



Received on 14 October, 2011; received in revised form 16 December, 2011; accepted 22 January, 2012

## ISOLATION OF *PENICILLIUM* SP. AND ITS ANTAGONISTIC ACTIVITY AGAINST DERMATOPHYTES FROM VOLCANO SOIL OF BARATANG ISLAND, ANDAMAN

A. Dhinakaran\*, M. Kalaiselvam, V. Sekar and G.V.B. Sethubathi

Centre of Advanced Study in Marine Biology, Faculty of Marine Sciences, Annamalai University, Parangipettai-608 502, India

### Keywords:

*Penicillium* sp.,  
Antagonistic activity,  
Dermatophytes,  
Baratang Island

### Correspondence to Author:

A. Dhinakaran

Centre of Advanced Study in Marine  
Biology, Faculty of Marine Sciences,  
Annamalai University, Parangipettai- 608  
502, India

### ABSTRACT

*Penicillium* sp. was isolated from the mud volcano of Baratang Island, Andaman, were tested for the growth inhibition against four fungal species of dermatophytes viz., *Epidermophyton floccosum*, *Microsporum gypseum*, *Tricophyton rubrum*, *Tricophyton mentagraphytes*. Three different concentration of fungal suspension (10µl, 25 µl and 50 µl) were tested using disc diffusion method. *Penicillium* sp. showed promising antidermatophytic activities against all four pathogens. Among the concentration tested maximum activity against *M. gypseum* (25mm) in the concentration of 50 µl and minimum activity expressed against *E. floccosum* (9mm) in the concentration of 10 µl. Further, the isolation and characterization of the novel anti-dermatophytic compound is in progress.

**INTRODUCTION:** Marine fungi are heterotrophic microorganisms, occurring commonly as saprotrophs and parasites. In general, they occur as saprotrophs in the large particulate detritus and also as parasites on plants and animals<sup>16</sup>. But, prevalence of their different taxonomic groups of fungi varies greatly with varying substrates and extends from the temperate oceans to the polar oceans. Although widespread in terrestrial habitats, they have remained under explored from various extreme marine habitats.

The isolation of these extremophilic fungi is very essential as they might be potential sources of enzymes with scientific and commercial interests. Among the various fungal species *Penicillium* sp. are important species of fungus which is known not only due to their widespread occurrence but also because of their ability to produce mycotoxins and other secondary metabolites<sup>9</sup>. The term "Dermatophyte" literally means "Skin Plant". The documented use of the word did not appear until 1882<sup>21</sup>.

The infections they cause have been existed for hundreds of thousands of years. Currently, dermatophytes are most common infectious agents of humans and are found among all peoples throughout the world. The most infectious elements of the dermatophytes are the arthroconidia that are found *in vivo*<sup>23</sup>.

They can be divided into three major groups on the basis of their natural habitat and host preferences<sup>1</sup> anthropophilic species, which make humans their primary host and rarely infect lower animals, (ii) zoophilic species, which make lower animals their primary hosts, but can also infect humans; (iii) geophilic species, which live as saprophytes in soil and occasionally infect humans or lower animals. The natural preferred ecology of the etiologic agent of human infection is clinically significant, as the fungi that are geophilic or zoophilic tend to produce more marked skin reaction than those that are anthropophilic<sup>12</sup>.

Generally, the less inflammatory infections are the more chronic ones<sup>22</sup>. Human infections, particularly those involving the skin and mucous membranes are increasing at an alarming rate, especially in tropical and subtropical developing countries, with the most common pathogens like dermatophytes and *Candida* sp. This increase is directly related to the growing population of immune compromised individuals, resulting from changes in medical practice such as the use of intensive chemotherapy and immune-suppressive drugs. HIV and other diseases causing immunosuppression have also contributed to this problem<sup>13,5</sup>.

With this back ground, the present study was aimed to isolate fungal species from volcano soil and to trail the antagonistic activity against dermatophytes. There is a scarcity of references that states the anti fungal activity of volcano soil samples. Keeping this in mind, the present study was under taken to isolate *Penicillium* sp. From the volcanic soil of Baratang Island at Andaman Nicobar, India and to test its anti-dermatophytic activity using crude extracts of *Penicillium* sp.

