(Review Article)

IJPSR (2023), Volume 14, Issue 11



INTERNATIONAL JOURNAL

Received on 22 February 2023; received in revised form, 09 May 2023; accepted, 31 May 2023; published 01 November 2023

A REVIEW ON NIOSOMAL SUSPENSION CONTAINING CEFPODOXIME PROXETIL

M. Sharma^{*} and K. Bains

Himalayan Institute of Pharmacy, Kala-Amb - 173030, Himachal Pradesh, India.

Keywords:

Cefpodoxime proxetil, Anti-biotic, cephalosporins, Niosomes, Niosomal suspension, Oral liquid dosage form

Correspondence to Author: M. Sharma

Research Scholar, Himalayan Institute of Pharmacy, Kala-Amb - 173030, Himachal Pradesh, India.

E-mail: manjeetsharma047@gmail.com

ABSTRACT: Cefpodoxime proxetil is an oral antibiotic belonging to 3rd generation Cephalosporin category. It has shown its antibiotic activity against both Gram positive and Gram-negative bacteria and has high stability in the presence of β -lactamase except *Enterococci* species. The drug has found its effectiveness in pediatric patients in treating respiratory tract infections. At the present time the most familiar form in which cefpodoxime proxitel is delivered is either oral tablet form or for pediatrics oral dry syrup form. The oral dry syrup form has to be administered twice or thrice a day in order to meet the required dosage regimen. Developing the sustained release formulation for cefpodoxime proxetil probably niosomal suspension can be effective in reducing the dosing frequency along with sustained release and targeted drug delivery. Niosomal suspensions are suspended form of niosomes in an aqueous phase to form liquid dosage form it is called niosomal suspension. It is a potential drug carrier for delivery of drugs orally in liquid form. The niosomal suspensions allow the targeted drug delivery with controlled/sustained release and lead to reduced dose frequency with improved bioavailability and higher therapeutic effectiveness. This could make the formulation most suitable with lesser side effects and higher patient compliance.

INTRODUCTION: Niosome is a sustained release dosage type which is designed to sustain a constant level of release of substance in the bloodstream of the patient by releasing the drug over a prolonged period of time maintaining constant blood levels of the drug in the bloodstream increases therapeutic efficacy of the drug $^{1, 2}$. The aim in the design of safe delivery systems is to minimize the frequency of dosing or increase the efficacy of the medication through localization at the site of the operation, to minimize the dose needed or to ensure consistent distribution of the drug ³.



Ideal drug delivery systems will allow, first, a single dose for the duration of treatment, whether for days or weeks, as with infection, or for the lifetime of the patient, such as hypertension or diabetes ⁴. Second, it will send the active agent directly to the site of the operation, reducing the side effects.

The basic goal in the design of the dosage form is to optimize the delivery of drugs in order to calculate the regulation of the therapeutic effect in the face of unpredictable variations in the *in-vivo* environment in which the medication is released ⁵.

Sustained release mode of drug administration has certain features that have an important impact on the magnitude of the pharmacologic response:

1. It minimizes fluctuation in blood drug concentrations (i.e., between peak and trough).

- 2. It produces a slow input rate which tends to minimize the body's counteraction to the drug's intervening effect on regulated physiological processes.
- 3. It provides a continuous mode of drug administration ⁶.

For many drugs with non-concentration dependent pharmacodynamics, the exposure time, rather than the AUC, is the relevant parameter and it can therefore be optimized by sustained release preparations. Sustained (modified)-release formulation provides higher maximum plasma concentrations with lower inter-patient variability than the conventional, immediate release, twicedaily formulation ⁷. Additionally, therapeutic drug levels with sustained release formulations are achieved rapidly and maintained over the course of 24 h, allowing once-daily dosing. The studies have also confirmed good tolerability and safety of sustained-release formulations similar to the immediate-release formulations⁸.

Cefpodoxime Proxetil: Cefpodoxime proxetil is an oral, broad spectrum third generation antibiotic belonging to the class Cephalosporins. Cefpodoxime has shown its anti-biotic activity against both Gram positive and Gram-negative bacteria and has high stability in the presence of β lactamase except *Enterococci* species Cefpodoxime has in-vitro activity against Staphylococcus aureus, Gram positive bacterial species such as Streptococcus pyogenes and Streptococcus agalactiae and also against several Gram-negative bacterial species such as E. coli, K. pneumonia, Moraxella catarrhalis and Н. influenzae. Cefpodoxime proxetil is only well tolerated, 3rd generation cephalosporin that is available in oral form and low concentration of drug is useful in inhibiting most of the respiratory pathogens¹¹. Cefpodoxime proxetil has found its therapeutic effectiveness especially in paediatrics against respiratory tract infections (bronchitis and pneumonia), urinary tract infections and in mild to moderate cases of gonorrhoea. Cefpodoxime has shown its activity against S. pneumoniae but less when compared to amoxicillin and amoxicillinclavulanic acid combination¹².

