IJPSR (2010), Vol. 1, Issue 8





INTERNATIONAL JOURNAL OF PHARMACEUTICAL SCIENCES AND RESEARCH



Received on 10 April, 2010; received in revised form 14 June, 2010; accepted 01 July, 2010

EXPERIMENTAL MODELS FOR ALZHEIMER'S DISEASE: A MECHANISTIC VIEW

Vivek Kumar Sharma

Department of Pharmacology, Govt. College of Pharmacy, Rohru, Shimla, Himachal Pradesh, India

Keywords:

Alzheimer's Disease, Streptozotocin, Colchicine, Ibotenic Acid

Correspondence to author:

Vivek Kumar Sharma

Department of Pharmacology, Govt. College of Pharmacy, Rohru, Shimla, Himachal Pradesh, India Email: viveksharma_pharma@yahoo.co.in

ABSTRACT

Alzheimer's disease (AD) is a leading cause of morbidity and mortality and its prevalence is continuously increasing. AD is characterized to varying degrees by Amyloid β, neurofibrillary tangles, gliosis, synaptic and neuronal loss leading to a decline in memory & apraxia agnosia and several neuropsychiatric changes like anxiety and depression etc. Increased age, oxidative stress and neuroinflammation are considered to be the major risk factors implicated in the progression of AD. Various signaling systems, such as vasoconstrictor peptides, inflammatory mediators, growth factors, are involved in the pathogenesis of AD. At present, no promising therapy is available due to lack of understanding of signaling culprits involved in the pathogenesis of AD. Animal models are being developed to better understand the disease pathogenesis and develop drugs for this ailment. In the present review, various common lab animal models for AD are discussed, which has been used, can be used and which will open new vistas for developing new drugs to treat this cognitive syndrome.

INTRODUCTION:

Alzheimer's Disease: Alzheimer's disease (AD) is an irreversible, progressive neurodegenerative disorder that destroys the higher structures of the brain. Prominent neuropathologic features of AD are senile plaques, neurofibrillatory tangles, synaptic, neuronal loss and volume loss (Atrophy) ^{1, 2, 3, 4}. There are around 35 million patients suffering from Alzheimer's disease all over the world, out of which United States of America alone has around 4.5 million patients ⁵. In India AD patents are estimated to be less than 3.5 million ⁶ and the number is increasing at a fast pace.

After heart diseases, cancer, and stroke it is the fourth leading cause of death in US 7 . Alzheimer's disease is accompanied by psychiatric manifestatations such as psychosis (delusions and hallucinations) and disruptive behaviors (e.g., psychomotor agitation and physical aggression), especially in the later stages of the disease ^{8, 9, 10}. AD usually begins with difficulties with memory and orientation, with subsequent gradual and progressive decline in visuo spatial skills, language and calculation, praxis (learned motor skills), gnosis (perception), and frontal and executive functions, such as reasoning, judgment, foresight, and insight ¹¹. Hippocampus, limbic system, and cortex are the primary areas affected in the pathophysiology of AD ^{1, 12}.

A small fraction of patients develop myoclonus, seizures, or spastic paraparesis, and many develop weight loss and extra pyramidal signs. Patients may become lost in their own homes and fail to recognize family members. In latter stages of the disease, basic activities of daily living such as dressing, grooming, bathing, mobility and transfers, toileting, and eating are progressively affected. After years of cognitive and functional decline, patients become vegetative, mute, unresponsive, incontinent, and bed-bound before death ensues- often from pneumonia and overwhelming infection ^{13, 14}.

Although several factors mediate the development and progression of AD, genetic factors (PS1, PS2 and APOE) and associated comorbid conditions, such as age, oxidative stress and hypertension, are considered to be independent risk factors and major determinants in the progression of AD in patients. Various experimental models are employed to induce dementia resembling Alzheimer's disease in order to identify the potential pharmacological targets. However, the literature for animal models of AD is currently inadequate. This review focuses on various common experimentally- developed animal models producing dementia of AD type.

Experimental Models: The animal models for Alzheimer's disease share many features which are common to human disease and have been delineated by targeting oxidative stress, neuroinflammation, cell death, formation of plaques, tangles, cognitive deficits and other behavioral changes.

• Intracerebroventricular Streptozotocin Induced Dementia: Streptozotocin is a glucosamine nitrosourea compound (STZ, $(C_8H_{15}N_3O_7, 2- \text{ deoxy- } 2 - (3- \text{ methyl- } 3$ nitrosoureido) - D- glucopyranose) discovered in a strain of the soil microbe *Streptomyces achromogene* in 1956 ^{15,16}. It was classified as an alkylating agent in the nitrosourea class of anti-cancer drugs and was used to treat cancer of Islets of Langerhans in the pancreas. STZ is toxic to the insulin producing beta cells of the Islets of Langerhans in the pancreas, and is now better known as a diabetogenic agent, it is used to induce diabetes under experimental conditions when injected i.p. in rats at a dose of 45 mg/kg,50, 55 and 60 mg/kg^{17,18,19}. The molecular weight of STZ is 265g/mol and the structure is composed of nitrosourea moiety with a methyl group attached at one end and a glucose molecule at the other end ²⁰. The molecular structure was first described by Herr et al. ²¹.