**Study Area:** Andaman and Nicobar islands located in east of Indian mainland which is the archipelago of 512 islands, islets and rocks (**Fig. 1**). This union territory stretches over a length of more than 700 kms from north to south with 36 inhabited islands.

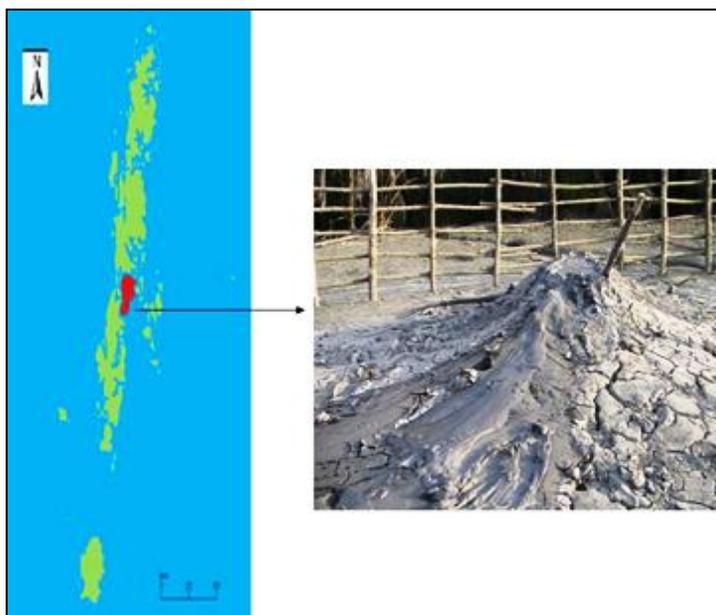


FIG. 1: THE MAP SHOWING STUDY AREA OF BARATANG ISLAND

On a hill range extending from Burma (Myanmar) to Indonesia, this indulating landscape is covered with dense forests and endless variety of exotic flora and fauna. The topography of this island in hilly and abounds in evergreen forests. The sandy beached on the edges of meandering coastline are fringed with coconut palms that sway to the rhythm of the sea. The rare flora and fauna, underwater marine life and corals crystal clear water and mangrove lined creeks, after a dream view of the rare gifts of nature.

## MATERIALS AND METHODS:

**Collection of Sediment Sample:** Sediment samples were collected from the mud volcano, Baratang Island (12° 07' N; 92° 47' E), Andaman Island. For Fungal analysis, pre-cleaned materials were used and samples were stored in sterilized polythene bags and bottles. Immediately after collection, samples were transported to the laboratory by keeping them in ice box and the samples were processed with 4 hrs.

**Isolation and Identification of Fungi:** Wet mud samples, weighing approximately one gram each, was placed in 100ml of sterile distilled water shake for 5 minutes. The resulting soil suspension was diluted in tenfold increment from 10<sup>2</sup> to 10<sup>3</sup>. One ml of suspension from each dilution was transferred into separate sterile petridishes in which 20 ml of sterilized and lukewarm mantn's SDA agar (pH7) was poured. The petridishes were gently rotated clockwise and anticlockwise to facilitate uniform spread of the inoculums in the medium.

The plates were incubated for a week at room temperature and then the fungal colonies were counted using a colony counter. Average number of fungal propagules per gram of soil and the average number of species were assessed for every collection. The fungal isolates were then transferred, cultivated and identified to species level applying standard mycological methods<sup>19</sup>. Most of the isolates were sub-cultured on Sabouraud Dextrose Agar (SDA) for further study. The classification outlined by smith<sup>19</sup> was followed here and cultures were examined at frequent intervals.

**Preparation of Fungal Crude Extract:** Well developed colonies were picked from the fungi culture slant. Then this colony were inoculated in the mass scale culture medium (SDA) and incubated at 37°C for three days. After the incubation the dense colonies were centrifuged at 5000rpm for 50mins. The crude filtrate were kept refrigerate at 4°C.