Molecular Weight: 557.6 [g/mol]

Molecular Formula: C₂₁H₂₇N₅O₉S₂

Functional Category: Anti-bacterial agent

IUPAC Name: (RS)-1-(isopropoxycarbonyloxy)ethyl (+)-(6R, 7R)-7-[2-(2- amino-4- thiazolyl)-2-{(Z) – methoxy - imino} acetamido] – 3 – methoxymethyl -8-oxo-5-thia-1- azabicyclo[4.2.0] oct-2-ene-2-carboxylate

Description: Cefpodoxime proxetil is a prodrug of Cefpodoxime and semi-synthetic cephalosporin. It is white to light brownish in colour, with odourless or faint odour. It has good solubility in dehydrated alcohol, acetonitrile, methyl alcohol, slight solubility in ether and water ¹³.

Structural Formula:



Mechanism of Action: Cefpodoxime proxetil is a prodrug form of Cefpodoxime. When administered orally, the drug cleaves enzymatically to 2propanol, CO_2 and acetaldehyde. The original form of drug Cefpodoxime gets absorbed rapidly from the gut wall. It reaches the body fluids after absorption Minimum exceeding Inhibitory Concentration (MIC). Then the drug undergoes metabolism process of de-esterification by intestinal esterase. Cefpodoxime acts by inhibiting the bacterial cell wall synthesis and hence works as an effective bactericidal agent. Thereafter, it gets excreted from kidneys unchanged. Also. Cefpodoxime has lower molecular weight which allows the passage of drug through the pores present in the bacterial cell wall. Then, it crosses the periplasmic space and gets bound to Penicillin binding proteins (PBP1 and PBP3) in bacterial cell wall membrane. This binding affects the synthesis of cell membrane and damages the cell ¹⁴⁻¹⁶.

Pharmacokinetics: Pharmacokinetics describes and predicts the time course of drug concentrations in body fluids.

Absorption: Cefpodoxime proxetil in its active form (Cefpodoxime) gets absorbed from the gut wall. The extent of absorption is independent of the food administered. But administration of this drug with other active agent can lead to increase in gastric pH which ultimately reduces the absorption from the gut wall.

Distribution: Cefpodoxime distributes rapidly in the body fluids after absorption.

Excretion: The drug excretes in the unchanged form from the kidneys.

Metabolism: The metabolism of Cefpodoxime proxetil depends on the de-esterification by intestinal esterase. Cefpodoxime acts by inhibiting the bacterial cell wall synthesis and hence works as an effective bactericidal agent.

Elimination half-life and clearance: Mean elimination half-life ranges from 1.7 to 3.3 hours in younger patients. The clearance of Cefpodoxime proxetil depends on the age of the patient which is 0.57 L/h/kg in paediatrics and 0.36 L/h/kg in children of age above 5 years.

Bioavailability: Cefpodoxime proxetil is a prodrug, which is orally administered cephalosporin with only 50% absolute bioavailability.

Cefpodoxime Proxetil for Pediatric use: The drug has found its effectiveness in pediatrics patients in treating respiratory tract infections. At the present time the most familiar form in which cefpodoxime proxetil is delivered is either oral tablet form or for pediatrics oral dry syrup form. The oral dry syrup form has to administered twice or thrice a day in order to meet the required dosage regimen.

Developing the sustained release formulation for cefpodoxime proxetil can be effective in reducing the dosing frequency along with sustained release and targeted drug delivery and also improve the bioavailability of drug. This makes the formulation most suitable with lesser side effects and higher patient compliance. **Niosomes:** Niosomes are vesicles composed from non-ionic surfactants. Niosomes are microscopic vesicles of nano-range ranging from 10-1000 nm^{17-¹⁹. Niosomes are prepared by hydration of synthetic non-ionic surfactants along with incorporation of lipids and cholesterol. Niosomes are microscopic lamellar structures formed on admixture of a nonionic surfactant, cholesterol, and diethyl ether with subsequent hydration in aqueous media. Niosomes are formulated by self-assembly of non-ionic surfactants in organic solvent (non-aqueous media) as microscopic, spherical, polyhedral, and multilamellar structures²⁰⁻²³.}

Several non-ionic surfactants are used in formation of niosomes and these include Spans and Tweens, Polyglycerol alkyl ethers, polyoxyethylene alkyl ethers, glucosyl dialkyl ethers, ester and steroid linked surfactants *etc*²⁴. Niosomes are categorised into 3 types based on size and bilayers. These are:

- Small Unilamellar Vesicles (SUV) 10-100nm.
- Large Unilamellar Vesicles (LUV) 100-3000nm.
- Multilamellar Vesicles (MLV) ^{25, 26}.



