



Received on 29 December, 2016; received in revised form, 28 February, 2017; accepted, 21 March, 2017; published 01 July, 2017

ASSESSMENT OF PHYTOCHEMICAL AND ANTIOXIDANT POTENTIAL OF UNDERUTILIZED PEAR (*PYRUS PYRIFOLIA*) AND PLUM (*PRUNUS DOMESTICA*) FROM INDIGENOUS HIMALAYAN REGION OF HIMACHAL PRADESH

Manjeet Singh ^{*1}, P. K. Chauhan ¹, Vikas Kumar ² and Janmeet Kour ¹

School of Bioengineering and Food Technology ¹, School of Applied Science and Biotechnology ², Shoolini University, Post Box No.9, Head Post Office, Solan, Himachal Pradesh - 173212, India.

Keywords:

Pyrus pyrifolia, *Prunus domestica*, Folin-Ciocalteu, Aluminium chloride, Antioxidants, IC₅₀

Correspondence to Author:

Manjeet Singh

Research Scholar
School of Bioengineering and Food Technology, Shoolini University,
Post Box No.9, Head Post Office,
Solan, Himachal Pradesh - 173212,
India.

Email: msdogra0007@gmail.com

ABSTRACT: The present investigation revealed the phytochemical constituent, antioxidant potential and correlation between antioxidant activities with total phenol and flavonoid contents of fruit extracts of *Pyrus pyrifolia* and *Prunus domestica* from local Himalayan region of Himachal Pradesh, India. The total phenolic content was estimated by using Folin-Ciocalteu reagent method, whereas, flavonoid content was quantified using aluminium chloride method. The antioxidant activity was evaluated by using DPPH, FRAP and Nitric oxide (NO) scavenging assay. The total phenolic content was found to be 159.86±2.91 and 145.56±2.43 µg/ml gallic acid equivalents. Similarly, flavonoid content was 48.69±1.90 and 284.27±2.16 µg/ml rutin equivalents for *P. pyrifolia* and *P. domestica*, respectively. The antioxidant activity values of DPPH scavenging for fruit extract were found to be (IC₅₀-17.37 and 9.47 µl/ml), FRAP activity values were (IC₅₀- 15.51 and 6.13 µM Fe (II) equivalents) and Nitric oxide scavenging values were (IC₅₀- 21.49 and 30.11 µl/ml) for *P. pyrifolia* and *P. domestica* respectively. The investigation revealed significant correlation between total phenolic and flavonoid contents with antioxidant activities of fruit extracts. This was the first report which provides insight towards development of value added products.

INTRODUCTION: Epidemiological studies have revealed that consumption of fruit, vegetables, and their byproducts have health benefits against chronic diseases including cardiovascular disease and certain types of cancer ¹. The local fruits of Himachal Pradesh can serve as an ideal source of dietary antioxidants. Due to seasonal constraints and low cost, these fruits are always within the reach of the normal populace.

Consumption of antioxidant rich fruits help to play a protective role against a number of seasonal diseases as well as improve general well-being. Currently, much attention is being focused on the consumption of fruits because of their valuable constituents which contribute towards prevention of degenerative diseases caused by oxidative stress ^{2, 3}. Fruits contain a wide array of dietary phytonutrients such as flavonoids, phenolic acids, carotenoids and vitamins with strong antioxidant capacities ⁴.

In the majority of fruit cultivars of the *Rosaceae* family, especially in the genera of *Malus*, *Pyrus* and *Prunus* species are reported to contain considerable amount of valuable natural antioxidants compounds such as phenolics,

<p>QUICK RESPONSE CODE</p> 	<p>DOI: 10.13040/IJPSR.0975-8232.8(7).2982-87</p> <p>Article can be accessed online on: www.ijpsr.com</p> <p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.8(7).2982-87</p>
---	--

flavonoids, carotenoids and anthocyanin that impart health-promoting effects to the consumers^{5, 6}. It has been proven that plum fruits have several times higher total antioxidant capacity than apples, the latter being one of the most commonly consumed fruits in our diet⁷. Plum fruits demonstrated very good scavenger activity against oxygen-derived free radicals, such as hydroxyl and peroxy radicals⁸. Pear fruits are good source of pectin, help in maintaining desirable acid balance in the body helpful in heart diseases, gastro intestinal tract disorders and recommended to patients suffering from diabetes because of low sucrose content; and included in low antigen content diets to alleviate the symptoms in the management of immune mediated diseases⁹.

