



Received on 11 September 2019; received in revised form, 27 December 2019; accepted, 17 August 2020; published 01 September 2020

EFFECT OF CHROMIUM ON *IN-VITRO* GROWTH AND ANTIOXIDANT POTENTIAL OF *DIANTHUS CARYOPHYLLUS* L.

Smita Purohit ^{*1}, Swati Gupta ² and Rohit Jain ²

Department of Botany ¹, The IIS University, Jaipur - 302020, Rajasthan, India.

Department of Biosciences ², Manipal University, Jaipur - 303007, Rajasthan, India.

Keywords:

Carnation, Chromium,
Abiotic stress, Antioxidant activity

Correspondence to Author:

Dr. Smita Purohit

Associate Professor,
Department of Botany, The IIS
University, Jaipur - 302020,
Rajasthan, India.

E-mail: smita.purohit@iisuniv.ac.in

ABSTRACT: Catalase, guaiacol peroxidase and polyphenol oxidase are antioxidant enzymes that are important in the metabolism of reactive oxygen species and can be induced by environmental stresses including chromium (Cr), heavy metal toxic to living organisms. *In-vitro* regenerated shoots of carnation (*Dianthus caryophyllus* L.) were exposed to $K_2Cr_2O_7$, and the activities of these antioxidant enzymes were analyzed. A significant reduction in the POD activity was observed at 2nd week of culture, followed by a slight increase in the activity in the 4th week of culture. Maximum PPO activity was detected during the 3rd week of culture, which was comparable to the control cultures. A significant reduction in CAT activity was found in the 4th week. The chlorophyll content did not change significantly in the shoot buds developed up to 4th week of culture.

INTRODUCTION: The genus *Dianthus* belongs to family Caryophyllaceae. It comprises around 300 species, of which *D. caryophyllus* (commonly known as carnation) is the commercially most important cut flower of the world. Due to its excellent keeping quality, a wide range of forms, the ability to withstand long-distance transportation and remarkable ability to rehydrate after continuous shipping, it is preferred by growers to roses and chrysanthemums in several flower-exporting countries ¹. To fulfill the increasing demand and to establish an efficient reproducible system for transformation studies, numerous micropropagation protocols have been devised *via* organogenesis ²⁻⁶ and somatic embryogenesis ⁷⁻¹⁰ in *Dianthus* sp.

Enhanced *in-vitro* growth is a multi-factorial event. Induction by diverse plant hormones is generally considered to be the most important factor and is by far the most widely studied. Another factor, which has been increasingly recognized in recent times, is the addition of heavy metals in culture medium for improved plant regeneration. A review of some exemplary evidence-based research and approaches resulted in wider acceptance of several heavy metals as culture medium components. Also, natural or artificial selection systems can be set to select the heavy-metal tolerant genotypes or populations adapted to toxic concentrations of metal ions. Especially for urban areas, ornamentals can beautify the environment and also resolve heavy metal pollution at the same time. It will have great and practical significance to screen out remediation plants from ornamental resources ¹¹.

According to an elementary experiment from herbaceous ornamentals, *Calendula officinalis* ¹², *Coronopus didymus* ¹³, and *Malva rotundifolia* ¹⁴ had strong Cd tolerance.

<p>QUICK RESPONSE CODE</p> 	<p>DOI: 10.13040/IJPSR.0975-8232.11(9).4420-25</p> <hr/> <p>This article can be accessed online on www.ijpsr.com</p> <hr/> <p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.11(9).4420-25</p>
---	---

Plant tolerance to a particular metal is governed by an inter-related network of physiological and molecular mechanisms, to be understood essentially for developing plants suitable for phytoremediation of the contaminated sites. Plant capability for hyperaccumulation can be improved by genetic manipulations or the chelator-regulated strategies. The problem of low-biomass phytoremediators can be overcome by increasing plant yield and metal uptake by engineering common plants with hyperaccumulating genes^{15,16}.

