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## ROLE OF TERPENOIDS AS HEPATOPROTECTIVE

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**ABSTRACT:** The liver is the most vital organ in our body, involved in several vital functions such as metabolism, secretion, storage as well as detoxification of several drugs and xenobiotic. Liver cell injury caused by various toxic chemicals like certain antibiotics, chemotherapeutic agents, carbon tetrachloride, thioacetamide, excessive alcohol consumption microbes and so, on. Due to liver disease worldwide, approximately 2 million deaths, 1million due to complications of cirrhosis, and 1 million due to viral hepatitis and hepatocellular carcinoma have been found every year. Now a day number of synthetic medicine available in the market for the treatment of liver disorders, but they have lots of side effects. So, the developed countries recently give their interest in herbal drugs. Many Ayurveda herbs, such as Andrographis, Punarnava, kokum, and soon have a long history of traditional uses in revitalizing the liver and treating liver dysfunction from the various literature survey this can be concluded that the plant containing a high amount of terpenoids possess good hepatoprotective effect. There are some plants enriched with a high amount of terpenoids such as Andrographolide, *Podophyllum hexanderum*, *Origanum vulgare*, and soon are very potential for hepatic disorders. The aim of this review is to enlighten the number of plants containing terpenoids and terpenoids for their hepatoprotective activity.

**INTRODUCTION:** The liver is the largest organ in the body, situated in the right of Hypochondrium, approximate weight 1400-1600 gm of the males and 1200-1400 gm of the females. It is located mainly in the upper right portion of the abdomen, beneath the diaphragm and above the stomach. The liver performs multifold functions such as metabolism, excretion of bile, manufacture of plasma protein like albumin, fibrinogen, vitamins (A, D, and B12), iron, and detoxification of toxic substances such as alcohol and drugs<sup>1, 2</sup>. Recently large number of people affected in liver disorders, about 20,000 deaths are found every year due to liver disorders.

Hepatocellular carcinoma is one of the ten most common tumors in the world 2, 50,000 new cases each year. The main causes of liver diseases are excessive drug therapy, environmental pollution, and alcoholic intoxication. The liver diseases include liver cirrhosis (cell destruction and increase in fibrous tissue), inflammatory diseases, and non-inflammatory diseases. There is some 200 million chronic carriers of the hepatitis B virus, of which 40% are accepted eventually to die of hepatocellular carcinoma and 15% of cirrhosis<sup>3</sup>.

**The Various Types of Liver Disease are given below:**

- Alcoholic steatosis (Fatty liver)
- Cirrhosis
- Hepatocellular carcinoma (HCC)
- Viral hepatitis – Hepatitis A, B, C, D, E
- Jaundice
- Hepatic failure
- Cholangitis

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- Hepatic tuberculosis
- Echinococcosis.
- Cholelithiasis (gallstones)
- Cholecystitis

**1. Alcoholic Steatosis:** Alcoholic steatosis is a fatty liver disease, and it is also known as hepatic steatosis. It is a condition of excess fat in the liver<sup>4-5</sup>. Hepatic steatosis is two types shown in **Fig. 1**.

- Non-alcoholic fatty liver disease (NAFLD)
- Alcoholic liver disease<sup>6</sup>.

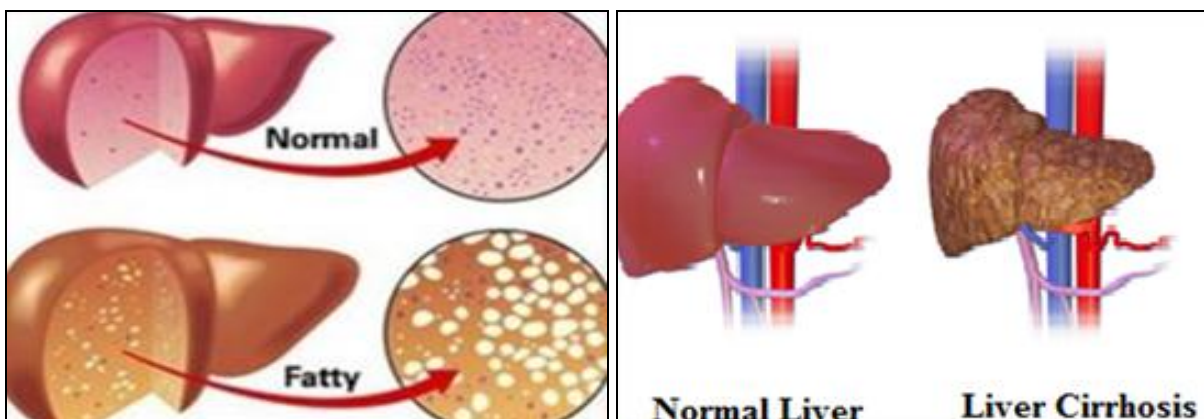
**2. Cirrhosis:** Cirrhosis is a condition of liver damage, and it is the last stage of scarring of the liver caused many diseases such as hepatitis, chronic alcoholism, hepatitis B, hepatitis C. Cirrhosis occur due to take alcohol more than 3 times (20-40%) per day and liver doesn't properly work. It is also known as hepatic or liver cirrhosis. The basic Symptoms for liver cirrhosis generally found are: weakness, nausea, vomiting, swelling in

the lower legs and fluid buildup in the abdomen, unconsciousness, yellow skin and so on<sup>7</sup> the liver cirrhosis shown in the **Fig. 1**.

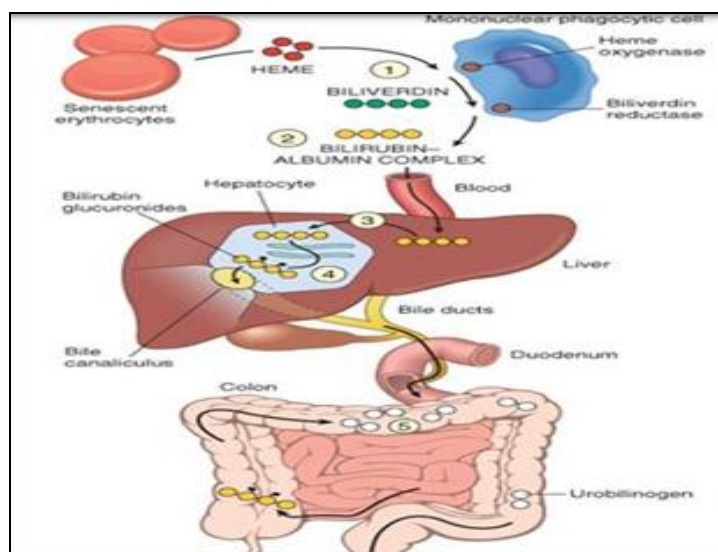
**3. Jaundice:** Jaundice is a basic very serious liver problem known as icterus. The normal range of bilirubin is 5-19  $\mu$  mole/liter, and in case of jaundice level of bilirubin becomes high. In the time of jaundice skin, sclera and mucous membranes of the skin become faded yellow color due to a raised plasma bilirubin. From the various literature surveys, it was found that the basic region of jaundice is imbalance production and clearance of bilirubin<sup>8</sup>, and it is shown in **Fig. 2**.

**Jaundice can be classified as:**

1. Pre-hepatic (resulting from excessive hemolysis).
2. Hepatic (due to congenital or acquired liver disorders causing impaired intra-hepatic bilirubin metabolism).



**FIG. 1: LIVER DISORDER A. FATTY LIVER B. LIVER CIRRHOSIS**

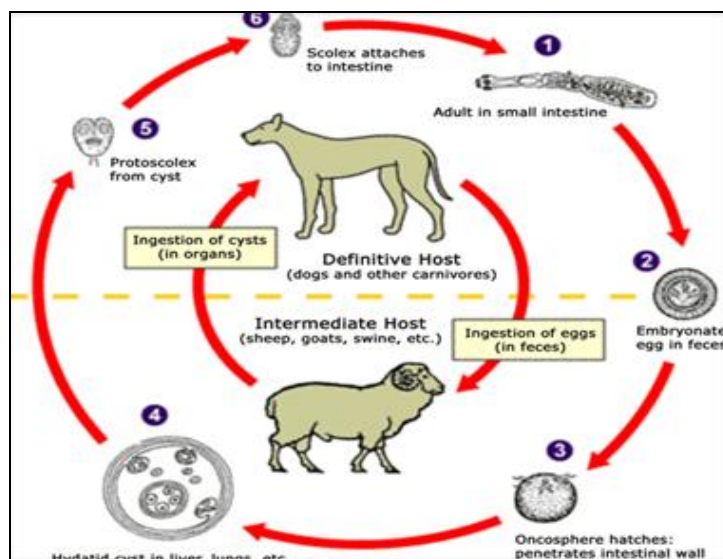


**FIG. 2: THE PRODUCTION AND METABOLISM OF BILIRUBIN ARE SHOWN IN FIGURE 1**

**4. Post-hepatic / Cholestatic:**<sup>9</sup>

**Echinococcosis:** It is a parasitic tapeworm infectious disease that affects the lungs, liver, brain, etc.<sup>11</sup>. It is spread contaminated animal faeces with

