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SEPARATION AND ESTIMATION OF COMPONENTS PRESENT IN *EMBLICA OFFICINALIS*: A PRELIMINARY INVESTIGATION ON ALLERGIC REACTIONS

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ABSTRACT

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Assistant Professor, Department of Chemistry, Sona college of Technology, Salem-636 005, Tamil Nadu, India Indian Gooseberry, *Emblica officinalis* is a, fruit widely accepted in Kerala for its medicinal & nutritive values. The fruit has dark green colored spherical seed surrounded by pale green colored tissue. Different varieties of food products, pickles being the major one, are used all over the world .The present work on gooseberry originated from a real medical case history. The medical reports say that a normal person who was eating gooseberry on a normal course all at a sudden became allergic only to this fruit since two years. This person was hospitalized two times for anti- dot injection as the allergic symptoms go beyond control. The major symptoms the patient showed are; (a) itching; (b) swelling of the whole bodily parts & (c) drowsiness. These allergic effects were due to gooseberry and which confirmed by medical examination. Based on our preliminary information, allergy due to Indian gooseberry was the first case reported in our locality. This made our interest to investigate further the chemistry & physiology of allergic reactions of gooseberry in this patient.

INTRODUCTION: An allergy is an abnormal reaction to certain substances called allergens. Food allergy is the most common type and its adverse reactions can be categorized into three types. They are non allergic causes, idiosyncratic causes and allergic causes. Of this three, last two can be grouped together as food intolerance.

An allergy to a food gives rise to symptoms very quickly, generally within minutes of eating or even touching the food ¹. Food allergies are usually due to the protein component of the offending food. For some reason some of the food protein is absorbed from the intestine intact, instead of being digested as most proteins are. Once the intact protein is in the blood stream, it is recognized as a foreign protein to the body, or in other words as an antigen; the body produces antibodies (usually immunoglobulin E) to this antigen; and the immunoglobulin binds the antigen to

form an antigen-antibody complex. This antigenantibody complex travels around the body & stimulates certain cells, called mast cells to burst open & release substance which mediate an allergic reaction. Histamine is an example of such a mediator. Histamine causes an inflammatory response in the cells that it reaches & this inflammatory response causes the symptoms of food allergy ².



If mast cells release chemicals in the nose & throat, the allergic person may experience an itching tongue or mouth & may have trouble in breathing or swallowing. If mast cells in the gastro intestinal tracts are involved a person may have diarrhea or abdominal pain. Skin mast cells can produce hives or intense itching. The food protein fragment responsible for an allergic reaction is not broken by cooking or by stomach acids or enzymes that digest food. These proteins can cross the gastro intestinal lining, travel through the blood stream & can cause allergic reactions throughout the body³. The timing & location of an allergic reaction to food is affected by digestion.

For e.g. an allergic person may first experience a severe itching of the tongue or tingling lips, vomiting, diarrhea may follow. Later, as the allergens enter the blood stream & travel throughout the body, they can cause a drop in blood pressure, hives or eczema or asthma when they reach the lungs. Indian gooseberry is a wonderful fruit & one the precious gifts of nature to man (**Figure 1**).

It is consumed as a fresh fruit or consumed in the form of food products like preserve. The fruit also forms an important constituent of many Ayurvedic preparations such as*chyvanprash* and *triphala* and is regarded as a best rejuvenating agent ⁴.



FIGURE 1: INDIAN GOOSEBERRY

A number of investigations have been previously reported on the separation of components of different medicinal fruits ⁵, but there is no information available in the literature in the past with allergic effects due to Indian gooseberry. Therefore the authors have attempted to study the allergic effects of this fruit

MATERIALS AND METHODS:

Plant materials: Different types of Indian gooseberries were collected from the Palayam vegetable market, Calicut, Kerala. Each fresh fruit was first washed under tap water and then with distilled water to remove chemicals and impurities in it.

General chemicals and Instrumentation: Sodium hydroxide, Trichloroacetic acid, Coppersulphate, Sodium carbonate, Acetone, Phenol, Sulphuric acid, Glucose, Oxalic acid, 2, 4-dinitrophenylhydrazine, Bromine, Ethanol, Ether, Potassium hydroxide, Hydrochloric acid, Ninhydrin (Merck), Thiourea, Phenolphthalein (Nice), Sodium potassium tartarate (Qualigens), Folin ciocalteau reagent (Sisco research laboratories Mumbai) and Ascorbic acid (Pfizer Limited) were used as such. The Citrate-phosphate buffer, Bovine's serum albumin, Molisch's reagent and Leishman's stain were freshly prepared using reported procedures⁶.

