



Received on 29 July 2023; received in revised form, 06 October 2023; accepted, 30 December 2023; published 01 March 2024

THERAPEUTIC EFFECT OF LEAVES OF *AYAPANA TRIPLINERVIS* IN INDOMETHACIN INDUCED MILD, MODERATE AND SEVERE GRADE GASTRIC ULCER

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Keywords:

Gastric ulcer, *Ayapana triplinervis*, Omeprazole, Oxidative stress, COX-2

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ABSTRACT: Peptic ulcer disease (PUD) is increasing gradually in India as well as worldwide. Excess use of non-steroidal anti-inflammatory agents (NSAID) and lifestyle factors cause gastric mucosal damage. This study was designed to search out the gastro-protective efficacy of hydromethanolic extract of *Ayapana triplinervis* on different graded ulcers. Mild to severe grade of gastric ulcer has been developed by oral administration of IND at the doses of 10mg, 20mg and 40mg/kg body weight for 15 days. Gastric ulcer were assessed by elevation of ulcer score, ulcer index, pepsin activity and gastric volume in IND-treated groups in respect to control group. Tumor necrosis factor-alpha (TNF- α), interleukin-6 (IL-6) levels were elevated in IND-treated groups in respect to control due to withdrawal of prostaglandin and cyclooxygenase (COX-2) enzyme induced suppression. Significant protection in the levels of these ulcerogenic parameters were observed after the treatment of *Ayapana triplinervis* (200mg/kg) when compared with the standard drug omeprazole (20mg/kg). The important antioxidant enzymes activities were decreased but lipid peroxidation levels elevated in all the ulcerated groups ($p < 0.05$). Gastric-occult test was positive in 40mg IND-treated group. Glutamate oxaloacetate transaminase (GOT) and glutamate pyruvate transaminase (GPT) activities in serum measured for toxicity assessment. All the parameters that altered in ulcerated groups were significantly protected towards the control level after *Ayapana triplinervis* or omeprazole treatment. This significant protective efficacy observed in histopathological evaluation of gastric tissue. *Ayapana triplinervis* significantly protect the mild, moderate and severe grade ulcers by its antioxidant activity and withdrawal of IND-induced suppression of prostaglandin and COX-2 levels.

INTRODUCTION: Gastric ulcer (GU) is a major public health burden in the present century¹ with a high morbidity of about 5–10 %. Annually 200,000 people are hospitalized with the diagnosis of ulcer and the financial cost of the treatment of this disease reaches about 4 billion dollars².

Infection of *Helicobacter pylori*, smoking, alcohol consumption, distress, haemorrhagic shock, sepsis, nutritional deficiencies and long-term use of NSAIDs are major causes of gastric ulceration³. Around 25% cases of gastric ulceration are caused by NSAIDs⁴.

Indomethacin (IND) is mainly used for the treatment of stroke, pain control, rheumatoid arthritis, osteoarthritis, tendonitis and other inflammatory diseases⁵. Experimentally, IND is widely used for the induction of GU due to its higher ulcerogenic potency compared to other NSAIDs^{6,7}.

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| <p>QUICK RESPONSE CODE</p>  | <p>DOI: 10.13040/IJPSR.0975-8232.15(3).788-99</p> <hr/> <p>This article can be accessed online on www.ijpsr.com</p> |
| <p>DOI link: https://doi.org/10.13040/IJPSR.0975-8232.15(3).788-99</p> | |

Gastric mucosal damage including erosion, ulceration, bleeding and perforation are the most significant adverse effects associated with NSAIDs⁸. Many synthetic anti-ulcer drugs have been developed For gastric ulcer treatment. However, these agents are facing major problems due to their partial effectiveness against gastrophelcosis and severe side effects, for instance, dizziness, diarrhoea, gynecomastia, hypoacidity, impotence, osteoporotic bone fracture, hypergastrinaemia, skin rash and cardiovascular disease risks^{9, 10, 11}. Thus, new drugs with high efficacy and low toxicity are desirable for the prevention and treatment of gastric ulcers. For safety and therapeutic efficacy, the World Health Organization (WHO) recommends medicinal plants¹². Medicinal plants and their extracts have an important therapeutic role against many diseases without significant side effects¹³. There are many plants with ethno-pharmacological

background that have been gastro-protective and used in traditional medicine^{14, 15}. The purpose of this study design is to develop different graded gastric ulceration in experimental model and to search out whether our selected plant *Ayapana triplinervis* (AT) has protective efficacy even on severe grade ulcers or not^{16, 17}. Rats were treated with three different doses of indomethacin 10mg, 20mg and 40mg/kg body weight/day for 15 days **Table 1** and after sacrifice accordingly to clinical manifestation of gastric environment and different gastric secreting parameters, we have divided these three different doses of indomethacin treated experimental rats in mild, moderate and severe grade ulcerated group. Now the gastro-therapeutic efficacy of hydromethanolic extract of *Ayapana triplinervis* was investigated on mild, moderate and severe grade ulcers.

TABLE 1: DIFFERENT DOSES OF INDOMETHACIN TREATMENT AND ULCER MANIFESTATION

| | Mild (IND10) | Moderate (IND20) | Severe (IND40) |
|--------------|-------------------------|------------------|--------------------------|
| Ulcer score | 10-15 | 15-30 | <30 |
| Ulcer lesion | Mild ulcer small lesion | Damaged tissue | Deep ulcerating bleeding |
| Smell | No smell | Bad smell | Foul smell |
| Discharge | No discharge | Mild discharge | Profuse discharge |
| Stomach wall | Smooth | Rough | Very Rough |

MATERIAL AND METHODS:

Chemicals: Indomethacin purchased from Tokyo Chemical Industry, Mumbai, India. Omeprazole procured from Alfa Aesar (UK). GOT, GPT kits were purchased from Span Diagnostic Limited, Surat, India. Prostaglandin E₂ and COX-2 ELISA Kit purchased from Elabscience biotechnology Co., Ltd, Wuhan, China. TNF- α and IL-6 ELISA kits obtained from Raybiotech Co., Peachtree Corners, United States. Alcian blue obtained from Loba Chemie Pvt. Ltd. (Mumbai, India). All other chemicals were of the highest purity grade commercially accessible.

Plant Collection: Fresh leaves of *Ayapana triplinervis* were collected from the local area of Medinipur, identified and authenticated by the taxonomist. The voucher number of the specimen is VU/CM/102.

Preparation of Extracts: Fresh leaves of *Ayapana triplinervis* were washed in normal distilled water and shed dried at room temperature and crushed. The crushed leaves were dissolved in hydro-

methanol (40:60) in a glass jar in the air tight condition for 7 days with occasional shaking. The filtrate was collected and dried at 40°C under reduced pressure in a rotary evaporator (N1200AS-W, EYELA, Japan). Finally, this was kept in an owing bottle and stored at 4°C to use in experiment¹⁸.

