Keywords: 
Centella asiatica, Evolvulus alsinoides L., Hydro-alcoholic extracts, Sub-acute, Biochemical, Hematological, Histopathological

COMPARATIVE ACUTE AND SUB-ACUTE TOXICITY STUDY OF HYDRO-ALCOHOLIC EXTRACTS OF CENTELLA ASIATICA AND EVOLVULUS ALSINOIDES IN SWISS ALBINO MICE

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INTRODUCTION: Centella asiatica (CA) L. urban belonging to family Apiaceae (Umbelliferae) is a psychoactive medicinal plant being used for centuries in the Ayurvedic system of medicine as a medhya rasayana. It has been reported to possess various pharmacological effects on the central nervous system such as stimulatory-nervine tonic, rejuvenant, sedative, anxiolytic, and intelligence promoting property 1,2. CA has been reported to contain triterpene derivatives in major amounts and the earliest examples of this compounds present in CA were identified in the late 1940s as “asiatic acid and madecassic acid” along with their heterosides named as “asiaticoside” and “madecassoside” constituting approximately up to 10% of the plant 3. Later on, compounds from various chemical classes have been also isolated, such as flavonoids 4, polyacetylenes 5, and phenolic acids 6.

Evolvulus alsinoides (EA) is an important plant that has been well documented in Ayurveda for its therapeutic values. Evolvulus alsinoides (Family: Convolvulaceae) commonly known as Shankhpuspi is found throughout India ascending to 6000ft in the Himalayas. It is well known for its therapeutic

ABSTRACT: In this study, the in-vivo toxicity of Centella asiatica & Evolvulus alsinoides were evaluated by acute and sub-acute toxicity assays according to the guidelines of OECD 423 & 407 respectively. For LD_{50} evaluation, a single dose of hydro-alcoholic extracts of both plants was orally administered to Swiss albino mice at doses of 200, 400, 800, 1600 and 2000 mg/kg. Then the animals were observed for 72 h. For acute toxicity evaluation, a single dose of hydro-alcoholic extracts of both plants was orally administered to mice at doses of 300, 600, 1200 and 2000 mg/kg. Then, the animals were observed for 14 days. In the sub-acute study, the extracts were orally administered to mice for 28 days at doses of 300, 600, 1200, and 2000 mg/kg. To assess the toxicological effects, animals were closely observed on general behavior, clinical signs of toxicity, body weight, food, and water intake. At the end of the study, it was performed biochemical and hematological evaluations, as well as histopathological analysis from the following organs: brain, heart, liver, and kidney.
effect on brain disorders like insanity, epilepsy, memory enhancement and nervous debility in the Indian Ayurvedic system of medicine. Recent pharmacological studies on leaves and whole plant of EA have indicated its anti-inflammatory, antipyretic and anti-diarrhoeal properties and immunomodulatory properties. Some in-vitro experiments have revealed the antioxidant properties of EA.

MATERIALS AND METHODS:

**Plant Material:** Whole plant material of *Centella asiatica* was collected from village Ramnapur, Varanasi, Uttar Pradesh, India in October 2015 and authentication was done by Department of Botany, Banaras Hindu University, India and also herbarium of *C. asiatica* (voucher specimen no. Apia/02/2015) and *Evolvulus alsinoides* (voucher specimen no. Convolvul./03/2015) of plants were deposited in the Department of Botany, Banaras Hindu University, India.

**Preparation of Extracts:** The extraction of both plants was done with Soxhlet method in hydroalcoholic (70: 30 ratio, ethanol: distilled water) solvent at 72-100 °C for 72 h. The Soxhlet extraction has widely been used for extracting valuable bioactive compounds from various natural sources. It is used as a model for the comparison of new extraction alternatives.

Generally, a small amount of dry sample is placed in a thimble. The thimble is then placed in distillation flask which contains the solvent of particular interest. After reaching an overflow level, the solution of the thimble-holder is aspirated by a siphon. Siphon unloads the solution back into the distillation flask. This solution carries extracted solutes into the bulk liquid. The solute remains in the distillation flask and solvent passes back to the solid bed of plant. The process runs repeatedly until the extraction is completed.