Interacerebro-STZ ventricular administration has become an established and most appropriate experimental model for Alzheimer's type of dementia²² as it is characterized by pathological changes similar to those observed in Alzheimer's disease. Streptozotocin, when injected intracerebroventricularly (ICV) in а subdiabetogenic dose (3mg/kg) in rat, causes prolonged impairment in learning and memory ^{22,23} by exerting profound and long lasting influence on brain biochemistry, metabolism and function ^{24, 25, 26}. After ICV administration of STZ severe abnormalities in brain glucose and energy metabolism have been found. Glucose utilization reduces in brain areas ²⁴ and the activities of glycolytic key enzymes are decreased markedly 27 causing diminished concentration of the energy rich compound ATP and creatinine phosphate ^{28,22}. This may form the biological basis for the marked reduction in learning and memory capacities ²².

Tau protein is hyperphosphorylated as a long-term consequence of STZ icv administration²⁹ and it also causes neuronal damage and cell loss as well as the accumulation of AB in the brain 30 . An increase in the total tau protein in the brain and increase in the β amyloid formation enhance inflammatory process and free radical formation ³¹. There is gradual increase in the levels of malondialdehyde (MDA) - the end product of lipid peroxidation. An increase in the levels of lipid proxidation in nerve cells results in apoptosis and cell death ^{31, 32}. There is a simultaneous decrease in glutathione levels too.

The increase in the levels of MDA and the decrease in the levels of glutathione, both directly associated with oxidative stress parallels with the diminution of learning and memory in rats ³³. Activities of glycolytic enzyme are also reduced ²⁷ leading to decreased formation of acetyl CoA and 34, 35, 36 thereby of acetylcholine Furthermore, ICV STZ application causes the down regulation of gene expression related to insulin signaling such as IGF-1 receptor. In contrast to this down regulation, an upregulation in gene expression related to potassium channels, GABA receptors and glutamate receptors is also observed ³⁷. After ICV administration, STZ it reaches the fornix and passes into the 3rd ventricle because of the flow of CSF in a rostrocaudal direction ³¹. STZ when administered ICV damages the septohippocampal system ³⁸ whereby memory impairment in rat could occur due to direct damage to the system. This is supported by reduced choline acetyl

transferase (ChAT) activity in the hippocampus ³⁵, reduction in the weight of septum by more than 40% ³⁹, decrease in the transport of nerve growth factor from the hippocampus to septum ⁴⁰, microglial activation and specific damage to myelinated tract in the fornix through generation of oxidative stress. This finally causes disruption in the connection between the septum and hippocampus ⁴¹.

 Intracerebroventricular Administration of Colchicine: Colchicine, an alkaloid extracted from some plants of the lily family, has been used for centuries to treat acute gouty arthritis. Since 1973, it has been recognized as an effective remedy for propphylaxis of attacks of familial Mediterranean fever ⁴². In animal models of central nervous system damage, colchiccine, а microtubuledisrupting agent, is used as a neurotoxin. Following its introduction into the brain, colchicine binds to tubulin, the principal structural protein of the microtubule, and induces microtubular depolymerization and destabilization, with subsequent block of axonal transport and mitosis, resulting in neuronal cell death 43-46.

The neurotoxicity is mediated through free radical production and the resultant oxidative stress ^{1, 45}. In addition, colchicine causes loss of cholinergic neurons, destruction of cholinergic pathways, and decrease in cholinergic turnover 47. The distribution of colchicine in the brain is unequal; its concentration in the hippocampus, the area most affected in Alzheimer's disease, is almost three times higher than in other brain regions ⁴⁸. The

central manifestations of colchicines neurotoxicity in the animal model closely simulate Alzheimer's disease in humans 45, 46, ⁴⁹. Both are characterized by oxidative stress, microtubule disruption, decrease in cholinergic activity, and progressive deterioration of cognitive functions 49, 44. Systemic administration of colchicine in rats also induced cognitive defects similar to those of Alzheimer's and characterized by amnesia of recent learning and loss of formerly established memories ⁴⁶.