Experimental Animals: Albino rats of Wistar strain having body weight 150±10g, were used for this study approved by the Institutional Animal Ethical Committee with the ethical number No.VU/IAEC-I/CM-1/3-6/19 dated 11/12/19. Rats were collected from Saha Enterprise, an authorized dealer of CPCSEA registered under the Ministry of Environment and Forest. Rats were divided into four cages and kept for 15 days for acclimatization in our animal houses with adequate temperature, humidity along with proper diet and water.

Induction of Gastric Ulcer: Gastric ulceration was induced in the animals by oral administration of indomethacin at the dose of 10mg, 20mg and 40 mg/kg body weight/day. They were deprived of

food before 2 hours of indomethacin treatment but had free access to water¹⁹. 2 hours after the indomethacin treatment rats were treated with AT (200mg/kg) or omeprazole (20mg/kg).

Animal Grouping and Treatments: Sixty male albino rats were randomized into ten groups equally.

Group I: Control group (CON): Rats of this group received only vehicles.

Group II: Indomethacin treated group (IND 10): Rats were given indomethacin at the dose of 10 mg/kg b.w /day.

Group III: Ayapana treated group (IND10AT): Rats were treated with indomethacin (10mg/kg/day) followed by *Ayapana triplinervis* at the dose of 200 mg/kg b.w.

Group IV: Omeprazole treated group (IND10OMZ): Rats were treated with indomethacin (10mg/kg/day) followed by omeprazole at the dose of 20 mg/kg b.w.

Group V: Indomethacin treated group (IND 20): Rats of this group were treated with indomethacin at the dose of 20mg/kg b.w.

Group VI: Ayapana treated group (IND20AT): These indomethacin (20mg/kg/day) treated rats were also administered with *Ayapana triplinervis* at the dose of 200 mg/kg b.w.

Group VII: Omeprazole treated group (IND20OMZ): These indomethacin (20mg/kg/day) treated rats were treated with omeprazole at the dose of 20 mg/kg b.w.

Group VIII: Indomethacin treated group (IND 40): Rats were treated with indomethacin at the dose of 40mg/kg b.w.

Group IX: Ayapana treated group (IND40AT): Rats were also treated with indomethacin at the dose of 40mg/kg b.w. After 2 hours these rats were again treated with *A. triplinervis* at the dose of 200mg/kg b.w.

Group X: Omeprazole treated group (IND40OMZ): These rats were also treated with indomethacin at the dose of 40mg/kg followed by omeprazole treatment at the dose of 20mg/kg b.w.

The experiment was continued for 15 days. On 16th day animals of all the groups were sacrificed and the abdomen was opened with clamping of the pyloric end for the collection of gastric juice. Stomach was collected and stored in -20°C for biochemical study. Pylorus ligation shows the possible changes of other gastric parameters like gastric content, volume, total acidity and pH. One piece of vertically incised stomach portion kept in bouins solution for histological study. Blood was collected and serum was separated for biochemical and immune assay.

Estimation of Gastric pH: After sacrifice, gastric content was collected and centrifuged for 10 min at 1000 rpm (4°C) then the clear supernatant was collected and used for the determination of pH by using pH meter²⁰.

Gastrocult Test (Gastric Occult Blood Test): The Gastrocult blood test is based on alpha guaiaconic acid with hydrogen peroxide reaction in the presence of heme to produce a highly conjugated blue quinone compound.

It is a qualitative screening method for detecting the existence of occult blood and determining the pH of gastric aspirate or vomitus. Results cannot be considered conclusive evidence of presence or absence of gastrointestinal bleeding²¹.

Ulcer Quantification: Rats were Sacrifice and stomachs were opened and gastric ulcer score was measured by standard protocol²². In the 10mg dose treated group 10-15 ulcerated areas are observed and we considered this group as mild ulcers. In the 20mg treated group 15-30 ulcerated areas are found. We consider these groups as moderate ulcerated groups and in the 40mg treated group above 30 ulcerated areas are observed and it is the severe ulcerated group (**Table 1**).

Determination of Pepsin Activity: Pepsin activity was determined by using hemoglobin as the substrate. Specific pepsin activity expressed by ug/ml²³.

Determination of Mucin Content: The conventional approach was used to determine the mucin content²⁴. Optical density of mucin content was determined at 605 nm and standard curve developed with various mucin concentrations.

Determination of Free Acidity and Total Acidity: Gastric free acidity and total acidity were measured by the standard method²³. Gastric juice was taken followed by addition of few drops of Topfer's reagent and phenolphthalein respectively for free and total acidity. Titrated against NaOH, and the volume of NaOH added was recorded. The formula for the calculation of gastric acidity was given below:

$$\text{Acidity} = (\text{Volume of NaOH} \times \text{actual normality of NaOH} \times 100) / 0.1 \text{ mEq/L.}$$

Determination of Prostaglandin E₂ (PGE₂) and Cyclooxygenase-2 (COX-2): Serum PGE₂ levels and COX-2 measured in gastric tissue were measured using ELISA kits²⁵ according to manufacturer protocols. Value was expressed as pg/ml and ng/ml²⁶.

Determination of Tumor necrosis Factor-alpha (TNF- α) and Interleukin-6 (IL-6) in Stomach: TNF- α and IL-6 in stomach tissue homogenate were measured by using standard ELISA kits according to the protocol provided by the manufacturer. The values were expressed as pg/ml²⁷.

Antioxidant Activity:

Biochemical Estimation Catalase (CAT) and Superoxide Dismutase (SOD), Glutathione peroxidase (GPx) and GSH: The stomach tissue was homogenized in ice cold 0.5 M Tris-HCl buffer (pH 7.0) and centrifuged. Supernatant was collected and catalase activity was measured by standard method at 240 nm²⁸.

The activity of superoxide dismutase was estimated using the standard method²⁹. The stomach tissue was homogenized and centrifuged. 10 mM pyrogallol and 50 mM Tris-HCl (pH 8.4) was mixed with the supernatant and absorbance was taken at 420 nm in a spectrophotometer using Tris-HCl as blank. The mucosal surface of the stomach was collected by scraping, and then weighed and homogenised in 2 mL of 50 mM Tris-HCl buffer containing 20 mM EDTA and 0.2 mM sucrose (pH 7.5). Then the homogenate was immediately precipitated with 0.1 mL of ice-cold 25% trichloroacetic acid, centrifuged at 4000 rpm for 40 min at 4°C. The supernatant was used for the determination of GSH levels. Absorption was

measured at 412 nm using a spectrophotometer. The GSH levels in the gastric mucosa were expressed as nmol/mg tissue^{30,31}.

Malondialdehyde (MDA) and Conjugated Diene (CD) Level: Phosphate buffer (pH 7.4) was used to homogenize the stomach tissue. Using thiobarbituric acid-trichloro acetic acid (TBA-TCA) mixture the homogenates were boiled for 10 minutes at 100°C and cooled at room temperature. The supernatant was collected after centrifugation at 4000×g for 10 min and OD was measured at 535 nm³². Phosphate buffer (pH 7.4) was used for homogenization of stomach tissue. Chloroform: methanol (2:1) mixture was used for the extraction of the lipid layer and centrifuged at 1000×g for 5 min then evaporated. Cyclohexane was used to dissolve the lipid residue and absorbance was measured at 233 in a UV-spectrophotometer³³.

