IJPSR (2013), Vol. 4, Issue 10







Received on 13 May, 2013; received in revised form, 17 June, 2013; accepted, 15 September, 2013; published 01 October, 2013

A VALIDATED RP-HPLC METHOD FOR THE ESTIMATION OF ANAGRELIDE IN CAPSULE DOSAGE FORMS

K. Sujatha*¹, C. Palavan², J.V.L.N. Seshagiri Rao³

Government Polytechnic¹, Visakhapatnam, Andhra Pradesh, India College of Pharmaceutical Sciences, Andhra University², Visakhapatnam, Andhra Pradesh, India Yalamarty College of Pharmacy³, Visakhapatnam, Andhra Pradesh, India

Keywords:

Anagrelide, Estimation, Capsules, HPLC

Correspondence to Author:

K. Sujatha

Senior Lecturer, Department of Pharmacy, Government Polytechnic, Visakhapatnam – 530 007, Andhra Pradesh, India

Email: sujikandi@yahoo.in

ABSTRACT: An accurate and precise high performance liquid chromatographic method was developed for quantitative estimation of anagrelide in capsule dosage forms. Chromatographic separation of the drug was achieved on a Kromosil C_{18} column (150 x 4.6 mm; 5µ) by isocratic elution using a mobile phase consisting of phosphate buffer (pH 2.5) and acetonitrile (75:25 v/v) at a flow rate of 1.2 mL/min. The drug in the eluate was monitored by UV detection at 250 nm. Under optimized conditions, the retention time obtained for the drug was 4.818 min. The calibration curve was linear over the range of 50-150 μ g/mL of the drug. The recovery of drug by the proposed method was found to be 98.95% to 100.45%. The performance of the method was validated as per ICH guidelines. The method was also found to be applicable for determination of anagrelide in its capsule dosage forms without any interference from the excipients. The proposed method is suitable for routine quality control analysis of anagrelide.

INTRODUCTION: Anagrelide hydrochloride (6, 7-dichloro-1, 5-dihydroimidazo [2, 1-b] quinazolin-2(3H)-one monohydrochloride monohydrate) is an antithrombotic and platelet-reducing agent used for the treatment of essential thrombocytosis ¹. The drug is especially useful in managing chronic myeloid leukemia². The mechanism by which anagrelide reduces blood platelet count is still under investigation. Studies in patients support a hypothesis of dose-related reduction in platelet production resulting from a decrease in megakaryocyte hypermaturation 3 .





FIGURE 1: STRUCTURE OF ANAGRELIDE

A literature survey revealed that very few analytical methods have been reported for the determination of anagrelide in biological samples and in pharmaceutical dosage forms using LC-MS⁴ and HPLC ⁵⁻⁷ techniques. We have developed a new accurate and precise RP-HPLC method with short retention and run times for the determination of anagrelide in raw drug samples and in capsule dosage forms. The developed method has been validated as per ICH guideline ⁸.

International Journal of Pharmaceutical Sciences and Research

MATERIALS AND METHODS:

Drugs, chemicals, and solvents: The reference sample of anagrelide was obtained from Aizant drugs, Hyderabad. The commercial capsule formulation of anagrelide (0.5 mg), manufactured by Cipla Ltd., Goa, India, was used for the study. Potassium dihydrogen phosphate, ortho phosphoric acid and HPLC grade acetonitrile, methanol and water used in the experimentation were purchased from Rankem Laboratories.

Equipment and chromatographic conditions: A Waters Alliance liquid chromatograph (Model

2695) fitted with a U.V. diode array detector (Model 2996) and running on Empower2 data handling system was employed in the study. A Kromosil C₁₈ column (150 x 4.6 mm; 5 μ m) was used for analyzing the drug. All chromatographic runs were carried out in isocratic mode by using a mobile phase consisting of phosphate buffer (pH 2.5) and acetonitrile at a flow rate of 1.2 mL/min. The injection volume of the samples was 10 μ L. The detector wavelength was set at 250 nm. Under these optimized conditions, the retention time obtained for anagrelide was 4.818 min.