Further, ICV infusion of colchicine significantly impaired the memory with decrease in norepinephrine, Dopamine and level cerebral serotonin in cortex, 50 hippocampus and caudate nucleus Dopamine, norepinehrine and Serotonin is involved in plural process supporting learning and memory 50, 51, 52. Decreased level of Dopamine and Serotonin is ^{52, 53, 54}. Central associated with AD administration of colchicines also causes loss of cholinergic neurons and cognitive dysfunction that is associated with excessive free radical generation ^{55, 56}. Cyclooxygenase isoforms are differentially regulated following colchicine injection 55 and it increases the expression of cyclooxygenases 57, 58

Cyclooxygenase isoforms are implicated in acute and chronic neurological diseases including neurodegenerative processes. Expression of cyclooxygenase- 1 and cyclooxygenase-2 is up regulated in cortical and hippocampal pyramidal circuits ⁵⁹. Increased expression of cyclooxygenase activity initiates a pathological cascade associated with elevated levels of inflammatory mediators ⁶⁰. Furthermore, Numerous findings suggest that cyclooxygenase and its products might be important mediators of neuronal injury ^{61, 62}. Furthermore, it has been reported that central administration of colchicines is also associated with an increase in free radical generation and growing body of evidences now further support the concept of reactive oxygen species and its involvement in oxidative pathway of memory impairment ^{56,} Indeed, Alzheimer's patients produce more glutathione peroxidase, an enzyme that helps to neutralize free radicals, as a defensive reaction against increased production of peroxides within the cells and have disturbances in the antioxidant balance which may predispose to increased oxidative stress ⁶⁴.

Central administration of colchicines also mediates a cascade of actions, particularly elevation of the GLU/GABA ratio in brain cortex ⁶⁵. The relative increase in glutamate activates intracellular metabolic events, including excitotoxicity, triggers the generation of free radicals, which overcome antioxidant defenses and provoke oxidative stress ^{66, 67} and hyperactivation of NMDA receptors leads to massive Ca²⁺ influx that triggers a rapid activation and over stimulation of Ca²⁺ dependent enzymes including phospholipase A2, proteases, protein phosphatases, endonucleases, nitric oxide synthase, protein kinase, xanthine oxidase and cyclooxygenase. This increase cyclooxygenase-2 activity that contributes to the neuronal and synaptic loss associated with neurodegeneration and increased

production of oxidative stress and the neurotoxic actions of prostaglandins ^{68, 69}. These observations are particularly relevant in view of evidences showing that oxidative stress is involved in the development and progression of Alzheimer's disease and that activation of cyclooxygenase- 1, cyclooxygenase- 2 isoforms or both are required for execution of oxidative neuronal death ⁵⁵.

Colchicine the also increases expression of inducible nitric oxide synthase and cyclooxygenase-2, thereby increasing nitric oxide (NO) and prostaglandin's synthesis, respectively 57, 67. Nitric oxide, a precursor for free radicals reacts with the superoxide anions produced as a result of excitotoxicity, peroxidase activity of cyclooxygenase and increased oxidative stress after central administration of colchicine that give rise to toxic intermediates (peroxynitrite, nitric dioxide) ^{66,67} and overproduction of nitric oxide is neurotoxic to cholinergic neurons too ^{68, 69}.

Acetylcholine in nerve terminals in hippocampus, govern vital aspects of memory and other cognitive functions. Central administration of colchicine cause a marked decrease in acetyl cholinesterase activity and the decrease in the acetyl cholinesterase levels that associated with loss of cholinergic neurons in brain might be colchicine inhibition of fast due to axoplasmic flow or a direct toxic effect on cholinergic terminals ^{70, 71}. It is also plausible that increased free radicals formation causes macromolecular changes in cholinergic neurons and leads to the reduction in acetyl

cholinesterase activity that contributes to learning and memory deficits ⁵⁵.

• Ibotenic Acid Lesion of Nucleus Basalis Magnocellularis: Ibotenic acid lesion of basalis magnocellularis is nucleus an validated model for Alzheimer's disease ^{72,73}. Learning and memory is closely associated with the functional status of the central cholinergic system. The basal forebrain provides the major source of cholinergic inputs to the neocortex and hippocampus. The main cholinergic pathways in the mammalian forebrain are the projections from the medial septal nucleus and the nucleus of the vertical limb (diagonal band of Broca) to the hippocampus via the fimbriafornix and the projection from nucleus basalis cellularis (nbm) to the neocortex. The nbm located in the ventromedial region of the globus pallidus accounts for 70-80% of the cholinergic innervation to the cortex ⁷⁴.