Still even though the consumption of pear juices was less than that of citrus and apple juices. Citrus and apple juice are in fashion due to their abundant nutrition especially with rich Vitamin C, good color and flavor^{10, 11}. But many researchers have indicated that the nutritional component of pear juice is as rich as those in citrus and apple juice¹². And we tried to enhance the color and flavor of pear juice products and processing technology according to consumers demand and reported that the nutrition of pear juice may be the next hotspot that will drive the global pear juice industry. The overall nutritional and functional food value of fruits can be better understood by assessing their antioxidants and bio-actives profile which in turn may depend on the type of fruits and their cultivation conditions¹³. Based on numerous evidences on the strong biological activity of phytonutrients and on the scarcity of data for their content in foods, the aim of current study was focused on to evaluate the phenolic content, flavonoid content and antioxidant activity of the underutilized Himalayan pear and plum in order to understand the health benefits of these fruits.

MATERIALS AND METHODS:

Chemicals and Reagents: Ascorbic acid, aluminum chloride, 2,2-diphenyl-2-picrylhydrazyl (DPPH), Sodium nitrite (NaNO₂), 2,4,6-tripyridyl-s-triazine (TPTZ) were purchased from Sigma Chemical Co., U.S.A. Ferric chloride, Folin-Ciocalteu reagent, Gallic acid and Rutin were procured from Loba Chemie Pvt. Ltd, Mumbai,

India. All the chemicals and reagents used in this study were of analytical grade.

Plant materials and extraction procedure: The fresh fruits of *Pyrus pyrifolia* and *Prunus domestica* were collected from Himalayan region (Altitude 1580m above mean sea level) Himachal Pradesh, India. Pear and Plum peels were cut off with a sharp knife to ensure minimum loss of pulp. Seeds from both fruit samples were disposed. The fruit extracts were prepared by adding equal weight of fruit pulp in 100 ml of water followed by mixing in thoroughly in a kitchen grade blender. For each fruit samples 100 ml of fiber free liquid extract was collected by passing the blended mixture through a muslin sieve. Photochemical analyses were done immediately with 50 ml of fiber free fruit extracts and rest of it was stored at 4 °C for further study.

Determination of total phenolic contents: The amount of total phenolic compounds in studied samples was determined according to the Folin-Ciocalteu procedure¹⁴. Total phenolic content was calculated from calibration curve of gallic acid (10-320 µg/ml) and expressed in terms of gallic acid equivalents (GAE) per ml of sample.

Determination of total flavonoid Content: The total flavonoids content in studied samples was determined by using aluminium chloride (AlCl₃) method¹⁵. The flavonoid content was calculated from standard curve of rutin (20-100 µg/ml) and expressed as rutin equivalents (RE) per ml of sample.

In-vitro antioxidant activity:

DPPH radical scavenging activity: DPPH radical scavenging activity of the samples was measured by modified method¹⁶.

Ferric Reducing Antioxidant Power (FRAP) assay: The ability to reduce ferric ions was dignified using the method¹⁷. Ascorbic acid was used as positive reference standard. The antioxidant capacity based on the ability to reduce ferric ions of samples was calculated from the linear calibration curve of FeSO₄ (2.5-20µM) and expressed as µmol FeSO₄ equivalents per ml of sample.

Nitric oxide (NO) scavenging assay: Nitric oxide scavenging assay was carried out using sodium

nitroprusside method with ascorbic acid as positive standard¹⁸.