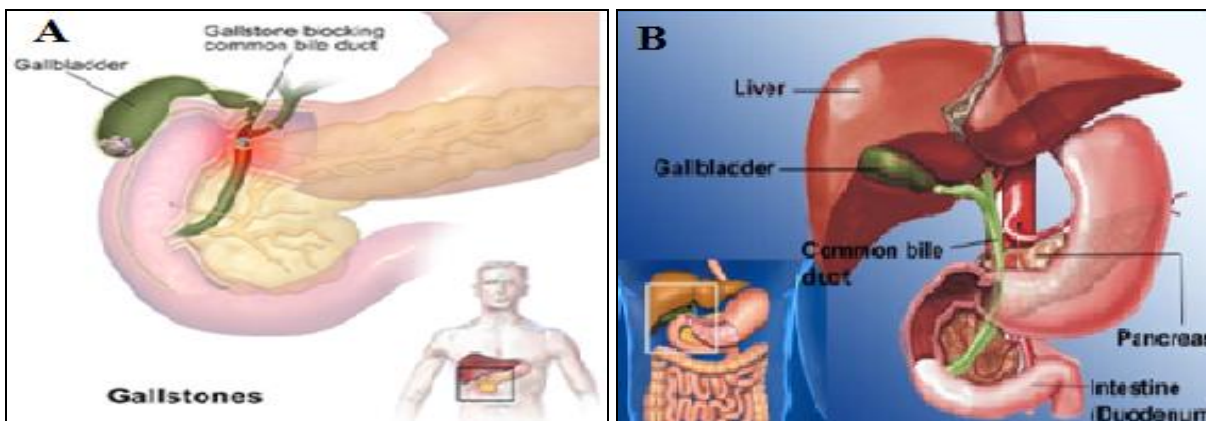
tapeworm eggs, through the contaminated food and water. And the spreading process of the disease is known as hydatid, hydatidosis<sup>12</sup>. The life cycle of echinococcosis is shown in **Fig. 3**.



**FIG. 3: ECHINOCOCCOSIS LIFE CYCLE**

**5. Cholangitis:** Cholangitis is an inflammation of the bile duct usually caused by bacteria upside from its junction<sup>13</sup> in case of ascending cholangitis the bile duct are infected by bacteria<sup>14</sup>. Symptoms of cholangitis are yellow discoloration of the skin or whites of the eyes, abdominal pain, confusion, low blood pressure and so on<sup>15</sup> and it is shown in **Fig. 4**.

**6. Cholelithiasis:** Cholelithiasis refers to the formation of gallstones in the gallbladder<sup>16</sup>. Gallstones are mainly composed of bilirubin, calcium salts, cholesterol, and small amounts of protein and other materials<sup>17</sup>. The Gallstone blocks the biliary fluid, as a symptom, a cramp-like pain in the upper part of the abdomen is found called gallbladder attack<sup>18</sup>, and it is shown in **Fig. 4**.



**FIG. 4A: ACUTE CHOLANGITIS B. FORMATION OF GALLSTONE IN GALLBLADDER**

**Role of Natural Products as Hepatoprotective Drug:** Till now the synthetic drug play the important role in the treatment of hepatotoxicity. From the clinical report and the various adverse effects such as of these drugs such as excessive bleeding, hemorrhage and difficulty breathing, dry mouth, dementia, and so on, since the present day,

the developing countries people gradually move the herbal drug for the treatment of the toxicity<sup>20</sup>. Herbs/medicinal plant/homemade remedies are less expensive than synthetic drugs, and majority peoples in rural/backward areas have blind faith in them. They are right because they can treat any disease by using them without any lethal side



effects<sup>21</sup>. Although herbal medicines are less potent in comparison to synthetic drugs in some cases, but these are still considered less toxic or having less side effects in contrast to synthetic drugs<sup>22</sup>. The ultimate norm for any medicine (human made or natural) is their non-toxicity, effectiveness, specificity, stability, and potency<sup>23-26</sup>. For the various literature surveys, it is clear that the terpenoids are the potent bioactive compounds for the treatment of ulcers. Terpenoids are the naturally occurring hydrocarbon compounds, and they are oxygenated derivatives like alcohols, aldehydes, ketones, and the terpenoids are called isoprenoids. Terpenoids are the derivative of polymers of isoprene unit (C<sub>5</sub>H<sub>8</sub>)<sup>27</sup>. Terpenoids is found in all volatile oils, resins combination of plant or animal origin. Terpenoids have the prevention and curative poverty for the of several diseases such as cancer, antimicrobial, antifungal, anti-parasitic, antiviral, anti-allergenic, hepatoprotective, antispasmodic, antihyperglycemic, anti-inflammatory, immune-modulatory properties and so, on<sup>28-31</sup> the classification of terpenoids is given in **Table 1**. Various plants and polyherbal formulations are used in the treatment of liver disease. Some herbal plants are providing protection from liver damage caused by toxic

chemicals and screening models of the drugs, oxidative mechanisms and so on. Screening plants for anti-hepatitis activity such as *P. Kurroa*, *Glycyrrhiza glabra*, *A. Paniculata* are likely to be active against in hepatitis virus and liver toxicity.

A combination of different herbal extracts is likely to provide desired activities to cure severe liver disease. To the importance of their use, we reviewed some popular herbal plants having hepatoprotective potential. The development of such medicines with the standard of safety and efficacy can revitalize treatment of liver disorders, and hepatoprotective activity of the medicinal plants is shown in **Table 2**.

**TABLE 1: CLASSIFICATION OF TERPENOIDS**

Name of Terpenoids	Unit of Terpenoids	Examples of Terpenoids
Hemiterpenoids	(C <sub>5</sub> H <sub>8</sub> )	Prenol
Monoterpenoids	(C <sub>10</sub> H <sub>16</sub> )	Geraniol, limonene
Sesquiterpenoids	(C <sub>15</sub> H <sub>24</sub> )	A-Bisabolbol, Dehydrocostuslactone
Diterpenoids	(C <sub>20</sub> H <sub>32</sub> )	Andrographis paniculata, Oridonin
Sesterterpenoids	(C <sub>25</sub> H <sub>40</sub> )	Geranylarnesol
Triterpenoids	(C <sub>30</sub> H <sub>48</sub> )	Actein, Ginsenoside
Tetraterpenoids (carotenoids)	(C <sub>40</sub> H <sub>64</sub> )	B-Carotene, Lycopene
Polyterpenoids	(C <sub>5</sub> H <sub>8</sub> ) <sub>n</sub>	Natural rubber

**TABLE 2: HEPATOPROTECTIVE ACTIVITY OF THE MEDICINAL PLANTS**

S. no.	Common Name	Botanical Name	Family	Model of Hepatoprotective Drug	Active Constituent	Plant Part used
1	Indian Rhododendron <sup>35</sup>	<i>Melastoma malabathricum L.</i>	(Melastomataceae)	Paracetamol-induced liver toxicity in rats.	Flavonoids, phenolic components	Leaves
2	Mimosa catechu, catechu, cachou, cutch tree, black cutch <sup>36</sup>	<i>Acacia catechu</i>	(Fabaceae)	Liver Damage Induced by Iron overload in mice	Saponins, tannins, flavonoids, phenols, alkaloidal	Heart wood
3	Kalmegh <sup>37</sup>	<i>Andrographis paniculata</i>	(Acanthaceae)	Against galactosamine or paracetamol induced hepatotoxicity in rats.	Terpenoids	Extract of the plant
4	Kutki <sup>38</sup>	<i>Picorrhiza kurroa</i>	(Scrophulariaceae)	Liver against CCl <sub>4</sub> intoxicated rats	Iridoid Glycosides	Extract of the plant
5	Sweet neem leaves, curry leaves <sup>39</sup>	<i>Murraya koenigii L.</i>	(Rutaceae)	CCl <sub>4</sub> treated hepatotoxic rats	Polyphenol, girinimbine	Leaf
6	Tulsi <sup>40-42</sup>	<i>Ocimum sanctum</i>	(Lamiaceae)	O.sanctum, against paracetamol, CCl <sub>4</sub> and lead induced liver damage	Phenolic components, anti-oxidant, oleanolic acid, urosolic acid.	Whole plant
7	Haridra, Haldi <sup>43-47</sup>	<i>Curcuma longa</i>	Zingiberaceae	hepatoprotective activity against CCl <sub>4</sub> and TAA induced toxicity	Diarylheptanoids, curcumin, zingiberene, germacrone	Different extracts of C. longa, rhizomes, stems, Roots
8	Punarnava <sup>48-52</sup>	<i>Boerhavia diffusa,</i>	(Nyctaginaceae)	induced by paracetamol and acetaminophen	Isoflavonoids, flavonoids, flavonoid glycosides, xanthene, purine nucleoside, lignans, steroids	Roots