FIGURE 2: A REPRESENTATIVE CHROMATOGRAM OF THE WORKING STANDARD SOLUTION OF ANAGRELIDE

Phosphate buffer (pH 2.5): The phosphate buffer was prepared by dissolving 6.8 g of Potassium dihydrogen ortho phosphate in 1000 mL of water and adjusting the pH of the solution to 2.5 using ortho phosphoric acid. It was then sonicated and filtered through a 0.45μ membrane filter.

The mobile phase: The mobile phase consisted of a mixture of the phosphate buffer (pH 2.5) and acetonitrile in the ratio of 75:25 v/v.

The diluent: A mixture of the phosphate buffer (pH 2.5) and acetonitrile in the ratio of 50:50 v/v was used as the diluent.

Working standard solution of anagrelide: 10 mg of anagrelide reference standard was accurately weighed and transferred into a 10 mL volumetric flask. To this, 2 mL of 0.1N NaOH solution was added and the contents shaken to dissolve the drug. The volume was made up with methanol. This was used as the standard stock solution.

The working standard solution (100 μ g/mL) was prepared by transferring 1.0 mL of the standard stock solution into a 10 mL volumetric flask and diluting to volume with water.

Calibration curve: Linearity study was done by preparing solutions of anagrelide at different concentration levels including the working standard concentration mentioned above. Ten micro litres of each concentration (n=3) was injected into the HPLC system. The response was read at 250 nm and the corresponding chromatograms were recorded. From these chromatograms, the mean peak areas at each concentration level were calculated and a linearity plot of the mean peak area over concentration was constructed.

Estimation of the drug from the capsule dosage forms: The contents of five capsules of Anagrelide (0.5 mg) were transferred into a 25 ml volumetric flask. To this, 15 ml of the diluent was added and sonicated for 25 min.

The volume was made up with the diluent and the contents were filtered through a 0.45μ nylon filter. Six replicate samples of this solution were then chromatographed. From the chromatograms

obtained, the average area of the drug peaks was noted. The drug content in the formulation was calculated from the calibration curve obtained with the standard drug dilutions.



FIGURE 3: A REPRESENTATIVE CHROMATOGRAM OF THE CAPSULE FORMULATION OF ANAGRELIDE

RESULTS AND DISCUSSION: During the method optimization studies, various combinations and proportions of the solvents and buffers were examined on the Kromosil C_{18} column for efficient separation of anagrelide. Using a mobile phase consisting of a mixture of phosphate buffer (pH 2.5) and acetonitrile in the ratio of 75:25 v/v, a good resolution and baseline separation of the drug

peak were obtained. All the chromatographic conditions were optimized after evaluating the column efficiency parameters like theoretical plates and tailing factor (**Table 1**). Under these optimized conditions, the retention time obtained for anagrelide was 4.818 min (**Figure 2**). The proposed method was also found to be applicable for the analysis of anagrelide in capsule formulations.

Stationary Phase	Kromasil C18 (150 x 4.6 mm, 5µm)		
Mobile Phase	Phosphate buffer : Acetonitrile =75:25 v/v		
Diluent	Phosphate buffer : Acetonitrile =50:50 v/v		
Flow Rate	1.2 mL/min		
Column Temperature	26°C		
Injection Volume	10 µL		
Detection Wavelength	250 nm		
Run Time	8 min		
Retention Time	4.818 min		

TABLE 1: OPTIMIZED CHROMATOGRAPHIC CONDITIONS

Specificity: A good analytical method should be able to measure the analytes accurately in the presence of probable interferences from matrix and excipients in the formulation. **Figure 2** shows good chromatographic baseline separation of anagrelide in working standard solution. **Figure 3** demonstrates that no interference was found at the retention time of anagrelide from the excipients in its dosage form.

Linearity: The calibration curve constructed for the drug was linear over the concentration range of $50 - 150 \ \mu\text{g/mL}$ (**Figure 4**). The regression of the plot was computed by the least squares method.

The relevant correlation coefficient is greater than 0.99 and the %RSD for each concentration studied (n=3) was less than 2.



International Journal of Pharmaceutical Sciences and Research

Accuracy and Precision: The accuracy of the method was determined by recovery experiments. The recovery studies were carried out and the percent recovery and standard deviation of the percent recovery were calculated and represented in **Table 2**. The high percentage of recovery indicates that the proposed method is highly accurate.