Improvement of plants by genetic engineering, *i.e.*, by modifying characteristics like metal uptake, transport, and accumulation and plant's tolerance to metals, opens up new possibilities of phytoremediation¹⁷⁻¹⁹. Considerable research has been focused on assessing metal tolerance in plants using tissue culture techniques²⁰⁻²⁴. Tissue cultures are useful in developing stress-tolerant cell lines in a relatively short period. Thus, metal toxicity appears to be an area where emerging methods in plant tissue culture may complement conventional crop improvement programs by providing additional means of screening and/or selection for improved levels of tolerance.

Therefore, the aim of this work was to identify the tolerance levels or the effect of Cr on *in-vitro* regenerated plantlets and to study the activity of certain antioxidant enzymes of *D. caryophyllus*.

MATERIALS AND METHODS:

Plant Material and Explant Preparation: Seeds of *D. caryophyllus* were surface sterilized with 0.1 % (w/v) mercuric chloride for 3 minutes, followed by four washes with sterile distilled water. Nodal segments of 0.5mm were aseptically dissected out from 21 days old aseptically raised seedlings on half-strength MS²⁵ medium. MS medium supplemented with 3% (w/v) sucrose and 1% (w/v) agar with pH 5.8±0.01 was used throughout the course of the study. Nodal segments of 0.5mm were aseptically dissected out from 21 days old seedlings and cultured on solid MS medium containing 2.2 µM BAP + 2.7 µM NAA supplemented with 3% (w/v) sucrose and 1% (w/v) agar with pH 5.8±0.01 for shoot induction. The cultures were incubated at 16 h photoperiod at 26±1 °C. The explants were cultured on shoot induction medium with 0, 2, 4, 8, 10, 25 µM of

K₂Cr₂O₇ for four weeks separately. Shoots regenerated from each treatment were collected to study the morphogenic and physiological responses. Pairs of leaves were used and cut into small pieces (3-4mm). The distal half of the leaf was removed, and only the proximal part was cultured into liquid MS medium containing 5 µM 2, 4-D supplemented with 3% (w/v) sucrose, and pH set at 5.8±0.01. The liquid culture was maintained in the dark for 28 days on a rotary shaker (120 rpm) in the growth chamber at 26±1 °C.

Enzymatic Activity: *In-vitro* shoots developed on 7, 14, 21, and 28th day were used for enzyme extraction. For total enzyme extracts, 1g tissue was homogenized with a mortar and pestle in 2ml of ice-cold 50 mM potassium phosphate buffer (pH 7.0) containing 1 mM EDTA, 1 mM sodium ascorbate and 0.5 M NaCl. After centrifugation (20,000 × g, 20 min) the supernatant was used for the determination of catalase, peroxidase and polyphenol oxidase activity.

Catalase Assay: Catalase (CAT, EC 1. 11. 1. 6) activity was assayed from the rate of H₂O₂ decomposition as measured by the decrease of absorbance at 240 nm, following the procedure of Aebi²⁶. The reaction mixture contained 0.0625 cm³ of 50mM sodium phosphate buffer (pH 7.0), 0.075 cm³ H₂O₂ (3%) and 0.01 cm³ enzyme extract.

Polyphenol Oxidase: Polyphenol oxidase (PPO, EC 1. 14. 18. 1) activity was determined according to the method of Raymond, Rakariyatham²⁷ at 40 °C. The reaction mixture contained 2.5cm³ 0.2M phosphate buffer (pH 6.8), 0.2cm³ 20mM pyrogallol and 0.02cm³ enzyme extract. The increase in absorbance was recorded at 430 nm.

Peroxidase Assay: Peroxidase (POD, EC 1. 11. 1. 7) activity was assayed by Guaiacol-H₂O₂ method²⁸. The reaction mixture was made by mixing 0.2 ml enzyme extract, 1 ml of 1% Guaiacol, 0.2 ml of 50 mM H₂O₂. Peroxidase activity was determined spectrophotometrically by monitoring the formation of tetraguaiacol at 470 nm after every 15 seconds. One unit of peroxidase activity corresponds to the levels of enzyme activity were expressed as moles of H₂O₂ destroyed/min/mg protein.

