



Received on 04 June 2020; received in revised form, 12 October 2020; accepted, 03 May 2021; published 01 June 2021

## SIMULTANEOUS ESTIMATION OF TEA TREE OIL AND NEEM SEED OIL IN BULK AND COSMECEUTICAL FORMULATION BY UV SPECTROPHOTOMETRY

B. P. Manjula <sup>\*1</sup>, V. G. Joshi <sup>1</sup>, S. Ramachandra Setty <sup>2</sup> and M. Geetha <sup>1</sup>

Department of Pharmaceutics <sup>1</sup>, Department of Pharmacology <sup>2</sup>, Government College of Pharmacy, Rajiv Gandhi University of Health Sciences, Bengaluru - 560027, Karnataka, India.

### Keywords:

Tea tree oil, Neem seed oil,  
Cosmeceutical, Simultaneous  
estimation, UV spectrophotometry

### Correspondence to Author:

**B. P. Manjula**

Associate Professor,  
Department of Pharmaceutics,  
Government College of Pharmacy,  
No. 2, P. Kalinga Rao Road, Subbaiah  
Circle, Bengaluru - 560027,  
Karnataka, India.

**E-mail:** karthikamogh@gmail.com

**ABSTRACT:** Tea tree oil, an essential oil, has claims for analgesic, antimicrobial, antiseptic, and anti-inflammatory activity. Its complex composition is governed by ISO 4730:2017. It is extensively used in skin and hair care cosmeceuticals, often in combination with neem seed oil. Neem seed oil has potent antimicrobial, skin rejuvenating, and conditioning action attributed to the presence of micro constituents and complex lipid composition of triglycerides, diglycerides, and monoglycerides of both saturated and unsaturated fatty acids. Although few methods are available for estimation of individual oils, simple spectrophotometric methods suitable for routine determination of either oil or for simultaneous estimation of both the oils are not reported. A novel, simple, validated UV spectrophotometric method for simultaneous estimation of tea tree oil and neem seed oil in bulk and cosmeceutical formulation has been developed after optimization and validation of UV spectrophotometric method for individual estimation of oils. The method relies on the measurement of absorbance at 267nm and 284nm,  $\lambda_{max}$  of tea tree oil, and neem seed oil, respectively. Linearity is established over a concentration range of 20-160mcg/ml and 0.2-2.0mg/ml for tea tree oil and neem seed oil, respectively. Recovery results are in the range of 101.3-109.8% for neem seed oil and 60.0-69.2% for tea tree oil, respectively. Percent relative standard deviation values for precision, robustness and ruggedness is less than 2%. The validated method confirmed to ICH: Q2 (R1) guidelines and was successfully applied for determining the oil content of a marketed face wash product containing both the actives.

**INTRODUCTION:** Tea tree oil (TTO), an essential oil obtained by steam distillation of aerial parts of *Melaleuca alternifolia* has claims for analgesic, antimicrobial, antiseptic, and anti-inflammatory activity <sup>1</sup>. Its complex composition is governed by ISO 4730:2017. Terpinene-4-ol is the principal constituent of the oil (35% - 48%) followed by  $\gamma$ -terpinene (14% -28%),  $\alpha$ -terpinene (6%-12%) and 1,8-cineole ( $\leq 15\%$ ). TTO is extensively used in the formulation of skin and hair care cosmeceuticals <sup>2</sup>.

Likewise, neem seed oil (NSO) obtained from the dried seed kernels of *Azadirachta indica* by mechanical expression or solvent extraction, has potent antibacterial, antifungal, anti-parasitic and anti-inflammatory properties besides wound healing, moisturizing, conditioning, and regenerating properties; hence, a key ingredient in cream, face wash, shampoo, toothpaste and soap formulations <sup>3</sup>.

OTC products containing neem oil or neem extracts are often used to treat simple cuts, burns, wounds, acne, and superficial skin infections (SSI). The chemical composition of NSO indicates the presence of both macro constituents and micro constituents. Macro constituents comprise triglycerides, diglycerides ( $C_{16}$  and  $C_{18}$ ) and monoglycerides ( $C_{18}$ ), and free fatty acids (FFA) ( $C_{16}$  and  $C_{18}$ ).

