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IN VITRO ANTHELMINTIC ACTIVITY OF DIFFERENT EXTRACTS OF ROOT OF *CARISSA SPINARUM*

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ABSTRACT

Modern medicines are gaining less attention due to their limited availability and affordability in human intestinal helminthiasis. Thus, most of the world's population depends to a greater extent on traditional medical remedies. *Carissa spinarum*, Linn. (*Apocynaceae*) is a small spinous, evergreen shrub growing throughout India in dry regions. Further, *C. spinarum* roots are traditionally used for their purgative properties as well as to treat worm infested wounds in animals. There is no report on pharmacologically evaluated antihelmintic activity of root extract of *C. spinarum* till date. Therefore, in the present study we have investigated the antihelmintic activity of methanolic, aqueous and chloroform extracts of root of *C. spinarum* on *Pheretima posthuma*. The fresh and dried root of *C. spinarum* were collected in the month of November from the Bilaspur region, Chhattisgarh state, India, and the antihelmintic activity was evaluated in terms of time taken to cause paralysis and death of the adult Indian earthworm *Pheretima posthuma*. Piperazine citrate (PC; 10 mg/ml) was included as reference compound. The present investigation revealed that the methanolic (100 mg/ml) and chloroform (50 and 100 mg/ml) extracts have equivalent potency compared to PC (10 mg/ml) in time taken for both paralysis and death of *Pheretima posthuma*. Standardization of each extracts and isolation of phytoconstituents in each extracts is required in the future. Furthermore, the pharmacological studies for antihelmintic activity should be undertaken in other parasites to mimic the exact human helminthiasis.

Keywords:

Anthelmintic,
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INTRODUCTION: Human intestinal helminthiasis is among the most common infectious diseases occurring throughout the developing world. These infections have been associated with low standard of sanitation and the worldwide prevalence lies between 500 million to one billion annually approximately ^{1, 2, 3, 4, 5}. The helminthes which infect the e. g. Tape worms (*Taenia solium*), nematodes e. g. hookworm (*Ancylostoma duodenale*), intestine are cestodes roundworm (*Ascaris lumbricoides*) and trematodes or flukes (*Schistosoma mansoni* and *Schistosoma hematobolium* ^{6, 7, 8, 9, 10, 11}. It has been well accepted that due to the limited availability and affordability of modern medicines most of the world's population depends to a greater extent on traditional medical remedies ^{12, 13, 14}. Further, the use of medicinal plant products for treatment of various acute and chronic diseases is gaining increasing importance around the globe ^{15, 16, 17, 18}. It has been well evidenced that the traditional medicines including plants and plant-derived preparations hold a great promise as source of easily available effective antihelmintic agents to the people ^{19, 20, 21, 22}.

Carissa spinarum, Linn. (*Apocynaceae*) is a small spinous, evergreen shrub growing throughout India in dry regions ^{23, 24}. It has been well investigated that *C. spinarum* receives less attention in academic literature. Literature survey revealed that *C. spinarum* contains lignans, sesquiterpenes of eudesmane type and several cardiac glycosides ^{25, 26}. Moreover, it has been reported that *C. spinarum* leaves contain urosolic acid and naringin, root contain caffeic acid and a new germacrane derivative, carenone is isolated from stem ²⁷. Further, *C. spinarum* produces edible fruits and its roots are traditionally used for their purgative properties as well as to treat worm infested wounds in animals. ^[28] Furthermore, pharmacological studies revealed that stem extracts of *C. spinarum* possess antioxidant and cardiotoxic activity ^{26, 27}. There is no report on

pharmacologically evaluated anthelmintic activity of root extract of *C. spinarum* till date. Therefore, in this present study we have evaluated the antihelmintic activity of methanolic, aqueous and chloroform extracts of root of *C. spinarum* on *Pheretima postuma*.

MATERIALS AND METHODS:

Plant Material: The fresh and dried root of *C. spinarum* were collected in the month of November from the Bilaspur region, Chhattisgarh state, India, and authenticated by Prof. S. D. Dubey, Department of Dravyaguna, Faculty of Ayurveda, Institute of Medical Science, Banaras Hindu University, Varanasi, India and a voucher specimen (No. 052) has been submitted to the Pharmacognosy Division, Department of Pharmaceutics, Institute of Technology, Banaras Hindu University, Varanasi, India for future reference.

