



Received on 23 February 2022; received in revised form, 29 November 2022; accepted 29 November 2022; published 01 December 2022

## ARSENIC-INDUCED ALTERATIONS IN GLYCOGEN AND LIPID DURING THE OVARIAN CYCLE OF A FRESHWATER SILUROID, *MYSTUS (M.) VITTATUS* (BL.)

Anuradha Shukla<sup>1</sup>, Yogendra Kumar Payasi<sup>\*2</sup> and J. P. Shukla<sup>2</sup>

P. G. Department of Zoology<sup>1</sup>, Jhun-Jhun Wala PG. College, Faizabad, Ayodhya - 224001, Uttar Pradesh, India.

Department of Zoology<sup>2</sup>, Indira Gandhi National Tribal University, Amarkantak - 484886, Madhya Pradesh, India.

### Keywords:

*Mystus (M.) vittatus*, Preparatory phase, Spawning phase, Post-spawning phase, Trivalent arsenic, Ovarian cycle, Glycogen, Lipid

### Correspondence to Author:

**Yogendra Kumar Payasi**

PG Department of Zoology, IGNTU  
Amarkantak - 484886, Madhya  
Pradesh, India.

**E-mail:** [yogendra.payasi@igntu.ac.in](mailto:yogendra.payasi@igntu.ac.in)

**ABSTRACT:** Heavy metals get their way into the environment through a wide spectrum of natural and anthropogenic sources. Arsenic (As) has been reported to be present in main too different oxidation states ( $As^{3+}$  and  $As^{5+}$ ). Trivalent arsenic ( $As^{3+}$ ) has been reported to be more toxic than the pentavalent ( $As^{5+}$ ) one. *Mystus (M.) vittatus*, a siluroid fish, were exposed to Sublethal concentration (SLC) of  $As^{3+}$  to observe alteration in the glycogen and lipid content during three different phases of the ovarian cycle preparatory, spawning and post-spawning. The increasing order in the glycogen and lipid content was observed during the preparatory and spawning phase. Still, decrease in these biochemical parameters was during post-spawning phase of ovarian cycle in *Mystus (M.) vittatus*, a freshwater siluroid. Less significant alteration in glycogen and lipid was noticed after 15 days of exposure, but a highly significant decrease was observed after 30 days in SLC of trivalent arsenic as  $AsCl_3$ . The causes for decline in these biochemical parameters have been discussed during the phases of ovarian cycle.

**INTRODUCTION:** Heavy metal pollution has become a serious environmental and public health hazard. It is because of the concentration of metallic pollutants released into the different sections of the environment from various industrial processes. These are often concentrated because of their bio-accumulative and non-biodegradable features. Heavy metals constitute a core group of aquatic pollutants<sup>1, 2</sup>. Their high toxicity even at low concentrations, may exert cumulative toxic effects in a wide variety of fish fauna and another aquatic biota<sup>3</sup>.

Heavy metals are introduced into the environment through a wide spectrum of natural sources such as volcanic activities, erosion and anthropogenic ones, including industrial waste release and leakage. Certain metallic pollutants such as chromium, arsenic, nickel, cadmium, mercury, *etc.*, exert toxic effects on living biota even at low concentrations, whereas zinc, manganese, copper, *etc.*, produce toxic effects on living biota only at higher concentrations<sup>4-6</sup>.

In the aquatic environment, fishes appear to be remarkable bio indicators of arsenic toxicity<sup>7-8</sup>. Allen and Rana<sup>9</sup> reported that toxicity of arsenical compounds depends upon species, sex, age, dose, duration of exposure, organic or inorganic form, valency state, *etc.* Arsenic has been reported to be present in two main different oxidation states (+3 and +5). Trivalent arsenic ( $As^{+3}$ ) has been observed to be more deleterious than pentavalent arsenic<sup>8, 10-</sup>

<p><b>QUICK RESPONSE CODE</b></p> 	<p><b>DOI:</b> 10.13040/IJPSR.0975-8232.13(12).5207-10</p> <hr/> <p>This article can be accessed online on <a href="http://www.ijpsr.com">www.ijpsr.com</a></p> <hr/> <p>DOI link: <a href="http://dx.doi.org/10.13040/IJPSR.0975-8232.13(12).5207-10">http://dx.doi.org/10.13040/IJPSR.0975-8232.13(12).5207-10</a></p>
---	--

<sup>14</sup>. Even though the toxicity of arsenic (+3) in aquatic biota, particularly fishes and terrestrial animals in general, has been enormously documented <sup>3, 12-13, 15</sup>. However, the deleterious effects of trivalent arsenic on the glycogen and lipid during different phases of fishes' ovarian cycle are scarce; hence, the present study has been undertaken. It is a well-known fact that lipid metabolism dysfunction is a risk factor for cardiovascular diseases and arsenic exposures affect lipid and glycogen metabolism.