**Inoculums preparation and standardization:** The four dermatophytic fungi namely *Epidermophyton floccosum*, *Micorosoprum gypseum*, *Tricophyton rubrum*, *Tricophyton mentagraphytes* were used for bioassays with fungal crude extracts. These dermatophytes were obtained from Department of Microbiology, Raja Muthiah Medical College and Hospital (RMMC), Annamalai University. The stock cultures were enriched in the Sabouraud Dextrose Agar (SDA) plates for 10-15 days. The grown culture was scrapped with sterile scalped and macerated in 10µl of sterile physiological saline. Inoculums standardization was done using standard procedure<sup>6</sup>. The fungal suspensions adjusted spectrophotometrically to an absorbance of 0.6 OD at 530 nm. In vitro fungal activity was determined by using agar well method in SDA medium.

**Anti-Dermatophytic assay with *Penicillim* sp. crude extract:** *In vitro* antifungal activity of *Penicillium* crude extracts was determined against four species of dermatophytic fungi. Stock cultures of fungi were maintained in Czapek Dox agar. For *Epidermophyton floccosum*, *Micorosoprum gypseum*, *Tricophyton rubrum*, *Tricophyton mentagraphytes* well - drained spores were distributed uniformly of the surface of the agar plates with the help of a sterile cotton swab. Other fungal strains were inoculated by taking a piece of fungal colony using a sterile cotton swab and gently swabbed on the surface uniformly. Agar was used as the medium for antifungal assay. On the surface of the medium, discs inoculated with the extract (20 µl/6mm disc) were placed. The inhibition zone was measured after 72 hours of incubation.

**RESULTS AND DISCUSSION:** The restricted fungal samples in volcano soil sample are difficult to identify as they are complicated due to lack of the knowledge. Based on the identification manual<sup>6</sup>, the colony morphology, spores, color of the mycelium and light

microscopic identification confirmed the organisms were *Penicillium* sp (fig. 2).

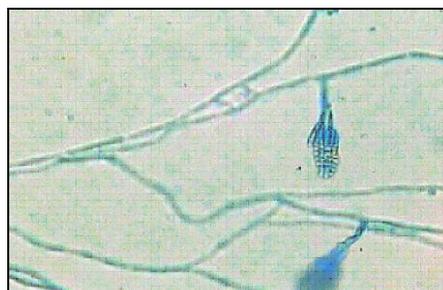
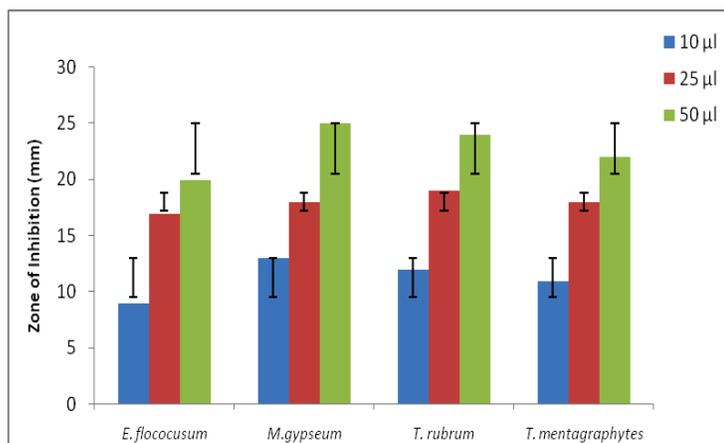


FIG. 2: MICROSCPIC VIEW OF *PENICILLIUM* SP.

In recent years, great attention has been paid to study the bioactivity of natural products because of their potential pharmacological utilization. The first attempt to locate antimicrobial activity in the marine organisms was initiated around 1950s<sup>2</sup>. Since this time, a large number of marine organisms from a wide range of phyla have been screened for antimicrobial activity<sup>18, 17</sup>. Many of these organisms have antimicrobial properties, although most of the antibacterial agents that have been isolated from marine sources have not been active enough to compete with classical antimicrobials obtained from microorganisms<sup>17</sup>. However, majority of marine organisms are yet to be screened for discovering useful antibiotics.