<p><b>QUICK RESPONSE CODE</b></p> 	<p><b>DOI:</b> 10.13040/IJPSR.0975-8232.12(6).3264-71</p> <p>This article can be accessed online on <a href="http://www.ijpsr.com">www.ijpsr.com</a></p>
<p>DOI link: <a href="http://dx.doi.org/10.13040/IJPSR.0975-8232.12(6).3264-71">http://dx.doi.org/10.13040/IJPSR.0975-8232.12(6).3264-71</a></p>	

Palmitic acid, palmitoleic acid, stearic acid, oleic acid, linoleic acid, arachidic acid, linolenic acid, gadoleic acid and behenic acid are the usual components and represent both saturated and unsaturated fatty acids. The micro constituents possess potent pharmacological activities of which the content of azadirachtin, a principal constituent, is about 250-3000 ppm. Nimbidin and gedunin notably possess antifungal activity<sup>4-7</sup>.

There are many cosmeceuticals such as face washes and creams which combine both these active cosmetic ingredient (ACI)s, TTO for its antimicrobial action, and NSO for its antimicrobial, skin conditioning, and rejuvenating properties; therefore, analysis of such products is a herculean task owing to the highly complex and varied composition of essential oils.

There are only a few simple validated methods for the estimation of TTO. A validated HPTLC method has been reported for the determination of TTO content of cosmeceutical formulations by estimating the principal constituent, terpinen-4-ol<sup>8</sup>. Likewise, the HPTLC method has been used to determine the terpinen-4-ol content of 5% TTO microemulsion<sup>9</sup>. A gas chromatographic method using a flame ionization detector was developed for assay of  $\alpha$ -pinene in TTO formulations<sup>10</sup>. Ethosomes loaded with TTO were analyzed for oil content by HPTLC method using terpinen-4-ol as reference standard<sup>11</sup>.

A few important and useful analytical methods for the estimation of NSO include a simple spectrophotometric method developed in order to determine the solubility of NSO<sup>12</sup>, which was successfully adopted for formulation and characterization of solid lipid nanoparticles loaded with NSO for topical treatment of acne<sup>13</sup>. A simple and robust spectrophotometric method based on the Hantzsch reaction was used for the determination of neem oil in neem oil coated urea (NOCU)<sup>14</sup>. Also, the entrapment efficiency of NSO in poly (3-hydroxybutyrate-co-3-hydroxyvalerate) prepared by supercritical fluid emulsion extraction (SFEE) technique was determined by measuring the absorbance at 287nm using chloroform as solvent<sup>15</sup>. A high-performance liquid chromatographic (HPLC) method has been reported for the analysis of azadirachtin in two commercial formulations and neem oil<sup>16</sup>.

In this scenario, it is imperative to develop a simple, validated UV spectrophotometric method for routine, simultaneous estimation of tea tree oil and neem seed oil, as simple analytical methods are not available for simultaneous estimation and many cosmeceuticals contain both the ACI. Therefore, an attempt was made to develop a novel, optimized, validated, and cost-effective UV spectrophotometric method for routine simultaneous estimation of TTO and NSO in bulk and a marketed face wash formulation.

## MATERIALS AND METHODS:

**Essential Oils and Solvents:** Tea tree oil and Neem seed oil were purchased from Messrs. Falcon Essential Oils, Bengaluru. Dichloromethane (SD Fine Chem) extra pure and methanol (Qualigens) HPLC grade were used for method development. Aroma magic neem and tea tree face wash was purchased online. Product details are mentioned in **Table 10**.

**Instruments:** Shimadzu UV 1800 Spectrophotometer and Shimadzu analytical balance were used for the study.

**Standard Solution of TTO and NSO:** TTO standard solution was prepared by dissolving 0.1 g of TTO in 100ml of the solvent system consisting of 54% dichloromethane (DCM) in methanol to obtain a stock solution of 1 mg/ml TTO. NSO standard solution was prepared by dissolving 1g of NSO in 100ml of the same solvent system to get a stock solution of 10mg/ml of NSO. TTO solution of 80mcg/ml and NSO solution of 1mg/ml were prepared by appropriate dilution of the stock solution with the solvent system and scanned individually in the range of 250-400 nm **Fig. 1a, 1b**, and the  $\lambda_{\max}$  value was recorded. TTO gave a principal peak at 267 nm, whereas NSO gave a prominent peak at 284nm and a second less significant one at 324nm.  $\lambda_{\max}$  284nm was selected for simultaneous estimation of the oils.