Lesions of the nbm have been proposed as an experimental model for AD, based on the observation that degenerative changes in nucleus basalis of Mynert (nbM), the human counterpart of nbm, are present in patients of AD. In addition, the nbm lesioned rat shows decreases in cholinergic markers, including Ach levels, release and turnover of Ach, choline uptake, ChAt and acetylcholinesterase activity, and number of muscarinic cholinergic receptors, in the frontal cortex ⁷⁴, similar to what reported in patients of AD ⁷⁵. In this model unilateral nbm lesion are induced by injecting ibotenic acid (10 μ g/rat), dissolved in 5 μ l of Artificial cerebrospinal fluid (CSF), in anaesthetised rats, using the stereotaxic co-ordinates 1.0

mm posterior to bregma, 2.6 mm right lateral and 7.9 mm below the cortical surface 73 .

 Lysophosphatidic Acid induced Tau Hyperphosphorylation: Microtubules are essential for neurite formation and maintenance. These structures are composed of tubulin and a variety of minor proteins denoted by microtubule-associated proteins (MAPs). The level of MAPs and, more importantly, their phosphorylation level appear to modulate microtubule stability and dynamics. In particular, Tau, one of the most abundant axonal MAPs, may play a key role in axonogenesis ³.

Таи phosphorylation is developmentally regulated, being lower in adult than in fetal brain. Additionally, Tau hyperphosphorylation occurs in the aberrant structures known as paired helical filaments (PHFs) that appear in neuronal disorders such as Alzheimer's disease where these structures are a neuropathological hallmark 8 The bioactive phospholipid lysophosphatidic acid causes growth cone collapse and neurite retraction in neuronal cells. These changes are brought about by the action of a cell surface receptor coupled to specific G proteins that control morphology and motility through the action of a group of small GTPases, the Rho family of proteins. Many studies have focused on actin reorganization modulated by Rho-GTPases, but almost no information has been obtained concerning microtubular network reorganization after LPA-induced Neurite retraction. It has been demonstrated that there is an increase in site-specific Alzheimer's disease-like *Tau* phosphorylation during LPA-induced neurite retraction in differentiated SY-SH5Y human neuroblastoma cells ⁷⁶.

• L-Methioine Induced Dementia: L-Methionine treatment for 4 weeks, significantly raises serum Homocysteine level ⁷⁷ and increased levels of homocysteine have been documented to produce changes in structure and function of cerebral blood vessels along with oxidative stress, which play a key role in cerebral vascular dysfunction ⁷⁸. Oxidative stress and vascular dysfunction are recognized as important contributing factors in the pathogenesis of AD and other dementia of vascular origin ⁷⁹. In AD and other neurodegenerative diseases, structural deformities in the cerebral capillaries lead to impairment of cerebral perfusion and produced a significant impairment of acquisition and retrieval of memory 77 with subsequent neuronal dysfunction and death.

Cerebral vascular endothelial dysfunction has also been shown to enhance progression of dementia of Alzheimer disease (AD) ⁸⁰. Enhanced levels of brain AChE activity and oxidative stress have also been noted in patients suffering form dementia of AD and other dementias⁸¹. Further hyperhomocysteinemia has also been shown to be neurotoxic, and the neurotoxicity may be due to overactivation of N-methyl-D-aspartate receptors or by enhanced vulnerability of hippocampal neuron to excitotoxic insults and amyloid β peptide toxicity^{82, 83}. Moreover, methionine rich diet in rats has been demonstrated to

enhance cholesterol concentration in the plasma and liver ⁸⁴. Several studies have also revealed high serum cholesterol level as another important risk factor of AD ⁸⁵ beside oxidative stress and inflammation ⁸⁶. Therefore L-Methionine induced memory dysfunction may be attributed to its multiple effects i.e. decrease in serum nitrite level (endothelial dysfunction), rise in oxidative stress level, enhancement of brain AChE activity, serum total cholesterol as well as direct neurotoxicity.

CONCLUSION: The morbidity and mortality due to Alzheimer's disease is continuously increasing worldwide and the therapeutic agents currently available are limited. During the past few decades, the use of animal models has provided new insights into understanding the complex pathogenesis of Alzheimer's disease. Important pathogenic mechanisms still remain active and unmodified by present therapeutic strategies. Identification of signaling culprits involved using various animal models may provide the lead in discovering novel therapeutic agents.

ACKNOWLEDGEMENT: Author is indebted to Mr. Rahul Deshmukh (Asst. Prof. ISF College of Pharmacy, Moga) for his moral support and encouragement. Author also acknowledges blessings of Mr. S.N Singh (Asst. Prof. and HOD, Pharmacy) and Mr. Rajendra Guleria (Asst. Prof. Govt. College of Pharmacy, Rohru, Distt. Shimla).

REFERENCES:

1. Kumar A, Dogra S and Parkash A: Neuroprotective Effects of Centella asiatica against Intracerebroventricular Colchicine-Induced Cognitive Impairment and Oxidative Stress. International Journal of Alzheimer's disease2009; Article ID 972178.