Statistical analysis: Each sample was analyzed individually in duplication and the results are expressed as the mean value ($n = 3$) \pm standard deviation. Antioxidants assay were analyzed by using two way ANNOVA in Graphpad Prism 5 software. The correlation coefficients between studied parameters were demonstrated by linear regression analysis.

RESULTS AND DISCUSSION:

Determination of total phenolic contents (TPC):

The total phenolic content of fruit extract of *P. pyrifolia* and *P. domestica* was determined by using Folin-Ciocalteu method. The phenolic content was calculated from standard curve of gallic acid (standard plot: $y=0.0042x$, $R^2=0.9932$) (Fig. 1). Fruit extract of *P. pyrifolia* (159.86 ± 2.91 $\mu\text{g/ml}$ gallic acid equivalents) possess higher amount of total phenolic content as compared to that of *P. domestica* fruit extract (145.56 ± 2.43 $\mu\text{g/ml}$ gallic acid equivalents).

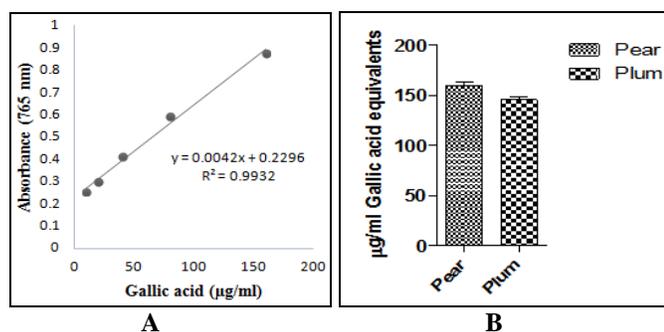


FIG. 1: TOTAL PHENOLIC CONTENT IN FRUIT EXTRACTS OF *P. PYRIFOLIA* AND *P. DOMESTICA* (SHOWN IN B) AND STANDARD CURVE OF GALLIC ACID (SHOWN IN A). PHENOLIC CONTENT OF FRUIT EXTRACTS REPRESENTED AS $\mu\text{g/ml}$ GALLIC ACID EQUIVALENTS. VALUES ARE EXPRESSED AS MEAN \pm STANDARD DEVIATION ($n = 3$)

Determination of Total Flavonoid Content (TFC):

The total flavonoids content of fruit extract of *P. pyrifolia* and *P. domestica* was determined by using aluminium chloride (AlCl_3) method. The flavonoids content was calculated from standard curve of rutin (standard plot: $y=0.0043x$, $R^2=0.9946$) (Fig. 2). Fruit extract of plum (284.27 ± 2.16 $\mu\text{g/ml}$ rutin equivalents) possess higher amount of total flavonoid content as compared to that of pear (48.69 ± 1.90 $\mu\text{g/ml}$ rutin equivalents).

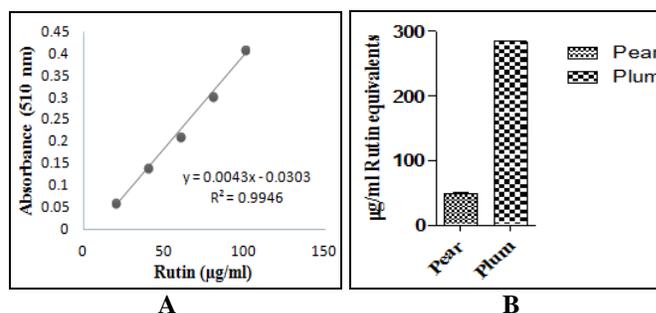


FIG. 2: TOTAL FLAVONOID CONTENT IN FRUIT EXTRACTS OF *P. PYRIFOLIA* AND *P. DOMESTICA* (SHOWN IN B) AND STANDARD CURVE OF RUTIN (SHOWN IN A). FLAVONOID CONTENT OF FRUIT EXTRACTS REPRESENTED AS $\mu\text{g/ml}$ RUTIN EQUIVALENTS. VALUES ARE EXPRESSED AS MEAN \pm STANDARD DEVIATION ($n = 3$)

In-vitro antioxidant activity: There are different methods to evaluate antioxidant characteristics of plants but none of them alone cannot be used for evaluating antioxidant property of extracts. Therefore, it is necessary to characterize the extract by different antioxidant mechanism^{19,20}.