**Determination of percent extinction coefficient of TTO and NSO:** Linearity of TTO and NSO were established separately at 267 nm for TTO and at 267 and 284nm for NSO in the concentration range of 20-160 mcg/ml and 0.2-2 mg/ml for TTO and NSO respectively **Fig. 2, 3, 4**. Mean percent extinction values for TTO and NSO at both 267nm

and 284nm were calculated from individual values obtained from linearity study using the formula **Table 1, 2, 3, 4.**

The developed method was validated for various parameters as per ICH: Q2 (R1) guidelines, and parameters of linearity are summarized in **Table 5.** The concentration of TTO and NSO in various aliquots and sample solutions were determined by the simultaneous equation as given below.

$$C_x = \frac{(A_{2y1} - A_{1y2})}{(a_{x2} a_{y1} - a_{x1} a_{y2})}$$

$$C_y = \frac{(A_{1x2} - A_{2x1})}{(a_{x2} a_{y1} - a_{x1} a_{y2})}$$

Where,

- $C_x$  and  $C_y$  are the concentrations of TTO and NSO,
- $A_1$  and  $A_2$  is the absorbance of the sample solution at 267 and 284nm,
- $a_{x1}$  and  $a_{x2}$  are the percent extinction coefficients of TTO at 267 and 284nm,
- $a_{y1}$  and  $a_{y2}$  are the percent extinction coefficients of NSO at 267 and 284nm, respectively.