Chlorophyll (Chl) Estimation: Chlorophyll was estimated in the periods for 0, 7, 14, 21, and 28 days of treatment by the method of Arnon²⁹. 1g regenerated shoots (4 weeks old) were chilled in deep freeze (-20 °C) and then homogenized in 80 % acetone, centrifuged at 10,000 rpm for 10 min. The supernatant obtained was used for chlorophyll estimation. The absorbance of the supernatant was read at 645 nm and 665 nm against 80% acetone solvent. The amount of chlorophyll was calculated according to the following equation and the total amount of expressed as mg chlorophyll/g fresh weight:

$$\begin{aligned} \text{Chl a} &= 12.7 (A_{663}) - 2.69 (A_{645}) \times \frac{V}{1000 \times W} \\ \text{Chl b} &= 22.9 (A_{645}) - 4.68 (A_{663}) \times \frac{V}{1000 \times W} \\ \text{Total Chl} &= \text{Chl a} + \text{Chl b} \end{aligned}$$

Where, A = absorbance at a specific wavelength

W = weight of tissue taken

V = volume taken for Chl estimation

Rooting and Acclimatization: Shoots upto 2 cm and more in length were excised and transferred on to rooting medium consisting of full-strength MS medium supplemented with 0.5mg/l IBA. Plantlets with a well-developed shoot and root systems were carefully taken out and washed with tap water to remove agar. These plantlets were then transferred to earthen pots containing garden soil and organic manure (1:1). Humidity was maintained initially by covering the pots with polythene bags. The experiment was repeated three times.

Statistical Analysis: Three repetitions per treatment were made, and enzyme activities were

assayed three times. The mean of all repetitions and standard deviation of each observation from the mean was calculated from the data obtained.

The cultures were examined after five weeks. Data were subjected to one-way analysis of variance (ANOVA) by Fischer's least significant difference (P = 0.05).

RESULTS: Nodal explants were cultured on MS medium supplemented with 2.2 μM BAP + 2.7 μM NAA and different concentrations of K₂Cr₂O₇ (0, 2, 4, 8, 10, 25μM). There was an obvious difference in the shoots between the treated and the untreated (control) cultures. Incorporation of K₂Cr₂O₇ in the medium induced positive response. The explants showed swelling during the first week of culture. The initiation of shoot bud induction occurred at the end of the second week of the culture. It was observed that the number of shoots increased with the increase in the concentration of K₂Cr₂O₇ from 2 to 10μM beyond which at 25μM, the number of shoots did not increase in **Table 1**. Hyperhydric condition of shoots was not observed in cultures grown in medium incorporated with K₂Cr₂O₇ **Fig. 1**, which is a major problem in *Dianthus* cultures.

A significant reduction in the POD activity was observed at 2nd week of culture, followed by slight increase in the activity in the 4th week of culture. Maximum PPO activity was detected during the 3rd week of culture, which was comparable to the control cultures. A significant reduction in CAT activity was found in the 4th week. The chlorophyll content did not change significantly in the shoot buds developed up to 4th week of culture.

TABLE 1: NUMBER OF SHOOTS INDUCED FROM NODAL SEGMENTS OF *DIANTHUS CARYOPHYLLUS* CULTURED ON MS MEDIUM SUPPLEMENTED WITH 2.2 μM BAP + 2.7 μM NAA AND DIFFERENT CONCENTRATIONS OF K₂Cr₂O₇. (CULTURE PERIOD: 6 WEEKS)