**Animals:** The experimental Swiss albino mice were issued by Animal house of Institute of Medical Sciences, Banaras Hindu University Varanasi, Uttar Pradesh.

Animals were divided into experimental groups, housed in plastic cages and maintained on a 12-h light and 12-h dark cycle. They were given standard food and water ad libitum. The Central Animal Ethical Committee of Banaras Hindu University approved all experimental procedures (CAEC/196).

**Preparation of Extract Samples:** Hydro-alcoholic extracts of *C. asiatica* (HACA) and *E. alsinoides* (HAEA) were solubilized in distilled water to obtain solutions of 30, 60, 120, and 200 mg/ml. The doses 300, 600, 1200, and 2000 mg/kg were evaluated for further study.

**Toxicity Assays:** The safety parameters assessed by conducting the acute and sub-acute toxicity study according to the OECD (Organization for Economic Co-operation and Development guideline) guidelines 423 & 407, respectively.

**Acute Toxicity Assay:** The animals were divided into nine experimental groups of 6 animals each. Group 1 received 10 μl/g of distilled water and served as control. Groups 2 to 5 treated with hydro-alcoholic extract of *C. asiatica* at the doses of 300, 600, 1200 and 2000 mg/kg and groups 6 to 9 were treated with the hydro-alcoholic extract of *E. alsinoides* at doses of 300, 600, 1200 and 2000 mg/kg respectively. All treatments were administered once by oral gavage. Animals were closely observed for 4 h following administration and once a day for 14 days on general behavior, clinical signs of toxicity, mortality, food, and water intake. Body weight was measured before and after administration on days 4, 7, 10, and 14. At the end of the experiment, animals were anesthetized with ketamine (20 mg/kg i.p.). After the anesthesia has reached depth, the cardiac puncture was performed to collect blood for biochemical and hematological evaluations.

**Sub-acute Toxicity Assay:** The animals were divided into nine experimental groups of 6 animals each. Group 1 received 10 μl/g of distilled water and served as control. Groups 2 to 5 treated with hydro-alcoholic extract of *C. asiatica* at the doses of 300, 600, 1200 and 2000 mg/kg and groups 6 to 9 were treated with hydro-alcoholic extract of *E. alsinoides* at doses of 300, 600, 1200 and 2000 mg/kg respectively. All treatments were administered once by oral gavage daily 7 days each week for 28 days. Animals were closely observed for 28 days on
general behavior, clinical signs of toxicity, mortality, food, and water intake. Body weight was measured before and after administration on days 7, 14, 21, and 28. At the end of the experiment, animals were anesthetized with ketamine (20 mg/kg i.p.). After the anesthesia has reached depth, the cardiac puncture was performed to collect blood for biochemical and hematological evaluations.

**Hematological Analysis:** The hematological evaluation was performed in all surviving animals at the end of the experiment. The complete blood count was performed using an automated hematology analyzer. Hematological evaluations included red blood cell count (RBC), hemoglobin concentration (HGB), platelet count (PLT), hematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC) and white blood cell count (WBC).

**Blood Serum Biochemistry Analysis:** The biochemical evaluation was performed in all surviving animals at the end of the experiment. The collected blood was transferred to tubes without anticoagulant and allowed to stand for 60 min at room temperature and centrifuged at 4000 rpm for 10 min. The serum from each blood sample was recovered and stored in cryogenic tubes at -80 °C deep freezer. Urea, creatinine, serum glutamic oxaloacetic transaminase (SGOT), serum glutamic pyruvic transaminase (SGPT), and alkaline phosphatase were evaluated.

**Histopathology:** The organs were collected from all surviving animals, washed with saline solution 0.9% (w/v), weight and fixed in 40% formaldehyde solution. Then organs were processed for paraffin embedding. 5 μm thick sections were prepared and stained with hematoxylin and eosin (H & E).

The tissues were analyzed in an optical microscope for their general structure, signs of inflammation, degenerative changes, and necrosis evidence. The images were captured with the microscope Motic B1 series, scanned through micro camera Moticam 480 using the Motic Images Plus 2.0ML Application Suite software.