**MATERIALS AND METHODS:** Adult specimens of *Mystus (M.) vittatus* (Bl.) were collected from a local lake having a weight of 92.38±4.48gm during the preparatory phase (January to April); Spawning phase (April to August) and post-spawning phase (September to December). They were acclimatized in laboratory tap water having pH=7.2±0.02; temperature = 22.4±2.2°C; DO= 6.2±40.52 mg/l; hardness as CaCO<sub>3</sub> = 126.62±3.6 mg/l. The methods outlined by APHA16 analyzed the physicochemical features of tap water. The sublethal concentration of analytic grade of trivalent arsenic (11.24 mg/l) as AsCl<sub>3</sub> was detected for long-term experimentation (15 and 30 days) as outlined by Shukla and Pandey <sup>17</sup>. The control and experimental media were aerated 3-5 h daily using stone diffuser, though *Mystus (M.) vittatus* is hardy air-breathing fish. 20 specimens were placed each in control and experimental media. The total glycogen and lipid in the ovary during preparatory, spawning and post-spawning phases were estimated by adapting the

methods outlined by Kemp and Pandey *et al* <sup>18-19</sup>. The data obtained in our study were statistically analyzed for significance by Student's 't' test as proposed by Fischer *et al.* <sup>20</sup>. A p-value of 0.05 or less was noticed as significant between the control and experimental groups.

**RESULTS & DISCUSSION:** Reproductive cycle includes the gonadal cycle, which is a dynamic process always in the state of gametogenesis. SLC of trivalent arsenic produced less significant dimension in the glycogen and lipid content during preparatory, spawning, and post-spawning phases under 15 days exposure than control. But a clear significant decrease in glycogen and lipid content was noticed after 30 days of exposure during all the phases. From **Table 1**, it becomes clear that the level of glycogen and lipid in the ovary during its control spawning phase of the ovarian cycle was maximum in comparison to a preparatory and post-spawning phase which indicates the possible supply of carbohydrate and lipid content in the form of glucose and lipid derivatives for active maturation of ova. The increasing order of decrease in glycogen content during preparatory and spawning phases of ovarian cycle of *Mystus (M.) vittatus*, a siluroid fish may be due to its enhance utilization as an immediate source to meet energy demands for maturation of ova under trivalent arsenic stress. It could also account for the prevalence of hypoxic or anoxic condition of the trivalent arsenic stress which generally enhances glycogen utilization in one way or another <sup>21-25</sup>.

**TABLE 1 TOTAL GLYCOGEN AND LIPID CONTENT IN MG/GM DRY WEIGHT OF OVARY DURING ITS DIFFERENT PHASES UNDER SLC OF TRIVALENT ARSENIC IN MYSTUS (M.) VITTATUS. EACH VALUE REPRESENTS MEAN +SE OF 5 OBSERVATIONS**

Content	Parameters	Control	15 days exposure	% Change	30 days exposure	% Change
Preparatory	Glycogen	24.12±1.02	22.04±0.84**	8.62	18.82±1.04***	21.97
	Lipid	76.22±1.02	70.82±1.38*	7.08	66.26±1.24**	13.06
Spawning	Glycogen	30.34±1.04	28.34±0.84**	6.59	22.22±1.04****	26.76
	Lipid	94.72±1.84	86.28±1.44**	8.91	80.52±1.34***	14.99
Post-spawning	Glycogen	14.12±0.60	12.46±0.52**	11.75	11.04±0.60***	21.81
	Lipid	40.08±1.06	37.68±1.36*	5.98	34.88±1.02**	12.97