- 2. Zilka N, Novak A: The tangled story of Alois Alzheimer. Bratisal Lek Listy2006;343-345.
- Zilka N, Ferencik M, Hulin I: Neuroinflamation in Alzheimer's disease; protector or promoter?" Bratisl Lek Listy2006;107(9-10):374-383.
- Leon D, Desanti S, Zinkowski R, MehtaP D, Pratico D, SegalS et al: MRI and CSF studies in the early diagnosis of Alzheimer's disease. Journal of internal medicine2004; 256:205-223.
- Hebert LE, Scherr PA, Bienias JL, Bennett, D:Alzheimer Disease in the U.S. Population: Prevalence Estimates Using the 2000 Census. *Arch. Neurol2003*; 60: 1119–1122.
- 6. Shaji S: Prevalence of dementia in an urban population in Kerla. India. *Brit. J. Psychiat2005;* 186: 136-140.
- 7. Stephen SF: Update on Alzheimer's disease.www.neurozone.org, 2006
- Reisberg R, Borenstein J, Salob SP: Behavioral symptoms in Alzheimer's disease: phenomenology and treatmen. J Clin Psychiatry1987; 48:9–15.
- Rubin EH, Morris JC, Berg L:The progression of personality changes in senile dementia of the Alzheimer's type. J Am Geriatr Soc; 35:721–725.
- Devanand DP, Jacobs DM, Tang MX: The course of psychopathologic features in mild to moderate Alzheimer disease. Arch Gen Psychiatry1997; 54:257–263.
- Knopman DS, Chui H, Corey-Bloom J, et al.: Practice parameter: Diagnosis of dementia (an evidence-based review). Neurology2001; 56: 1143–1153.
- 12. McIlroy S and Craig D. Neurobiology and genetics of behavioural syndromes of Alzheimer's disease. *Current Alzheimer Research2004;* 1: 135–142.
- Turner AJ & .NalivaevaNN: Targetting amyloid degrading enzymes as therapeutic strategies in neurodegeneration. AnnNY Acad Sci2004;1035:1-20.
- Turner RS: Neurologic Aspects of Alzheimer's disease," c01.qxd 5/15/03
- 15. LewisC, Barbiers AR: Streoptozotocin, a new antibiotic In vitro and in vivo evaluation. Antibiot Ann1960; 22: 247-54.
- WileyPF: Isolation and chemistry of streptozotocin. Streptozotocin: Fundamental and therapy Amsterdam, the Netherlands: Elsevier North-Holland biomedical press1981; 3-18.
- Casey GR, JoyceM, NagleRG, Chen G:Pravastatin modulates early diabetic nephropathy in an experimental model of diabetic renal disease. J Surg Res2004; 123:176-81.
- HaidaraMA, MikhailidiscDP, RatebaMA : Evaluation of the effect of oxidative stress and vitamin E supplementation on renal function in rats with streptozotocin-induced type 1 diabetes. J Diabetes Complications2008;521:231-235.
- 19. GojoA, UtsunomiyaK, TaniguchiK; The Rhokinase inhibitior, fasudil, attenuates diabetic nephropathy in strepazotocininduced diabetic rats. *Eur J Pharm2007;* 568:242-7.