Preparation of Extracts: The dried root of *C. spinarum* were powdered, defatted with petroleum ether and subjected to successive solvent extraction with chloroform, methanol in Soxhlet extractor. Aqueous extract was obtained by cold maceration for 24 hours. All the extracts were further evaporated to dryness before investigation to be carried out. The preliminary phytochemical investigation was carried out for methanolic (MCS), aqueous (ACS) and chloroform (CCS) extracts of root of *C. spinarum*. Further, the percentage of yield of MCS, ACS and CCS were found to be 10.17, 6.21 and 8.79 respectively.

Animals: Indian adult earthworms (*Pheretima postuma*) were collected from water logged areas and were identified by Dr. R. Kundu, Department of Zoology, Guru Ghasidas University, Bilaspur.

Evaluation of Antihelmintic Activity: The anthelmintic activity was evaluated on adult Indian earthworm *Pheretima postuma* due to its anatomical and physiological resemblance with the intestinal round worm parasites of

human beings. Three different concentrations, each of crude extract of methanolic, aqueous and chloroform (25, 50, 100 mg/ml in distilled water) were prepared and six worms (identical to each other) were placed in it. Observations were made for the time taken to cause paralysis and death of the individual worms. Mean time for the paralysis in min was noted when no

movement of any sort could be observed, except when the worm was shaken vigorously; time of death in min was recorded after ascertaining the worms neither moved when shaken vigorously nor when dipped in warm water (50°C) and Piperazine citrate (PC; 10 mg/ml)^{29, 30} was included as reference compound as shown in **Table 1**.

TABLE 1: ANTHELMINTIC ACTIVITY OF DIFFERENT EXTRACTS OF ROOTS OF *C. SPINARUM* ON *PHERETIMA POSTHUMA*

GROUPS	CONCENTRATION (MG/ML)	TIME TAKEN FOR PARALYSIS (MIN)	TIME TAKEN FOR DEATH (MIN)
MCS	25	113.85±2.74*	438.04±3.57*
	50	92.28±1.91*	357.78±1.11*
	100	26.22±0.84	62.90±1.56
ACS	25	97.98±2.68*	465.98±2.21*
	50	90.95±0.97*	435.86±2.57*
	100	86.19±1.86*	396.50±3.00*
CCS	25	90.30±0.98*	286.43±4.74*
	50	29.10±1.25	73.38±2.65
	100	22.95±1.65	63.14±3.32
PC	10	22.46±0.45	63.77±1.57
Control	-	-	-

All the values are Mean±SEM (n = 6). *P<0.05 compared to PC

Chemicals and Reagents: All the chemicals including PC and reagents were procured from local suppliers and were of analytical grade.

Statistical Analysis: The data were analyzed with GraphPad Prism 4 (San Diego, CA). Statistical analysis of data was done by One-way ANOVA, followed by Newman Keuls test. Data are expressed as Mean ± Standard error of mean (S.E.M.). A level of P<0.05 was accepted as statistically significant.

RESULTS: Table 1 illustrates the effect of different extracts of root of *Carissa spinarum* (25, 50 and 100 mg/ml) in time for paralysis and death of *Pheretima posthuma*. Statistical analysis by One-way ANOVA showed that there was significant difference in time taken for paralysis [F (9, 50) = 473.11, P<0.05] of Indian earthworm among groups. Post-hoc test revealed that MCS (100 mg/ml), CCS (50 mg/ml) and CCS (100 mg/ml) groups were not significantly different compared to PC in time

taken for paralysis of Indian earthworm, indicating equivalence in potency. Further, all the treated groups except MCS (100 mg/ml), CCS (50 mg/ml) and CCS (100 mg/ml) groups showed significant difference compared to PC (10 mg/ml) in time taken for paralysis of earthworm. Furthermore, statistical analysis by One-way ANOVA showed that there was significant difference in time taken for death [F (9, 50) = 3957.2, P<0.05] of Indian earthworm among groups. The post-hoc test indicated that the time taken for death of *Pheretima posthuma* was similar to that of the effect observed in time taken for paralysis of earthworm, indicating equivalent potency while compared to PC.