\* = p>0.05 (insignificant) \*\* = p<0.05 \*\*\* = p<0.01 \*\*\*\* = p <0.001

Arsenic interferes with phosphate binding sites in adenosine triphosphate (ATP) resulting in the formation of ADP-arsenate which inhibits metabolic pathways and requires ATP. Glucose 6-phosphate is an essential mediator for glycolysis, glycogenesis, gluconeogenesis, glycogenesis and

the pentose phosphate pathways (PPP), also called an HMP shunt. Acute arsenic toxicity may be associated with hepatic necrosis and elevated levels of liver enzymes. The dimension in the lipid content during different phases of the ovarian cycle of *Mystus (M.) vittatus* (Bl.), a freshwater siluroid

might be partly due to its utilization in cell repair and tissue organization with the formation of lipoprotein, which is a salient constituent of the cell membrane and cytoplasmic organelles<sup>26-29</sup>.

Our findings may well be correlated with the observation made by Vutukuru<sup>1-2</sup>, Shukla, et al.<sup>25</sup>, and confirms that long-term exposure to trivalent arsenic interferes with the fishes' ovarian physiology.

**CONCLUSIONS:** Least significant alteration ( $P>0.05$ ) was observed in the biochemical parameters (Glycogen and lipid) after 15 days of exposure to SLC of trivalent arsenic during different phases (preparatory; spawning and post-spawning) of ovarian cycle of *Mystus (M.) vittatus* (Bl.). However, more or less significant alteration ( $P<0.05-0.001$ ) in glycogen and lipid content has been observed during selected different phases of ovarian cycle when exposed to SLC of trivalent arsenic after 30 days. Our finding reveals that trivalent arsenic may interfere in Oogenesis by interfering with the lipid, glycogen, and lipid metabolism.

**ACKNOWLEDGEMENTS:** The author thanks CST (UP) for financial assistance vide letter no. CST/D-265 dt. 14.5.2015.

**CONFLICTS OF INTEREST:** Nil

## REFERENCES:

1. Vutukur SS: Chromium induced alterations in some biochemical profiles of the Indian major carp, *Labao rohita* (Ham). Bull. Environ Contam Toxicol 2003; 70(1): 118-123.
2. Vutukur SS: Acute effects of hexavalent chromium on survival, oxygen consumption, hematological parameters and some biochemical profiles of Indian major carp, *Labao rohita*. Int J Environ Res Public Health 2005; 2(3): 456-462.
3. Storelli MM, Isabarove G, Storelli A and Macrotrigiano GO: Trace metals in tissue of Mugilids (*Mugil aratus*, *Mugil capito* and *Mugil labrus*) from the Mediterranean sea. Bull Env Conam Toxicol 2006; 77: 43-50.
4. Cohen T, Hee S and Ambrase R: Trace metals in fish and invertebrate of three California coastal wetlands. Mal Pollu Bull 2001; 42: 224-232.
5. Karadede AH and Unlu E: Heavy metal concentration in water, sediment, fish and some benthic organism from Tigris river. Turkey Env Monit Assess 2007; 131: 323-327.
6. Yilmaz AB, Cemal T and Tashin T: Uptake and distribution of hexavalent chromium in tissue gill, skin and muscle of a freshwater fish, Tilapia, (*Oreochromis aureus*). Env Chem & Ecotoxicol 2010; 2(3): 28-33.