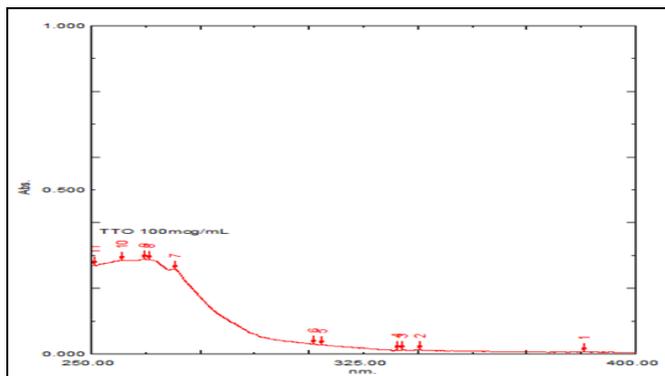


FIG. 1A: UV SPECTRUM OF TTO

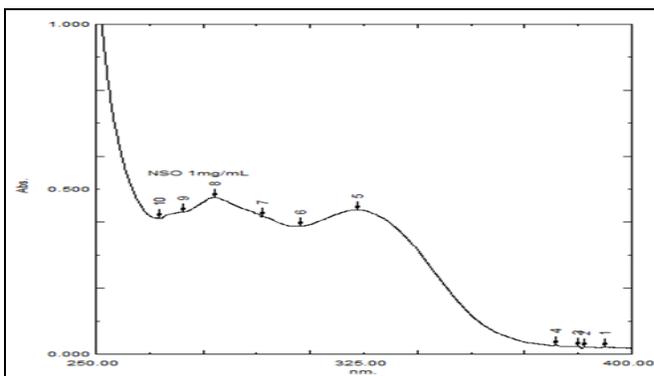


FIG. 1B: UV SPECTRUM OF NSO

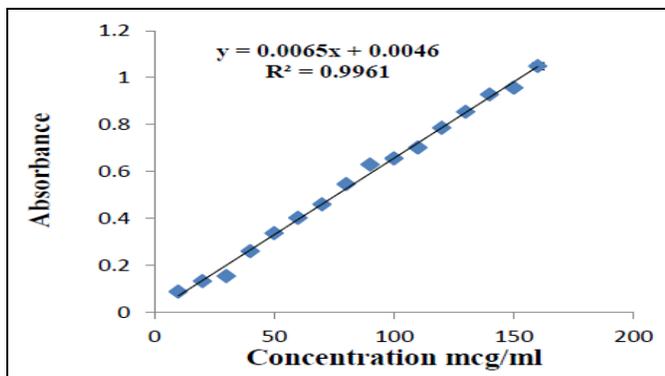


FIG. 2: CALIBRATION CURVE OF TTO AT 267 nm

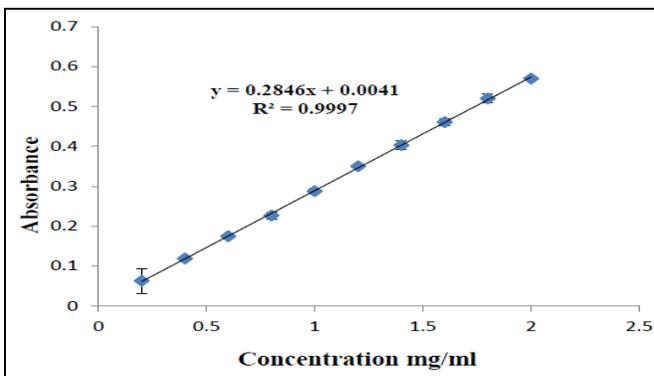


FIG. 3: CALIBRATION CURVE OF NSO AT 267 nm

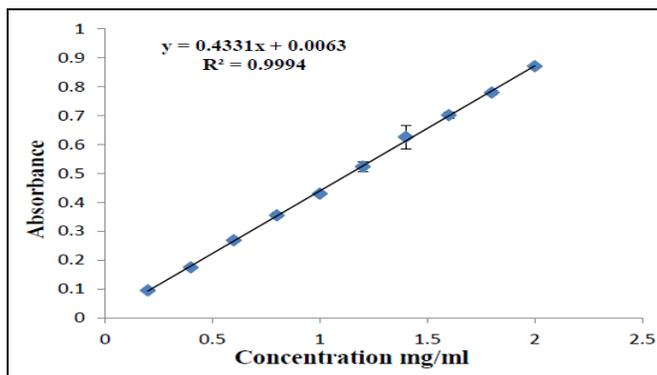


FIG. 4: CALIBRATION CURVE OF NSO AT 284 nm

**TABLE 1: DATA OF CALIBRATION CURVE AND PERCENT EXTINCTION COEFFICIENT OF TTO**

S. no.	Concentration mcg/ml	Average absorbance $\pm$ SD	$\epsilon_{1\% 1cm}$ at $\lambda_{max}$ 267nm
1	10	0.087 $\pm$ 0.002	87
2	20	0.132 $\pm$ 0.002	66
3	30	0.154 $\pm$ 0.002	51.3
4	40	0.26 $\pm$ 0.004	65
5	50	0.337 $\pm$ 0.001	67.4
6	60	0.402 $\pm$ 0.002	67
7	70	0.46 $\pm$ 0.006	65.7
8	80	0.546 $\pm$ 0.003	68.3
9	90	0.63 $\pm$ 0.002	70
10	100	0.656 $\pm$ 0.003	65.6
11	110	0.702 $\pm$ 0.002	63.8
12	120	0.786 $\pm$ 0.004	65.5
13	130	0.854 $\pm$ 0.003	65.7
14	140	0.928 $\pm$ 0.002	66.3
15	150	0.957 $\pm$ 0.008	63.8
16	160	1.049 $\pm$ 0.016	65.6

Average percent extinction coefficient=66.5

**TABLE 2: DATA OF CALIBRATION CURVE AND PERCENT EXTINCTION COEFFICIENT OF NSO**

S. no.	Concentration mg/ml	Average absorbance $\pm$ SD	$\epsilon_{1\% 1cm}$ at $\lambda_{max}$ 267nm
1	0.2	0.063 $\pm$ 0.031	3.13
2	0.4	0.119 $\pm$ 0.002	2.97
3	0.6	0.174 $\pm$ 0.001	2.91
4	0.8	0.226 $\pm$ 0.009	2.83
5	1.0	0.288 $\pm$ 0.007	2.88
6	1.2	0.35 $\pm$ 0.002	2.91
7	1.4	0.403 $\pm$ 0.012	2.88
8	1.6	0.46 $\pm$ 0.008	2.88
9	1.8	0.52 $\pm$ 0.011	2.89
10	2.0	0.57 $\pm$ 0.002	2.85