K ₂ Cr ₂ O ₇ in Primary Culture (μM)	% Response	No. of shoot buds/explants Mean ± S.D.	K ₂ Cr ₂ O ₇ in second stage subculture (μM)	% response	No. of shoots proliferated (Mean ± S.D.)
0 ^c	46	4.0 ± 0.5	0	52	9.5 ± 0.5
2.0	53	5.2 ± 0.7	2.0	56	8.0 ± 0.5
			0	73	7.4 ± 0.7
4.0	80	6.2 ± 1.4	4.0	84	6.9 ± 1.4
			0	89	5.1 ± 0.5
8.0	66	8.6 ± 0.8	8.0	58	10.7 ± 0.6
			0	67	13.1 ± 1.1
10.0	60	8.8 ± 0.7	10.0	62	12.4 ± 0.5
			0	74	12.8 ± 0.7
25.0	40	3.1 ± 0.7	25.0	57	3.7 ± 0.4
			0	78	4.4 ± 0.8

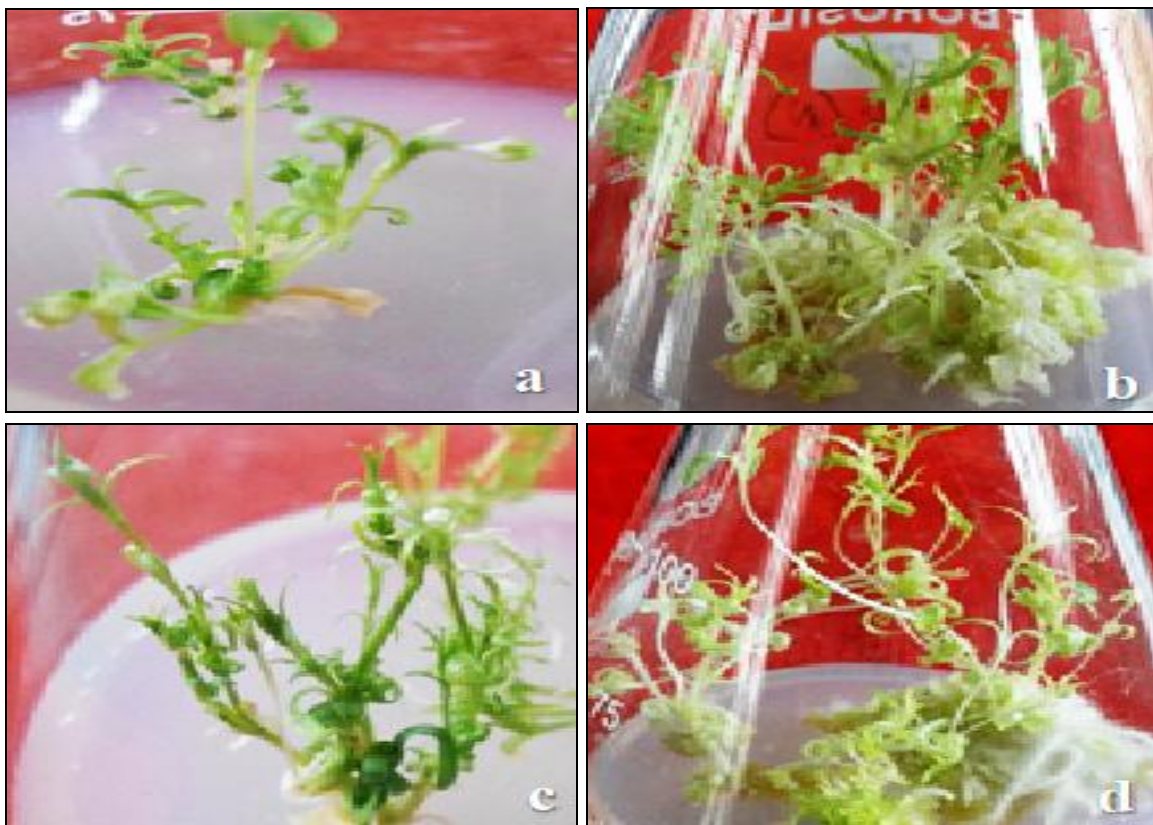


FIG. 1: SHOOT BUDS INDUCED FROM NODAL EXPLANTS OF *D. CARYOPHYLLUS* ON MEDIUM SUPPLEMENTED WITH 2.2μM BAP + 2.7μM NAA AND (A) WITHOUT $K_2Cr_2O_7$, (B) WITH 10μM $K_2Cr_2O_7$, (C) PROLIFERATED ON WITHOUT $K_2Cr_2O_7$ MEDIUM INDUCED ON SIMILAR MEDIUM AND (D) PROLIFERATED ON 10μM $K_2Cr_2O_7$ INDUCED ON SIMILAR MEDIUM