Histological Study: Stomach tissue processing was done according to the standard method and the section was prepared at 5 μ m using a Leica RM2245 microtome³⁴.

Statistical Analysis: Inhibition against ulceration was expressed in percentage. Other results were expressed as Mean \pm SEM. One-way analysis of variance (ANOVA) followed by multiple comparisons two tailed 't test' using the SPSS software package for differences between means was used to detect any significant difference (p<0.05) between the treatment groups in this study²³.

RESULTS:

Behavioral Changes: Remarkable physical and behavioral changes were observed in indomethacin treated all the ulcerated groups. We have carefully observed the activities, weaknesses and movement of the experimental animals. It has been found that in mild ulcerated groups, weakness, abnormal movement was not observed but activities were reduced remarkably. In moderate and severe ulcerated groups remarkable signs of weakness, abnormal activities were noted and normal activities were reduced. Sign of weakness and abnormal movement and reduced normal activities were most remarkably observed in the severely ulcerated group than the other ulcerated groups (Table 2).

TABLE 2: BEHAVIORAL CHANGES IN MILD, MODERATE AND SEVERE ULCERATED RAT

| Observation | Mild (IND 10) | Moderate (IND 20) | Severe (IND 40) |
|--|---------------|-------------------|-----------------|
| Indomethacin dose(mg/kg bw/day) | 10 | 20 | 40 |
| Sign of weakness | - | + | + |
| Number of rats with signs of weakness | - | ++ | +++++ |
| Abnormal movement | - | + | + |
| Number of rats with abnormal movement | - | + | +++ |
| Reduced activities | + | + | + |
| Number of rats with reduced activities | + | ++ | ++++ |

Key: + signifies present while – signifies absent.

Ulcer Score, Ulcer Index, Preventive Index and occult Blood Test: The ulcer score and ulcer index were significantly elevated in 10mg, 20mg and 40mg doses of indomethacin treated group in comparison to control group. Ulcer score and ulcer index were significantly reduced towards the control level in all the *Ayapana triplinervis* (AT) and omeprazole (OMZ) treated groups. Ulcer scores in all the indomethacin treated groups significantly differ from each other and it is highest in 40mg/kg indomethacin treated groups.

Preventive index was highest in IND10AT and IND10OMZ treated groups i.e., 80.82% and 82.19% respectively. In case of IND20AT and IND20OMZ treated groups, preventive indexes are 64.06% and 64.84%. Preventive indexes in IND40AT and IND40OMZ treated groups are 61.95% and 59.51%.

Occult blood test of gastric content showed positive in 20mg/kg and 40mg/kg indomethacin treated groups **Table 3**.

TABLE 3: EFFECT OF AYAPANA TRIPLINERVIS ON ULCER SCORE, ULCER INDEX, PREVENTIVE INDEX AND OCCULT BLOOD TEST

| Group | Treatment | Ulcer scores | Ulcer index | Preventive index (%) | Occult Blood (OB) |
|----------|-----------|-------------------------|-------------|----------------------|-------------------|
| | CON | 0 | 0 | - | -VE |
| Mild | IND 10 | 12.16±0.75 ^a | 1216.66 | - | -VE |
| | IND10AT | 2.33±0.42 ^b | 233.33 | 80.82 | -VE |
| | IND10OMZ | 2.16±0.40 ^b | 216.66 | 82.19 | -VE |
| Moderate | IND 20 | 21.33±1.92 ^c | 2133.33 | - | +VE |
| | IND20AT | 7.66±0.66 ^d | 766.67 | 64.06 | -VE |
| | IND20OMZ | 7.5±0.42 ^d | 750 | 64.84 | -VE |
| Severe | IND 40 | 34.16±1.13 ^e | 3416.67 | - | +VE |
| | IND40AT | 13±0.51 ^f | 1300 | 61.95 | -VE |
| | IND40OMZ | 13.83±0.65 ^f | 1383.33 | 59.51 | -VE |

Values are expressed as Mean ± SEM, n= 6. ANOVA followed by two-tail t-test. Different superscripts (a,b,c,d,e and f) indicate the significant difference (p<0.05).

Gastric pH, Free Acidity, Total Acidity and Pepsin Activity: Gastric pH were reduced but gastric free acidity, total acidity and pepsin activity were significantly elevated in 10mg, 20mg and 40mg doses of indomethacin treated groups when compared with control group (p<0.05). These parameters were significantly recovered towards the control level in all the AT and OMZ treated groups. More significant restoration was noted after *Ayapana triplinervis* and OMZ treatment in mild (IND10AT, IND10OMZ) ulcerated groups than other ulcerated groups **Table 4**.

Mucin Content: Alcian blue is an indicator of the mucus content in gastric mucosa. Regular treatment of indomethacin at different doses causes significant degeneration of gastric mucosal layer, as well as a significant reduction in alcian blue binding capacity of gastric mucosa in comparison to the normal control group. Whereas treatment of AT and OMZ significantly increase the alcian blue binding with the mucosal layer in mild, moderate and severe ulcerated groups. More significant recovery was observed in the mild ulcerated group than moderate and severe ulcerated group **Table 4**.

TABLE 4: EFFECT OF AYAPANA TRIPLINERVIS ON GASTRIC SECRETORY PARAMETERS IN DIFFERENT GRADED ULCERATED RATS

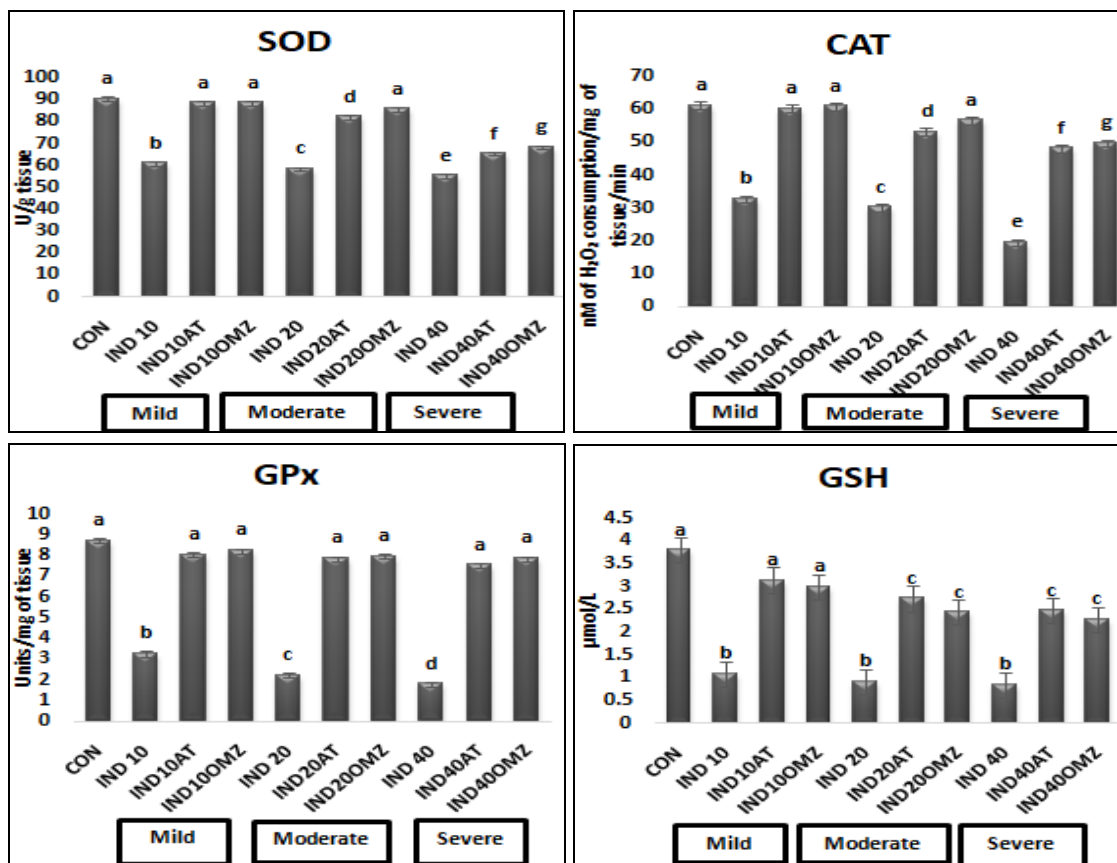
| Groups | Parameters | Gastric volume | Gastric pH | Mucin content (µg alcian blue/ | Pepsin activity | Free acidity (µEq/L) | Total acidity (µEq/L) |
|--------|------------|----------------|------------|--------------------------------|-----------------|----------------------|-----------------------|
|--------|------------|----------------|------------|--------------------------------|-----------------|----------------------|-----------------------|