Average percent extinction coefficient = 2.91

**TABLE 3: DATA OF PERCENT EXTINCTION COEFFICIENT OF TTO**

S. no.	Concentration mcg/ml	Average absorbance $\pm$ SD	$\epsilon_{1\% 1cm}$ at $\lambda_{max}$ 284nm
1	10	0.004 $\pm$ 0.001	4
2	20	0.009 $\pm$ 0.001	4.5
3	30	0.010 $\pm$ 0.001	3.3
4	40	0.014 $\pm$ 0.001	3.5
5	50	0.017 $\pm$ 0.001	3.4
6	60	0.022 $\pm$ 0.001	3.7
7	70	0.046 $\pm$ 0.002	6.6
8	80	0.075 $\pm$ 0.001	9.4
9	90	0.109 $\pm$ 0.002	12.1
10	100	0.165 $\pm$ 0.002	16.5
11	110	0.188 $\pm$ 0.002	17.1
12	120	0.213 $\pm$ 0.001	17.8
13	130	0.240 $\pm$ 0.002	18.5
14	140	0.271 $\pm$ 0.002	19.4
15	150	0.466 $\pm$ 0.002	31.1
16	160	0.499 $\pm$ 0.003	31.1

Average percent extinction coefficient=12.6

**TABLE 4: DATA OF CALIBRATION CURVE AND PERCENT EXTINCTION COEFFICIENT OF NSO**

S. no.	Concentration mg/ml	Average absorbance $\pm$ SD	$\epsilon_{1\% 1cm}$ at $\lambda_{max}$ 284nm
1	0.2	0.095 $\pm$ 0.004	4.77
2	0.4	0.175 $\pm$ 0.007	4.38
3	0.6	0.269 $\pm$ 0.003	4.48
4	0.8	0.355 $\pm$ 0.009	4.44
5	1.0	0.43 $\pm$ 0.003	4.30
6	1.2	0.524 $\pm$ 0.017	4.37
7	1.4	0.626 $\pm$ 0.040	4.47
8	1.6	0.702 $\pm$ 0.012	4.39
9	1.8	0.78 $\pm$ 0.007	4.33
10	2.0	0.871 $\pm$ 0.006	4.36

Average percent extinction coefficient=4.43

**TABLE 5: SUMMARY OF LINEARITY PARAMETERS**

Parameter	TTO	NSO	NSO
	$\lambda_{max}$ 267nm	$\lambda_{max}$ 267nm	$\lambda_{max}$ 284nm
Linearity range	20-160 mcg/ml	0.2-2.0 mg/ml	0.2-2.0 mg/ml
Equation	$y = 0.0065x + 0.0046$	$y = 0.2846x + 0.0041$	$y = 0.4331x + 0.0063$
R <sup>2</sup> value	0.9961	0.9997	0.9994
LOD	0.1277 mcg/ml	0.3595 mg/ml	0.1259 mg/ml
LOQ	4.195 mcg/ml	1.0892 mg/ml	0.3815 mg/ml

**Preparation of Sample Solution:** 0.1g of the face wash was weighed accurately into a 25 ml volumetric flask, dissolved and volume made up using the solvent system.

The absorbance of the resulting solution was measured at both wavelengths.

### Method Development and Validation:

**Linearity:** Linearity was established over the range of 20-160mcg/ml for TTO at  $\lambda_{max}$  267nm and 0.2-2.0 mg/ml for NSO at 267nm and  $\lambda_{max}$  284nm by

appropriate dilution of the stock solution with the solvent system and measuring the absorbance of the solutions at the respective wavelength. All determinations were done in triplicate. Limit of detection (LOD) and limit of quantification (LOQ) values were calculated using the formula

$$LOD = \frac{3.3 \times \sigma}{\text{slope}} \quad \text{and,}$$

$$LOQ = \frac{10 \times \sigma}{\text{slope}}$$

respectively, where  $\sigma$  is the standard deviation of the response at low concentration.

**Accuracy:** Accuracy of the method was determined by the method of standard addition at

80%, 100%, and 120% levels of the select concentration of TTO/NSO. Recovery values are given in **Table 6**.

**TABLE 6: ACCURACY OF THE METHOD**

Concentration Levels %	Amount added		Amount recovered		Recovery %	
	TTO mg	NSO mg	TTO mg	NSO mg	TTO	NSO
80	0.8	8.0	0.51	8.69	63.75	108.6
100	1.0	10.0	0.60	10.98	60.0	109.8
120	1.2	12.0	0.83	12.15	69.2	101.3

**Precision:** Intraday precision or repeatability was established by measuring the absorbance of the standard mix solution containing 100mcg/ml TTO and 1mg/ml NSO, respectively, in triplicate. Inter-day precision or intermediate precision was

determined by measuring the absorbance of the solution in triplicate on three consecutive days by following the proposed method. Percent RSD was calculated and the data presented in **Table 7**.