- 20. Weiss RB: Streptozotocin: a review of its pharmacology, efficacy and toxicity. *Cancer treat Rep1982*; 66: 427-38.
- 21. Herr RR, Jahnke HK, Argondelis AD: The structure of streptozotocin. J. Am. Chem. Soc1967; 89: 4808-09.
- LannertH, HoyerS: Intracerebroventricular administration of streptozotocin causes long term diminiution in learning and memory abilities and cerebral energy metabolism in adult rats. *Behav. Neurosci1998;* 112: 1199-1208.
- 23. Blockland A and Jolles J: Spatial learning deficits and reduced hippocampal ChAT activity in rats after an i.c.v. injection of streptozotocin. *Pharmacol. Biochem. and Behav1993;* 44: 491-94.
- Duelli R, Schrock H, KuschinskyW, Hoyer S. Intracerebroventricular injection of streptozotocin induces discrete local changes in cerebral glucose utilization in rats. Int. J. Dev. Neurosci1994; 12: 737–43.
- Muller D, Nitsch R, Wurtman, Hoyer S: Streptozotocin increases free fatty acids and decreases phospholipids in rat brain. J. Neural. Transm1998; 105:1271-81.
- Hoyer S and Lannert H: Inhibition of the neuronal insulin receptor causes Alzheimer-like disturbances in oxidative/ energy brain metabolism and in behavior in adult rats. Ann. N. Y. Acad. Sci1999;893: 301-03.
- Plaschke K and Hoyer S: Action of the diabetogenic drug streptozotocin on glycolytic and glycogenolytic metabolism in adult rat brain cortex and hippocampus. Int. J. Dev.Neurosci1993; 11: 477-83.
- Nitsch R, Hoyer S: Local action of the diabetogenic drug, streptozotocin, on glucose and energy metabolism in the brain cortex. Neurosci. Lett1991;128: 199–202.
- Grunblatt E, Salkovic-Petrisic M, Osmanovic J, Riederer P and Hoyer S: Brain insulin system dysfunction in streptozotocin intracerebroventricularly treated rats generates hyperphosphorylated tau protein. Journal of Neurochemistry2007;101: 757–770.
- Lester-Coll, E, RiveraSJ, SosciaK, Doiron JR: Intracerebral streptozotocin model of type 3 diabetes: relevance to sporadic Alzheimer's disease. J. Alzheimers Dis2006; 9:13– 33.
- Baluchnejadmojarad T, Roghani M: Effect of Naringenin on Intracerebroventricular Streptozotocin-Induced Cognitive Deficits in Rat: A Behavioural Analysis. *Pharmacol*2006; 78:193–97.
- 32. Grunblatt S, Koutsilieri E, Hoyer S, Riederer P: Gene expression alterations in brain areas of intracerebroventricular streptozotocin treated rat. *J. Alzheimers Dis2006;* 9: 261–71.
- Sharma M, Gupta YK: Intracerebroventricular injection of streptozotocin rats produces both oxidative stress in the brain and cognitive impairment. Life Sciences2001; 68: 1021–29.

Available online on www.ijpsr.com

- Blokland, A, Jolles J: Behavioral and biochemical effects of an ICV injection of streptozotocin in old Lewis rats. Pharmacol. Biochem. Behav2004;47: 833–37.
- BloklandA, Jolles J: Spatial learning deficit and reduced hippocampal ChAT activity in rats after an icv injection of streptozotocin. Pharmacol. Biochem. Behav1993; 44:491– 94.
- Weinstock M, Kirschbaum-SlagerN, LazaroviciP, BejarC, Youdim MB, ShohamS: Neuroprotective effects of novel cholinesterase inhibitors derived from rasagiline as potential anti-Alzheimer drugs. *Ann. N. Y. Aca. Sci*2001; 939:148–61.
- Grunblatt E, Hoyer S, Riederer P: Gene expression profile in streptozotocin rat model for sporadic Alzheimer's disease. J. Neural Transm2004; 111:367–86.
- Prickaerts J, Fahrig J, BloklandA: Cognitive performance and biochemical markers in septum, hippocampus and striatum of rats after an i.c.v. injection of streptozotocin: a correlation analysis. *Behav. Brain Res1999;* 102: 73–88.
- Terwel D, Prickaerts J, Meng F, Jolles J: Brain enzyme activities after intracerebroventricular injection of streptozotocin in rats receiving acetyl-L-carnitine. Eur. J. Pharmacol1995; 287:, 65–71.
- Hellweg R, Nitsch R, Hock, Jaksch, Hoyer S: Nerve growth factor and choline acetyltransferase activity levels in the rat brain following experimental impairment of cerebral glucose and energy metabolism. Journal of Neuroscience Research1992; 31: 479–86.
- Shoham S, Bejar C, Kovalev E, Weinstock M: Intracerebroventricular injection of streptozotocin causes neurotoxicity to myelin that contributes to spatial memory deficits in rats. Experimental Neurology2003;184: 1043–52.
- 42. Leibovitz A, Lidar M, Baumoehl Y, Livneh A and Segal R: Colchicine Therapy and the Cognitive Status of Elderly Patients with Familial Mediterranean Fever. IMAJ2006;,8:69–472.
- James F, Dennis WL: Long term memory: disruption by inhibitors of protein synthesis and cytoplasmic flow. *Pharmacol Biochem Behav1981;*15:289–96.
- Walsh TJ, Schulz DW, Tilson TA, Schmechel DE: Colchicine induced granule cell loss in rat hippocampus: selective behavioral and histological alterations. *Brain Res1986*;398:23–36.
- 45. Kumar V and Gupta YK: Intracerebroventricular administration of colchicine produces cognitive impairment associated with oxidative stress in rats. *Pharmacol Biochem Behav2002;*73:565–71.
- BensimonG, ChermatR: Microtubule disruption and cognitive defects: effecut of colchicine on learning behavior in rats. *Pharmacol Biochem Behav1991;38:141–5*.
- MeyersCA, Kudelka AP, Conrad CA, Gelke CK, Grove W, Pazdur P: Neurotoxicity of CI-980, a novel mitotic inhibitor. *Clin Cancer Res1992*;3419–22.