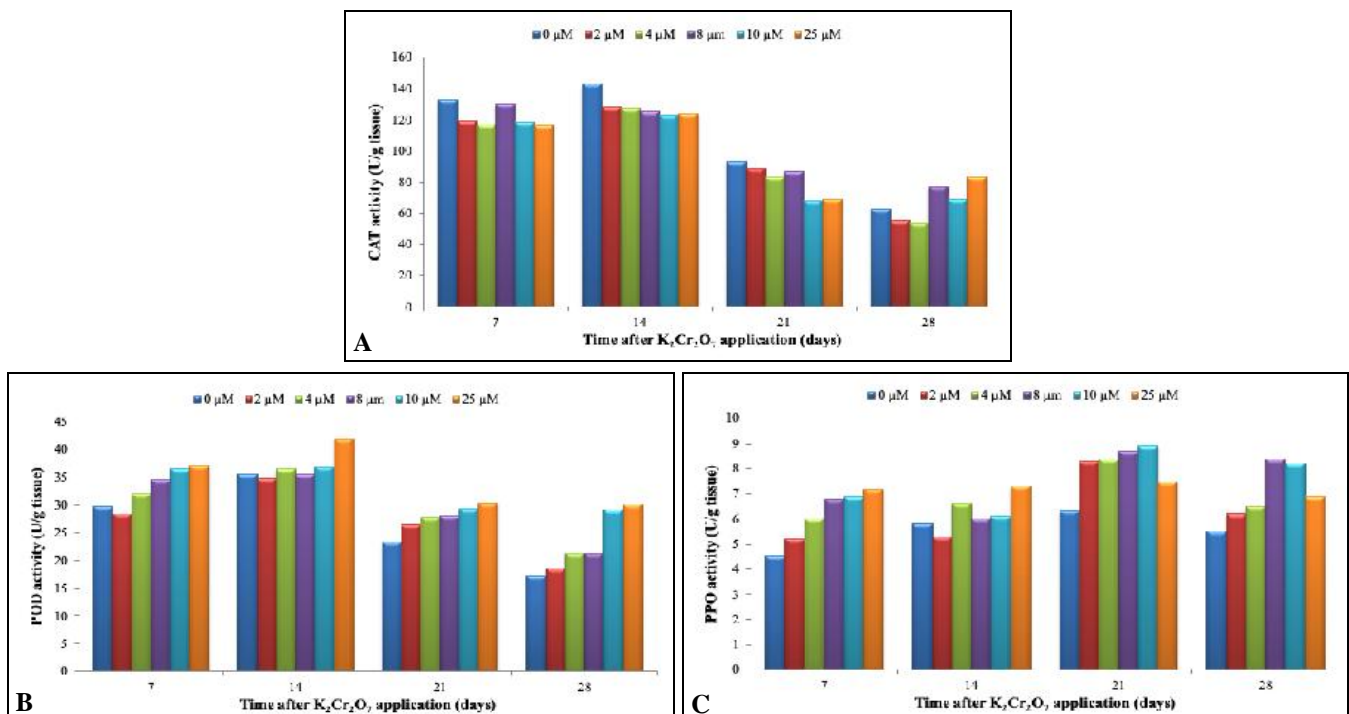


FIG. 2: ENZYME ACTIVITY DURING SHOOT BUD INDUCTION FROM NODAL SEGMENTS TAKEN FROM ASEPTICALLY RAISED MATURE PLANTS OF *DIANTHUS CARYOPHYLLUS* CULTURED ON MS MEDIUM SUPPLEMENTED WITH BAP (0.5 mg l⁻¹) + NAA (0.5 mg l⁻¹) + K₂Cr₂O₇ (0-25 μM) (a) CAT ACTIVITY, (B) POD ACTIVITY AND (C) PPO ACTIVITY