9	Himalayan mayapple <sup>53</sup>	<i>Podophyllum hexandrum</i>	(Berberidaceae)	CCl <sub>4</sub> -induced hepatotoxicity in rats	Tannins, terpenoids, alkaloids, flavonoids, phenols, steroids	Rhizome
10	Milkvetch, goat's thorn, locoweed <sup>54</sup>	<i>Astragalus kahiricus</i>	(Fabaceae)	Ethanol-induced liver apoptosis in rats	flavonoids, phenolic compounds	Roots
11	Wild marjoram <sup>55</sup>	<i>Origanum vulgare</i>	(Lamiaceae)	Carbon tetrachloride-induced hepatotoxicity in rats	Terpenoids, tannin, phenolic compounds flavonoids, saponins	Leaves
12	Cup shaped sori, little cup, cyathea <sup>56</sup>	<i>Cyathea gigantea</i>	(Cyatheaceae)	Paracetamol induced hepatotoxicity in rats	Phenolic compounds, tannins & flavonoids	Leaves
13	Pineapple guava, guavas teen <sup>57</sup>	<i>Feijoa sellowiana</i>	(Myrtaceae)	3, 4-methylene dioxymethamphetamine (MDMA or ecstasy) induced liver damage	Polyphenols, carbohydrates, vitamin A	Fruits peel
14	Kokum <sup>58</sup>	<i>Garcinia Indica</i>	(Clusiaceae)	Ethanol-induced hepatotoxicity in rats	Xanthones, flavonoids, benzophenones, lactones & phenolic acids	Fruit rind
15	sacred fig, Bo-Tree, Pippal <sup>59</sup>	<i>Ficus religiosa</i>	(Moraceae)	Isoniazid rifampicin & paracetamol-induced hepatotoxicity	Flavonoids	Leaves
16	Sunset musk mallow, sunset hibiscus, hibiscus Manihot <sup>60</sup>	<i>Abelmoschus Manihot (L.) Medic</i>	(Malvaceae)	Carbon tetrachloride (CCl <sub>4</sub> ) induced hepatocyte damage	Total Flavonoids	Flowers
17	Gum Arabic tree, Babul, Kikar <sup>61</sup>	<i>Acacia nilotica Linn</i>	(Fabaceae)	Acetaminophen-induced hepatic damage in Wistar rats	Carbohydrate cardiac glycoside, saponin, tannins	Aerial parts
18	wild carrot, bird's nest, bishop's lace, Queen <sup>62</sup>	<i>Daucus carota</i>	(Apiaceae)	Thioacetamide induced Oxidative stress in rat liver	Monoterpenoids, flavonoids, quercetin, limonene	Seeds
19	Lelom leaves, Lelompata <sup>63</sup>	<i>Premna esculenta Roxb.</i>	(Verbenaceae)	CCl <sub>4</sub> -induced liver toxicity in rats.	Polyphenols, flavonoids	leaves
20	Veldt Grape or Devil's Backbone <sup>64</sup>	<i>Cissus quadrangularis</i>	(Vitaceae)	Rifampicin-induced hepatotoxicity in rats.	β-carotene	Stem
21	Oleander <sup>65</sup>	<i>Nerium oleander</i>	(Apocynaceae)	CCl <sub>4</sub> -induced hepatotoxicity in rats	Terpenoids, cardiac glycosides, tannin, flavonoids, saponins, phenolic	Flower
22	Rose mallow, bharadwaji, bankapas <sup>66</sup>	<i>Hibiscus vitifolius Linn.</i>	(Malvaceae)	Anti-tubercular drug-induced hepatotoxicity in rats	Flavonoids, phenolic compounds	Roots
23	Sweet neem leaves, curry leaves <sup>67</sup>	<i>Murraya koenigii L.</i>	(Rutaceae)	CCl <sub>4</sub> treated hepatotoxicity in rats	Polyphenol	Leaf
24	Yellow Berried Nightshade <sup>68</sup>	<i>Solanum xanthocarpum</i>	(Solanaceae)	CCl <sub>4</sub> -induced liver injury in rats.	Steroidal alkaloid, Solasonine, fatty & resinous substances	Fruits
25	Chiretta <sup>69</sup>	<i>Swertia chirayita</i>	(Gentianaceae)	Paracetamol induced hepatotoxicity in Swiss albino mice	Tannins, glycosides	Whole plant
26	Cilantro, Chinese parsley or dhania <sup>70</sup>	<i>Coriandrum sativum (Linn.)</i>	(Apiaceae)	carbon tetrachloride (CCl <sub>4</sub> ) induced hepatotoxicity	Alkaloids, phenolic compound, flavonoids,	Whole plant
27	Conkerberry or Bush Plum, Currant Bush <sup>71</sup>	<i>Carissa opaca</i>	(Apocynaceae)	CCl <sub>4</sub> -induced damage in rat	isoquercetin, quercetin Flavonoids, tannins, terpenoids, alkaloids, anthraquinones & cardiac glycosides	Leaves
28	GendaPhul (Marigold) <sup>72-73</sup>	<i>Tagetes erecta Linn</i>	(Compositae)	Targets erecta against Carbon tetrachloride-induced hepatic damage in rats	Quercetagenin, glucoside, quercetagenin, Phenolics, syringic acid, methyl-3,5-dihydroxy-4-Methoxy benzoate, quercetin, thienyl and ethyl gallate	Flower
29	Arar <sup>74</sup>	<i>Juniperus procera</i>	Cupressaceae	Against carbon tetrachloride	Diterpenes,	Bark and

30	Hadidi <sup>75-77</sup>	<i>Fagonia schweinfurthii</i>	(Zygophyllaceae)	induced liver injury. Carbon tetrachloride (CCl <sub>4</sub> ) induced hepatotoxicity in HepG2 cell line and rats.	sesquiterpenes Flavonol glycosides and terpenoid glycosides	leaves Whole plant
31	Almecega, breubranco <sup>78-80</sup>	<i>Protium heptaphyllum</i>	(Burseraceae)	Against acetaminophen-induced liver injury in mice	triterpenoids like oleanolic acid, ursolic acid, hederin, and glycyrrhizin	trunk wood resin
32	Bee Sting Bush 81	<i>Azima tetracantha</i>	(Salvadoraceae)	Paracetamol induced	Flavonoids, triterpenoids	Leaves
33	Bitter leaf <sup>82-84</sup>	<i>Vernonia amygdalina</i>	(Compositae)	Against acetaminophen-induced hepatotoxicity and oxidative stress in mice <i>in-vivo</i> .	Terpenoids	Leaf extract
34	Christmas tree <sup>85</sup>	<i>Alchornea cordifolia</i>	(Euphorbiaceae)	Carbon tetra chloride-induced hepatic damage in rats	Alkaloids, flavonoids, saponins and tannins	Leaves
35	Abuta, velvet leaf <sup>86</sup>	<i>Cissampelos pareira</i>	(Menispermaceae)	Carbon-tetra chloride induced hepatic damage	Alkaloids, essential oil, sterol, leno	Roots
36	Sweetgum <sup>87-88</sup>	<i>Liquidambar styraciflua</i> L.	(Altingiaceae)	CCl <sub>4</sub> -induced hepatic damage in rats,	Triterpenoids	Leaves
37	Celery <sup>89-91</sup>	<i>Apium graveolens</i> L.	(Umbelliferae)	against CCl <sub>4</sub> -induced hepatotoxicity in albino rats.	flavonoids, phenolic compounds, betacarotene, vitamin C, sesquiterpene	Seeds
38	Sickle bush, Bell mimosa, Chinese lantern tree <sup>92</sup>	<i>Dichrostachys cinerea</i>	(Fabaceae)	CCL <sub>4</sub> induced hepatotoxicity	Flavonoids, polyphenols, tannins	Leaves
39	Grains of Paradise, Melegueta pepper <sup>93</sup>	<i>Aframomum melegueta</i>	(Zingiberaceae)	Ethanol-induced Liver toxicity in male Wistar rats	Alkaloids, tannins, saponin, steroids, cardiac glycoside, flavonoid, terpenoids	Whole plant
40	Jiwanti <sup>94</sup>	<i>Leptadenia reticulata</i> (Retz.)	(Asclepiadaceae)	Carbon tetrachloride-induced hepatotoxicity in rats	Glycosides, flavonoids, tannins, phytosterols, phenolic	Stems

### Active Compound Terpenoids used for Hepatoprotective Activity:

**1. C-Methylflavone:** It is obtained from the dried herb *Boerhavia diffusa* belongs to the family (Nyctaginaceae). It contains the phenolic compounds, anthocyanin, antho-xanthin, flavonoids, and so on. Flavones have been shown to have a wide range of biological and pharmacological activities are *in-vitro* studies. Examples include are anti-bacterial, antifungal, anti-viral, antioxidant, anti-microbial, anti-cancer,

anti-diarrheal, hepato-protective, antiinflammatory, anti-allergic activities<sup>95-96</sup>. The structure of C- Methyl flavone is given in Fig. 1.

**2. Borhavine:** It is obtained from the *Boerhavia diffusa* belongs to the family (Nyctaginaceae). It is used in Ayurveda of anti-diabetic, diuretic, anti-fibrinolytic agent, anti-inflammation, jaundice, dyspepsia, and diuretic properties<sup>97</sup>. The structure of Borhavine is given in Fig. 2.