Fresh blood sample from the allergic person was freshly collected as and when needed using a syringe. Ordinary porcelain mortar was used to ground the tissue. Cold centrifuge (Rota 4R – V/F M), Electronic balance (K- Roy classics), Ordinary centrifuge (REMI laboratory R4C), Magnetic stirrer (REMI equipments), Rotex Vortex mixer (cyclo mixer), Photo colorimeter (Sistronics), Compound microscope (K-Roy classics) and Digital camera (Nikon) were used as common instruments.

RESULTS & DISCUSSION:

Identification of Components:

1. Identification of Proteins and lipids: The tissue was agitated in an electric mixer & the juice was extracted with ether. The ethereal layer was analyzed for the presence of lipids. The aqueous layer was filtered to remove the fibrous & insoluble parts. This water extract was divided in to two portions. One portion is subjected to the tests for proteins. Biuret & Ninhydrin spray tests were carried out, which confirmed the presence of proteins.

The ether layer contains most of the lipid present in the fruit. This is because lipids are hydrophobic & well soluble in organic solvents like ether. The ether layer was evaporated to give oily deposits at the bottom of the tube. This is then refluxed with KOH, followed by the addition of HCl gave oily appearance on the top of the aqueous layer. This test confirmed the presence of lipid in the fruit 5 .

- 2. Identification of Carbohydrates: The second portion of the water extract was subjected to Molisch's test. This test gave violet color at the junction of two liquids, also confirmed the presence of carbohydrates. The detailed analyses for their characterization of carbohydrates were not presented in this report. Hence it can be concluded that the carbohydrate present in gooseberry is a reducing sugar moiety ⁸.
- 3. Identification of Vitamin C (Ascorbic acid): The raw tissue was mixed thoroughly with 4% oxalic acid in a mortar & centrifuged to separate the aqueous layer. The selective dissolution of vitamin C in oxalic acid is due to the presence of acidic pH, which hindered the co-dissolution of proteins & carbohydrates. This aqueous layer, which mainly contains vitamin C, was analyzed by standard test to confirm its presence ⁸.

Estimation of Various Components:

1. Estimation of Proteins-Lowry's Method: The protein present in Indian gooseberry was quantitatively estimated by the well- known Lowry's methods. A definite weight of fruit was homogenized in 25ml 0.2N NaOH in a mortar. This was then filtered & the filtrate is ultra centrifuged to remove other suspended impurities, followed by treatment with ammonium sulphate. The protein present in this solution was precipitated by adding trichloroacetic acid, which then dissolved in NaOH to prepare the stock solution.

Definite volume of this solution was mixed with the coloring reagents & the optical density was monitored at lambda 720nm, using colorimeter. This was then compared with the value of standard protein solution prepared using "Bovine's Serum Albumin" protein sample.

The final result showing the protein content per gram of dry tissue was calculated to be 4.28 mg which is slightly less than the result published earlier (~5 mg). This may be partly due to the fact that our analysis was based on raw tissue where as the earlier analysis was on dry tissue ⁸.

2. Estimation of Total Carbohydrate Content-Phenol Sulphuric acid Method: The total carbohydrate present in fruit was calculated by the phenolsulphuric acid method. A definite weight of the sample was hydrolyzed with HCl, when the long carbohydrate chains are broken down in to smaller fragments. The content was filtered & neutralized with a base. From this suitable volumes were pipetted & mixed with the coloring reagents at lambda 490 nm was measured ⁸.

A standard glucose solution was also prepared and the unknown concentration was extrapolated from the absorbance v/s concentration graph (**Figure 2**). Apparently the amount of total carbohydrate obtained by this method was found to be 135 mg per gram of tissue. This value is very much in agreement with the literature value of 137 mg per gram ⁸.



FIGURE 2: ABSORBANCE VS. CONCENTRATION OF GLUCOSE COMPLEX AT 490 nm

3. Estimation of Vitamin C (Ascorbic Acid): Indian gooseberry was known for its rich vitamin C content. This was extracted from the fruit using oxalic acid. After centrifugation, the supernatant solution was collected & reacted with bromine water. All the enolic hydrogens in ascorbic acid were replaced by bromine. Known volume of this solution is pipetted & mixed with the coloring reagents & the absorbance was recorded at 540 nm.