DISCUSSION: The tissue culture studies can augment the study of uptake behavior of nutrient and non-nutrient elements by providing intact plants for the quantitative estimate of the potential

of heavy metal tolerance and also to study certain kinetics involved in the root absorption and subsequent transfer to shoots while maintaining the physiological and structural integrity of the system. The present investigations have shown the tolerance levels of *in-vitro* regenerated *D. caryophyllus* plantlets to increasing levels of potassium dichromate.

The most widespread visual evidence of metal toxicity is a reduction in plant growth as metal toxicity increases. However, as different metals have different sites of action within the plant, the overall visual toxic response differs between metals. Chromium compounds are highly toxic to plants and are detrimental to their growth and development. Although some crops are not affected by low Cr concentration³⁰, Cr is toxic to most of the higher plants at 100 $\mu\text{M.kg}^{-1}$ dry weight³¹. In the present study, the highest level non-toxic to carnation plants was found to be 10 μM . Hence the minimum level of intolerance incarnation was found to be 25 μM *in-vitro*.

Chromium is a non-essential element and considered toxic to plants at very high concentrations³¹. The range of high Cr concentrations in plant tissues before toxicity symptoms was observed was from about 5 ppm for barley, corn, oats, and citrus to 175 ppm for tobacco³². In the present study, *Dianthus* cultures could grow at minimal levels of chromium, and increased amounts too led to regeneration but with pale-colored shoots. The excess of metals has been reported to have deleterious effects on the content and functionality of the photosynthetic pigments³³. This can be caused by the inhibition of the pigment synthesis, the formation of metal-substituted chlorophylls of reduced functionality, or the direct oxidative damage of the pigments³⁴. Several authors have reported damages on the reaction centers or the peripheral antennae complexes of PSII in response to high concentrations of metals³⁵.

Tissue culture responses are greatly influenced by three main factors *viz.* whole plant physiology of donor, *in-vitro* manipulation, and *in vitro* stress physiology. *In-vitro* cultures are also good models for studying the effect of different metals on the oxidative stress and antioxidants that follow. Under Cr heavy metal stress, the overall antioxidant

activity was found maximum at the highest concentration (25 μM). The activity of CAT, POD, and PPO increased significantly in cultures exposed to Cr supplemented media. The function of these enzymes is to scavenge the toxic radicals produced during oxidative stress and thus help plants to survive through such conditions. POD induction is a general response of vascular plants to the uptake of a toxic amount of metals³⁶. Also, PPO has been reported to function in oxygen scavenging by oxidation of phenols³⁷.

CONCLUSION: Although this study did not take into consideration the whole plant, the observations made herewith *in-vitro* cultures may supplement the known facts and can be beneficial for the production of metal tolerant carnation plants.

ACKNOWLEDGEMENT: Authors are grateful to The IIS University for providing all the laboratory facilities required to conduct this work. Financial assistance in the form of Enhanced Seed Grant by Manipal University Jaipur is also highly acknowledged.

CONFLICTS OF INTEREST: Authors have no conflict of interest.

REFERENCES:

1. Kanwar J and Kumar S: Influence of growth regulators and explants on shoot regeneration in carnation. Horticultural Science 2010; 36: 140-46.
2. Jain A, Kantia A and Kothari S: De novo differentiation of shoot buds from leaf-callus of *Dianthus caryophyllus* L. and control of hyperhydricity. Scientia Horticulturae 2001; 87: 319-26.
3. Bora A, Talukdar M and Hazarika B: Direct organogenesis in *Dianthus caryophyllus* Linn. Journal of Ornamental Horticulture 2006; 9: 164-68.
4. Ahmadian M, Babaei A, Shokri S and Hessami S: Micropropagation of carnation (*Dianthus caryophyllus* L.) in liquid medium by temporary immersion bioreactor in comparison with solid culture. Journal of Genetic Engineering and Biotechnology 2017; 15: 309-15.
5. Thakur K and Kanwar K: *In-vitro* plant regeneration by organogenesis from leaf callus of Carnation, *Dianthus caryophyllus* L. cv. 'Master'. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences 2018; 88: 1147-55.
6. Jorapur S, Jogdande N and Dhumale D: Petal callus mediated de novo regeneration of shoots in carnation (*Dianthus caryophyllus* L.). The Pharma Innovation Journal 2018; 7: 218-22.
7. Pareek A and Kothari S: Direct somatic embryogenesis and plant regeneration from leaf cultures of ornamental species of *Dianthus*. Scientia Horticulturae 2003. 98: 449-59.