| | | (ml) | | g wet stomach) | (µg/ml) | | |
|----------|----------|-------------------------|-------------------------|--------------------------|--------------------------|-------------------------|-------------------------|
| Mild | CON | 0.93±0.01 ^a | 4.53±0.10 ^a | 245.66±1.64 ^a | 99.66±1.38 ^a | 25.65±0.92 ^a | 37.53±0.75 ^a |
| | IND 10 | 1.463±0.10 ^b | 4.14±0.05 ^b | 100.33±0.76 ^b | 225.66±1.33 ^b | 35.22±0.91 ^b | 66.75±0.56 ^b |
| | IND10AT | 0.96±0.01 ^a | 4.373±0.07 ^a | 243.05±0.94 ^a | 100.61±0.70 ^a | 26.98±0.76 ^a | 39.43±0.39 ^a |
| Moderate | IND100MZ | 0.95±0.004 ^a | 4.52±0.10 ^a | 242.37±0.84 ^a | 101.27±0.46 ^a | 26.72±0.91 ^a | 39.38±0.68 ^a |
| | IND 20 | 1.73±0.07 ^c | 3.25±0.07 ^c | 74.16±1.16 ^c | 262.33±1.56 ^c | 39.77±0.89 ^c | 68.61±0.56 ^c |
| | IND20AT | 1.05±0.05 ^d | 3.59±0.16 ^d | 223.5±0.76 ^d | 138.08±0.75 ^d | 33.15±0.83 ^d | 53.23±0.81 ^d |
| Severe | IND200MZ | 0.98±0.003 ^a | 4.33±0.09 ^a | 219.87±1.03 ^a | 137.08±0.75 ^a | 27.85±0.64 ^a | 40.06±0.45 ^a |
| | IND 40 | 2.33±0.08 ^e | 2.45±0.06 ^e | 51.33±0.49 ^e | 282.66±0.95 ^e | 48.60±0.65 ^e | 73.53±0.77 ^e |
| | IND40AT | 1.34±0.13 ^f | 3.37±0.08 ^f | 198.5±2.36 ^f | 169.33±0.49 ^f | 37.94±0.75 ^f | 59.87±0.75 ^f |
| | IND400MZ | 1.32±0.01 ^g | 3.49±0.12 ^g | 199.83±0.87 ^g | 167.30±0.89 ^g | 40.05±0.77 ^g | 63.20±0.69 ^g |

Values are expressed as Mean ± SEM, n= 6. ANOVA followed by two-tail t-test. Different superscripts (a,b,c,d,e,f and g) indicate the significant difference (p<0.05).

Activities of CAT, SOD, GPx and GSH in Stomach Tissue: Indomethacin treatment significantly reduced the activities of CAT, SOD, GPx and GSH in stomach tissue. These enzyme activities were significantly recovered in all the hydromethanolic extract of leaf of AT and OMZ treated groups in comparison to the control groups. More significant protection of these parameters were noted to the control level after the treatment of AT in the mild ulcerated group **Fig. 2** than other ulcerated groups.

MDA and CD Levels in Stomach Tissue: Gastric MDA and CD levels were significantly elevated in all the indomethacin treated ulcerated groups when compared with the control group. These parameters were significantly recovered towards the control level in all the AT and OMZ treated groups in comparison to control groups. More significant recovery was noted to the control level after the treatment of AT and OMZ in mild ulcerated groups **Fig. 2**.



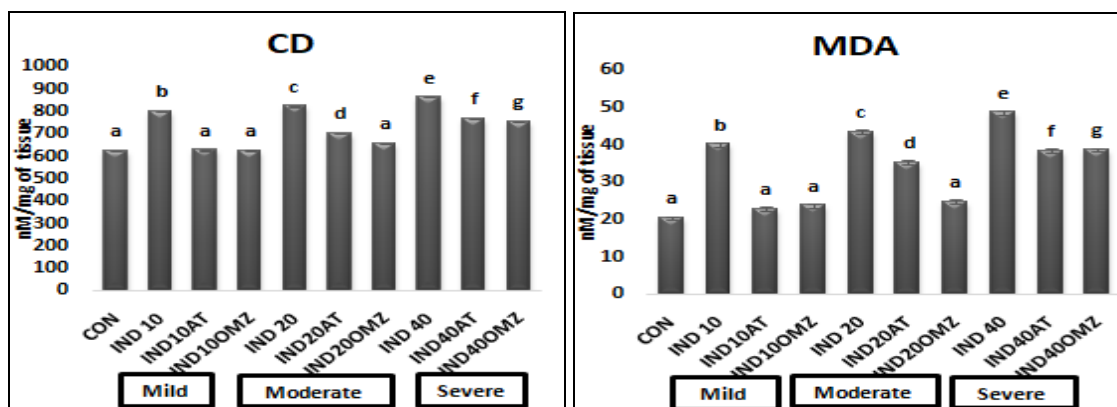


FIG. 2: AYAPANA TRIPLINERVIS ON ANTIOXIDANT ACTIVITIES AND LIPID PEROXIDATION LEVELS IN DIFFERENT GRADED ULCERATED RATS. Values expressed as Mean ± SEM, n= 6. ANOVA followed by two-tail t-test. Different superscripts (a, b, c, d, e, f and g) indicate the significant difference (p<0.05).

