



Received on 06 June, 2010; received in revised form 22 August, 2010; accepted 14 September, 2010

PREVENTIVE EFFECT OF DIOSMIN, A BIOFLAVONOID, ON GLYCOPROTEIN CHANGES IN STREPTOZOTOCIN-NICOTINAMIDE-INDUCED TYPE 2 DIABETIC RATS

Leelavinothan Pari*¹, Subramani Srinivasan¹ and Mohammed Saddiq²

Department of Biochemistry and Biotechnology¹, Faculty of Science, Annamalai University, Annamalainagar, Tamilnadu, India

Department of Biochemistry², Adiparasakthi College of Arts and Science, Kalavai, Vellore (Dt), Tamilnadu, India

ABSTRACT

Keywords:
Plasma glucose,
Diosmin,
Diabetes mellitus,
Glycoprotein components

Correspondence to Author:

Dr. L. Pari

Department of Biochemistry and
Biotechnology, Faculty of
Science, Annamalai University,
Annamalainagar, Tamil Nadu,
India

The present study was designed to investigate the effect of diosmin on the levels of glycoprotein components in plasma and tissues of streptozotocin (STZ)-nicotinamide (NA) induced diabetic rats. Diabetes was induced in male Wistar rats by a single intraperitoneal injection of STZ (45 mg/kg b. w). Diosmin (DS) (100 mg/kg b. w) was administered orally for 45 days. The effects of diosmin on plasma glucose, plasma insulin, plasma and tissue glycoproteins were studied. The levels of plasma glucose and plasma glycoproteins were increased significantly whereas the level of plasma insulin was significantly decreased in diabetic rats. There was a significant decrease in the level of sialic acid and elevated levels of hexose, hexosamine and fucose in the liver and kidney of diabetic rats. Oral administration of diosmin to diabetic rats led to decreased levels of plasma glucose and plasma glycoproteins. The levels of plasma insulin and tissue sialic acid were increased, whereas the levels of tissue hexose, hexosamine and fucose were near normal. The present study indicates that diosmin possesses a significant beneficial effect on glycoproteins in addition to its antidiabetic effect.

INTRODUCTION: Diabetes Mellitus is a serious, complex metabolic disorder of multiple etiologies, characterized by chronic hyperglycemia with disturbances of carbohydrate, fat and protein metabolism resulting from defects in insulin secretion (β -cell dysfunction), insulin action (insulin resistance) or both ¹. It is of particular concern since the disease incidence is expected to increase worldwide by more than 100% between 2000 and 2030 ². Hyperglycaemia may perturb cellular antioxidant defense systems and damage cells. Free radicals are formed disproportionately in diabetes by glucose oxidation, non-enzymatic glycation of proteins, and the subsequent oxidative degradation of glycated proteins ³, which occurs in various tissues. This process leads to long-term damage, dysfunction, and failure of various organs, especially the eyes, kidneys, nerves, heart, and blood vessels, and creates a huge economic burden related to the management of diabetic complications ⁴.

Glycoproteins are carbohydrate linked protein macromolecules found in the cell surface, which form the principal component of animal cells. Hexose, hexosamine, fucose and sialic acid are the basic components of the glycoproteins. They play an important role in membrane transport, cell differentiation and recognition, the adhesion of macromolecules to the cell surface, and the secretion and absorption of macromolecules. Impaired metabolism of glycoproteins plays a major role in the pathogenesis of diabetes mellitus ⁵. In recent times, many traditionally important medicinal plant phytochemicals have been tested for their efficacy against impaired glycoprotein levels in diabetes ⁶.