FIG. 1: STRUCTURE OF NIOSOMES

Niosomes are vesicular drug delivery systems which are formulated in case of delivery of amphiphilic, hydrophobic or lipophilic drugs ²⁷. Niosomes have similar structure and activity as that of liposomes but niosomes are highly preferred for following reasons:

- Higher chemical stability.
- Highly suitable for lipophilic drugs or categorically for BCS Class II and IV.
- Easy to manufacture.
- Economical.
- Higher efficacy.
- Doesn't require special handling or storage conditions.
- Précised.
- Promising drug carrier²⁸.

Niosomal Suspension: When the niosomes are suspended in an aqueous phase to form liquid dosage form it is called niosomal suspension. It is a potential drug carrier for delivery of drugs orally in liquid form. The niosomal suspensions allow the targeted drug delivery with controlled/ sustained release and lead to reduced dose frequency with improved bioavailability and higher therapeutic effectiveness^{29, 30}.

Advantages of Niosomes/ Niosomal Suspension:

- Nontoxic and non-immunogenic formulations.
- Improved patient compliance.
- Higher therapeutic efficacy.
- Enhanced oral bioavailability of drug.
- Biocompatible.
- Good carrier option for amphiphilic and lipophilic drugs.
- Controlled and sustained release drug delivery.
- Osmotically active and stable.
- Drug protection against enzymatic metabolism.

- Higher stability.
- Targeted drug delivery.
- Enhanced skin permeability.
- Easy to manufacture, handle, store and transport.
- Can be administered orally, topically as well as through parenteral route ³¹.

Components of Niosomes: The essential components of niosomes are non-ionic surfactants, lipids such as cholesterol and hydration or aqueous medium.

Non-ionic Surfactants: The non-ionic surfactants as the name indicates are non charged surfactants which do not have charged groups in hydrophilic heads. Non-ionic surfactants are amphiphilic molecules that consist of 2 regions: Hydrophilic region (water soluble) and Hydrophobic region (lipid soluble) ³²⁻³⁵. Non-ionic surfactants are used for the preparation of niosomes because of several reasons:

- Non-toxic.
- Stable.
- Biocompatible.

Lipids: During the formulation of niosomes, the cholesterol forms hydrogen bond with the hydrophilic head of the non-ionic surfactant. Several properties of niosomes are dependent on the choice and content of cholesterol. The rigidity of the vesicles and stability of the vesicles improves with the increase in cholesterol content and permeability of the vesicles for entrapped drug decreases and hence inhibits leakage from the vesicles. The higher content of cholesterol also enhances entrapment efficiency ³⁶. Examples of lipids used for niosomes formulation are cholesterol and $1-\alpha$ -Soya phosphatidyl choline.

| TADLE 1, EXAMILLES OF MONTONIC SUNFACTANTS | | | | |
|--|--|---|--|--|
| S. no. | Non-ionic surfactant | Examples | | |
| 1 | Alkyl ethers (Alkyl glycerol ethers, Polyoxyethylene | Hexadecyl diglycerol ether, Brij 30, 52, 72, 76, 78 | | |
| | glycol alkyl ethers) | | | |
| 2 | Alkyl esters (Span and Tween) | Span 20, 40, 60, 80, 85 and Tween 20, 40, 60, 80, 85 | | |
| 3 | Alkyl amides (Glycosides and Alkyl polyglycosides) | C-Glycoside, Octyl-decyl polyglycoside | | |
| 4 | Fatty alcohols and Fatty acids | Stearyl alcohol, Cetyl alcohol, Myristyl alcohol; Stearic | | |
| | | acid, Palmitic acid, Myristic acid | | |
| 5 | Block copolymer | Pluronic L64, 105 | | |

TABLE 1: EXAMPLES OF NON-IONIC SURFACTANTS

Charged Molecules: Apart from lipids and nonionic surfactants, charged molecules are important components of niosomes that increase the stability of niosomes. The charged molecules increase surface charge density and prevent the aggregation of vesicles. Both negatively and positively charged molecules can be used for formulation of niosomes. Appropriate amount of charged molecules (2.5-5 mol %) is important as the higher concentration of charged molecules inhibits the formulation of niosomes 37 .