- Evrard PA, Ragusi C, Boschi G, Verbeeck VK, Scherrmann JM: Simultaneous microdialysis in brain and blood of the mouse:extracellular and intracellular brain colchicine disposition. *Brain Res1998*;786:122–7.
- 49. NakayamaT, SawadaT: Involvement of microtubule integrity in memory impairment caused by colchicines. *Pharmacol Biochem Behav2002;* 71:119–38.
- Ganguly R & Guha D: Alteration of brain monoamines & EEG wave pattern in rat model of Alzheimer's disease & protection by *Moringa oleifera*. Indian J Med Res2008;128:744-751.
- 51. Trond M: Neurotransmitter systems involved in learning and memory in the rat: a meta-analysis based on studies of four behavioural tasks. *Brain Res Rev2003; 41:* 268-87.
- 52. Ha TM, Cho DM, Park SW, Joo MJ, Lee BJ, Kong BG: Evaluating associations between 5-HTTLPR polymorphism and Alzheimer's disease for Korean patients. *Dement Geriatr Cogn Disor2005; 20:* 31-4.
- 53. DaberkowRP, KesnerKA: Relation between methamphetamine- induced monoamine depletions in the striatum and sequential motor learning. *Pharmacol Biochem Behav2005,81:* 198-204.
- 54. Joyce JN, MurrayAM, et al. Loss of dopamine D2 receptors in Alzheimer's disease with Parkinsonism but not Parkinson's or Alzheimer's disease. *Neuropsychopharmacology1998; 19* : 472-80.
- Kumar A, Seghal N, Padi SV, Naidu PS: Differential effects of cyclooxygenase inhibitors on intracerebroventricular colchicine-induced dysfunction and oxidative stress in rats. European Journal of Pharmacology2006; 551:58–66.
- Kumar V, Gupta YK: Intracerebroventricular administration of colchicine produces cognitive impairment associated with oxidative stress in rats. Pharmacol. Biochem. Behav2002; 73: 565–571.
- HoL, Osaka S, Aisen PS, Pasinetti GM: Induction of cyclooxygenase-2 but not cyclooxygenase-1 gene expression in apoptotic cell death. J. Neuroimmunol1998; 89: 142–149.
- Subbaramaiah K, Hart JC, Norton L, Dannenberg AJ: Microtubule interfering agents stimulate the transcription of cyclooxygenase-2. Evidence for involvement of ERK 1/2 and p38 mitogen-activated protein kinase pathways. J. Biol. Chem2002;275: 14838–14845.
- 59. Kitamura Y, Shimohama S, Koike H, Kakimura J, MatsuokaY, Nomura Y:Increased expression of cyclooxygenase and peroxisome proliferator-activated receptor-gamma in Alzheimer's disease brains. Biochem. Biophys. Res. Commun1999; 254: 582–586.
- Montine TJ, Sidell KR, Crews BR, Markesbery R, Marnett WR, Roberts IILJ, Morrow LJ: Elevated CSF prostaglandin E2 levels in patients with probable Alzheimer's disease. Neurology1999; 53: 195–198.

Available online on www.ijpsr.com

- Candelario-Jalil E, Gonzalez-FalconA, Garcia-Cabrera M, Alvarez D et al.Assessment of the relative contribution of cyclooxygenase-1 and cyclooxygenase-2 isoforms to ischemia-induced oxidative damage and neurodegeneration following transient global cerebral ischemia.J. Neurochem2003; 86: 545–555.
- Madrigal JL, Garcia-Bueno B, Moro MA, Lizasoain I, Lorenzo P: Relationship between cyclooxygenase-2 and nitric oxide synthase-2 in rat cortex after stress.Eur. J. Neurosci2003; 18: 1701–1705.
- Bruce-Keller AJ, Lovell MA, Kraemer PJ, Gary DS, Brown R et al.: 4-Hydroxynonenal, a product of lipid peroxidation, damages cholinergic neurons and impairs visuospatial memory in rats. J. Neuropathol. Exp. Neurol1998; 57: 257– 267.
- 64. Sinclair AJ, Bayer AJ, Johnston J, Warner C, MaxweelCR: Altered plasma antioxidant status in subjects with Alzheimer's disease and vascular dementia. Int. Geriatr. Psychiatry1998; 13: 840–845.
- 65. Yu Z, Cheng G, Hu B:Mechanism of colchicine impairment on learning and memory, and protective effect of CGP36742 in mice. Brain Res 1997; 750: 53–58.
- Beal MF: Aging, energy, and oxidative stress in neurodegenerative disease. Ann. Neurol1995;38: 357–366.
- 67. Bondy CS: The relation of oxidative stress and hyperexcitation to neurological disease. Proc. Soc. Exp. Biol. Med1995; 208: 337–345.
- Bezzi P, Carmignoto G, Pasti L: Prostaglandins stimulate calcium dependent glutamate release in astrocytes. Nature1998; 39: 281–285.
- 69. Kukreja RC, Kontes HA, Hess ML, Ellis EF: PGH synthase and lipooxygenase generate superoxide in the presence of NADH or NADPH. Circ. Res1986; 59: 612–619.
- Leist M, Volbracht C, KuhnleH, Fava E, Ferrando ME: Caspase-mediated apoptosis in neuronal excitotoxicity triggered by nitric oxide. Mol. Med1997; 3: 750–764.
- 71. Dawson R, Beal MF, Bondy SC, Di Monte, Isom GE: Excitotoxins, aging and environmental neurotoxins: implications for understanding human neurodegenerative diseases. Toxicol. Appl. Pharmacol1995; 134: 1–17.
- Fass U, Panickar K, Personett D, Bryan D, Williams K, Gonzales,K, Sugaya M. McKinn: Differential vulnerability of primary cultured cholinergic neurons to nitric oxide excess. NeuroReport2003; 11: 931–936.
- Emerich DF, Walsh TJ: Cholinergic loss and cognitive impairment following intraventricular or intradentate injections of colchicines. Brain Res1990; 517: 157–167.
- 74. Nakagawa YS, Nakamura S, Kase Y, Noguchi T, Ishihara T: Colchicine lesions in the rat hippocampus mimic the alterations of several markers in Alzheimer's disease. Brain Res1987; 408, 57–64.