**Statistical Analysis:** The relative hematological and biochemical data were expressed as the mean ± standard error of the mean (SEM). Data were submitted to analysis of variance (one-way ANOVA) followed by Dunnett multiple comparison test. The software GraphPad Prism 6.0 (GraphPad Software, USA) was used for statistical analysis. P values < 0.05 were considered statistically significant.

**RESULTS:**

**Acute Toxicity & Sub-acute Toxicity:**

**General Signs and Mortality:** No signs of toxicity or behavioral changes were observed after the treatment with HAEA and HACA. No deaths were recorded within 72 h after administration of the extracts and the control in mice.

**Body Weight, Relative Organ Weight, Food and Water Intake:** Female mice treated with HAEA and HACA at the three evaluated doses showed weight gain throughout the entire experiment duration. The increase was the same in treated and control group animals, and the treatment did not affect relative organs weights, food, and water intake.

**Hematological Parameters:** Treatment with HAEA and HACA at all doses did not produce any changes on animal (female) hematological parameters, and the result found statistically significant at P<0.01 Table 1. Hematological parameters of Swiss mice treated for 28 days with different doses (300, 600, 1200 and 2000 mg/kg) of hydro-alcoholic extracts of *E. alsinoides* (HAEA) and *C. asiatica* (HACA).

**Biochemical Parameter:** Treatment with HAEA and HACA at the all doses did not produce any statistically significant changes on urea, creatinine, SGOT, SGPT and alkaline phosphatase Table 2 and the result found statistically significant at P<0.01.

**Histopathological Analysis:** The oral administration of HAEA and HACA did not produce significant dose-dependent histopathological alterations. At the evaluated doses, it was not observed any tissue damage on the brain, heart, lungs, and liver of female mice. Fig. 1-4 microscopic histological slides from different organs of Swiss albino mice treated with hydro-alcoholic extracts of *E. alsinoides* & *C. asiatica* i.e. HAEA & HACA, respectively.
TABLE 1: HEMATOLOGICAL PARAMETERS

<table>
<thead>
<tr>
<th>Groups</th>
<th>HGB (g/dL)</th>
<th>RBC (10^6/µL)</th>
<th>PLT (10^3/µL)</th>
<th>HCT (%)</th>
<th>MCV (fL)</th>
<th>MCH (pg)</th>
<th>MCHC (g/dL)</th>
<th>WBC (10^3/µL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>13.1 ± 0.45</td>
<td>6.78 ± 0.29</td>
<td>881.97 ± 32.4</td>
<td>40.8 ± 0.86</td>
<td>55.7 ± 0.82</td>
<td>19.2 ± 0.57</td>
<td>31.8 ± 1.00</td>
<td>5.78 ± 0.08</td>
</tr>
<tr>
<td>HAEA 300</td>
<td>12.7 ± 0.28</td>
<td>6.24 ± 0.24</td>
<td>936.40 ± 15.7</td>
<td>41.7 ± 0.75</td>
<td>57.4 ± 1.27</td>
<td>20.4 ± 0.51</td>
<td>33.7 ± 0.98</td>
<td>5.89 ± 0.18</td>
</tr>
<tr>
<td>HAEA 600</td>
<td>13.2 ± 0.75</td>
<td>7.11 ± 0.69</td>
<td>858.24 ± 17.2</td>
<td>45.9 ± 1.64</td>
<td>56.2 ± 1.21</td>
<td>18.6 ± 0.52</td>
<td>32.5 ± 0.63</td>
<td>5.41 ± 0.61</td>
</tr>
<tr>
<td>HAEA 1200</td>
<td>12.9 ± 0.73</td>
<td>6.60 ± 0.43</td>
<td>881.57 ± 49.0</td>
<td>42.8 ± 2.10</td>
<td>55.7 ± 1.51</td>
<td>18.0 ± 0.80</td>
<td>30.7 ± 1.24</td>
<td>5.14 ± 0.58</td>
</tr>
<tr>
<td>HAEA 2000</td>
<td>13.2 ± 0.75</td>
<td>7.11 ± 0.69</td>
<td>858.24 ± 17.2</td>
<td>45.9 ± 1.64</td>
<td>56.2 ± 1.21</td>
<td>18.6 ± 0.52</td>
<td>32.5 ± 0.63</td>
<td>5.41 ± 0.61</td>
</tr>
<tr>
<td>HACA 300</td>
<td>12.7 ± 0.28</td>
<td>6.24 ± 0.24</td>
<td>936.41 ± 41.0</td>
<td>41.7 ± 1.06</td>
<td>57.4 ± 1.56</td>
<td>20.4 ± 1.06</td>
<td>33.7 ± 1.61</td>
<td>5.89 ± 0.42</td>
</tr>
<tr>
<td>HACA 600</td>
<td>13.5 ± 0.46</td>
<td>7.25 ± 0.56</td>
<td>892.21 ± 58.9</td>
<td>42.2 ± 1.89</td>
<td>55.8 ± 1.75</td>
<td>19.9 ± 1.38</td>
<td>32.9 ± 0.73</td>
<td>5.27 ± 0.52</td>
</tr>
<tr>
<td>HACA 1200</td>
<td>13.5 ± 0.46</td>
<td>7.25 ± 0.56</td>
<td>892.21 ± 58.9</td>
<td>42.2 ± 1.89</td>
<td>55.8 ± 1.75</td>
<td>19.9 ± 1.38</td>
<td>32.9 ± 0.73</td>
<td>5.27 ± 0.52</td>
</tr>
<tr>
<td>HACA 2000</td>
<td>14.1 ± 0.26</td>
<td>7.91 ± 0.33</td>
<td>810.10 ± 17.6</td>
<td>45.2 ± 1.95</td>
<td>56.3 ± 1.63</td>
<td>19.9 ± 1.38</td>
<td>32.7 ± 0.57</td>
<td>5.67 ± 0.11</td>
</tr>
</tbody>
</table>