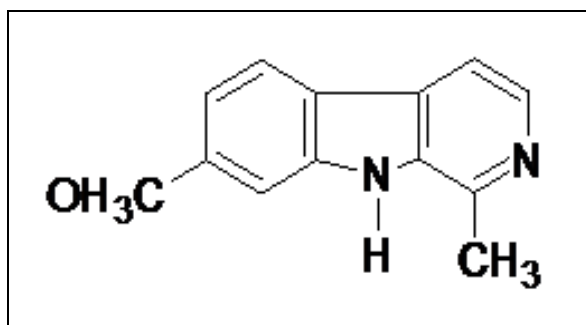


FIG. 1: CHEMICAL STRUCTURE OF BORHAVINE

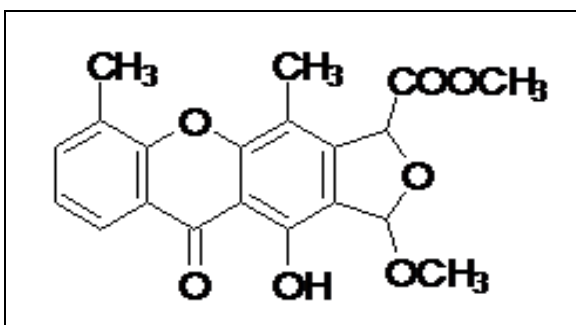


FIG. 2: CHEMICAL STRUCTURE OF C-METHYLFLAVONE

**3. Kutkoside:** The drug found dried roots and rhizomes of *Picrorhiza kurroa* Royle belongs to the family (Scrophulariaceae). Uses of these drugs are hepatoprotective activity, anti-inflammatory, bitter tonic, stomachic, purgatives preparations, jaundice, hepatitis, and picrorhiza as an antidote for dog-bite<sup>98</sup>. The structure of kutakoside is given in Fig. 3.

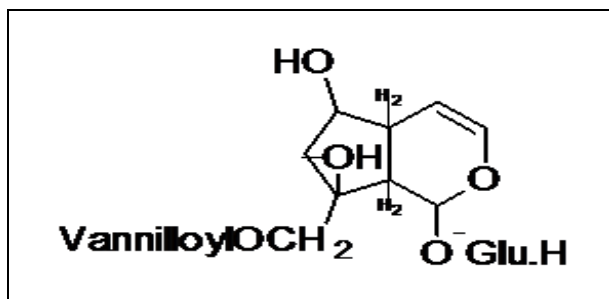


FIG. 3: CHEMICAL STRUCTURE OF KUTKOSIDE

**4. Kutkin:** It is obtained from the *Picrorhiza kurroa* belongs to the family (Plantaginaceae). It contains a bitter glycoside that contains two C-9 iridoid glycosides Picrosidei and kutakoside. It is used in the treatment of digestive problems, liver damage, cirrhosis, wound healing, vitiligo, and so on. The structure of kutkin is given in Fig. 4.

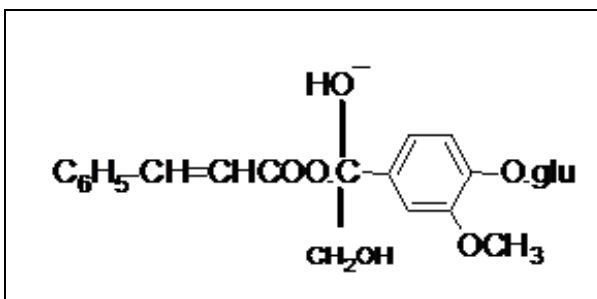


FIG. 4: CHEMICAL STRUCTURE OF KUTKIN

**5. Beta-Eudesmol:** It is the dried rhizome of *Atractylodes lanceae* that belongs to the family (Asteraceae)<sup>99</sup>.  $\beta$ . Eudesmol consists of monoterpenoids, phenolic acids, steroids, and the major constituents include are atractylodin (14%), Beta- eudesmol (6%), hinesol (1%). Other minor constituents include are atractyloside, atractyloquinone, atractylochromene<sup>100-101</sup>. Uses of the  $\beta$ . Eudesmol is hepatoprotective, night blindness, optic atrophy, to relieve stagnant liver, reducing stress and relieving depression. The structure of beta-eudesmol is given in Fig. 5.

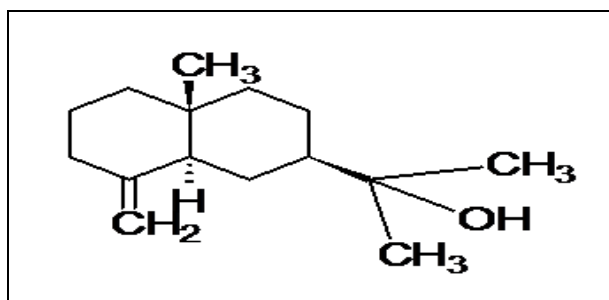


FIG. 5: CHEMICAL STRUCTURE BETA-EUDESMOL

**6. Andrographolide:** Andrographolide consists of leaves or entire aerial parts of *Andrographis paniculata* Nees. Belongs to the family (Acanthaceae). It consists a diterpene lactone, andrograpanin, flavonoids, and phenols. And the roots of kalmegh consist of monohydroxy-trimethyl flavone, panicolin, 5-hydroxy tetramethoxy flavone, and it is used in febrifuge, anthelmintic, astringent, anodyne, and it is useful in debility, cholera, piles, immune-modulator and jaundice<sup>102</sup>. The structure of andrographolide is given in Fig. 6.

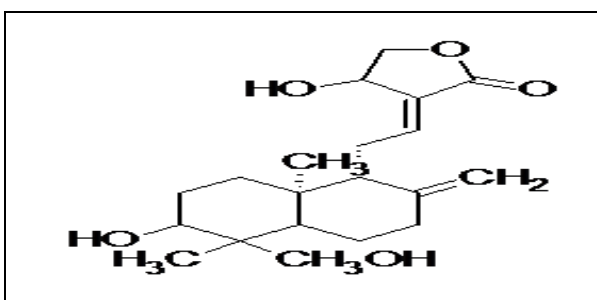


FIG. 6: CHEMICAL STRUCTURE OF ANDROGRAPHOLIDE

**7. Andrograpanin:** Andrograpanin is a minor compound of *Andrographis paniculata* belong to the family (Acanthaceae).

Andrograpanin consists of diterpene lactone, polyphenols, flavonoids, triacylglycerol's, lupeol and it is used in the anti-inflammatory, anti-infectious function, cold, fever, and diarrhea. The structure of andrograpanin is given in Fig. 7.

**8. *Lindera strychnifolia*:** It is obtained from the roots of *Lindera aggregate* belonging to the family (Lauraceae). *L. strychnifolia* consists of sesquiterpenes lactones, hydrocarbons, alkaloids, hydroxyi sogerma furen olide, lindenone, lauro-litsine. It is used in the hepatoprotective, anti-inflammatory, antioxidant, anti-cancer activity for lungs<sup>103</sup>. The structure of lindera strychnifolia is given in Fig. 8.

**9. Cucurbitacin:** It is obtained from the fruit of Cucurbita pepo belong to the family (Cucurbitaceae). It consists of triterpenes, alkaloids, flavonoids, palmitic, oleic acid and linoleic acid, 5-hydroxytryptophan, cucurbitacin. These are useful

in the anti-inflammatory, analgesic, urinary disorders, anti-ulcer, hepato-protection, anti-oxidant, antidiabetic<sup>104-105</sup>. The structure of cucurbitacin is given in Fig. 9.

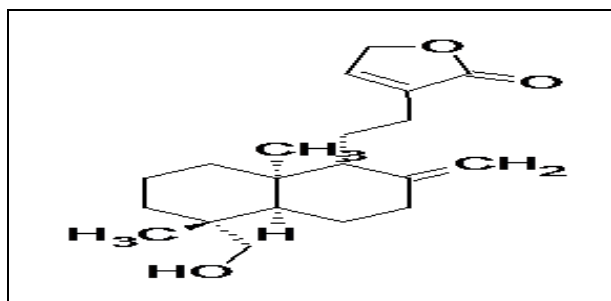


FIG. 7: CHEMICAL STRUCTURE OF ANDROGRAPANIN

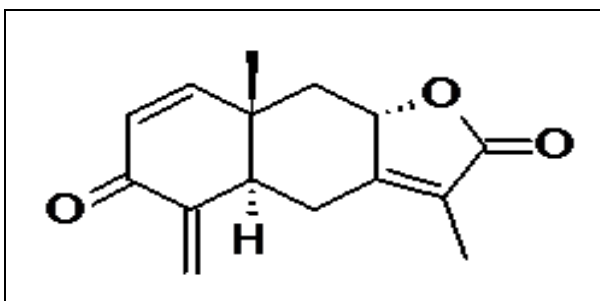


FIG. 8: CHEMICAL STRUCTURE OF LINDERA STRYCHNIFOLIA

**10. Secologanin:** It is obtained from the genus of flowering plant Ecballium elaterium belong to the family (Cucurbitaceae). Secologanin consists of triterpenoids glycosides, proteins, lipids, glycosyl cucurbitacin. It is used in the treatment of epilepsy, treatment of malaria, rhinosinusitis, prevention of CCl<sub>4</sub>-induced hepato-toxicity, abortifacient, immunomodulator. The structure of Secologanin is given in Fig. 10.

family (Nyctaginaceae). It consists of pentacyclic triterpenoids hydroxy monocarboxylic acid, oleanolic acid, betulinic acid.