Scavenging activity on 2, 2-diphenyl-1-picrylhydrazyl (DPPH) radical:

Activity on scavenging DPPH radical is a well-established and relatively quick method. Stable DPPH free radical shows strong absorbance at 517 nm which is effectively scavenged by antioxidants. DPPH radical accepts an electron or hydrogen radical from antioxidants present in the samples to be analyzed and become a stable diamagnetic molecule which can be visualized as a noticeable discoloration from purple to yellow²¹.

The percentage scavenging activity of the analyzed fruit extracts were determined using different concentrations (2.5-20 $\mu\text{l/ml}$) which have been taken to compare with the standard ascorbic acid. The DPPH radical scavenging capacity in this study has been reported after 30 minutes of incubation for all samples analyzed. *P. domestica* methanolic extract is the only sample which showed observable antioxidant activity in all different concentrations analyzed. Extent of DPPH radical scavenged was determined by increase in intensity of violet color in the form of IC_{50} values, defined as the concentration of antioxidant required for 50% scavenging of DPPH radicals in specified time period. IC_{50} of the both methanolic extracts along with standard ascorbic acid has been shown in Fig 3. IC_{50} value of *P. domestica* extract

(9.47 $\mu\text{l/ml}$) is lower than that of *P. pyrifolia* (17.37 $\mu\text{l/ml}$), whereas ascorbic acid has IC_{50} -14.52 $\mu\text{l/ml}$.

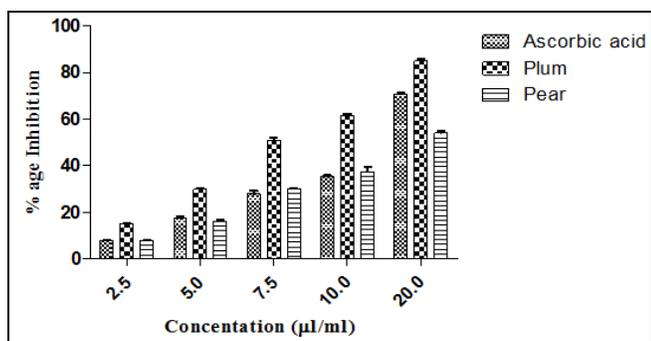


FIG. 3: DPPH RADICAL SCAVENGING ACTIVITY OF FRUIT EXTRACT OF *P. DOMESTICA* AND *P. PYRIFOLIA* IN CONCENTRATION DEPENDENT MANNER WITH STANDARD ASCORBIC ACID. VALUES ARE EXPRESSED AS MEAN \pm STANDARD DEVIATION ($n = 2$).

Ferric Reducing Antioxidant Power (FRAP) assay:

FRAP assay measures the reducing potential of an antioxidant reacting with a ferric tripyridyltriazine [Fe^{3+} -TPTZ] complex and producing a colored ferrous tripyridyltriazine [Fe^{2+} -TPTZ] ¹⁷. Generally, the reducing properties are associated with the presence of compounds which exert their action by breaking the free radical chain by donating a hydrogen atom. Hence the FRAP assay provides a direct estimation of the level of antioxidants or reductants present in a sample. The fruit extracts of *P. pyrifolia* and *P. domestica* showed increased ferric reducing power with the increased concentration as compared to ascorbic acid (Fig. 4). The IC_{50} values were found to be 6.13 $\mu\text{l/ml}$, 15.51 $\mu\text{l/ml}$ and 24.60 $\mu\text{l/ml}$ Fe (II) equivalents for ascorbic acid, *P. domestica* and *P. pyrifolia* extract respectively.