n = 6 Swiss albino mice (females), One-way AVOVA, followed by Dunnett's multiple comparison test. *p<0.05, **p<0.01, ***p<0.001. Hemoglobin concentration (HGB), Red blood cell count (RBC), platelet count (PLT), hematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC) and white blood cell count (WBC).

TABLE 2: BIOCHEMICAL PARAMETERS

<table>
<thead>
<tr>
<th>Groups</th>
<th>Urea Normal value= 25-30 mg/dl</th>
<th>Creatinine Normal value= 0.2-0.9 mg/dl</th>
<th>SGOT Normal value= 54-298 mg/dl</th>
<th>SGPT Normal value= 17-77 mg/dl</th>
<th>Alkaline phosphatase Normal value= 35-96 mg/dl</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>27.47 ± 0.94</td>
<td>0.3 ± 0.03</td>
<td>60.64 ± 1.68</td>
<td>49.09 ± 2.07</td>
<td>49.32 ± 3.08</td>
</tr>
<tr>
<td>HAEA 300</td>
<td>26.42 ± 0.67</td>
<td>0.5 ± 0.08</td>
<td>66.48 ± 2.42</td>
<td>34.80 ± 2.16</td>
<td>59.16 ± 1.86</td>
</tr>
<tr>
<td>HAEA 600</td>
<td>27.12 ± 0.95</td>
<td>0.4 ± 0.08</td>
<td>56.06 ± 1.71</td>
<td>54.59 ± 2.61</td>
<td>49.72 ± 3.78</td>
</tr>
<tr>
<td>HAEA 1200</td>
<td>28.36 ± 0.67</td>
<td>0.6 ± 0.05</td>
<td>58.64 ± 2.26</td>
<td>57.65 ± 2.90</td>
<td>50.37 ± 1.28</td>
</tr>
<tr>
<td>HAEA 2000</td>
<td>28.22 ± 0.55</td>
<td>0.6 ± 0.13</td>
<td>63.50 ± 2.58</td>
<td>42.03 ± 1.55</td>
<td>53.21 ± 2.99</td>
</tr>
<tr>
<td>HACA 300</td>
<td>27.72 ± 1.08</td>
<td>0.4 ± 0.11</td>
<td>57.5 ± 1.20</td>
<td>56.59 ± 2.11</td>
<td>52.73 ± 3.67</td>
</tr>
<tr>
<td>HACA 600</td>
<td>28.00 ± 1.23</td>
<td>0.8 ± 0.12</td>
<td>57.20 ± 0.49</td>
<td>51.61 ± 2.59</td>
<td>57.81 ± 3.03</td>
</tr>
<tr>
<td>HACA 1200</td>
<td>26.70 ± 1.18</td>
<td>0.8 ± 0.11</td>
<td>61.91 ± 2.75</td>
<td>48.83 ± 1.98</td>
<td>51.08 ± 2.58</td>
</tr>
<tr>
<td>HACA 2000</td>
<td>27.57 ± 0.53</td>
<td>0.5 ± 0.08</td>
<td>57.97 ± 1.52</td>
<td>49.01 ± 2.78</td>
<td>52.30 ± 1.88</td>
</tr>
</tbody>
</table>