8. Aalifar M, Arab M, Aliniaiefard S, Dianati S, Mehrjerdi M, Limpens E and Serek M: Embryogenesis efficiency and genetic stability of *Dianthus caryophyllus* embryos in response to different light spectra and plant growth regulators. *Plant Cell, Tissue and Organ Culture* 2019; 139: 479-92.
9. Lantcheva A: Somatic embryogenesis and genetic transformation of Carnation (*Dianthus caryophyllus* L.). Springer: Place 2016: 107-20.
10. Kumar S, Kumari R, Baheti T, Thakur M and Ghani M: Plant regeneration from axillary bud, callus and somatic embryo in carnation (*Dianthus caryophyllus*) and assessment of genetic fidelity using RAPD-PCR analysis. *Indian Journal of Agricultural Sciences* 2016; 86: 1482-88.
11. Liu J, Zhou Q, Sun T, Ma L and Wang S: Identification and chemical enhancement of two ornamental plants for phytoremediation. *Bulletin of Environmental Contamination and Toxicology* 2008; 80: 260-65.
12. Saffari V and Saffari M: Effects of EDTA, citric acid, and tartaric acid application on growth, phytoremediation potential, and antioxidant response of *Calendula officinalis* L. in a cadmium-spiked calcareous soil. *International Journal of Phytoremediation* 2020; 1-11.
13. Sidhu G, Singh H, Batish D and Kohli R: Tolerance and hyperaccumulation of cadmium by a wild, unpalatable herb *Coronopus didymus* (L.) Sm. (Brassicaceae). *Ecotoxicology and Environmental Safety* 2017; 135: 209-15.
14. Wu M, Luo Q, Liu S, Zhao Y, Long Y and Pan Y: Screening ornamental plants to identify potential Cd hyperaccumulators for bioremediation. *Ecotoxicology and Environmental Safety* 2018; 162: 35-41.
15. Clemens S: How metal hyperaccumulating plants can advance Zn biofortification. *Plant and Soil* 2017; 411: 111-20.
16. Wani W, Masoodi K, Zaid A, Wani S, Shah F, Meena V, Wani S and Mosa K: Engineering plants for heavy metal stress tolerance. *Rendiconti Lincei. Scienze Fisiche e Naturali* 2018; 29: 709-23.
17. Kumari P, Rastogi A, Shukla A, Srivastava S and Yadav S: Prospects of genetic engineering utilizing potential genes for regulating arsenic accumulation in plants. *Chemosphere* 2018; 211: 397-406.
18. Liu H, Zhao H, Wu L and Xu W: A genetic transformation method for cadmium hyperaccumulator *Sedum plumbizincicola* and non-hyperaccumulating ecotype of *Sedum alfredii*. *Frontiers in Plant Science* 2017; 8: 1047.
19. Shaheen S, Mahmood Q, Asif M and Ahmad R: Genetic control of metal sequestration in hyper-accumulator plants. Springer: Place 2017: 343-68.
20. Muszyńska E, Hanus-Fajerska E and Koźmińska A: Differential tolerance to lead and cadmium of micropropagated *Gypsophila fastigiata* ecotype. *Water, Air, & Soil Pollution* 2018; 229: 42.
21. Wiszniewska A, Muszyńska E, Hanus-Fajerska E, Smoleń S, Dziurka M and Dziurka K: Organic amendments enhance Pb tolerance and accumulation during micropropagation of *Daphne jasminea*. *Environmental Science and Pollution Research* 2017; 24: 2421-32.
22. Vuksanović V, Kovačević B, Katanić M, Orlović S and Miladinović D: *In-vitro* evaluation of copper tolerance and accumulation in *Populus nigra*. *Archives of Biological Sciences* 2017; 69: 679-87.
23. Sychta K, Słomka A, Sliwiska E, Migdalek G and Kuta E: From cells highly tolerant to Zn and Pb to fully fertile plants—Selection of tolerant lines with *in-vitro* culture. *Plant Physiology and Biochemistry* 2020; 146: 231-37.
24. Soleimani S, Bernard F, Amini M and Khavari-nezhad R: Cadmium accumulation and alkaloid production of *Narcissus tazetta* plants grown under *in-vitro* condition with cadmium stress. *Plant Physiology Reports* 2019. 1-7.
25. Murashige T and Skoog F: A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum* 1962; 15: 473-97.
26. Aebi H: Catalase. *Methods of Enzymatic Analysis* 1974: 673-84.
27. Raymond J, Rakariyatham N and Azanza J: Purification and some properties of polyphenoloxidase from sunflower seeds. *Phytochemistry* 1993. 34: 927-31.
28. Racusen D and Foote M: Protein synthesis in dark-grown bean leaves. *Canadian Journal of Botany* 1965; 43: 817-24.
29. Arnon D: Copper enzymes in isolated chloroplasts. Polyphenoloxidase in *Beta vulgaris*. *Plant Physiology* 1949. 24: 1.
30. Huffman E and Allaway W: Growth of plants in solution culture containing low levels of chromium. *Plant Physiology* 1973; 52: 72-75.
31. Davies F, Puryear J, Newton R, Egilla J and Grossi JS: Mycorrhizal fungi increase chromium uptake by sunflower plants: influence on tissue mineral concentration, growth, and gas exchange. *Journal of Plant Nutrition* 2002; 25: 2389-2407.
32. Sinha P, B Dube and Chatterjee C: Amelioration of chromium phytotoxicity in spinach by withdrawal of chromium or iron application through different modes. *Plant Science* 2005. 169: 641-46.
33. Mathur S, Kalaji H and Jajoo A: Investigation of deleterious effects of chromium phytotoxicity and photosynthesis in wheat plant. *Photosynthetica* 2016. 54: 185-92.
34. Rai R, Agrawal M and Agrawal S: Impact of heavy metals on physiological processes of plants: with special reference to photosynthetic system. Springer: Place 2016: 127-40.
35. Sharma A, Kumar V, Shahzad B, Ramakrishnan M, Sidhu G, Bali A, Handa N, Kapoor D, Yadav P and Khanna K: Photosynthetic response of plants under different abiotic stresses: a review. *Journal of Plant Growth Regulation* 2019; 1-23.
36. Bucker-Neto L, Paiva A, Machado R, Arenhart R and Margis-Pinheiro M: Interactions between plant hormones and heavy metals responses. *Genetics and Molecular Biology* 2017; 40: 373-86.
37. Jaiswal A and Srivastava J: Changes in reactive oxygen scavenging systems and protein profiles in maize roots in response to nitric oxide under water logging stress. *Indian Journal of Biochemistry & Biophysics* 2018; 55: 26-33.

How to cite this article:

Purohit S, Gupta S and Jain R: Effect of chromium on *in-vitro* growth and antioxidant potential of *Dianthus caryophyllus* L. *Int J Pharm Sci & Res* 2020; 11(9): 4420-25. doi: 10.13040/IJPSR.0975-8232.11(9).4420-25.

All © 2013 are reserved by the International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **Android OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Play store)