In the present investigation, *Penicillium* sp. was isolated from the Baratang Island. The *Penicillium* Sp. crude extracts were assessed antidermatophytic activity against four species (*Epidermophyton floccosum*, *Micorosoprum gypseum*, *Tricophyton rubrum*, *Tricophyton mentagraphytes*) using disc diffusion method. This extract showed promising antidermatophytic activites against all four pathogens.

Among the concentration tested the crude extract showed maximum activity against *M. gypseum* (25mm) pathogen in the concentration of 50 µl and minimum activity expressed against *E. floccosum* (9mm) in the concentration of 10 µl. In 50 µl concentration, maximum activity against *M. gypseum* (25mm) and minimum activity expressed against *E. floccosum* (20mm). In 25 µl concentration, maximum activity against *T. rubrum* (19mm) and minimum activity expressed against *E. floccosum* (17mm). In 10 µl concentration, maximum activity against *M. gypseum* (13mm) and minimum activity *E. floccosum* (9mm) (Fig. 3).



**FIG. 3: DIFFERENT CONCENTRATION OF CRUDE EXTRACTS AND ZONE OF INHIBITION IN (mm)**

There are many studies on anti-fungal activity of many organisms. In 1969, it was reported that the anti-fungal activity in marine algae against *Candida albicans* and towards fungal strains<sup>4</sup>. Many habitats in which thermophilic fungi are likely to occur and grow have not yet been investigated or have received only cursory study<sup>20</sup>. Thermophilic fungi have been isolated from many habitats in the past few years. Most of these are heated habitats or have a past history of being heated, many habitats in which thermophilic fungi are likely to occur and grow have not yet been investigated or have received only cursory study, some habitats for example aquatic ones, are especially deserving of exploration<sup>20</sup>.

Researchers isolated a number of the thermophilic and thermotolerant fungi and actinomycetes from mushroom compost during peak heating<sup>7</sup>. They demonstrated that certain thermophilic fungi have been associated with the process of microbial thermogenesis<sup>20</sup>. It has been reported that anti-dermatophytic effect of (+) – crucephenol against the human pathogenic fungus *Trichophyton mentagrophytes*, as dermatophyoses are the most common form of fungal infection found in most countries, affecting skin, hair and nails<sup>22</sup>.

However, the present study is the first attempt made to study the fungi from volcanic soil as a novel anti-dermatophytic agent. The crude extract for *Penicillium* sp. was evaluated against the dermatophytes at different concentrations of 10µl, 25 µl and 50 µl and result shows that all the test concentration crude extract inhibited the growth of the dermatophytes. Hence the present investigation proved the *Penicillium*

sp. isolated from volcanic soil contains novel anti-dermatophytic compound.

The present investigation provides basic information on the anti- dermatophytic effects of the *Penicillium* sp. are isolated from extreme conditions. However, much detailed clinical trials are very essential for its commercial application. The extracts must also be performed to ensure the safety of the extracts.

**ACKNOWLEDGEMENTS:** Authors thank to Dean and Director, Faculty of Marine Sciences, Annamalai University, Parangipettai and to Dr. K. Sivakumar, for the facility given throughout the study period.