Serum Prostaglandin E₂ (PGE₂) and COX-2 Level: Serum PGE₂ and COX-2 levels were significantly decreased in all the indomethacin induced mild, moderate and severe ulcerated groups in respect to the control group. These two parameters were significantly recovered towards the control level in hydro-methanol extract of leaf of AT and OMZ treated mild, moderate and severe ulcerated groups. In case of PGE₂ at IND10AT (96.84%), IND20AT (91.24%) and IND40AT (78.45%); and in COX-2 the recovery rate were noted IND10AT (97.24%), IND20AT (91.36%)

and IND40AT (90.59%) Fig. 3. More significant recovery were observed to the control level in the mild ulcerated groups.

Tissue Tumor Necrosis Factor (TNF-α) and Interleukin 6 (IL-6): Gastric tissue TNF-α and IL-6 levels were elevated significantly in all the indomethacin treated groups in respect to the control group. These parameters were significantly recovered in AT and OMZ treated mild, moderate and severe ulcerated groups Fig. 3.

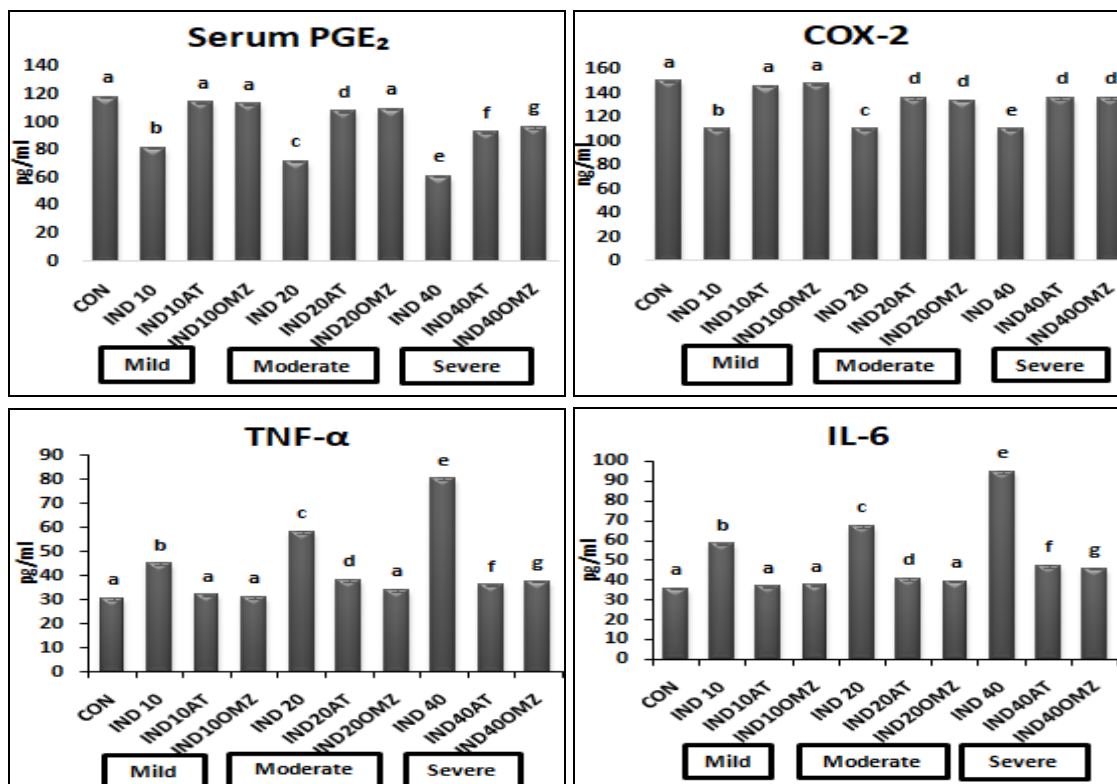


FIG. 3: EFFECT OF AYAPANA TRIPLINERVIS ON TNF-α, IL-6, PGE₂ AND COX-2 IN INDOMETHACIN TREATED DIFFERENT ULCERATED GROUPS. Values expressed as Mean ± SEM, n= 6. ANOVA followed by two-tail t-test. Different superscripts (a, b, c, d, e, f and g) the significantly differ from each other (p<0.05).

More significant recovery of these parameters was observed in the mild ulcerated group in respect to the moderate and severe ulcerated group.

In TNF- α , IND10AT (70.95%), IND20AT (65.74%) and IND40AT (44.87%); and in case of IL-6, IND10AT (62.83%), IND20AT (60.38%) and IND40AT (49.85%) respectively in compared to different ulcerated group such as mild (IND 10), moderate (IND 20) and severe (IND 40).

Activities of SGPT and SGOT: There was significant elevation in SGPT and SGOT activities observed in all indomethacin treated groups when compared with control groups. A significant restoration was noted towards control level in hydromethanolic extract of leaf of AT and OMZ treated all the ulcerated groups **Fig. 4**. More significant recovery was noted in mild ulcerated groups in respect to moderate, severe ulcerated groups.

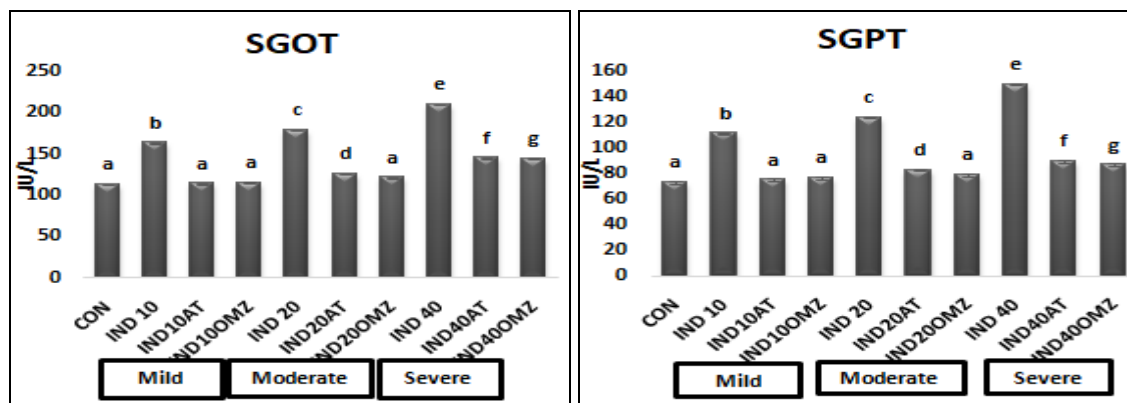


FIG. 4: EFFECT OF AYAPANA TRIPLINERVIS ON SERUM GOT AND GPT ACTIVITIES IN INDOMETHACIN TREATED ULCERATED RATS. Values expressed as Mean \pm SEM, n= 6. ANOVA followed by two-tail t-test. Different superscripts (a, b, c, d, e, f and g) indicate the significant difference from indomethacin treated groups (p<0.05).

Histological Studies: Histomorphological study of gastric tissue carried out by haematoxylin and eosin (H & E) staining. Indomethacin treated groups resulted in marked changes in gastric tissue morphology in respect to control. AT treated rats

showed significant recovery of the mucosal layer, thereby indicating gastroprotective effect of *Ayapana triplinervis* against indomethacin induced gastric ulcerative damage in rats **Fig. 5**.