Method of Preparation: The method of preparation for niosomes is classified as follow:

| TABLE 2: METHOD | OF | PREPARATION OF NIOSOMES |
|-----------------|----|-------------------------|
| | _ | |

| S. no. | Type of niosomes | Method of preparation |
|--------|----------------------------------|--|
| 1 | Small unilamellar vesicles (SUV) | Sonication method Microfluidization method |
| 2 | Multilamellar vesicles (MLV) | Hand shaking method (Thin film hydration technique) Transmembrane pH |
| | | gradient technique |
| 3 | Large unilamellar vesicles (LUV) | Ether injection method. Reverse phase evaporation technique |
| 4 | Miscellaneous | Multiple membrane extrusion method. Emulsion method. Lipid injection |
| | | method |

Ether Injection Method: In this method, the nonionic surfactant along with other additives is added into diethyl ether (organic solvent) and further this solution is injected slowly through needle into aqueous drug solution which is maintained at constant temperature throughout. Consequently, the organic solvent is evaporated using rotary evaporator and single layered vesicles (niosomes) are formed ³⁷⁻⁴⁰.

Hand Shaking Method (Thin Film Hydration Technique): In this method, all the niosomal components are mixed in organic solvent in a RBF. Then, the organic solvent is evaporated using rotary evaporator and a thin film is obtained inside the walls of RBF. Furthermore, drug aqueous solution is added to RBF and the film is hydrated for specific time with constant shaking. This method of preparation is highly recommended for its ease of application and simplicity. This method allows the formation of multilamellar vesicles (MLV) Niosomes ⁴¹⁻⁴⁶.

Transmembrane pH Gradient Method: In this method of niosome preparation, the non-ionic surfactants and cholesterol are dissolved in organic solvent (chloroform) and then evaporated to form thin film on the walls of RBF. This film is then hydrated by addition of citric acid solution and the resulting mixture is freeze thawed. The above mixture is then hydrated by addition of aqueous drug solution and the pH is maintained in between 7-7.2^{9,11,37}.

Reverse Phase Evaporation Technique: In this method, the niosomes are prepared by dissolving

the niosomal components in organic solvent (ether and chloroform) and further adding aqueous drug solution. This mixture is sonicated until the organic solvent evaporates and an emulsion is formed. This method is useful in formulating Large unilamellar vesicles (LUV)³⁸⁻⁴⁰.

Microfluidization Technique: This method is performed in an interaction chamber whereby the non-ionic surfactant and drug is interacted at super high velocity. This high-speed impingement allows the formation of précised niosomes. This method is useful in formation of uniform, highly reproducible, small sized, and unilamellar vesicles ⁴¹.

Heating Method: In this method, the surfactant and cholesterol are hydrated separately in buffer solution and heated upto 120° C with constant stirring to dissolve cholesterol. The temperature of above cholesterol mixture is reduced, and both the solutions are mixed together with continuous stirring to form Niosomes^{8-11, 38}.

Bubble Method: In this method, the niosomal components and buffer are added to glass flask. All the components are dispersed at 70° C and mixed in homogenizer. Thereafter, the flask is kept in water bath and nitrogen gas is bubbled or passed through the flask in order to form large unilamellar vesicles (LUV)⁴²⁻⁴⁵.

Sonication Method: In this method, the niosomal components (surfactant and cholesterol) and aqueous solution of drug are mixed and sonicated at 60° C for 3 minutes in order to form niosomes.

This method is useful in formation of uniform sized and small unilamellar vesicles $(SUV)^{46}$.

Multiple Membrane Extrusion Method: In this method, the niosomal components and diacetyl phosphate are added into chloroform and evaporated to form thin films.

This film is hydrated with aqueous drug solution and the mixture is extruded through polycarbonate membrane to obtain small controlled and uniform sized Niosomes ^{17-20, 56}.

Emulsion Method: In this method, oil in water (o/w) emulsion is formed using organic solvent, surfactant, cholesterol and aqueous drug solution. Then, the organic solvent is evaporated resulting in formulation of niosomes in aqueous phase ¹⁸.

Lipid Injection Method: In this method, the lipid and surfactant mixture is melted and injected into hot aqueous drug solution which is agitated. The drug is further dissolved in above mixture and niosomes are formed. The organic solvent is not used in this method ¹⁸.

Proniosomes: Proniosomes are dry formulations which are prepared by coating water soluble carriers such as sorbitol and mannitol with surfactant. The proniosomes are prepared because they are highly stable when compared to niosomes.

The proniosomes are hydrated before use to form niosomes. This method allows the formulation of physically stable proniosomes that do not leak, aggregate, and are easier to manufacture, store and transport when compared to Niosomes^{17, 18, 57}.