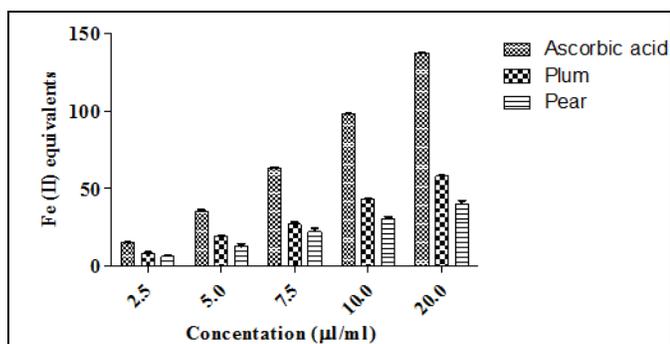


FIG. 4: FERRIC REDUCING ANTIOXIDANT POWER OF FRUIT EXTRACT OF *P. DOMESTICA* AND *P. PYRIFOLIA* IN DIFFERENT CONCENTRATIONS WITH STANDARD ASCORBIC ACID. VALUES ARE EXPRESSED AS MEAN \pm STANDARD DEVIATION ($n = 2$).

Nitric oxide scavenging assay: Nitric oxide radical is well known as it has an important role in various types of inflammatory processes. The production of nitric oxide radical at a sustained levels result in direct tissue toxicity and contribute to the vascular collapse associated with septic shock, whereas chronic expression of nitric oxide radical is associated with various carcinomas and inflammatory conditions including juvenile diabetes, multiple sclerosis, arthritis and ulcerative colitis ²². The nitric oxide generated from sodium nitroprusside reacts with oxygen to form nitrite. Fruit extract also moderately inhibits nitrite formation by directly competes with oxygen to react with nitric oxide. The scavenging of NO by the fruit extract was increased in concentration dependent manner. The fruit extract of *P. domestica* showed maximum activity of 61.48% at 25 $\mu\text{l/ml}$ as compared to that of *P. pyrifolia* and ascorbic acid at the same concentration exhibited 57.72% and 40.61% inhibition respectively (Fig. 5). IC_{50} values were found to be 30.11 $\mu\text{l/ml}$, 21.49 $\mu\text{l/ml}$ and 23.80 $\mu\text{l/ml}$ for ascorbic acid, *P. domestica* and *P. pyrifolia* fruit extracts respectively.

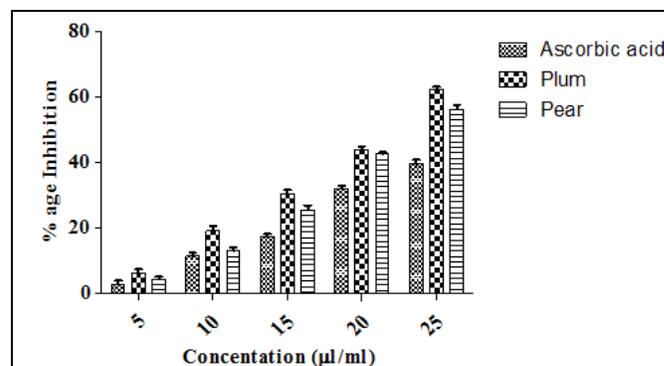


FIG. 5: NITRIC OXIDE SCAVENGING ACTIVITY OF FRUIT EXTRACT OF *P. DOMESTICA* AND *P. PYRIFOLIA* IN CONCENTRATION DEPENDENT MANNER WITH STANDARD ASCORBIC ACID. VALUES ARE EXPRESSED AS MEAN \pm STANDARD DEVIATION ($n = 2$).