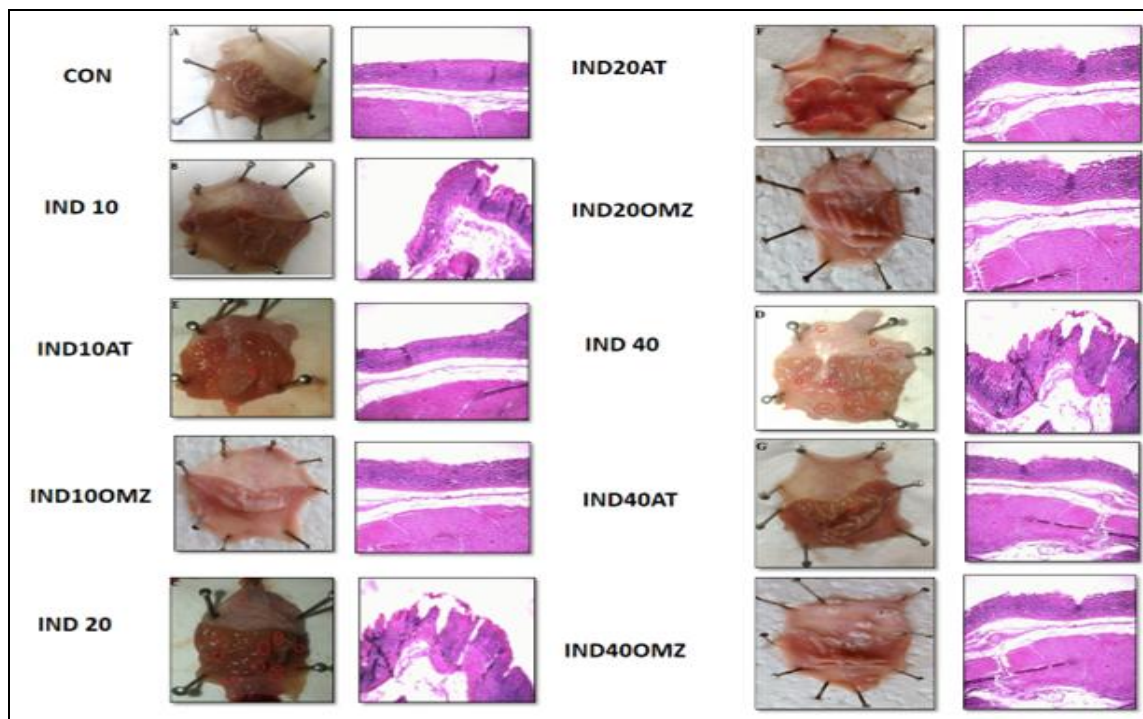


FIG. 5: MAGNIFICATION VIEW OF GASTRIC MUCOSAL SURFACE

CON indicate the normal stomach histoarchitecture of control group. IND 10 showed a very few number of black spots or ulcerated areas. IND 20 showed a significant elevation of black spot or ulcerated areas. IND 40 showed the increasing number of black spot or ulcerated areas. Extract treated groups (IND20AT and IND40AT) shows significant recovery and a decreasing number of the ulcerated area. The IND10AT and omeprazole groups show significant recovery and very less number of ulcerated areas.

DISCUSSION: Stomach ulcers are open sores formed due to the damage of the lining of the stomach. Indomethacin is a potent ulcerogenic agent³⁵ used for the induction of gastric ulcers in an experimental model²³. A significant increase in ulcer index²² and gastric volume²² was noted in all the indomethacin treated mild, moderate and severe ulcerated groups may be due to either generation of free radicals³⁶, or inhibition of prostaglandin synthesis³⁷ due to inhibition of COX³⁴. Ulcer index, ulcer score and gastric volume are significantly different in mild, moderate and severe ulcerated groups. Depletion of prostaglandin level stimulates gastric acid secretion and impairs gastric protection that causes mucosal lesion and gastric ulceration³⁸. Gastric pH increased in indomethacin treated groups which indicate the decreased hydrogen ion concentration in gastric juice that causes pathogenicity of ulcer³⁹. Pepsin activity also increased in indomethacin induced mild, moderate and severe ulcerated groups²². Gastric free acidity, total acidity was elevated by indomethacin induced depletion of serum prostaglandin level^{34, 38}. In all ulcerated groups, increased pepsin activity with decrease in mucin secretion indicated altered hydrophobicity and reduced protective ability of the mucosal membrane against hemorrhagic erosions, thus resulting in tissue damage. In AT treated group significant restoration was observed due to recovery of prostaglandin levels as well as COX levels.

Gastric mucus forms a continuous adherent layer and gives important gastric defense against various aggressive factors such as pepsin, gastric acid, swallowed material and alcohol⁴⁰. Gastric mucus is also important for gastric ulcer healing⁴¹. A decrease in gastric mucosa was observed due to

exposure of acid and other aggravating factors, further slowing ulcer healing. Furthermore, mucus functions as an antioxidant, which reduces the damaging effects of reactive oxygen species and lubricates gastric mucosal surface⁴². Finally gastric mucus is also essential for tissue regeneration during gastric ulcer healing; it provides a suitable pH environment for gastric epithelial cell restitution and enhances the binding of epithelial growth factor and other growth factors to their receptors⁴³. These would promote healing by increasing cell proliferation, granulation tissue formation and re-epithelialization⁴⁴. In moderate and severe ulcerated groups more gastric mucosal lesion was observed due to more acid formation and pepsin activity than the mild ulcerated group. After the AT treatment significant complete recovery in the mucin content and pepsin activity was observed in mild ulcerated groups but partial recovery was observed in severe ulcerated groups. Partial recovery is also observed in OMZ treated groups.

Indomethacin reduces CAT, SOD, GPx and GSH activities and these enzymes reduce intracellular levels of free radicals. SOD acts as the first-line defense system against reactive oxygen species⁴⁵. SOD catalyzes the superoxide anion into H₂O₂ which then converts into water by catalase. Elevation of CD and MDA levels in gastric tissue in indomethacin treated rats indicates the involvement of oxidative injury^{46, 47}. Hydro-methanol extract of leaves of AT have the potent antioxidant activity that scavenged the free radicals and protects the rats from indomethacin induced oxidative damage²⁰.

The important inflammatory markers like TNF- α and IL-6 levels in gastric tissue were elevated in indomethacin induced ulcerated rats through activating neutrophil infiltration²³ due to withdrawal of down regulated control of prostaglandin as prostaglandin were decreased in ulcerated groups. AT extract has also exhibited an anti-inflammatory effect that causes significant decrease in TNF- α and IL-6 levels by stimulating prostaglandin. From histo-architectural point of view it may be stated that significant damage and erosion was noted in mucosa and submucosa layer of stomach tissue in all the indomethacin treated groups and severe lesion observed in severe

ulcerated groups⁴⁸. Hydro-methanol extract of AT treatment gives significant protection. Serum GOT and GPT levels also restored towards control in AT treated rats which indicate the AT has no toxic effect in general.