Characterization of Niosomal Suspension

Appearance: Prepared formulation will be visually checked for its colour, clarity and phase separation ⁵⁷.

Drug Content: The drug content of formulation will be determined by diluting 1ml of the formulation with 100 ml methanol, further diluted 5ml to 50ml with methanol followed by analysis with UV-visible spectrophotometer ⁵⁸.

Rheological Study: The viscosity of Niosomal suspension of different formulation will be measured at 10 rpm for 3 min at 25°C by Brookfield type rotary viscometer with spindle 63.

SEM: The suspension will be determined for the morphology using Scanning Electron microscopy.

Entrapment Efficiency: The Niosomal suspension is sonicated and centrifuged, and the drug is separated. Assay is done at optimum wavelength using UV spectrophotometer.

In-vitro drug release: The *in-vitro* drug release study is carried out in two medium: Phosphate buffer and gastrointestinal pH medium ^{59, 60}.

Activation of Cellophane Membrane: The cellophane membrane is activated by dipping firstly in hot water and then in ethanol for 1 hour and 30 minutes respectively. Further, the membrane is kept in acetate buffer (pH 4) overnight.

In-vitro **Drug Release in Gastrointestinal pH Medium:** 3ml of Niosomal suspension is taken on activated cellophane membrane tied to open ended cylinder. The apparatus is kept in 30ml of 0.1N HCl and stirred at 37°C. Samples are withdrawn after 5 minute interval for 2 hours and analysed using UV spectrophotometer.

In-vitro **Drug Release in Phosphate bBuffer Medium:** The above mentioned procedure is used for determination using phosphate buffer solution pH 7.4.

In-vitro Anti-microbial Activity: The *in-vitro* antimicrobial activity is performed using agar diffusion assay method. The microorganism employed is E. coli. The activity is performed on optimized formulation and plain Niosomal suspension. Petri plate is incubated at 37°C for 24h. The sensitivity of test organism to the formulation will be indicated by clear zone of inhibition around the disc and the diameter of the zone of inhibition is measured.

Stability Studies: Samples (triplicate) will be placed in flasks and air tightened completely. The samples will be submitted to a thermostable hot air oven at $45 \pm 2^{\circ}$ C for 90 days.

Control samples will be kept at room temperature for the same period of time. The evaluation of the samples are performed initially at time zero and after 15, 30, 60, and 90 days and evaluated for organoleptic parameters (colour, odor, and appearance) and drug content $^{61-63}$.

CONCLUSION: In a nutshell to conclude, 3^{rd} proxetil is generation cefpodoxime cephalosporin used as an antibiotic. It has found its effectiveness in the treatment of respiratory tract infection in pediatrics. The oral dry syrup form of drug available in market is therapeutically active but has to be administered twice a day depending on the severity of the disease. Here, the approach is the niosomal suspension studied that of cefpodoxime proxetil could be an effective approach in delivering the drug with sustained or controlled release. The niosomal suspension is an effective approach with appropriate delivery strategies and further research can lead to a novel approach in the antibiotics category.

ACKNOWLEDGEMENT: I would like to express my special thanks of gratitude to my guide Mrs. Kaushalya Bains for their able guidance and support in completing this review.

CONFLICTS OF INTERESTS: Declared None

REFERENCES:

- 1. Adepu S and Ramakrishna S: Controlled Drug Delivery Systems: Current Status and Future Directions. Molecules 2021; 26(19): 5905.
- 2. Habet S: Narrow Therapeutic Index drugs: Clinical pharmacology perspective. Journal of Pharmacy and Pharmacology 2021; 73(10): 1285-91.
- 3. Karna S, Chaturvedi S, Agrawal V and Alim M: Formulation approaches for sustained release dosage forms: A review. Asian Journal of Pharmaceutical and Clinical Research 2015; 8(5): 34-41.
- 4. Kube RS, Kadam VS, Shendarkar GR, Jadhav SB and Bharkad VB: Sustained release drug delivery system: review. Indian Journal of Research in Pharmacy and Biotechnology 2015; 3(3): 246-51.
- Mali RR, Goel V and Gupta S: Novel Study in Sustained Release Drug Delivery System: A Review. International Journal of Pharmaceutical and Medicinal Research 2015; 3(2): 204-15.
- Pareek S, Kumawat S, Sharma V, Sharma D, Rathore D, Agarwal M and Pawan S: Review on Sustained Release Technology. International Journal of Pharmaceutical and Biological Science Archive 2019; 7(6): 29-38.
- 7. Patel D and Patel HK: A Review on Sustain Release Drug Delivery System. International Journal of Pharmacy and Pharmaceutical Research 2020; 18(1): 412-27.
- 8. Ojha AK and Verma S: A Review Sustained Release Drug Delivery Technology. World Journal of Pharmacy and Pharmaceutical Sciences 2018; 7(5): 250-60.
- 9. Zhang G, Fu Z & Sun X: Research progress of Cefpodoxime proxetil application in pediatrics. Chinese Journal of Applied Clinical Pediatrics 2019; 1429-1433.