Correlation of total phenolic content and flavonoid content with antioxidant activity:

Phenolic compounds are the secondary metabolites which act as natural antioxidants. Phenolic content of the plants are directly correlated with their antioxidant activity ²³. Several studies have reported on the strong relationships between phenolic content and the antioxidant activity ^{24, 25}. Our findings also showed that total phenolics and

flavonoids were directly correlated with antioxidant capacity of both *P. pyrifolia* and *P. domestica* fruit extracts as shown in **Table 1**.

TABLE 1: CORRELATION BETWEEN TOTAL PHENOLICS AND FLAVONOIDS WITH ANTIOXIDANT ACTIVITIES OF FRUIT EXTRACTS *P. PYRIFOLIA* AND *P. DOMESTICA*

Antioxidant Assays	Correlation coefficient (R ²)			
	Total phenolics		Total flavonoids	
	Pear	Plum	Pear	Plum
DPPH radical scavenging activity	0.9832	0.9658	0.9019	0.9959
Ferric Reducing Antioxidant Power (FRAP) assay	0.9783	0.9044	0.9737	0.9844
Nitric oxide scavenging activity	0.9767	0.9575	0.9385	0.9931

The present study has contributed to considerate the holistic and the impending applications of *Pyrus pyrifolia* and *Prunus domestica* as a source of phytonutrients and antioxidants based on both TPC and TFC values along with their free radical scavenging activities and IC₅₀ values. The results attained from various assays exhibited high antioxidant activity of the fruit extracts even at lower concentrations, which in accordance with good correlation to total phenolics and flavonoids content. Therefore, the prominence of the phytochemical constituents and antioxidant activities of these fruits in the maintenance of health is strengthened as trend of the imminent and can offer tremendous opportunities to develop functional foods and nutraceuticals.

ACKNOWLEDGEMENTS: The authors acknowledge the infrastructure and financial support provided by the Shoolini University.

CONFLICT OF INTEREST: The authors declare no competing financial interest.

REFERENCES:

1. Khoo HE, Azlan A, Kong KW and Ismail A. Phytochemicals and medicinal properties of indigenous tropical fruits with potential for commercial development. Evidence-Based Complementary and Alternative Medicine 2016. (<http://dx.doi.org/10.1155/2016/7591951>).
2. Manzoor M, Anwar F, Bhatti IA, Jamil A. Variation of phenolics and antioxidant activity between peel and pulp parts of pear (*Pyrus communis* L.) fruit. Pakistan Journal of Botany 2013; 45(5):1521-5.
3. Reddy KR, Nurdijati SB and Salleh B: Efficacy of aqueous medicinal plant extracts on growth and citrinin production by *Penicillium citrinum* isolated from rice grains African Journal of Microbiology Research 2010; 4(23):2562-65.
4. de Oliveira AC, Valentim IB, Silva CA, Bechara EJ, de Barros MP, Mano CM and Goulart MO. Total phenolic content and free radical scavenging activities of methanolic extract powders of tropical fruit residues. Food Chemistry 2009; 115(2):469-75.
5. Gil MI, Tomás-Barberán FA, Hess-Pierce B and Kader AA: Antioxidant capacities, phenolic compounds, carotenoids, and vitamin C contents of nectarine, peach, and plum cultivars from California. Journal of Agricultural and Food Chemistry 2002; 50(17):4976-82.
6. Kim DO, Chun OK, Kim YJ, Moon HY and Lee CY: Quantification of polyphenolics and their antioxidant capacity in fresh plums. Journal of Agricultural and Food Chemistry 2003; 51(22):6509-15.
7. Wang H, Cao G and Prior RL. Total antioxidant capacity of fruits. Journal of Agricultural and Food Chemistry 1996; 44(3):701-5.
8. Murcia MA, Jiménez AM and Martínez-Tomé M: Evaluation of the antioxidant properties of Mediterranean and tropical fruits compared with common food additives. Journal of Food Protection 2001; 64(12): 2037-46.
9. Challice JS and Wood MNW: Phytochemistry. 1972; 11: 37-44.
10. Yao YX: Study on nutritional value and health function of apple and apple juice. China fruit and vegetables 2002; 4-13.
11. Wu GX, Zhou HW and Wang JM: Biochemical mechanism and substances determination of enzymic browning of yali pear (*Pyrus Bretschneideri* Rehd). Acta Horticulturae Sinica 1992; 19(3):198-202.
12. Xie D, Zhong HY, Mo J, Li ZH, Cui T and Yi CP: Nutritional and medicinal quality of pear juice: next hotspot. Food 2007; 1(1):41-8.
13. Scalzo J, Politi A, Pellegrini N, Mezzetti B and Battino M: Plant genotype affects total antioxidant capacity and phenolic contents in fruit. Nutrition 2005; 21(2):207-13.
14. Singleton VL, Orthofer R and Lamuela-Raventos RM: Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Methods in enzymology 1999; 299:152-78.
15. Dhingra NA, Sharma RA, Kar AN. Evaluation of the antioxidant activities of *Prunus domestica* whole fruit: An *in vitro* study. International Journal of Pharmacy and Pharmaceutical Sciences 2014; 6:271-6.
16. Barros L, Baptista P and Ferreira IC: Effect of *Lactarius piperatus* fruiting body maturity stage on antioxidant activity measured by several biochemical assays. Food and Chemical Toxicology 2007; 45(9):1731-7.
17. Benzie IF and Strain JJ: The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. Analytical biochemistry 1996; 239(1):70-6.
18. Najafabad AM, Jamei R. Free radical scavenging capacity and antioxidant activity of methanolic and ethanolic extracts of plum (*Prunus domestica* L.) in both fresh and dried samples. Avicenna journal of phytomedicine. 2014; 4(5):343.
19. Chanda S, Dave R: *In vitro* models for antioxidant activity evaluation and some medicinal plants possessing antioxidant properties: An overview. African Journal of Microbiology Research 2009; 3(13):981-96.

