochemical Constituents from Methanolic Extracts of Fruit Rinds of *Punica granatu*. Chinese J of Natural Med 2011; 9 (6): 414-420.
13. Rajesh Kumar, Murugananthan G., Nandakumar K., Sahil Talwar, Isolation of anxiolytic principle from ethanolic root extract of *Cardiospermum halicacabum*. Phytomed 2011; (18): 219-23.
14. Courtney N, Sanjay B, Melissa A.V, Oliver H. V, Mikhail A. G, Mark D. W, Denis C. G, Erich G and Andrea I. D, Apigenin Bocks Lipopolysaccharide-induced lethality in-vivo and pro-inflammatory cytokines expression by inactivating NF- κ B through the suppression of p65 Phosphorylation. J Immunol 2007; 179:7121-7127.
15. Omayma A. E, and Samar S. A, Anti-inflammatory Effect of Apigenin-7-neohesperidoside (Rhoifolin) in Carrageenan-Induced Rat Oedema Model. J of Applied Pharma Sci 2012; 02(08): 74-79
16. Kavon R.Z, Jared E, Yun B, Paul R S, Paula B, Jun T and Douglas S R, Apigenin and luteolin modulate microglial activation via inhibition of STAT1-induced CD40 expression. J of Neuroinflam 2008; 5 (41): 01-10.

How to cite this article:

Murugananthan G and Shamanna M: Phytochemical studies on *Delonix elata* bark. *Int J Pharm Sci Res* 2014; 5(3): 982-88. doi: 10.13040/IJPSR.0975-8232.5(3).982-88

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)