DISCUSSIONS: The present study revealed that the MCS (100 mg/ml) and CCS (50 and 100 mg/ml) have equivalent potency compared to PC (10 mg/ml) in time taken for both paralysis and death of *Pheretima posthuma*. Preliminary phytochemical screening of the extracts

revealed that the presence of terpenoids, flavonoids, alkaloids, tannins, saponins and steroids. It has been well established that PC by increasing chloride ion conductance of worm muscle membrane produces hyperpolarization and reduced excitability that leads to muscle relaxation and flaccid paralysis^{31, 32} thus, our drug may have the similar profile of mechanism of action. Further, it has been reported that tannins which are polyphenolic compounds produce antihelmintic activity by binding to glycoprotein on the cuticle of the parasite and thus leads to death of the worm³¹. Therefore, standardization of each extracts and isolation of phytoconstituents in each extracts for antihelmintic activity is required in the future. Furthermore, the pharmacological studies for antihelmintic activity should be undertaken in other parasites to mimic the exact human helminthiasis.

CONCLUSIONS: The methanolic and chloroform extract of root of *C. spinarum* showed antihelmintic activity on *Pheretima posthuma*. Therefore, standardization of each extracts and isolation of phytoconstituents in each extracts for antihelmintic activity is required in the future. Furthermore, the pharmacological studies for antihelmintic activity should be undertaken in other parasites to mimic the exact human helminthiasis.

REFERENCES:

- Allen H, Crompton DW T, Silwa ND, loverde PT, and Old GR: New policy for using antihelmintic in high risk groups. Trends in parasitology 2002; 18:381-382.
- Peters W. Medical aspects: Comments and discussions II. In: A. E. R. Taylor, R. Muller (Eds.). The relevance of parasitology to human welfare today. Symposium of the British Soc for Parasitol 1978; 16:25-40.
- WHO: Prevention and control of intestinal parasitic infections. WHO Technical Report Series 1987; 749.
- Agbolade OM, Akinboye DO, and Awolajama: Intestinal helminthiasis and Urinary schistosomiasis in some villages of Ijebu North, Ogun State, Nigeria. African journal of Biotech 2004; 3:206-209.
- Ademola LO, Fagbami BO and Idows SO: Evaluation of the antihelmintic activity of Khaya Senegalensis extract against gastrointestinal nematodes of sheep: in vitro and in vivo studies. Veterinary Parasitology 2004; 122:151-164.
- Borges FA, Silva HC, Buzzulini C, Soares VE, Santos E, Oliveira GP and Costa AJ: Endoctoside activity of new long action formulation containing 2.25% ivermectin and 1.25% abamectin in cattle. Veterinary Parasitology 2008; 155:299-307.
- Kar PK, Tandon V and Saha N: Antihelmintic efficacy of Flemingia vestita: genesterininduced effect on the activity of nitric oxide synthase and nitric oxide in the trematode parasite Fasciolopsis buski. Parasitology International 2002; 51:249-257.
- Tariq KA, Chisti MZ, Ahmad F and Shawl AS: Antihelmintic activity of extract of Artemisia absinthiam against oxine nematodes. Veterinary Parasitology 2009; 160:83-88.
- Grover J K, Vats v, Uppal G, Yadav S: Antihelmintic Activity. Trop Gastroenterol 2001; 22:180-9.
- Grade JT, Tabuti JR, Van DP: Ethnoveterinary knowledge in pastoral Karmoja, Uganda. J. Ethnopharmacol 2009; 18:273-93.
- Vasconcelos ALFC, Bevilaqua CML, Morais SM, Marciel MV, Costa CTC, Macedo ITF, Oliveira LMB, Braga RR, Silva RA, Vieira LS: Antihelmintic activity of croton zehntneri and Lippia sidoides essential oils. Veterinary Parasitology 2007; 148:288-294.
- Satyavati GV: Use of plant drugs in Indian Traditional System of Medicine and their relevance to primary health care. In: Economic and Medicinal Plant Research by N. R. Farnworth, H. Wagner (Eds.), Academic Press Ltd. London 1990; 190-210.
- Kamboj VP: Herbal Medicine. Current Science 2000; 78:1.
- Teklehaymanot T and Giday M: Ethno botanical study of medicinal plants used by people in Zegie Peninsula, Northwestern Ethiopia. Journal of ethnobiology and Ethnomedicine 2007; 3:12.
- Mali GR, Mehta AA: A Review on Antihelmintic Plants. Nat Prod Rad 2008; 7:466-75.
- Rastogi T, Bhutdav V, moon K, Aswar PB and Khadabadi SS: Comperative studies on antihelmintic activity of moringa olifera and vitex negunda. Asian J. Research Chem 2009; 2:2.
- Deore SL, Khadabadi SS, Kamadi KS, Ingle VP, Kawlkar NG, Sawarkar PS, Patil VA, Vyas AJ: In vitro antihelmintic activity of cassia tora. Int. J. Of Chem. Tech. Rsearch 2009; 1:177-179.
- Nirmal SA, Malwadkar G and Laware RB: Antihelmintic activity of pongamia glabra. Segklankari J. Sci. Technol 2007; 29:755-757.
- Gbolade AA and Adeyemi AA: Antihelmintic activity of three medicinal plants from Nigeria. Fitoterapia 2008; 79:223-225.
- Enwerem NM, Okogun JL, Wambebe CO, Okorie DA and Akah PA: Antihelmintic activity of the stem bark extracts of Berlina grandiflora and one of its active principles Betulinic acid. Phytomedicine 2001; 8:12-114.
- Yoganandan GP, Gouri R, Biswas D: Evaluation of Wedelia Biflora (Linn) D. C. for antihelmintic and antimicrobial activity. Journal of pharmacy research 2009; 2.