CONCLUSION: From this result it may be concluded that hydro-methanol extract of AT shows potent gastro-protective activity in mild, moderate and severe ulcerated groups by stimulating the PGE₂ level and by scavenging the free radicals generation. Most protective effect was observed in the mild ulcerated group. This study was conducted on crude extract further research will be required for identification of active ingredient(s) of leaf extract of *Ayapana triplinervis* for the protection of gastric ulcer.

ACKNOWLEDGMENTS: We are thankful to Vidyasagar University authority for providing us with research facilities to conduct this work.

Declarations:

Funding: This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Author Contributions: RM: Performed experiment, data analysis, writing-reviewing and editing, PM: Extract preparation, PRB: Biochemical analysis, SM: Biochemical analysis, CM: Supervision, writing-reviewing and editing, formal analysis. All authors revised and approved the final manuscript for publication.

Data Availability Statement: The data that support the findings of this study are available from the corresponding author upon request.

CONFLICT OF INTEREST: The author declares that there is no conflict of interest.

REFERENCES:

1. Lanas A and Chan FK: Peptic ulcer disease. *The Lancet* 2017; 390(10094): 613-624.
2. Tijani AS, Temitayo MJ and Farombi OE: Cytoprotective effect of methanol extract of *Laportea aestuans* on acidified ethanol-induced gastric ulcer in male wistar rats. *Trends in Sciences* 2022; 19(13): 4648.
3. Selmi S, Rtibi K, Grami D, Sebai H and Marzouki L: Protective effects of orange (*Citrus sinensis* L.) peel aqueous extract and hesperidin on oxidative stress and peptic ulcer induced by alcohol in rat. *Lipids in Health and Disease* 2017; 16(1): 1-2.
4. McEvoy L, Carr DF and Pirmohamed M: Pharmacogenomics of NSAID-induced upper gastrointestinal toxicity. *Frontiers in Pharmacology* 2021; 12: 684162.
5. Danisman B, Cicek B, Yildirim S, Bolat I, Kantar D, Golokhvast KS, Nikitovic D, Tsatsakis A and Taghizadehghalehjoughi A: Carnosic Acid Ameliorates Indomethacin-Induced Gastric Ulceration in Rats by Alleviating Oxidative Stress and Inflammation. *Biomedicines* 2023; 11(3): 829.
6. Shaik RA and Eid BG: Piceatannol affects gastric ulcers induced by indomethacin: association of antioxidant, anti-inflammatory, and angiogenesis mechanisms in rats. *Life* 2022; 12(3): 356.
7. Harakeh S, Saber SH, Akefe IO, Shaker S, Hussain MB, Almasaudi AS, Saleh SM and Almasaudi S: Saudi honey alleviates indomethacin-induced gastric ulcer via improving antioxidant and anti-inflammatory responses in male albino rats. *Saudi Journal of Biological Sciences* 2022; 29(4): 3040-3050.
8. McEvoy L, Carr DF and Pirmohamed M: Pharmacogenomics of NSAID-induced upper gastrointestinal toxicity. *Frontiers in Pharmacology* 2021; 12: 684162.
9. Kulikova OI, Stvolinsky SL, Migulin VA, Andreeva LA, Nagaev IY, Lopacheva OM, Kulichenkova KN, Lopachev AV, Trubitsina IE and Fedorova TN: A new derivative of acetylsalicylic acid and carnosine: synthesis, physical and chemical properties, biological activity. *DARU Journal of Pharmaceutical Sciences* 2020; 28:119-130.
10. Chakravarty K and Gaur S: Role of probiotics in prophylaxis of *Helicobacter pylori* infection. *Current Pharmaceutical Biotechnology* 2019; 20(2): 137-145.
11. Singh AK, Singh SK, Singh PP, Srivastava AK, Pandey KD, Kumar A and Yadav H: Biotechnological aspects of plants metabolites in the treatment of ulcer: A new prospective. *Biotechnology Reports* 2018; 18: e00256.
12. Hervé EE, Bernard GN, Léandre KK, Paul YA and Etienne EE: Acute toxicity and gastric anti-ulcer activity of an aqueous extract of the leaves of *Macaranga barteri* Mill. (Euphorbiaceae) on rat models. *Journal of Medicinal Plants Research* 2018; 12(9): 96-105.
13. De Sales IR, Formiga RD, Machado FD, Nascimento RF, Pessoa MM, Barros ME, Vieira GC, Gadelha FA, Marinho AF, Barbosa Filho JM and Júnior RF: Cytoprotective, antioxidant and anti-inflammatory mechanism related to antiulcer activity of *Cissampelos sympodialis* Eichl. in animal models. *Journal of Ethnopharmacology* 2018; 222: 190-200.
14. Park JU, Kang JH, Rahman MA, Hussain A, Cho JS and Lee YI: Gastroprotective effects of plants extracts on gastric mucosal injury in experimental sprague-dawley rats. *BioMed Research International* 2019; 2019: 8759708.
15. De Araújo ER, Guerra GC, Andrade AW, Fernandes JM, Da Silva VC, De Aragão Tavares E, De Araújo AA, de Araújo Júnior RF and Zucolotto SM: Gastric ulcer healing property of *Bryophyllum pinnatum* leaf extract in chronic model *in-vivo* and gastroprotective activity of its major flavonoid. *Frontiers in Pharmacology* 2021; 12: 744192.
16. Sugumar N and Karthikeyan Gowdhami T: Preliminary photochemical screening on the leaf extract of *Eupatorium triplinerve* Vahl. *International Journal of Pharmaceutical and Biological Archives* 2014; 5(5): 141-144.
17. Kawase M, Sakagami H, Motohashi N, Hauer H, Chatterjee SS, Spengler G, Vigiyanne AV, Molnár A and Molnár J: Coumarin derivatives with tumor-specific