7. Gertofer M, Poert M. Schramm Muller M & Rriebskorn R: Ultra-structural bio-markers as tools to characterize the health states of fish in contaminated streams. J Aqua Ecosyst Stress Res 2001; 8: 241-260.
8. Ghosh D, Bhattacharya S and Mazumdar S: Long term exposure to arsenic effects to head kidney and impair humoral immune response of *Clarias sarrachus*. Aquat Toxicol 2007; 81: 79-89.
9. Allen T and Rana SV: Effect of arsenic on glutathione dependent enzymes in liver and kidney of fresh water fish *Channa punctatus*. Biol Trace El Res 2004; 100: 39-48.
10. Bears H, Chards R and Chitti JGPM: Arsenic exposure alters hepatic and stress mediated gene expression in the common killfish, *Fundulus heteroclitus*. Aquat Toxicol 2006; 77: 257-266.
11. Kovandon KS, Jananathan S and Saranamam M: Expression of malathion in liver and Kidney of freshwater Vineer fish, cyprinus carpio var communis (Linn) exposed to arsenic. trioxide Ame J Sci Indus Res 2013; 4(1): 1-10.
12. Shukla Anuradha and Shukla JP: Toxic impact of arsenic on the blood pyruvate level in the fingerlings of a freshwater siluroid, *Mystus (M.) vittatus* (Bl). The GI J Env Sci & Res 2016; 3: 59-62.
13. Shukla Anuradha and Shukla JP: Succinate dehydrogenase activity as an index of trivalent arsenic toxicity in fingerlings of tropical fresh water siluroid *Mystus M. vittatus* (Bl). Int J Curr Trends in Sci & Tech 2017; 8(1): 20482-20486.
14. Shukla Anuradha and Shukla JP: Distillery effluent induced alterations in the nucleic acids and protein during testicular cycle of *Colisa fasciata* (Bl. & Schn.) a tropical freshwater perch. Int J Pharma & Bio Sci 2011; 3(1): 532-537.
15. Vankataram Raddy V, Vutukutu SS and Tchounnam PB: Ecotoxicology of hexavalent chromium in freshwater fish. Rev Environ Health 2009; 24(2): 129-145.
16. APHA, Standard Methods for examinations of water and waste water, 21st Edition, Washington DC 2005.
17. Shukla JP and Pandey K: Toxicity and long-term effects of a sublethal concentration of cadmium on the growth of the fingerlings of *Ophiocephalus punctatus* (Bl.). Acta Hydrochim Hydrobiol 1988; 16(5): 537-540.
18. Kemp AJM and Kits VH: A calorimetric micromethod for the determination of glycogen. J Biochem 1994; 56: 640-648.
19. Pandey SV, Khan AP and Subhranmanayam TAV: Micro-determination of lipids and serum fatty acids. Anal Biochem 1963; 6(5): 120-125.
20. Fischer RA: Statistical methods for research work. 13 Eds. Oliver and Boyd, London. 1983; 122-125.
21. Dezwaan A and Zondee DI: Body distribution and seasonal changes in glycogen content of the common sea mussel, *Mytilus edulis*, Comp. Biochem Physiol 1973; 43:53-55.
22. Geetha S, Suryanarayan HBI and Nair NB: On the nature of variations of biochemical constituents during the breeding cycle in *Heteropneustes fossilis* Proc. Nat Acad Sci India 1991; 61(2): 311-315
23. Ozretic B and Ozretic KM: Plasma sorbitol dehydrogenase, glutamate dehydrogenase, and alkaline phosphatase as potential indicators of liver intoxication in grey mullet (*Mugil auratus*). Bull Environ Contam Toxicol 1993; 50: 586-592.
24. Shukla JP, Tripathi S, Chandel BS and Dwivedi ND: Deleterious effects of arsenic on blood pyruvate level in a

- tropical freshwater fish *Colisa fasciatus*. National Journal of Life Sciences 2005; 1: 125-126.
25. Shukla JP, Shukla Anuradha and Dubey RK, Deleterious effects of hexavalent chromium on the blood puruvate level in the fingerlings of *Channa punctatus* (BI) a Freshwater murrel. International Journal of Pharma and Bio Sciences 2012; 3(4) 789-794.
26. Harper AH: Review of biochemistry. 20th Eds. Large Medical Publication Co. California 1983; 10 12.
27. Y annum Z, Meng Li, Xiaolim T, Ji axin X, Peng hul L and Xiaodong Y: Effects of Arsenic Exposure on Lipid metabolism, A systematic review and Meta-analysis. Toxicol Mechanism and Method 2021; 31(3): 188-196.
28. Dong Z, Li C, Yin, C, Xu M, Lu S, Gao M and LncRNA PU: 1 AS regulated arsenic induced lipid metabolism through EZ H2/Sirt6/SREBP-1c pathway. J Envorn Sc 2019; 85: 138-146.
29. Kermi UT, Sunkam NR and Das B: Effects of Arsenic and Lead and glycogen content and on the activities of selected enzymes invoved in carbohydrarte metabolism in fresh water cat fish, heteropneustes Follsilis. Int Acuatic Reas 2019; 11: 253-266.

**How to cite this article:**

Shukla A, Payasi YK and Shukla JP: Arsenic induced alterations in glycogen and lipid during ovarian cycle of a freshwater siluroid, *Mystus (M.) vittatus* (BL.). Int J Pharm Sci & Res 2022; 13(12): 5207-10. doi: 10.13040/IJPSR.0975-8232.13(12).5207-10.

All © 2022 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **Android OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)