**TABLE 7: DATA OF PRECISION**

Parameter	Trial/Day	Amount present		Amount present%		RSD%	
		TTO mg	NSO mg	TTO	NSO	TTO	NSO
Intraday precision	I	0.63	12.50	63	125	0.51	0.75
	II	0.63	12.69	63	126.9		
	III	0.63	12.61	63	126.1		
Inter day precision	I	0.69	11.66	69	116.6	0.15	0.56
	II	0.69	11.53	69	115.3		
	III	0.69	11.60	69	116.0		

**Ruggedness:** Ruggedness of the method was determined by measuring the absorbance of the standard solution in triplicate using two different

instruments and analysts. Percent RSD was calculated, and the data is presented in **Table 8**.

**TABLE 8: DATA OF RUGGEDNESS**

Parameter		Amount present		Amount present %		RSD %	
		TTO mg	NSO mg	TTO	NSO	TTO	NSO
Analyst	I	0.69	11.60	69	116.0	0.15	0.56
	II	0.66	11.24	66	112.4	0.68	0.11
Instrument	I	0.66	11.24	66	112.4	0.68	0.11
	II	0.72	10.89	72	108.9	0.70	0.51

**Robustness:** Robustness of the method was evaluated by measuring the absorbance of the standard solution at  $267 \pm 1\text{nm}$  and  $284 \pm 1\text{nm}$  in

triplicate, and percent RSD values are presented in **Table 9**.

**TABLE 9: DATA OF ROBUSTNESS**

Wave length $\pm 1\text{nm}$		Amount present		Amount present %		RSD %	
$\lambda_{\text{max}1}$	$\lambda_{\text{max}2}$	TTO mg	NSO mg	TTO	NSO	TTO	NSO
267nm	284nm						
268	285	0.66	10.77	66	107.7	1.04	0.15
267	284	0.72	10.89	72	108.9	0.70	0.51
266	283	0.66	11.26	66	112.6	0.19	0.39

**Assay of Marketed Face Wash Formulation:** An average of six determinations was taken for estimating the content of TTO and NSO present in

the face wash formulation. The content of the oils in the face wash product is presented in **Table 10**.

**TABLE 10: CONTENT OF TTO AND NSO IN FACE WASH**

Formulation		Composition	Content of TTO % w/w	Content of NSO % w/w
Aroma magic neem and tea tree face wash 100mL		Aqua, Lauryl glucoside, Glycerol oleate, <i>Melaleuca alternifolia</i> essential oil, <i>Azadirachta indica</i> extract, Hydroxypropyltrimonium honey, Fruit derived beta carotene, <i>Lavendula angustifolia</i> essential oil and <i>Citrus medica</i> limonum extract	2.05	7.35
Mfg lic no	16/C/UA/2009			
Mfg date	NOV 2019			
Best before	OCT 2022			
Mfg by	Blossom Kochhar Beauty Products Pvt Ltd, Haridwar, India			

**RESULTS AND DISCUSSION:** An initial screening study was carried out to select a suitable common solvent to dissolve NSO and TTO to get a stable solution. Solvents chosen for the study were n-hexane, cyclohexane, toluene, dichloromethane and methanol; binary solvent systems like chloroform-methanol (2:1v/v) and dichloromethane-methanol (3:2v/v). NSO is highly non-polar and insoluble in common solvents such as chloroform, carbon tetrachloride, n-hexane, toluene, and chloroform-methanol (2:1v/v). It was, however, soluble in cyclohexane and freely soluble in dichloromethane-methanol (DCM-Me OH) (3:2v/v). TTO contains both polar and nonpolar constituents, and the dichloromethane-methanol (3:2v/v) solvent system yielded a stable solution. Dichloromethane-methanol (3:2v/v) was a suitable common solvent system for both NSO and TTO. DCM, widely used for the extraction of lipids, has excellent solvency for various organic compounds. It forms a binary solution with methanol, in which both the oils are readily soluble to yield a stable solution. Spectral scans from 400-250nm were run for NSO and TTO solutions, respectively. Distinct peaks were obtained for both the oils, and also the absorbance remained stable, which may be attributed to the reduced volatility of the solvent system due to the presence of methanol; DCM has a boiling point of 40 °C and Me OH, 64.7 °C, respectively.