- Arumugham VB, Gujarathi R and Cascella M: Third Generation Cephalosporins. In: StatPearls. Treasure Island (FL): StatPearls 2022. Available from: https://www.ncbi.nlm.nih.gov/books/NBK549881/
- 11. Bui T & Preuss CV: Cephalosporins. In StatPearls [Internet]. StatPearls Publishing 2022.
- 12. Frampton JE, Brogden RN, Langtry HD and Buckley MM: Cefpodoxime proxetil: A review of its antibacterial activity, pharmacokinetic properties and therapeutic potential. Drugs 1992; 44(5): 889-917.
- 13. Pahwa R, Rana A, Dhiman S, Negi P and Singh I: Cefpodoxime Proxetil: An Update on Analytical, Clinical And Pharmacological Aspects. Journal of Current Chemical and Pharmaceutical Sciences 2015; 5(1): 56-66.
- 14. Makabenta JMV, Nabawy A, Li CH, Schmidt-Malan S, Patel R & Rotello VM: Nanomaterial-based therapeutics for antibiotic-resistant bacterial infections. Nature Reviews Microbiology 2021; 19(1): 23-36.
- Babu NR, Latha K., Padmavathi Y, Padmavathi R, Preethi, Y, Rama A & Ashwini V: Development and Validation of Bioanalytical Spectrophotometric Method for Pharmacokinetic Study of Cefpodoxime Proxetil Microemulsion in Rats. YMER 2022; 22(11): 948-957.
- Pahwa R, Rana A, Dhiman S, Negi P and Singh I: Cefpodoxime Proxetil: An Update On Analytical, Clinical And Pharmacological Aspects. J Curr Chem Pharm Sci 2015; 5(1): 56-66.
- 17. Bhardwaj P, Tripathi P, Gupta R & Pandey S: Niosomes: A review on niosomal research in the last decade. Journal of Drug Delivery Science and Technology 2020; 56: 101581.
- Yasamineh S, Yasamineh P, Kalajahi HG, Gholizadeh O, Yekanipour Z, Afkhami H & Dadashpour M: A state-ofthe-art review on the recent advances of niosomes as a targeted drug delivery system. International Journal of Pharmaceutics 2022; 121878.
- Ge X, Wei M, He S & Yuan WE: Advances of non-ionic surfactant vesicles (niosomes) and their application in drug delivery. Pharmaceutics 2019; 11(2): 55.
- 20. Ag Seleci D, Seleci M, Walter J, Stahl F and Scheper T: Niosomes as nanoparticular drug carriers: fundamentals and recent applications. J Nanomater 2016; 2016(3): 1-16.
- 21. Kumavat S, Sharma PK, Koka SS, Sharma R, Gupta A & Darwhekar GN: A review on niosomes: potential vesicular drug delivery system. Journal of Drug Delivery and Therapeutics 2021; 11(5): 208-212.
- 22. Durak S, Esmaeili Rad M, Alp Yetisgin A, Eda Sutova H, Kutlu O, Cetinel S and Zarrabi A: Niosomal drug delivery systems for ocular disease-recent advances and future prospects. Nanomater Basel 2020; 10(6): 1191.
- 23. Kalpesh C, Ashara K, Paun J, Soniwala M, Chavada J, Nathawani S, Mori N and Mendapara V: Vesicular Drug Delivery System: A Novel Approach. Mintage J Pharm Med Sci 2014; 3(3): 1-14.
- 24. Chen S, Hanning S, Falconer J, Locke M and Wen J: Recent advances in non-ionic surfactant vesicles (niosomes): Fabrication, characterization, pharmaceutical and cosmetic applications. Eur J Pharm Biopharm 2019; 144: 18-39.
- 25. Ge X, Wei M, He S & Yuan WE: Advances of non-ionic surfactant vesicles (niosomes) and their application in drug delivery. Pharmaceutics 2019; 11(2): 55.
- Verma S, Singh SK, Syan N, Mathur P and Valecha V: Nanoparticle vesicular systems: a versatile tool for drug delivery. J Chem Pharm Res 2010; 2(2): 496–509.