- cytotoxicity and multidrug resistance reversal activity. *In-vivo* 2005; 19(4): 705-711.
18. Ghosh C, Maity R, Roy A and Mallick C: Dose-Dependent Protective Effect of *Hygrophila auriculata* Seeds on Cyproterone Acetate-Induced Testicular Dysfunction. *Reproductive Sciences* 2023; 1-3.
 19. Bauerova K, Nosalova V, Mihalova D and Navarova J: Contribution to safe antiinflammatory therapy with indomethacin. *Central European Journal of Public Health* 2004; 12(SUPP): S8-10.
 20. Maity R, Mondal P, Giri MK, Ghosh C and Mallick C: Gastroprotective effect of hydromethanolic extract of Ayapana triplinervis leaves on indomethacin-induced gastric ulcer in male Wistar rats. *Journal of Food Biochemistry* 2021; 45(8): 13859.
 21. Godkar PB and Godkar DP: Textbook of medical laboratory technology 2003.
 22. Sabiu S, Garuba T, Sunmonu TO, Sulyman AO and Ismail NO: Indomethacin-induced gastric ulceration in rats: Ameliorative roles of *Spondias mombin* and *Ficus exasperata*. *Pharmaceutical Biology* 2016; 54(1): 180-186.
 23. Suleyman H, Albayrak A, Bilici M, Cadirci E and, Halici Z: Different mechanisms in formation and prevention of indomethacin-induced gastric ulcers. *Inflammation* 2010; 33: 224-234.
 24. Corne SJ: A method for the quantitative estimation of gastric barrier mucus. *J Physiol London* 1974; 242: 116-117.
 25. Adhikary B, Yadav SK, Roy K, Bandyopadhyay SK and Chattopadhyay S: Black tea and theaflavins assist healing of indomethacin-induced gastric ulceration in mice by antioxidative action. *Evidence-based Complementary and Alternative Medicine* 2010; 2011.
 26. AlKreathy HM, Alghamdi MK and Esmat A: Tetramethylpyrazine ameliorates indomethacin-induced gastric ulcer in rats: Impact on oxidative, inflammatory, and angiogenic machineries. *Saudi Pharmaceutical Journal* 2020; 28(8): 916-926.
 27. Katary MA and Salahuddin A: Gastroprotective effect of vanillin on indomethacin-induced gastric ulcer in rats: protective pathways and anti-Secretory mechanism. *Clinical and Experimental Pharmacology* 2017; 7(2): 2161-1459.
 28. Guzmán-Gómez O, García-Rodríguez RV, Pérez-Gutierrez S, Rivero-Ramírez NL, García-Martínez Y, Pablo-Pérez SS, Pérez-Pastén-Borja R, Cristóbal-Luna JM and Chamorro-Cevallos G: Protective effect of the phycobiliproteins from *arthrospira maxima* on indomethacin-induced gastric ulcer in a Rat model. *Plants* 2023; 12(8): 1586.
 29. Hafez HM, Morsy MA, Mohamed MZ and Zenhom NM: Mechanisms underlying gastroprotective effect of paeonol against indomethacin-induced ulcer in rats. *Human & Experimental Toxicology* 2019; 38(5): 510-518.
 30. Garabadu D, Singh S and Gautam T: *Manilkara hexandra* (Roxb.) Dubard Ameliorates Acetic Acid-induced Rat Gastric Ulcer. *Journal of Dietary Supplements* 2021; 18(3): 278-292.
 31. Mahmoud YI and Abd El-Ghffar EA: *Spirulina ameliorates* aspirin-induced gastric ulcer in albino mice by alleviating oxidative stress and inflammation. *Biomedicine & Pharmacotherapy* 2019; 109: 314-321.
 32. Devasagayam TP and Tarachand U: Decreased lipid peroxidation in the rat kidney during gestation. *BBRC* 1987; 145(1): 134-138.
 33. Slater TF: Overview of methods used for detecting lipid peroxidation. In *Methods in Enzymology* 1984; 105: 283-293.
 34. Ghosh C and Mallick C: Protective effect of ethanolic extract of *Hygrophila auriculata* seeds in cyproterone acetate-induced sexual dysfunction in male albino rats. *Andrologia* 2020; 52(2): 13482.
 35. Jarosz M, Szkaradek N, Marona H, Nowak G, Młyniec K and Librowski T: Evaluation of anti-inflammatory and ulcerogenic potential of zinc-ibuprofen and zinc-naproxen complexes in rats. *Inflammopharmacology* 2017; 25: 653-663.
 36. Shahin NN, Abdelkader NF and Safar MM: A novel role of irbesartan in gastroprotection against indomethacin-induced gastric injury in rats: targeting DDAH/ADMA and EGFR/ERK signaling. *Scientific Reports* 2018; 8(1): 4280.
 37. Ugan RA, Un H: The protective roles of butein on indomethacin induced gastric ulcer in mice. *The Eurasian Journal of Medicine* 2020; 52(3): 265-270.
 38. Beck PL, Xavier R, Lu N, Nanda NN, Dinauer M, Podolsky DK and Seed B: Mechanisms of NSAID-induced gastrointestinal injury defined using mutant mice. *Gastroenterology* 2000; 119: 699-705.
 39. El-Ashmawy NE, Khedr EG, El-Bahrawy HA and Selim HM: Gastroprotective effect of garlic in indomethacin induced gastric ulcer in rats. *Nutrition* 2016; 32(7-8): 849-854.
 40. Herath M, Hosie S, Bornstein JC, Franks AE and Hill-Yardin EL: The role of the gastrointestinal mucus system in intestinal homeostasis: implications for neurological disorders. *Frontiers in Cellular and Infection Microbiology* 2020; 10: 248.
 41. Zhang C, Gao F, Gan S, He Y, Chen Z, Liu X, Fu C, Qu Y and Zhang J: Chemical characterization and gastroprotective effect of an isolated polysaccharide fraction from *Bletilla striata* against ethanol-induced acute gastric ulcer. *Food and Chemical Toxicology* 2019; 131: 110539
 42. Zhu K, Yang X, Yang C, Ye X and Zhang H: Gastroprotective actions of *Aloe barbadensis* Miller mitigate ethanol-induced gastric injury in rats. *Tropical Journal of Pharmaceutical Research* 2020; 19(12): 2645-2650.
 43. Saremi K, Rad SK, Tayeby F, Abdulla MA, Karimian H and Majid NA: Gastroprotective activity of a novel Schiff base derived dibromo substituted compound against ethanol-induced acute gastric lesions in rats. *BMC Pharmacology and Toxicology* 2019; 20(1): 1-3.
 44. Kim YS, Park HJ, Kim H, Song J and Lee D: Gastroprotective effects of paeonia extract mixture HT074 against experimental gastric ulcers in rats. *Evidence-based Complementary and Alternative Medicine* 2019; 2019.
 45. Tijani AS, Olori DO and Farombi EO: Manganese abated indomethacin-induced gastrohepatorenal toxicities in rats via suppression of oxidative stress, polyamine catabolism, inflammation and activation of caspase-3. *Advances in Redox Research* 2023; 100070.
 46. Tai A, Sawano T, Yazama F and Ito H: Evaluation of antioxidant activity of vanillin by using multiple antioxidant assays. *Biochimica et Biophysica Acta (BBA)-General Subjects* 2011; 1810(2): 170-177.
 47. Zaghlool SS, Abo-Seif AA, Rabeh MA, Abdelmohsen UR and Messiha BA: Gastro-protective and anti-oxidant potential of *Althaea officinalis* and *Solanum nigrum* on pyloric ligation/indomethacin-induced ulceration in rats. *Antioxidants* 2019; 8(11): 512.
 48. Hamza SE, Wahdan SA and El-Demerdash E: Effect of phylloquinone on indomethacin-induced gastric ulceration in rats: Role of SIRT-1. *Clinical and Experimental Pharmacology and Physiology* 2023; 50(5): 369-379.

How to cite this article:

Maity R, Mondal P, Bera PR, Midya S and Mallick C: Therapeutic effect of leaves of *Ayapana triplinervis* in indomethacin induced mild, moderate and severe grade gastric ulcer. Int J Pharm Sci & Res 2024; 15(3): 788-99. doi: 10.13040/IJPSR.0975-8232.15(3).788-99.

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