It is used in beneficial effects, which include anti-inflammatory, anti-oxidant, anti-apoptotic, anti-carcinogenic.

Ursolic acid can be used in the treatment and prevention of obesity, diabetes, cardiovascular disease, brain disorder, and liver disease. The structure of ursolic acid is given in Fig. 11.

**11. Ursolic Acid:** Ursolic acid is present in many plants such as Mirabilis Jalapa belongs to the

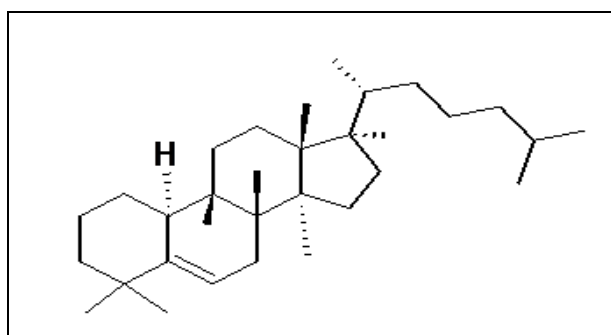


FIG. 9: CHEMICAL STRUCTURE OF CUCURBITACIN

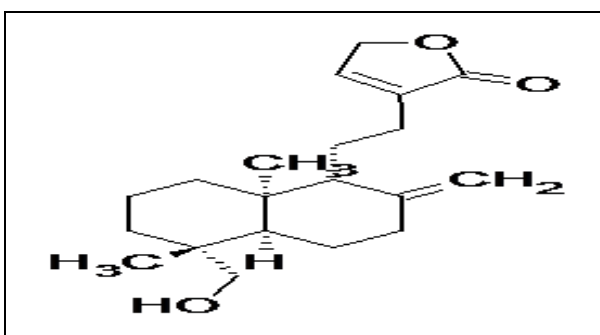


FIG. 10: CHEMICAL STRUCTURE OF SECOLOGANI

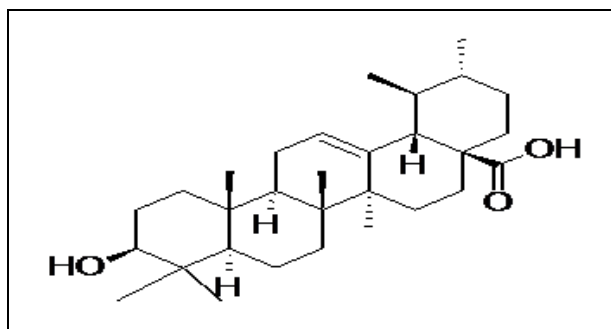


FIG. 11: CHEMICAL STRUCTURE OF URSOLIC ACID

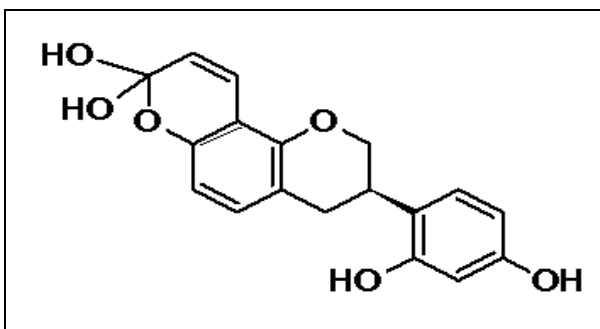


FIG. 12: CHEMICAL STRUCTURE OF GLABRIDIN



**12. Glabridin:** It is obtained from the extract of the unpeeled root of *Glycyrrhiza glabra* belong to the family (Leguminosae). It consists of triterpenoids, saponin glycosides, coumarins, flavonoids, isoliquiritigenin, and it is used in the demulcent, expectorant, anti-inflammatory, spasmolytic agent, hepatitis C and psoriasis<sup>111-112</sup>. The structure of glabridin is given in Fig. 12.

**13. Glabrene:** It is obtained from the roots of *Glycyrrhiza glabra* belong to the family (Fabaceae). Glabrene is used in the treatment of hepatitis C, Eczema, stomach infection and ulcers and so on. The structure of glabrene is given in Fig. 13.

**14. Harmine:** It is obtained from seeds of *Peganum harmala* L. belongs to the family (Zygophyllaceae). It consists of triterpenoids, flavonoids, monoamine oxidase inhibitors, alkaloids. It is used in the psychoactive effect, inhibits the formation of bone-resorbing cells, antitumor, antidiabetic, hepatoprotective, Par-

kinson's disease, anti-microbial<sup>113-115</sup>. The structure of harmine is given in Fig. 14.

**15. Harmaline:** It is an alkaloid from *Passiflora incarnate* belong to the family (Nitrariaceae). It consists of alkaloids, Beta-carbolines, saturated fatty acid, tetra decanoic acid, tridecanoic acid, hexadecanoic acid, and so on. Harmaline is used in the analgesic, emmenagogue, abortifacient and anthelmintic, anti-tumor. The structure of harmaline is given in Fig. 15.

**16. Iridomyrmecin:** It is obtained from the plant of *Actinidia polygama* belong to the family (Actinidiaceae). Iridomyrmecin consists of alkaloids, iridoids, crocetin, glycosides.

It is used as an antioxidant, to reduce swelling, constipation, gallbladder diseases, high cholesterol, high blood pressure, bladder infection, wound healing, swelling of the pancreas, rheumatoid arthritis<sup>116</sup>. The structure of Iridomyrmecin is given in Fig. 16.

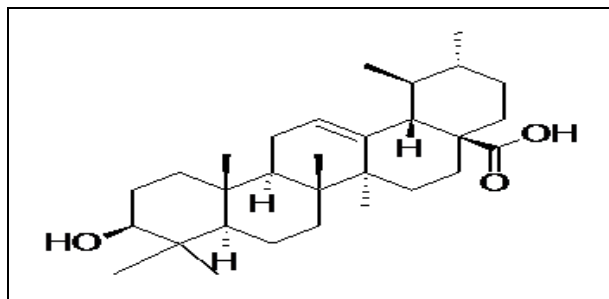


FIG. 13: CHEMICAL STRUCTURE OF GLABRENE

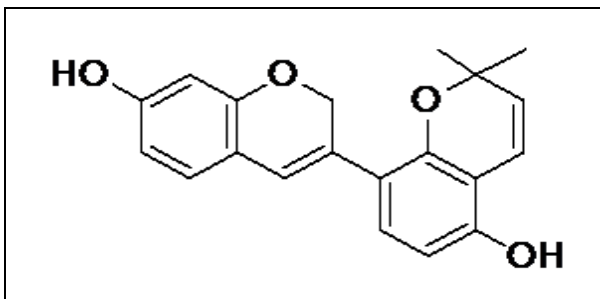


FIG. 14: CHEMICAL STRUCTURE OF HARMINE

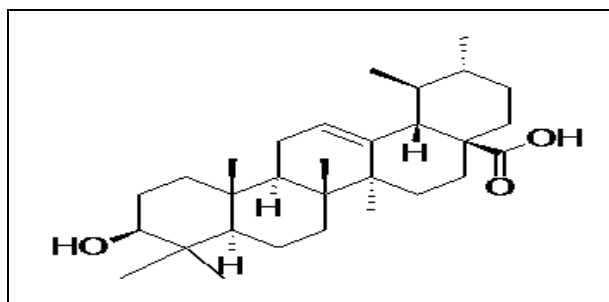


FIG. 15: CHEMICAL STRUCTURE OF HARMALINE

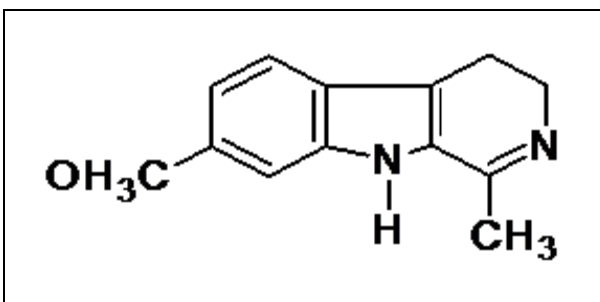


FIG. 16: CHEMICAL STRUCTURE OF IRIDOMYRMECIN

**17. Genipin:** It is obtained from the fruit of *Gardenia jasminoides* belong to the family (Rubiaceae). Genipin is an excellent natural cross-linker for proteins, collagen, and gelatin, and it is used in the treatment of cholestasis and hepatitis, wound dressing, jaundice, and so on<sup>117</sup> the structure of Genipin is given in Fig. 17.