- 27. John G, Sinha P, Rathnam G, Ubaidulla U & Aravind R: A Review on Future Prospects of Niosomes towards Drug Delivery Applications. IOSR J Pharm 2021; 11: 1-9.
- 28. Kaur D and Kumar S: Niosomes: present scenario and future aspects. J Drug Deliv Ther 2018; 8(5): 35-43.
- Khan R & Irchhaiya R. Niosomes: a potential tool for novel drug delivery. J Pharm Investig 2016; 46(3): 195-204.
- Kumavat S, Sharma PK, Koka SS, Sharma R, Gupta A and Darwhekar GN: A Review on Niosomes: Potential Vesicular Drug Delivery System. J Drug Del Therap 2021; 11(5): 208-12.
- 31. Makeshwar K and Wasankar S: Niosome: a Novel Drug Delivery System. Asian J Pharm Res 2013; 3(1): 16-20.
- 32. Jiao J: Polyoxyethylated nonionic surfactants and their applications in topical ocular drug delivery. Adv Drug Del Rev 2008; 60(15): 1663-73.
- García-Manrique P, Machado ND, Fernández MA, Blanco-López MC, Matos M & Gutiérrez G: Effect of drug molecular weight on niosomes size and encapsulation efficiency. Colloids and Surfaces B: Biointerfaces 2020; 186: 110711.
- 34. Ge X, Wei M, He S and Yuan WE: Advances of Non-Ionic Surfactant Vesicles (Niosomes) and Their Application in Drug Delivery. Pharma 2019; 11(2): 55-70.
- 35. Elmi N, Ghanbarzadeh B, Ayaseh A, Sahraee S, Heshmati, MK, Hoseini M & Pezeshki A: Physical properties and stability of quercetin loaded niosomes: stabilizing effects of phytosterol and polyethylene glycol in orange juice model. Journal of Food Engineering 2021; 296: 110463.
- 36. Agarwal S, Bakshi V, Vitta P, Raghuram AP, Pandey S and Udupa N: Effect of cholesterol content and surfactant HLB on vesicle properties of niosomes. Indian J Pharma Sci 2004; 66(1): 121-23.
- 37. Barani M, Mirzaei M, Torkzadeh-Mahani M & Adeli-Sardou M: Evaluation of carum-loaded niosomes on breast cancer cells: Physicochemical properties, *in-vitro* cytotoxicity, flow cytometric, DNA fragmentation and cell migration assay. Scientific reports 2019; 9(1): 7139.
- Biswal S, Murthy PN, Sahu J, Sahoo P and Amir F: Vesicles of Non-ionic Surfactants (Niosomes) and Drug Delivery Potential. Int J Pharm Sci Nanot 2008; 1(1): 1-8.
- Ge X, Wei M, He S & Yuan WE: Advances of non-ionic surfactant vesicles (niosomes) and their application in drug delivery. Pharmaceutics 2019; 11(2): 55.
- Budhiraja A and Dhingra G: Development and characterization of a novel antiacne niosomal gel of rosmarinic acid. Drug Del 2015; 22(6): 723-30.
- Kumar BS, Krishna R, Ps L, Vasudev DT and Nair SC: Formulation and evaluation of niosomal suspension of cefixime. Asian J Pharm Clin Res 2017; 10(5): 194-201.
- Liu T, Guo R, Hua W and Qiu J: Structure behaviors of hemoglobin in PEG 6000/Tween 80/Span 80/H₂O niosome system. Colloids Surf. A: Physiochem Eng Asp 2007; 293(1-3): 255–61.
- 43. De Silva L, Fu JY, Htar TT, Muniyandy S, Kasbollah A, Wan Kamal WHB & Chuah LH: Characterization, optimization, and *in-vitro* evaluation of Technetium-99mlabeled niosomes. International Journal of Nanomedicine 2019; 1101-1117.
- 44. Zoghroban HS, El-Kowrany SI, Aboul Asaad IA, El Maghraby GM, El-Nouby KA & Abd Elazeem MA: Niosomes for enhanced activity of praziquantel against Schistosoma mansoni: *in-vivo* and *in-vitro* evaluation. Parasitology Research 2019; 118: 219-234.
- 45. Moghassemi S, Parnian E and Hakamivala A: Uptake and transport of insulin across intestinal membrane model

using trimethyl chitosan coated insulin niosomes. Mater Sci Eng C 2015; 46(1): 333-40.