An optimized and validated UV spectrophotometric method for the determination of NSO and TTO was successfully developed by applying statistical design of experiments (unpublished data). The optimum composition of the solvent system suggested for the method was 54% DCM in methanol. It was observed that TTO exhibits a prominent spectral peak at  $\lambda_{\max}$  267nm alone, whereas NSO exhibits two prominent spectral peaks at  $\lambda_{\max}$  284nm and 324nm, respectively. TTO showed negligible absorbance at 284nm and

324nm, whereas NSO showed significant absorbance at 267nm. Hence absorbance at 267nm and 284 nm respectively were selected for routine simultaneous estimation of both the oils in bulk and a marketed formulation. Simultaneous equation method was applied for calculating the content of TTO and NSO in standard solutions and sample solutions. All determinations were carried out in triplicate. The method was validated as per ICH: Q2 (R1) guidelines for accuracy, precision, ruggedness, and robustness.

Linearity was observed for 20-160 mcg/ml of TTO at 267nm and 0.2-2.0 mg/ml of NSO at 267nm and 284nm, respectively. The linear regression equation and  $R^2$  values indicate a good correlation between concentration and the measured absorbance at the respective wavelength. LOD and LOQ for NSO determined at 267nm were high, although  $R^2$  value of 0.9997 indicated an excellent correlation between concentration and absorbance. This is acceptable considering the fact that  $\lambda_{\max}$  for NSO is 284nm but shows good absorbance at 267nm.

Accuracy of the method was established by carrying out recovery studies at 80%, 100%, and 120% levels of mid-concentration of Beer's law range of TTO and NSO, which is 1mg TTO and 10mg of NSO, respectively. Results for recovery are tabulated in table 5 and lies in the acceptable range of 80%-120% for NSO. It was observed that recovery of TTO was less compared to NSO *i.e.*, 60.0% - 69.2%, probably due to the presence of volatile components in TTO as observed during the validation of the method for various parameters. In spite of lower recovery values of TTO, the method can still be considered for routine determination in view of unavailable simple analytical methods.

Precision of the method was determined by carrying out intra-day and inter-day determinations,

the results of which are given in **Table 6**. RSD values less than 2% indicate that the method has adequate precision.

Ruggedness of the method was determined by carrying out the analysis by two different analysts and by using two different instruments, the results of which are given in **Table 7**. RSD values less than 2% indicate that the method is rugged.

Robustness of the method was established by deliberately varying the wavelengths by  $\pm 1$ nm of the selected wavelengths viz, 267nm, and 284nm, respectively. Results are presented in table 8. RSD values less than 2% imply that the method is robust enough to be used for routine analysis.

Finally, the method was applied to determine the content of oils in a marketed face wash formulation. The content of TTO and NSO was found to be 2.05% w/w and 7.35% w/w, respectively **Table 9**. Verification with label claim was not possible since the current labeling requirements for cosmetics and personal care products do not impose a limit on the amount of active cosmetic ingredients to be included in a formulation nor specify the quantity of ACIs present in the product. TTO, a potent antimicrobial, and anti-inflammatory agent is also a potential skin irritant and must be used with caution<sup>10</sup>. TTO content in the product is well within the acceptable limit of 1-5%. TTO, although generally not suitable for dry and sensitive skin owing to its irritancy, is counterbalanced by the presence of a high concentration of neem extract, which helps to address this issue by soothing, moisturizing, and rejuvenating the skin.

**CONCLUSION:** An attempt has been made to develop a novel UV spectrophotometric method for routine, simultaneous estimation of TTO and NSO in bulk and cosmeceutical formulations. The developed method is simple, rapid, and economical; furthermore, it does not require sophisticated instrumentation. The method, of course, has certain limitations in that; it is non-specific and less sensitive, considering the variable and highly complex composition of the oils. Thus, simultaneous analysis of both oils can be performed in any simple analytical laboratory but only after necessary standardization. However, this study may be of significance to various regulatory

bodies in the pursuit of maintaining the highest possible quality standards for cosmeceuticals since it helps in the development and standardization of products containing essential oils.

**ACKNOWLEDGEMENT:** The authors wish to thank the Government College of Pharmacy and Drugs Control Department, Government of Karnataka, for providing the necessary facilities to carry out research.