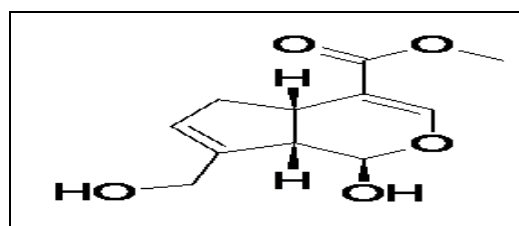


FIG. 17: CHEMICAL STRUCTURE OF GENIPIN

**CONCLUSION:** The present study synthesized the most accurate evidence for the hepatoprotective effects of some plants, fruits, and natural resin against different toxic compounds that cause hepatic damage. In general, this article identified and provided evidence of some phytochemicals with hepato-protective activity; the mechanism of action was related to their antioxidant potential and evaluated to determine their safety of the hepatoprotective activity. Now a day terpenoids get very important in the field of phytochemistry. Terpenoids act as antioxidants, and allopathic agents are used in hepatoprotection. Several leads obtained from terpenoids containing plants potential hepatoprotective agents, andrographolide, silymarin, oleander, *Daucus carota*, wild marjoram have been established to have potent hepatoprotective properties. Andrographolide is very effective on the treatment of hepatitis, jaundice, liver failure. Despite inspiring data on the possibility of discoveries in the future, evidence on the treatment of chronic liver diseases by natural medications is not sufficient. Therefore, medications discovered from natural sources should recommend to conducted more clinical trials. More confidence, better training, and little bit awareness for the natural medicine are necessary for both patients and physicians.

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#### REFERENCES:

- Ivan D: A Textbook of pathology, Jaypee brother's Medical Publishers, Eight Edition 2019.
- Michael C: A Textbook of Clinical medicine. Saunders Elsevier publishers, Seventh Edition 2009.
- Mohammed A: A Text Book of Pharmacognosy & plant cultivation. CBS Publishers & Distributors, Second Edition 2009:
- Razzak MA, Rahman M. Non-alcoholic Fatty Liver Disease: An Emerging Silent Killer. Journal of Armed Forces Medical College, Bangladesh 2018; 14(1): 1-3.
- Singh S, Osna NA and Kharbanda KK: Treatment options for alcoholic and non-alcoholic fatty liver disease. World Journal of Gastroenterology 2017; 23(36): 6549.
- Iser D and Ryan M: Fatty liver disease a practical guide for GPs. Australian Family Physician 2013; 42-44.
- Cirrhosis: National institute of diabetes and digestive and kidney diseases. April 23, 2014. Archived from the original on 9 June 2015. Retrieved 19 May 2015.
- Boyer JL: Bile formation and secretion comprehensive physiology 201; 1035-78.
- Anciaux ML, Pelletier G, Attali P, Meduri B, Liguory C and Etienne JP: Prospective study of clinical and biochemical features of symptomatic choledocholithiasis, Digestive Diseases and Sciences 1986; 31: 449-53.
- Kumar VAK and Jon C: Aster robbins basic pathology, Ninth Edition Elsevier 2013; 704-14.
- Echinococcosis Fact sheet N°377: World Health Organization, March 2014 Archived from the original on 21 February 2014. Retrieved 19 March 2014.
- Echinococcosis: CDC, 29 November 2013. Archived from the original on 20 March 2014. Retrieved 20 March 2014.
- Kinney TP: Management of ascending cholangitis. Gastrointestinal Endoscopy Clinics of North America 2007; 289-306.
- Oddsottir M and Hunter JG: Gallbladder and extrahepatic biliary system. Schwartz's Principles of Surgery. Ninth Edition. United States of America: The McGraw-Hill Education. 2010: 821-44.
- Oyama LC, Marx JA, Hock Berger RS and Walls RM: Disorders of the liver and biliary tract in (Eds): Rosen's Emergency Medicine: Concepts and Clinical Practice, 8 Edition 2010; 1186-05.
- Tahir AA, Kamal S, Sultana N, Khan MT and Khan A: CRP as an indicator of severity of acute cholecystitis, Journal of Medical Sciences 2017; 25(4): 429-32.
- Gallstones (Cholelithiasis) Clinical Presentation: History, Physical Examination medicine, Archived from the original on 2016-11-14. Retrieved 2016-11-14.
- Ansaloni L, Pisano M, Coccolini F, Peitzmann AB, Fingerhut A, Catena F, Agresta F, Allegri A, Bailey I, Balogh ZJ and Bendinelli C: WSES guidelines on acute *Calculous cholecystitis*, World Journal of Emergency Surgery 2016; 11(1): 25.
- Mishra BB and Tiwari VK: Natural products an evolving role in future drug discovery. European Journal of Medicinal Chemistry 2011; 46(10): 4769-807.
- Shah BA, Qazi GN, Taneja SC, Boswellic acids: a group of medicinally important compounds, Natural Product Reports 2009; 26(1): 72-89.
- Sherman PW and Billing J: Darwinian gastronomy: why we use spices. Spices taste good because they are good for us Bio Science 1999; 49(6): 453-63.
- Rosenblum A, Marsch LA, Joseph H and Portenoy RK: Opioids and the treatment of chronic pain controversies, current status and future directions, Experimental and Clinical Psychopharmacology 2008; 16(5): 405.
- Rabadia J, Hirani U, Kardani D and Kaneria A: Hepatoprotective activity of aqueous extract of digitalis purpurea in carbon tetra chloride induced hepatotoxicity in albino rats. Asian Journal of Biomedical and Pharmaceutical Sciences 2014; 4(34): 64.
- Zhang D, Yang R, Wang S and Dong Z: Paclitaxel: new uses for an old drug drug design. Development and Therapy 2014; 8: 279.
- Smith PM: When your hormones go haywire. Solutions for Women Over 2005; 40:109.
- Tanne J: Paracetamol causes most liver failure in UK and US, BMJ 2006; 332(7542): 628.
- Hay EM, Paterson SM, Lewis M, Hosie G and Croft P: Pragmatic randomized controlled trial of local corticosteroid injection and naproxen for treatment of