In a similar way commercially available ascorbic acid also converted in to the dehydro form using bromine water & was treated with the colouring reagent ⁹. From the graph of known concentration v/s optical density, unknown concentration was calculated by extrapolation (**Figure 3**).



FIGURE 3: ABSORBANCE VS. CONCENTRATION OF VITAMIN C COMPLEX AT 540 nm

4. Estimation of Lipids: Lipid present in the fruit was derived from the weight of oil obtained from a definite weight of tissue on extracting with diethyl ether and subsequent evaporation of the solvent. This value was found to be 0.875 mg/gram of wet tissue, which was found to be exactly same as that of the literature value.

The saponification value of the oil present in gooseberry was determined to deduce the molecular weight of lipid ⁹. For this a definite weight of the fruit was mixed thoroughly with water in a mortar. It was then treated with diethyl ether in a separating funnel & the ethereal layer was collected and evaporated to yield lipid. This lipid was then weighed and transferred to an RB flask & refluxed with alcoholic KOH using a water condenser ¹⁰.

The unreacted KOH was estimated by titrating with standard HCl. A blank experiment is also conducted with same volume of KOH ¹¹. From these two titer values (**Table 1**), the saponification value was found to be 230g.

TABLE 1: THE READINGS OF ACID-BASE TITRATION

Number	Burette r	Volume of HCI	
Number	Initial	Final	used in ml
Sample	0	19.8	19.8ml
Blank	0	23.9	23.9ml

Isolation of Components:

- 1. **Isolation of Vitamin C:** Ground 500 mg sample in 25-50ml 4% oxalic acid solution & centrifuged to collect vitamin C.
- Isolation of Carbohydrate: 100mg of the sample was taken in a boiling tube, hydrolyzed by keeping it in a boiling water bath for 3 hours, with 5ml of 2.5N HCl & cooled at room temperature. Neutralized with solid sodium carbonate & centrifuged to extract the carbohydrate.
- 3. **Isolation of Protein:** 500mg of the sample was weighed & homogenized in a mortar with 3ml phosphate buffer at pH 7.2 & centrifuged. This was then used for the interaction studies with human blood.
- 4. Isolation of Lipids: 3g of the sample was weighed & homogenized in a mortar with 8ml of phosphate buffer of pH 7.2, filtered through a muslin cloth & the residue is collected. To this, 10ml of fat solvent was added, filtered through a Whatman no. 1 filter paper. It was then allowed to evaporate at room temperature. In addition to this, Calcium, Phosphorus and iron content were also analyzed using reported procedure. The values obtained were in good agreement with literature values ¹².

Sample No.	Proteins	Carbohydrate	Vitamin C	Lipids	Calcium	Phosphorus	Iron
1	0.48%	13.5%	590mg	0.875%	42 mg	13.8mg	1.15mg
T	(0.5%)	(13.7%)	(600mg)	(0.1%)	(50mg)	(20mg)	(1.2mg)
2	0.53%	13.2%	585mg	0.876%	46mg	13.5mg	1.28mg
3	0.39%	13.8%	583mg	0.834%	40mg	13.1mg	1.18mg
4	0.27%	13.0%	612mg	0.838%	48.5mg	12.9mg	1.74mg
5	0.61%	12.9%	498mg	0.856%	43.3mg	13.6mg	1.25mg
6	0.47%	13.9%	578mg	0.768%	40.2mg	13.0mg	1.16mg
7	0.50%	12.7%	595mg	0.788%	46.9mg	12.5mg	1.13mg
8	0.46%	14.1%	589mg	0.878%	52.1mg	12.6mg	1.21mg
9	0.40%	14.5%	621mg	0.793%	47.5mg	13.3mg	1.23mg
10	0.30%	13.3%	593mg	0.790%	41.8mg	13.9mg	1.17mg

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The literature values are given in brackets

Interaction of Blood with Proteins & Lipids: Blood sample (2 mL) was taken in a watch glass and 1ml diluting fluid was added to prevent clotting. It was mixed with1ml protein & taken in a slide (4 cm long & 2 cm wide) and stained with Leishman's stain. This was allowed to dry and then washed with water & imaged the interaction with blood using a microscope having high magnification (45 X'). With the help of a digital camera photograph of the sample was recorded. Similarly interaction of lipid with blood was also recorded.