**CONFLICTS OF INTEREST:** Nil

## REFERENCES:

1. International organization for standardization. Essential oil of Melaleuca, terpinen-4-ol type (Tea tree oil). ISO4730: 2017 and AS 2782:2017 Standards.
2. Carson CF, Hammer KA and Riley TV: *Melaleuca alternifolia* (Tea Tree) Oil: a Review of Antimicrobial and Other Medicinal Properties. Clin Microbiol Rev 2006, 19(1): 50-62.
3. Kausik B, Chattopadhyay I, Banerjee RK and Bandyopadhyay U: Biological activities and medicinal properties of Neem (*Azadirachta indica*). Current Science 2002; 82(11): 1336-45.
4. Kumar PS, Mishra D, Ghosh G and Panda CS: Biological action and medicinal properties of various constituent of *Azadirachta indica* (Meliaceae)" an Overview. Annals Biol Res 2010; 1(3): 24-34.
5. Rahman SZ and Jairajpuri MS: Neem in Unani Medicine. Neem Research and Development Society of Pesticide Science, India, New Delhi, edition 2, 1996; 208-19.
6. Khanam Z, Al-Yousef HM, Singh O and Bhat IUH: Neem oil. Green Pesticides Handbook: Essential Oils for Pest Control. CRC Press, 2017; 383-87.
7. Sandanasamy J, Nour AH, Tajuddin SNB and Nour AH: Fatty Acid Composition and Antibacterial Activity of Neem (*Azadirachta indica*) Seed Oil. The Open Conference Proceedings Journal 2013; 4(Suppl-2, M11): 43-48.
8. Biju SS, Ahuja A, Rafiullah MRM and Khar RKA: Validated HPTLC method for determination of tea tree oil from cosmeceutical formulations. J Pharm Biomed Anal. 2005; 38: 41-44.
9. Sonia K and Anupama D: Microemulsion based transdermal drug delivery of tea tree oil. Int J Drug Dev Res 2011; 13: 191-98.
10. Gulati N, Goyal AK, Rajesh KS, Mittal K and Rath G: Method development and validation for the GC assay of  $\alpha$ -pinene in tea tree oil formulation. Pharmacia 2012; 1(3): 102-06.
11. Venugopal V, Goh R, Ping TY and Jin TJ: Formulation development and characterization of tea tree oil loaded ethosomes. Indonesian J Pharm 2016; 27: 44-52.
12. Kulkarni AR, Soppimath KS and Aminabhavi TM: Solubility Study of *Azadirachta indica* A. Juss. (Neem) Seed Oil in the Presence of Cosolvent/Nonionic Surfactant at (298.15, 303.15, 308.15, and 313.15) K. J Chem Eng Data 1999; 44: 836-38.
13. Vijayan V, Aafreen S, Sakthivel S and Reddy RK: Formulation and characterization of solid lipid nanoparticles loaded Neem oil for topical treatment of acne. J Acute Dis 2013; 282-86.

14. Kumar R, Devakumar C, Kumar R and Gupta AK: A simple and robust method for determination of neem (*Azadirachta indica* A. Juss) oil in neem oil coated urea. Toxicol Environ Chem 2012; 94(4): 641-49.
15. Mendonc FMR, Pollonia AE, Junges A, Silva RS, Rubira, AF, Borges GR, Dariva C and Franceschi E: Encapsulation of neem (*Azadirachta indica*) seed oil in poly (3-hydroxybutyrate-co-3-hydroxyvalerate) by SFEE

- technique. The J Supercrit Fluids. 2019, //doi.org/10.1016 / j.supflu.2019.104556 0896-8446/2019.
16. Sundaram KMS and Curry J: High performance liquid chromatographic method for the analysis of azadirachtin in two commercial formulations and neem oil. J Environ Sci Health, Part B: Pesticides, Food Contaminants, and Agricultural Wastes 1993; 28(2): 221-41.

**How to cite this article:**

Manjula BP, Joshi VG, Setty SR and Geetha M: Simultaneous estimation of tea tree oil and neem seed oil in bulk and cosmeceutical formulation by UV spectrophotometry. Int J Pharm Sci & Res 2021; 12(6): 3264-71. doi: 10.13040/IJPSR.0975-8232.12(6).3264-71.

All © 2013 are reserved by the International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **Android OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)