- 46. Mohanty A and Dey J: Enantioselectivity of vesicleforming chiral surfactants in capillary electrophoresis. Role of the surfactant headgroup structure. J Chromatogr A 2006; 1128(1-2): 259-66.
- 47. Chen S, Hanning S, Falconer J, Locke M & Wen J: Recent advances in non-ionic surfactant vesicles (niosomes): Fabrication, characterization, pharmaceutical and cosmetic applications. European Journal of Pharmaceutics and Biopharmaceutics 2019; 144: 18-39.
- 48. Erfani-Moghadam V, Aghaei M, Soltani A, Abdolahi N, Ravaghi A, Cordani M & Balakheyli H: ST8 micellar/niosomal vesicular nanoformulation for delivery of naproxen in cancer cells: Physicochemical characterization and cytotoxicity evaluation. Journal of Molecular Structure 2020; 1211: 127867.
- 49. Zaid Alkilani A, Hamed R, Abdo H, Swellmeen L, Basheer HA, Wahdan W & Abu Kwiak AD: Formulation and evaluation of azithromycin-loaded niosomal gel: optimization, *in-vitro* studies, rheological characterization, and cytotoxicity study. ACS Omega 2022; 7(44): 39782-39793.
- 50. Pardakhty A, Varshosaz J and Rouholamini A: *In-vitro* study of polyoxyethylene alkyl ether niosomes for delivery of insulin. Int J Pharm 2007; 328(2): 130-41.
- 51. Puras G, Mashal M, Zárate J, Agirre M, Ojeda E, Grijalvo S, Eritja R, Diaz-Tahoces A, Martínez Navarrete G, Avilés-Trigueros M, Fernández E and Pedraz JL: A novel cationic niosome formulation for gene delivery to the retina. J Control Release 2014; 174: 27-36.
- 52. Rai AK, Alam G, Singh AP and Verma NK: Niosomes: An approach to current drug delivery-A Review. Int J Adv Pharm 2017; 6(2): 41-48.
- 53. Sharma R, Dua JS, Prasad DN & Hira S: Advancement in novel drug delivery system: niosomes. Journal of Drug Delivery and Therapeutics 2019; 9(3): 995-1001.
- 54. Rao B, Reddy K, Mounika B, Fathima S and Tejaswini A: Vesicular Drug Delivery System: A Review. Int J Chem Tech Res 2019; 12(5): 39-53.
- 55. Miatmoko A, Faradisa AA, Jauhari AA, Hariawan BS, Cahyani DM, Plumeriastuti H & Hendradi E: The effectiveness of ursolic acid niosomes with chitosan coating for prevention of liver damage in mice induced by n-nitrosodiethylamine. Scientific Reports 2022; 12(1): 21397.
- 56. Sguizzato M, Pepe A, Baldisserotto A, Barbari R, Montesi, L, Drechsler M & Cortesi R: Niosomes for Topical Application of Antioxidant Molecules: Design and In Vitro Behavior. Gels 2023; 9(2): 107.
- 57. Afreen U, Fahelelbom KM, Shah SNH, Ashames A, Almas U, Khan SA & Murtaza G: Formulation and evaluation of niosomes-based chlorpheniramine gel for the treatment of mild to moderate skin allergy. Journal of Experimental Nanoscience 2022; 17(1): 467-495.
- 58. Sharma S and Chauhan M: Span-60 niosomal oral suspension of fluconazole: formulation and *in-vitro* evaluation. J Pharm Res Health Care 2009; 1(1): 142-56.
- Naderi R, Pardakhty A, Abbasi MF, Ranjbar M & Iranpour M: Preparation and evaluation of crocin loaded in nanoniosomes and their effects on ischemia–reperfusion injuries in rat kidney. Scientific Reports 2021; 11(1): 23525.
- Kumari P, Bhandari DD & Kondal N: Formulation and characterization of niosomal suspension of Valsartan for the treatment of hypertension. Materials Today: Proceedings 2023.

- 61. Natsheh H, Vettorato E & Touitou E: Ethosomes for dermal administration of natural active molecules. Current Pharmaceutical Design 2019; 25(21): 2338-2348.
- 62. Yadavar-Nikravesh MS, Ahmadi S, Milani A, Akbarzadeh I, Khoobi M, Vahabpour R & Bakhshandeh H: Construction and characterization of a novel tenofovir-

loaded pegylated niosome conjugated with tat peptide for evaluation of its cytotoxicity and anti-hiv effects. Advanced Powder Technology 2021; 32(9): 3161-3173.

63. Bhaskaran S and Lakshmi PK. Comparative evaluation of niosome formulations prepared by different techniques. Acta Pharm Sci 2009; 51(1): 27-32.

How to cite this article:

Sharma M and Bains K: A review on niosomal suspension containing cefpodoxime proxetil. Int J Pharm Sci & Res 2023; 14(11): 5141-49. doi: 10.13040/IJPSR.0975-8232.14(11).5141-49.

All © 2023 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to Android OS based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)