- lateral epicondylitis of elbow in primary care Bmj 1999; 319(7215): 964-8.
28. Agarwal OP: Chemistry of organic natural products, Krishna prakashan, Edition 34<sup>th</sup> 2007; 312-313.
  29. Kokate CK, Purohit AP and Gokhale SB: Pharmacognosy. Nirali Prakashan Edition 50<sup>th</sup> 2009; 14.1-14.6
  30. Rabi T and Bishayee A: Terpenoids and breast cancer chemoprevention Breast cancer research and treatment. 2009; 115(2): 223-39.
  31. Wagner KH and Elmadfa I: Biological relevance of terpenoids. Anna of Nut and Meta 2003; 47(3-4): 95-106.
  32. Sultana N and Ata A: Oleanolic acid and related derivatives as medicinally important compounds. J of Enzyme Inhibition and Medi Chemistry 2008; 6: 739-56.
  33. Hill RA: Dictionary of terpenoids. Mono and Sesquiterpenoids 1991; 1-2156.
  34. Croteau R, Kutchan TM and Lewis NG: Natural products (secondary metabolites). Biochemistry and Molecular Biology of Plants 2000; 24:1250-19.
  35. Mamat SS, Kamarolzaman MF, Yahya F, Mahmood ND, Shahril MS, Jakius KF, Mohtarrudin N, Ching SM, Susanti D, Taher M and Zakaria ZA: Methanol extract of *Mela stoma malabathricum* leaves exerted antioxidant and liver protective activity in rats. BMC Complementary and Alternative Medicine 2013; 13(1): 326.
  36. Hazra B, Sarkar R, Ghate NB, Chaudhuri D and Mandal N: Study of the protective effects of Katha (Heartwood Extract of *Acacia catechu*) in liver damage induced by iron overload. Journal of Environmental Pathology Toxicology and Oncology 2013; 32(3): 324.
  37. Handa SS: Natural products and plants as liver protecting drugs, Fitoterapia 1986; 57(5): 307-51.
  38. Bonati A and Mustich G: Patent Chem Abs 1978; 88: 22625.
  39. Desai SN, Patel DK, Devkar RV, Patel PV and Ramachandran AV: Hepatoprotective potential of polyphenol rich extracts of *Murraya koenigii* L. an *in-vivo* study. Food and Chemical Toxicology 2012; 50(2): 310-4.
  40. Akilavalli N, Radhika J and Brindha P: Hepatoprotective activity of *Ocimum sanctum* Linn. against lead induced toxicity in *albino* rats. As J Pha Clin Res 2011; 4(2): 84-7.
  41. Lahon K, Das S: Hepatoprotective activity of *Ocimum sanctum* alcoholic leaf extract against paracetamol-induced liver damage in *Albino* rats. Pharmacognosy Research 2011; 3(1): 13.
  42. Bhargava KP and Singh N: Anti-stress activity of *Ocimum sanctum* Linn. The Indian J of Medic Rese 1981; 73: 443.
  43. Phansawan B and Pongbangpho S: Antioxidant capacities of *Pueraria mirifica*, *Stevia rebaudiana Bertonii*, *Curcuma longa* Linn. *Andrographis paniculata* (Burm. f.) Nees and *Cassia alata* Linn. for the Development of Dietary Supplement Kasetsart J 2007; 41(3): 407-13.
  44. Salama SM, Abdulla MA, AlRashdi AS, Ismail S, Alkiyumi SS and Golbabapour S: Hepatoprotective effect of ethanolic extract of *Curcuma longa* on thioacetamide induced liver cirrhosis in rats. BMC Complementary and Alternative Medicine 2013; 13(1): 56.
  45. Ho CH, Noryati I, Sulaiman SF and Rosma A: *In-vitro* antibacterial and antioxidant activities of *Orthosiphon stamineus* Benth. extracts against food-borne bacteria. Food Chemistry 2010; 122(4): 1168-72.
  46. Kadir FA, Othman F, Abdulla MA, Hussain F and Hassan Darvish P: Effect of *Tinosporacrispa* on thioacetamide-induced liver cirrhosis in rats. Indian Journal of Pharmacology 2011; 43(1): 64.
  47. Sengupta M, Sharma GD and Chakraborty B: Hepatoprotective and immunomodulatory properties of aqueous extract of *Curcuma longa* in carbon tetra chloride intoxicated Swiss albino mice. Asian Pacific Journal of Tropical Biomedicine 2011; 1(3): 193-9.
  48. Mishra S, Aeri V, Gaur PK and Jachak SM: Phytochemical, therapeutic and ethnopharmacological overview for a traditionally important herb. *Boerhavia diffusa* Linn. Bio Med Research International 2014.
  49. Venkata Lakshmi P, Eazhisai VD and Netaji S: Hepatoprotective Activity of *Boerhavia diffusa* against paracetamol induced toxicity in rats. Journal of Chemical & Pharmaceutical Research 2011; 3: 229-32.
  50. Olaleye MT, Akinmoladun AC, Ogunboye AA and Akindahunsi AA: Antioxidant activity and hepatoprotective property of leaf extracts of *Boerhavia diffusa* Linn. against acetaminophen-induced liver damage in rats. Food and Chemical Toxicology 2010; 48(8-9): 2200-5.
  51. Rawat AK, Mehrotra S, Tripathi SC and Shome U: Hepatoprotective activity of *Boerhavia diffusa* L. roots-a popular Indian ethnomedicine. Journal of Ethnopharmacology 1997; 56(1): 61-6.
  52. Rajpoot K and Mishra RN: *Boerhavia diffusa* roots (Punarnava)-review as rasanya (rejuvenator/antiaging). International Journal of Pharmaceutical and Biomedical Research 2011; 2(4):1451-60.
  53. Ganie SA, Zargari BA, Masood A and Zargari MA: Hepatoprotective and antioxidant activity of rhizome of *Podophyllum hexandrum* against carbon tetra chloride induced hepatotoxicity in rats. Biomedical and Environmental Sciences 2013; 26(3): 209-21.
  54. Allam RM, Selim DA, Ghoneim AI, Radwan MM, Nofal SM, Khalifa AE, Sharaf OA, Toaima SM, Asaad AM and El-Sebakhy NA: Hepatoprotective effects of *Astragalus kahiricus* root extract against ethanol-induced liver apoptosis in rats. Chine J of Nat Me 2013; 11(4): 354-61.
  55. Sikander M, Malik S, Parveen K, Ahmad M, Yadav D, Hafeez ZB and Bansal M: Hepatoprotective effect of *Organum vulgare* in Wistar rats against carbon tetrachloride-induced hepatotoxicity. Protoplasma 2013; 250(2): 483-93.
  56. Kiran PM, Raju AV and Rao BG: Investigation of hepatoprotective activity of *Cyathia gigantea* (Wall. ex. Hook.) leaves against paracetamol-induced hepatotoxicity in rats. Asian Pacific J of Trop Biomed 2012; 2(5): 352-6.
  57. Karami M, Saeidnia S and Nosrati A: Study of the hepatoprotective activity of methanolic extract of *Feijoa sellowiana* fruits against MDMA using the isolated rat liver perfusion system. Iranian Journal of Pharmaceutical Research IJPR 2013; 12(1): 85.
  58. Panda V, Ashar H and Srinath S: Antioxidant and hepatoprotective effect of *Garcinia Indica* fruit rind in ethanolinduced hepatic damage in rodents. Interdisciplinary Toxicology 2012; 5(4): 207-13.
  59. Parameswara SA, Chetty CM and Chandrasekhar KB: Hepatoprotective activity of *Ficus religiosa* leaves against isoniazid+ rifampicin and paracetamol induced hepatotoxicity. Pharmacognosy Research 2013; 5(4): 271.
  60. Ai G, Liu Q, Hua W, Huang Z and Wang D: Hepatoprotective evaluation of the total flavonoids extracted from flowers of *Abelmoschus Manihot* (L.) Medic: *in-vitro* and *in-vivo* studies. Journal of Ethnopharmacology 2013; 146(3): 794-802.
  61. Mohan S, Thiagarajan K, Chandrasekaran R and Arul J: *In-vitro* protection of biological macromolecules against oxidative stress and *in-vivo* toxicity evaluation of *Acacia nilotica* (L.) and ethyl gallate in rats. BMC Complementary and Alternative Medicine 2014; 14(1): 257.



62. Ottu OJ, Atawodi SE and Onyike E: Antioxidant, hepatoprotective and hypolipidemic effects of methanolic root extract of *Cassia singueana* in rats following acute and chronic carbon tetrachloride intoxication. *Asian Pacific Journal of Tropical Medicine* 2013; 6(8): 609-15.
63. Mahmud Z, Bachar S and Qais N: Antioxidant and hepatoprotective activities of ethanolic extracts of leaves of *Premna esculenta* Roxb. against Carbon Tetrachloride-Induced Liver Damage in Rats. *J Young Pharm* 2012; 4(4): 228-34.
64. Swamy AV, Kulkarni RV, Koti BC, Gadad PC, Thippeswamy AH and Gore A: Hepatoprotective effect of *Cissus quadrangularis* stem extract against rifampicin-induced hepatotoxicity in rats. *Indian Journal of Pharmaceutical Sciences* 2012; 74(2): 183.
65. Balkan İA, Dogan HT, Zengin G, Colak N, Ayaz FA, Goren AC, Kirmizibekmez H and Yeşilada E: Enzyme inhibitory and antioxidant activities of *Nerium oleander* L. flower extracts and activity guided isolation of the active components. *Industr Crops and Products* 2018; 112: 24-31.
66. Samuel AJ, Mohan S, Chellappan DK, Kalusalingam A and Aria Muthu S: *Hibiscus vitifolius* (Linn.) root extracts shows potent protective action against anti-tubercular drug induced hepatotoxicity. *J of Ethno* 2012; 141(1): 396-02.
67. Desai SN, Patel DK, Devkar RV, Patel PV and Ramachandran AV: Hepatoprotective potential of polyphenol rich extract of *Murraya koenigii* L. an *in-vivo* study. *Food and Chemical Toxicology* 2012; 50(2): 310-4.
68. Gupta RK, Hussain T, Panigrahi G, Das A, Singh GN, Sweetey K, Faiyazuddin M, Rao CV: Hepatoprotective effect of *Solanum xanthocarpum* fruit extract against CCl<sub>4</sub> induced acute liver toxicity in experimental animals. *Asian Pacific Journal of Tropical Medicine* 2011; 4(12): 964-8.
69. Pandey A, Bigoniya P, Raj V and Patel KK: Pharmacological screening of *Coriandrum sativum* Linn. for hepatoprotective activity. *Journal of Pharmacy and Bio Allied Sciences* 2011; 3(3): 435.
70. Nwozo SO and Oyinloye BE: Hepatoprotective effect of aqueous extract of *Aframomum melegueta* on ethanol-induced toxicity in rats. *Acta Biochi Polonica* 2011; 58(3).
71. Sahreen S, Khan MR and Khan RA: Hepatoprotective effects of methanol extract of *Carissa opaca* leaves on CCl<sub>4</sub>-induced damage in rat. *BMC Com and Alte Medi* 2011; 11(1): 48.
72. Kirtikar KR: *Indian Medicinal Plants*, Lalit Mohan Basu, Allahabad. Jayyd Press New Delhi India 1987; 2: 146.
73. Meyer BN, Ferrigni NR, Putnam JE, Jacobsen LB, Nichols DJ and McLaughlin JL: Brine shrimp: a convenient general bioassay for active plant constituents. *Plantamedica* 1982; 45(05): 31-4.
74. Colonette S: *Wild flowers of Saudi Arabia*, East Anglian Engraving Co. Ltd., Norwich 1999; 110: 274-5.
75. El-Negoumy SI, Al-Wakeel SAM, El-Hadidi MN and Saleh NAM: The flavonoid of the *Fagonia arabica* Complex. *Phytochemistry* 1986; 25: 2423-34.
76. Al-Wakeel SA, El-Negoumy SI, El-Hadidi MN and Saleh NA: Flavonoid patterns in *Fagonia mollis* complex. *Biochemical Systemati and Ecology* 1987; 15(4): 459-60.
77. El-Hadidi MN, Al-Wakeel SA and El-Graf IA: Systematic significance of the flavonoid constituents in the *Fagonia Indica*-complex. *Bi Sy and Ec* 1988; 16(3): 293-7.
78. Oliveira FA, Chaves MH, Almeida FR, Lima Jr RC, Silva RM, Maia JL, Brito GA, Santos FA and Rao VS: Protective effect of  $\alpha$ - and  $\beta$ -amyrin, a triterpene mixture from *Protium heptaphyllum* March. trunk wood resin, against acetaminophen-induced liver injury in mice. *Journal of Ethnopharmacology* 2005; 98(1-2): 103-8.
79. Hinson JA, Roberts DW and James LP: Mechanisms of acetaminophen-induced liver necrosis. In *Adverse drug reactions* Springer. Berlin Heidelberg 2010; 369-405.
80. Liu J, Liu Y, Parkinson A and Klaassen CD: Effect of oleanolic acid on hepatic toxicant-activating and detoxifying systems in mice. *Journal of Pharmacology and Experimental Therapeutics* 1995; 275(2): 768-74.
81. Arthika S, Shanthammal Y, Sheryl Igal1 N, Elankini P, Pramod Reddy G, Gaidhani SN and Ganesan R: Hepatoprotective activity of the ethanolic extract of *Azima tetracantha* against paracetamol-induced hepatotoxicity in Wistar albino rats. *Journal of Advances in Pharmacy and Healthcare Research* 2011; 1(2): 14-20.
82. Kambizi L and Afolayan AJ: An ethnobotanical study of plants used for the treatment of sexually transmitted diseases in Guruve District, Zimbabwe. *Journal of Ethnopharmacology* 2001; 77(1): 5-9.
83. Regassa A: The use of herbal preparations for tick control in western Ethiopia. *Journal of the South African Veterinary Association* 2000; 71(4): 240-3.
84. Oliver BE: *Medicinal plants in Nigeria*, Nigerian College of Arts Science and Technology 1960; 21(37): 52-3.
85. Osadebe PO, Okoye FB, Uzor PF, Nnamani NR, Adiele IE and Obiano NC: Phytochemical analysis, hepatoprotective and antioxidant activity of *Alchornea cordifolia* methanol leaf extract on carbon tetrachloride-induced hepatic damage in rats. *Asian Pacific Journal of Tropical Medicine* 2012; 5(4): 289-93.
86. Singh BK, Pillai KK, Kohli K and Haque SE: Effect of *Cissampelo spareira* root extract on isoproterenol-induced cardiac dysfunction. *Journal of Natural Medicines* 2013; 67(1): 51-60.
87. Huxley CA: *Dictionary of gardening*. The Macmillan Press Limited London New York 1992; 3: 94-5.
88. Motawi TK, Hamed MA, Shabana MH, Hashem RM and Naser AF: Zingiber officinal acts as a nutraceutical agent against liver fibrosis. *Nutriti & Metabol* 2011; 8(1): 40.
89. Kooti W, Ghasemiboroon M, Asadi-Samani M, Ahangarpour A, Noori Ahmad Abadi M, Afrisham R and Dashti N: The effects of hydroalcoholic extract of celery on lipid profile of rats fed a high fat diet. *Advances in Environmental Biology* 2014; 8(9): 325-30.
90. Nagella P, Ahmad A, Kim SJ and Chung IM: Chemical composition, antioxidant activity and larvicidal effects of essential oil from leaves of *Apium graveolens*. *Immunopharmacology and Immunoto* 2012; 34(2): 205-9.
91. Asif HM, Akram M, Usmanghani K, Akhtar N, Shah PA, Uzair M, Ramzan M, Shah SA and Rehman R: Monograph of *Apium graveolens* Linn. *Journal of Medicinal Plants Research* 2011; 5(8): 1494-6.
92. Ahmed B, Alam T, Varshney M and Khan SA: Hepatoprotective activity of two plants belonging to the Apiaceae and the Euphorbiaceae family. *Journal of Ethnopharmacology* 2002; 79(3): 313-6.
93. Babu PS, Krishna V, Maruthi KR, Shankarmurthy K and Babu RK: Evaluation of acute toxicity and hepatoprotective activity of the methanolic extract of *Dichrostachys cinerea* leaves. *Pharma Rese* 2011; 3(1): 40.
94. Nema AK, Agarwal A and Kashaw V: Hepatoprotective activity of *Leptadenia reticulata* stems against carbon tetrachloride-induced hepatotoxicity in rats. *Indian Journal of Pharmacology* 2011; 43(3): 254.
95. *Boerhavia diffusa* was originally described and published in *Species Plantarum*. *Boerhavia diffusa*: Germplasm Resources Information Network (GRIN), Agricultural Research Service (ARS), United States Department of Agriculture (USDA), Retrieved 2013; 1: 1753.