#### REFERENCES:

1. Aly, R: Ecology and epidemiology of dermatophyte infections. *J Am Acad Dermatol* 1994; 31:21-25.
2. Berkholder, P.R. and L.M. Burholder: Antimicrobial activity of horny corals. *Science* 1958; 127: 1174.
3. Brock: Thermophilic microorganisms and life at high temperatures. Springer – Verlag, New York 1978.
4. Burkholder, P.R. and Sharman, G. M: Antimicrobial agents from the se. *Lyloydia* 1969; 32: 40-51.
5. Denning, D. W., E. G. V. Evans, C.C. Kibbler, M.D. Richardson, M. M. Robertss, T.R. Rogers, D.W. Warnock and R.E. Warren: Guidelines for the investigation of invasive fungal infections in haematological malignancy and solid organ transplantation. *Eur J Clin Microbial Infect Dis* 1997; 16: 424-436.
6. Espinel-Ingroff. A., Kerkering. T.M., Goldson. P.R., Shadomy. S: Comparison study of broth macrodilution and broth microdilution antifungal susceptibility testing of yeast isolates. *Diagn Microbiol Infect Dis* 1991; 19:9–13.
7. Fergus, C.L: Thermophilic and thermotolerant molds and actinimycetes of mushroom compost during peak heating. *Mycologia* 1964; 56:267-284.
8. Frisvad, J.C: Physiological criteria and mycotoxin production as aids in identificastion of common asymmetric penicillia. *Appl Environ Microbiol* 1981; 41: 568-579.
9. Frisvad, J.C: Mycotoxins and Mycotoxigenic fungi in storage In: Sotred – grain Ecosystems (Jayys, D.S., White, N.D.G. and Muir, W.E., Eds.), Marcel Dekker, New York 1995; 251-288.
10. Frisvad, J.C. and O. Filtenborg: Terverticillate Penicillia; chemotaxonomy and mycotoxin production. *Mycologia* 1989; 81:837-861.
11. Frisvad, J.C., Thrane, U. and O. Filtenborg: Role and use of secondary metabolites in fungal taxonomy In: Chemical Fungal Taxonomy (Frisvad, J.C., Bridge, P.D. and Arora, D.K., Eds.), Marcel Dekker, New York 1998.289-319.
12. Gdom, R: Pathology of dermatophyte infections. *J Am Acad Dermatol* 1993; 28: 2-7.
13. Groll A.H., P.M. Shah, C. Mentzel., M. Schneider, G. Just-Neubling and K. Huebner: Trends in the postmortem epidemiology of invasive fungal infecdtion at a university hospital. *J Infect* 1996; 3.23-32.
14. Howard, d: Dermatomycoses. IN Barom, E(Ed), Medical Microbiology A Short Course. Wiley – Liss, New York 1994; 154.

15. Larsen, T.O. and J.C. Frisvad: Chemosystematics of *Penicillium* based on profiles of volatile metabolites. *Mycol Res* 1995; 99:1167-1174.
16. Raghukumar, S., C. Raghukumar and A. Rajendran: Abundance of Thraustochytrid fungi in the Arabian Sea. *Esur Coast Shelf Sci* 1990; 31:351-358.
17. Rinehart, K.L., P.D. Shaw, L.S. Shield, J.B. Gloer, G.C. Harbour, M.E.S Koker, D. Samain, R.E. Schwartz, A.A. Tymiak, E.G. Swynenberg, D.A. Stringfellow, J.J. Vavva, J.H. Coats, G.E., Zurenko, S.L. Kuentzel, L.H.Li, G.J Bakus, R.C. Brasca, I.I. Craft, D.N. Young and J.L. Connot: Marine natural products as a source of Antiviral, Antimicrobial and Antineoplastic agents. *Pur Appl Chem* 1981; 53: 795-817.
18. Shaw, P.D., W.O, McClure, G. Van Blaricom, J. Sims, W. Fenical and J. Rude: Antimicrobial activities from marine organisms, In *Food – drug from sea* 1976; 25:55-60.
19. Smith, G: An introduction to industrial mycology. Edward Arnold (Publishers) Ltd., London 1969; pp 329.
20. Tansey, M.R. and Brock, T.D: Microbial life at high temperatures: ecological aspects. In: *Microbial Life in Extreme Environments* (ed. D. Kushner). Academic Press, London 1978; 159-216.
21. Tanaka, S: Advance in dermatophytes and dermatophytosis j. *Med vet mycol*, vol. 30 (Suppl. 1) 1992; pp 29-39
22. Wagner, D. K. and P.G. Sohnle: Cutaneous defenses against dermatophytes and yeasts. *Clin microbial Rev*, 1995; 8: 317-335.
23. Weitzman, I. and Summerbell R.C: The dermatophytes. *Clin microbial Rev*, Vol. 8, 1995; 240 – 259.

\*\*\*\*\*