Blood serum biochemical parameters of swiss albino mice treated with dosage (300, 600, 1200 & 2000 mg/kg) of hydro-alcoholic extracts of C. asiatica & E. alsinoides. n = 6 swiss albino mice (females), One-way AVOVA followed by Dunett’s multiple comparison test. *p<0.05, **p<0.01, ***p<0.001.

FIG. 1: HISTOPATHOLOGICAL SLIDES OF KIDNEY ORGAN SHOWN NO CHANGES IN SWISS ALBINO MICE TREATED WITH (B) HAEA & (C) HACA EXTRACTS

FIG. 2: HISTOPATHOLOGICAL SLIDES OF LIVER ORGAN SHOWN NO CHANGES IN SWISS ALBINO MICE TREATED WITH (B) HAEA & (C) HACA EXTRACTS
DISCUSSION: Animal models are widely used to assess the preliminary toxicity because the early identification of side effects is usually predictive of the toxicity in humans and can save time, resources, and efforts.

In this study, several parameters evaluated after the in-vivo acute and sub-acute administration of hydro-alcoholic extracts from *E. alsinoides* & *C. asiatica*. In toxicological evaluation, mortality is an important criteria, and there was no mortality seen in the both acute and sub-acute evaluation of extracts. For LD$_{50}$ no death was recorded in 72 h of administration of extracts. In acute toxicity, no death was recorded in 14 days extracts administration and in sub-acute toxicity study also no death recorded for 28 days extract administration. Clinical signs of toxicity were observed after the acute administration and during the sub-acute evaluation for all extract dosage.

It is known that several toxic compounds accumulate in the liver where the detoxification occurs. Liver damage is usually assessed by the determination of SGOT, SGPT, and alkaline phosphatase. It was not observed any significant alterations in serum levels of these three markers of liver function after acute and sub-acute administration of *E. alsinoides* and *C. asiatica* extracts, and histopathological analysis of liver did not cause liver damage.

The kidneys receive about 25% of the cardiac blood flow, and any substance that reaches the systemic circulation will reach this organ. So these are considered to be frequent targets of toxicity. Renal function was evaluated by serum levels of urea, creatinine, and by histological analysis. The histopathological evaluation did not reveal alterations in this organ of any treated groups of sub-acute toxicity. Also, no tissue alterations found in the heart and brain of animals treated with 28 days *E. alsinoides* & *C. asiatica* extracts.

CONCLUSION: In this study, we had evaluated that *E. alsinoides* & *C. asiatica* extracts at different dosages cause no toxicity in signs in acute and sub-acute administration. Also, histopathology of kidney, liver, heart, and brain showed no alterations in tissues morphology. Since, the *E. alsinoides* & *C. asiatica* are already used in Indian traditional medicine as the neuroprotective agent and also found promising effects over inflammatory diseases, wound healing, immunomodulatory
activity and these data on the toxicity profile of *E. alsinoides* & *C. asiatica* will serve as a guide to future studies for theses both medicinally important plants.

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**CONFLICT OF INTEREST:** The authors declare that there is no conflict of interest.

**REFERENCES:**