22. Dornetshuber R, Kamyar MR, Rawnduzi P, Baburin I, Kouri K, Pilz E, Hornbogen T, Zocher R, Berger W and Gruber RL: Effect of the antihelmintic drug PF1022A on mammalian tissue and cells. *Biochemical Pharmacology* 2009; 77:1437-1444.
23. CSIR: The Wealth of India A Dictionary of Indian Raw Materials and Industrial Products-Raw Materials Council of Scientific and Industrial Research New Delhi 1950; 2:82.
24. Flora of china: 1995; 16:146-147.
25. Mishra RM, Gupta P: Frugivory and seed dispersal of *Carissa spinarum* (L.) in a tropical deciduous forest of central India. *Tropical ecology* 2005; 46:151-156.
26. Vohra MM, De NN: Comparative Cardiotonic activity of *Carissa carandas* and *Carissa spinarum* A.D.C. *Indian J. Med. Res* 1963; 51:937-40.
27. Rao RJ, Kumar US, Reddy SV, Tiwari AK, Rao JM: Antioxidants and a new germacrane sesquiteroene from *Carissa spinarum*. *Nat Prod Res* 2005; 19:763-9.
28. Teklehaymanot T, Giday M: Ethno botanical study of medicinal plants used by people in Zegie Peninsula, Northwestern Ethiopia. *J Ethnobiol and Ethnomed* 2007; 3:1-12.
29. Ajaiyeoba EO, Onocha PA, Olarenwaju OT: In vitro anthelmintic properties of *Buchholzia coriacea* and *Gynandropsis gynandra* extract. *Pharm Biol* 2001; 39:217-20.
30. Mali GR, Hundiwale CJ, Sonawane SR, Patil NR, Hatapakki CB: Evaluation of *Capparis decidua* for antihelmintic and antimicrobial activities. *Ind J Nat Prod* 2004; 20:10-3.
31. Kane RS, Mohite KS, Shete SJ: Antihelmintic activity of aqueous and methanolic extracts of *euphorbia thymifolia* linn. *Int J Pharm Tech Res* 2009; 1:666-9.
32. Martin RJ: *c*-Aminobutyric acid and piperazine activated single channel current from *Ascaris suum* body muscle. *Br J Pharmacol* 1985; 84:445-61.