96. Bhowmik D, Sampath KK, Srivastava S, Paswan S and Sankar A: Traditional Indian herbs Punarnava and its medicinal importance. *J Pharmacognosy Phytochemical* 2012; 1(1): 52-8.
97. Sreeja S and Sreeja S: An *in-vitro* study on antiproliferative and antiestrogenic effects of *Boerhavia diffusa* L. extracts. *J of Ethnopharm* 2009; 126(2): 221-5.
98. Handa SS and Kapoor VK: Textbook of pharmacognosy. Vallabh Prakashan, reprint 2005; 206-07.
99. Robertshaw P, Hou JP and Jin Y: The Healing Power of Chinese Herbs and Medicinal Recipes. *J of the Australian Traditional-Medicine Society* 2010; 16(1): 41-2.
100. Ouyang Z, Yang L, Su SL, Feng X and Wang M: Fingerprint of volatile oil of *Atractylodes lancea* by GC-MS. *Yao Xue Bao-Acta Pharm Sinic* 2007; 42(9): 968-72.
101. Chen YM, Chou GX and Wang ZT: Determination of beta-eudesmol in rhizome of *Atractylodes lancea* by RP-HPLC. *China J of Chine Mat Medi* 2007; 32(21): 2265-7.
102. Ali Mohammed: Textbook of pharmacognosy CBS Publishers & Distributors, Second Edition 1998; 369-371.
103. Makino T: Revised Makino's new illustrated flora of Japan. Hokuryukan Tokyo 1989; 163-4.
104. Benson MD, Yang Q, Ngo D, Zhu Y, Shen D, Farrell LA, Sinha S, Keyes MJ, Vasan RS, Larson MG and Smith JG: Genetic architecture of the cardiovascular risk proteome *Circulation* 2018; 137(11): 1158-72.
105. Gutierrez PRM: Review of *Cucurbita pepo* (pumpkin) its phytochemistry and pharmacology. *Medicinal Chemistry* 2016; 6(1): 12-21.
106. <http://eprints.ugd.edu.mk/11963/12/Ecbalium%20elaterium.pdf>
107. Lavie D, Szinai S: The Constituents of *Ecballium elaterium* L. *J of the Ameri Che Soci* 1958; 80(3): 707-10.
108. Erciyas AT, Karaosmanoglu F and Civelekoglu H: Fruit oils of four plant species of Turkish origin. *Journal of the American Oil Chemists' Society* 1989; 66(10): 1459-64.
109. Heitz A, Chiche L and Castro B: Proton 2D NMR and distance geometry study of the folding of *Ecballium elaterium* trypsin inhibitor, a member of the squash inhibitors family. *Biochemistry* 1989; 28(6): 2392-8.
110. Salhab AS: Human exposure to *Ecballium elaterium* fruit juice fatal toxicity and possible remedy. *Pharmacology & Pharmacy* 2013; 4(05): 447.
111. Andrade RJ, Medina-Caliz I, Gonzalez-Jimenez A, Garcia-Cortes M and Lucena MI: Hepatic damage by natural remedies. In *Seminars in Liver Disease* Thieme Medical Publishers 2018; 38: 21-40.
112. Yu JJ, Zhang CS, Coyle ME, Du Y, Zhang AL, Guo X, Xue CC and Lu C: Compound glycyrrhizin plus conventional therapy for psoriasis vulgaris: a systematic review and meta-analysis of randomized controlled trials. *Current Medical Resear and Opinion* 2017; 33(2): 279-87.
113. He JH, Niu YF, Li JX, Wang LB, Zi TP, Yu S and Tao J: Studies on terpenoids from *Zygophyllum fabago*. *Zhongguo Zhongyaozazhi= Zhongguozhongyaozazhi= China Journal of Chinese Materiamedica* 2015; 40(23): 4634-8.
114. Barzegar R, Safaei HR, Nemati Z, Ketabchi S and Talebi E: Green synthesis of silver nanoparticles using *Zygophyllum Qatarse* Hadidi leaf extract and evaluation of their antifungal activities. *Journal of Applied Pharmaceutical Science* 2018; 8(03): 168-71.
115. Mnafigui K, Kchaou M, Ben Salah H, Hajji R, Khabbabi G, Elfeki A, Allouche N and Gharsallah N: Essential oil of *Zygophyllum album* inhibits key-digestive enzymes related to diabetes and hypertension and attenuates symptoms of diarrhea in alloxan-induced diabetic rats. *Pharmaceutical Biology* 2016; 54(8): 1326-33.
116. Hammel BE: Three new species of *Pentagonia* (Rubiaceae) from southern Central America, one foreseen. *Two Surprising Phytoneuron* 46: 1-13.
117. Xi-xun Y, Fei L, Yuan-ting X, Chang-xiu W: *In-vitro* study in the endothelial cell compatibility and endothelialization of genipin-crosslinked biological tissues for tissue-engineered vascular scaffolds. *Journal of Materials Science. Material in Medic* 2010; 21(2): 777-85.

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