The cylindrical ones are blood cells. The number of eosnophill cells is limited and one of such cell is marked in **Figure 4**. The violet color of the cell in Figure 4 is due to staining. A dilute solution of protein was prepared & mixed with a fresh blood sample before staining. This was also imagined by transferring on to a glass slide. The image shown in **Figure 5** indicates clumped blood cells forming island like morphology. In a similar way another photograph was taken with lipid and was presented in **Figure 6**. These observations were confirmed by repeated analysis.



FIGURE 4: STAINED IMAGE OF BLOOD CELLS AND THE EOSNOPHILL CELL



FIGURE 5: STAINED IMAGE OF BLOOD CELLS AFTER MIXING WITH PROTEIN



FIGURE 6: STAINED IMAGE OF BLOOD CELLS AFTER MIXING WITH LIPID

It can be seen that the interaction with lipid is different from that with protein. In the former case cellular structures were almost completely lost as evident from Figure 6, where as in the case of proteins the cellular structure is retained except for the clumping (Figure 5).

Though, we cannot draw any solid conclusion out of this observation, it is indeed an indication that the protein in the fruit is responsible for the allergic reactions in the said patient. In addition, there are reports to support this observation, that in a few other cases of food allergy, it is the proteins present in these food materials trigger the allergic reactions.

CONCLUSION: Based on our studies on Indian gooseberry, organic constituents like proteins, lipids, carbohydrates & vitamin C were identified estimated & isolated. Only proteins & lipids were isolated for the possible interaction studies with fresh blood samples from the person who is allergic to Indian gooseberry. Optical micrographs of blood cells before & after interaction with proteins & lipids were recorded using a high resolution microscope.

Here, the blood cells were completely collapsed and could not able to see them distinctly, in the case of lipids, where as in the case of proteins the cells were visible but clumped as islands. Nevertheless, clear evidence cannot be generated to confirm the fact that allergic reaction occur due to proteins. Allergic reaction occurs due to proteins in Indian gooseberry. But at the same time the said person is not allergic to other food materials, which contain vitamin C, lipids, carbohydrates & proteins. It can bee seen that it is the protein part that is very much different in fruit to fruit. Hence it may be concluded that the allergic reaction occur in that person's body due to some type of association by proteins in Indian gooseberry with blood cells. However, the possibility of fragmented proteins, some chemicals present in Indian gooseberry or synergistic effects cannot be ruled out.

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REFERENCES:

- 1. Frederic M Dearborn: Diseases of the skin. B. Jain publishers, New Delhi, Second Edition 1998.
- Lancet: The International Study of Asthma & allergies in Childhood (ISAAC) Steering Committee. Worldwide variation in prevalence of symptoms of asthma, allergic rhinoconjuctivitis, and atopic eczema 1998: 351:1225-32.
- 3. Bock SA, Muncoz-Furlong A and Sampson HA.Fatalities: *Due to Anaphylactic Reaction to Foods.* Journal of Allergy & Clinical Immunology 2003: 112: 1203-1207.
- 4. Kaviratana AC and Sharma P (translators): Caraka-Samhita. Indian Books Centre, New Delhi, Second Revised Edition 1996.
- 5. Ghosal S, Triethi VK and Chauhan S: *Active x constituents of Emblica officinalis*: Part 1: The Chemistry and antioxidative effects of two new hydrolysable tannis, Emblicanin A and *B.* Indian Journal of chemistry 1996: 35B:941-948.
- 6. Sadasivam & Manikam: Biochemical methods. New Age International (P), New Delhi, Second Edition 1996.
- David T. Plummer: An Introduction to Practical Biochemistry. TataMcGraw-Hill Publishing Company, New Delhi, Third Edition 1988.
- 8. Keith Wilson & John Walker: Practical Biochemistry Principles & Techniques Cambridge University Press. Fourth Edition 1994.
- 9. J-H, Weil: General Biochemistry. New Age International Limited, New Delhi, Sixth Edition. 1996.
- Edward Staunton West, Wilbert R.Todd, Howard S Mason & John T.Van: Bruggen.Text Book of Biochemistry. Oxford & IBH Publishing Company, New Delhi, Fourth Edition1966.
- 11. Max E. Rafelson & Stephen B.Binkley: Basic Biochemistry. The MacMillian Company, New Delhi, Second Edition 1968.
- 12. Vogel: Text Book of Practical Organic Chemistry. ELBS, Fourth Edition1978

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