20. Chanda S, Amrutiya N and Rakholiya K: Evaluation of antioxidant properties of some Indian vegetable and fruit peels by decoction extraction method. *American Journal of Food Technology* 2013; 8:173-82.
21. Hu C and Kitts DD: Studies on the antioxidant activity of Echinacea root extract. *Journal of Agricultural and Food Chemistry* 2000; 48(5):1466-72.
22. Taylor BS, Kim YM, Wang QI, Shapiro RA, Billiar TR and Geller DA: Nitric oxide down-regulates hepatocyte-inducible nitric oxide synthase gene expression. *Archives of Surgery* 1997; 132(11):1177-83.
23. Orak HH, Yagar H and Isbilir SS: Comparison of antioxidant activities of juice, peel, and seed of pomegranate (*Punica granatum* L.) and inter-relationships with total phenolic, Tannin, anthocyanin, and flavonoid contents. *Food Science and Biotechnology* 2012; 21(2):373-87.
24. Kumar, V.; Dev, K.; Sourirajan, A.; Khosla, P. K. Comparative Antioxidant Potential of Bark and Leaves of *Terminalia arjuna* (Roxb) Wight & Arn from Himachal Pradesh. *International Journal of Pharmaceutical and Phytopharmacological Research* 2016, 6, 27-33.
25. Borneo R, León AE, Aguirre A, Ribotta P and Cantero JJ: Antioxidant capacity of medicinal plants from the Province of Córdoba (Argentina) and their *in vitro* testing in a model food system. *Food Chemistry* 2009; 112(3):664-70.

How to cite this article:

Singh M, Kumar P, Chauhan PK and Kour J: Assessment of phytochemical and antioxidant potential of underutilized pear (*Pyrus pyrifolia*) and plum (*Prunus domestica*) from indigenous Himalayan region of Himachal Pradesh. *Int J Pharm Sci Res* 2017; 8(7): 2982-87. doi: 10.13040/IJPSR.0975-8232.8(7). 2982-87.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)