- 75. Takahashi M, Sugaya K, Kubota K: I methioninephysiological role in human physiology. Japan J. Pharmacol1987; 60; 307-18.
- Bhattacharya SK, Kumar A and Jaiswal AK: Effect of Mentat, a Herbal Formulation, on Experimental Models of Alzheimer's Disease and Central Cholinergic Markers in Rats. Fitoterapia,(LXVI)1995;3: 216-219.
- 77. Dekker AJ, Cornor AM, Thal DJ, Neurosci. Biobehav. Res1991; 15: 299-301
- 78. Katzman RN: Biochemical alterations in alzheimer's patients. Engl. J. Med1986; 314: 964-971.
- 79. Sayas CL, Moreno-Flores MT, Avila J and Wandosell F: The Neurite Retraction Induced by Lysophosphatidic Acid Increases Alzheimer's Disease-like *Tau* Phosphorylation. The journal of biological chemistry1999;274:52:37046–37052.
- Koladiya UR, Jaggi AS, Singh N and Sharma BK: Ameliorative role of Atorvastatin and Pitavastatin in L-Methionine induced vascular dementia in rats. BMC Pharmacology2008,8:14.
- Dayal S, Devlin AM, McCawJ, Liu ML, Arning E, Bottiglieri T, Shane B, Faraci FM, Lentz SR: Cerebral Vascular Dysfunction in Methionine Synthase Deficient Mice. *Circulation2005*; 112;737-44.
- Corzo L, Zas R, Rodr'ýguez S, Fern'andez-Novoa L, Cacabelos L:Decreased levels of serum nitric oxide in different forms of dementia. *Neurosci Lett2007;* 420:263-7.
- Zhu X, Smith MA, Honda K, Aliev G, Moreira PI, Nunomura A: Vascular oxidative stress in Alzheimer disease. J Neurological Sciences2007; 257:240-6.
- 84. Gauthier S: Alzheimer's disease: current and Future therapeutic perspectives. *Pmg Nem-Psychopharmnco1 and Bid Psychia2001*, 25:73-89.
- Lipton SA, Kim WK, Choi S, Kumar S, D'emilia DM, RayuduPV, ArnelleDV,Stamler JS. Neurotoxicity associated with dual actions of homocysteine at the N-methyl-Daspartate receptor. *Proc Natl Acad Sci1997*; 94:5923-8.
- Herrmann R: Mechanisms of homocysteine neurotoxicity in neurodegenerative diseases with special reference to dementia. *FEBS Lett2006;* 580:2994-3005.
- Hirche F, SchröderA, KnothB, StangGI, Eder K: Effect of dietary methionine on plasma and liver cholesterol concentrations in rats and expression of hepatic genes involved in cholesterol metabolism. *Br J Nut2006;* 95:879-88.
- WolozinB,BehlC: Mechanisms of neurodegenerative disorders: Part 1: protein aggregates. Arch Neurol2000;57:793-6.
- Deshmukh R, Vivek S, Mehan S, Sharma N, BediKL: Amelioration of intracerebroventricular streptozotocin induced cognitive dysfunction and oxidative stress by vinpocetine — a PDE1 inhibitor. European Journal of Pharmacology2009; 620:49–56.

Available online on www.ijpsr.com