The precision of the method was demonstrated by inter-day and intra-day variation studies. Six replicate injections of sample solutions were made and the percent RSD was calculated and presented in **Table 3**. From the data obtained, the proposed RP-HPLC method was found to be precise.

|--|

Analyte	Amount of the analyte taken (µg/mL)	Mean recovery (µg/mL) ± SD	% Mean recovery ± SD
Anagrelide	50	50.23 ± 0.047	100.45 ± 0.094
	100	100.02 ± 0.935	100.02 ± 0.935
	150	148.43 ± 0.793	98.95 ± 0.529

S. No.	Intra-day precision	Inter-day precision
1	1239846	1216598
2	1223570	1225784
3	1219789	1217859
4	1219985	1235478
5	1224567	1224875
6	1231256	1244965
Average	1226502	1227593
SD	7752.927	10865.18
%RSD	0.63	0.88

System suitability parameters: System suitability parameters were studied with six replicates of the standard sample solution and the parameters are presented in **Table 4**.

ACKNOWLEDGEMENT: The authors are thankful to M/s Aizant Drugs, Hyderabad, for providing the reference sample of anagrelide.

REFERENCES:

 TABLE 4: SYSTEM SUITABILITY PARAMETERS OF

 THE PROPOSED METHOD

Parameter	Value
Retention time (min)	4.818
Tailing factor	1.1
Theoretical plates	4586
HETP	0.03271

Method suitability: Studies for recovery of anagrelide from its commercial capsule formulation (Agrylin) were carried out by the proposed method. The average value obtained from six replicate samples (99.6 % recovery) was found to be in good agreement with the labeled amount. This confirms the suitability of the method for the analysis of anagrelide in capsule dosage form.

CONCLUSION: The proposed RP -HPLC method is sensitive, accurate and precise and can be used for the routine quality control analysis of anagrelide in its capsule dosage forms. 1. Hussar DA: New drugs of 1997. Journal of the American Pharmaceutical Association (Washington, D.C.: 1996) 1998; 38:155-195.

- Voglová J, Maisnar V, Beránek M and Chrobák L: Combination of imatinib and anagrelide in treatment of chronic myeloid leukemia in blastic phase. Vnitřní Lékařství 2006; 52:819 – 822.
- 3. Petro E Petrides: Anagrelide: What was new in 2004 and 2005. Seminars in Thrombosis and Hemostasis 2006; 32:399-408.
- 4. Zhu Z, Gonthier R and Neirinck L: Highperformance liquid chromatography–mass spectrometry method for determination of anagrelide in human plasma. Journal of Chromatography B 2005; 822: 238-243.
- Kalaichelvi R and Jayachandran E. Development 5. and validation of a reversed-phase HPLC method for the determination of anagrelide in capsule. International Journal of Pharmaceutical Chemistry. Available at: <http://ijpc.ssjournals.com/index.php/journal/article/ view/30>. Date accessed: 15 Jun. 2013. doi:10.7439/ijpc.v3i1.30.

- 6. Venugopal V, Ramu G, Malleswara Rao NNV and Rambabu C: A new RP-HPLC method development for the estimation of an anti-thrombocythemic agent anagrelide in pure and pharmaceutical formulations. Der Pharma Chemica 2012; 4:1716-1722.
- 7. Sudhakar SP, Khader AMA and Seetharamappa J: Development and validation of a stability-indicating RP-HPLC method for the quantitative analysis of

How to cite this article:

anagrelide hydrochloride. Scientia Pharmaceutica 2012; 80: 567–579.

8. ICH Harmonized Tripartite Guidelines (Q2R1). Validation of analytical. Validation of analytical procedures: Text and Methodology. International Conference on Harmonization, European commission, Japan and USA :2005.

Sujatha K, Palavan C, Seshagiri Rao JVLN: A validated RP-HPLC method for the estimation of Anagrelide in capsule dosage forms. *Int J Pharm Sci Res* 2013: 4(10); 3920-3924. doi: 10.13040/IJPSR. 0975-8232.4(10).3920-24

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)