Streptozotocin, a monofunctional nitrosourea derivative, is one of the most commonly used substances to induce diabetes in experimental animals ⁷. Evidences suggest that the diabetogenic capacity of streptozotocin may depend on its ability to damage β -cell and induce oxidative stress ⁸. In the diabetic state,

glucose is utilized by the insulin independent pathways leading to the synthesis of glycoproteins, and even a mild deficiency of insulin influences the thickening of the basement membrane. At the cell surface or inside the cells, the glyco-components such as fucose and sialic acid form specific structures, called glycanic chains covalently linked to lipids or proteins. An important in the metabolism of glycoproteins could be related to the deposition of these materials in the basal membrane of pancreatic cells ⁹.

Diosmin was first isolated in 1925 from *Scrophularia nodosa*. Today, it can be manufactured by extracting hesperidin from citrus rinds, followed by conversion of hesperidin to diosmin ¹⁰. Diosmin possesses antioxidant ¹¹ and blood lipid lowering ¹² activities. It enhances venous tone and microcirculation and protects capillaries ¹³, mainly by reducing systemic oxidative stress ¹⁴. In our previous studies, we reported the effect of diosmin on rate-limiting enzymes of glycolysis, HMP shunt and gluconeogenesis in the liver and circulation of diabetic rats ¹⁵. To our knowledge, there are no available reports on the effect of this flavonoid on glycoprotein levels in diabetic rats. Hence, the present study was carried out to determine the effect of diosmin on plasma and tissue glycoproteins in STZ-NA-induced diabetic rats.

MATERIALS AND METHODS:

Animals: Male albino Wistar rats, weighing 180–220 g, bred in the Central Animal House, Rajah Muthiah Medical College, Annamalai University, were obtained and housed in polypropylene cages in a pathogen free environment at an ambient temperature of $28\pm 2^{\circ}\text{C}$ and 45–55% relative humidity with 12 h each of dark and light cycle. Rats were fed pellet diet (Hindustan Lever Ltd., India) and water ad libitum. The animals were maintained in accordance with the guidelines of the National Institute of Nutrition, Indian Council of Medical Research, Hyderabad,

India and approved by the Institutional Animal Ethical Committee, Annamalai University (Reg. No. 160/1999/CPCSEA; vide No. 566, /2008).

Chemicals: Diosmin and streptozotocin were purchased from Sigma Chemical Company, St Louis, MO, USA and all other chemicals used were of analytical grade obtained from E. Merck or Himedia, Mumbai, India.

Induction of diabetes: Non-insulin dependent DM was induced in over night fasted experimental groups by a single intraperitoneal (i. p.) injection of freshly prepared STZ (45 mg/kg b. w.) dissolved in 0.1 mol/l citrate buffer (pH 4.5) 15 min after the i. p. administration of NA (110 mg/kg b. w.). The animals were allowed to drink 20% glucose solution overnight to overcome drug-induced hypoglycemia. Control rats were injected with the same volume of isotonic saline. After 72 h, plasma glucose was determined and those rats with fasting glucose levels greater than 250 mg/dL were served as diabetic rats and used in the present study.

Experimental design: The animals were randomly divided into four groups of six animals in each group (12 diabetic surviving and 12 normal). Diosmin was dissolved in vehicle solution of 0.6% dimethylsulfoxide (DMSO) and diosmin (100 mg/kg b. w) were administered orally using an intragastric tube for a period of 45 days¹⁵.

- Group I: Normal control (vehicle treated; DMSO: 1 ml/kg b. w)
- Group II: Normal + diosmin (100 mg/kg b. w)
- Group III: Diabetic control
- Group IV: Diabetic + diosmin (100 mg/kg b. w)

Analytical procedure: Measurements of plasma glucose and plasma insulin were estimated colorimetrically using commercial diagnostic kits (Sigma Diagnostics (I) Pvt. Ltd, Baroda, India). Plasma insulin was assayed by ELISA using a

Boehringer–Mannheim kit with an ES300 Boehringer analyzer (Mannheim, Germany).

Determination of glycoprotein components: For the estimation of glycoprotein components, the tissues were defatted by the method of Folch *et al*¹⁶. And the defatted tissues were treated with 0.05 M H₂SO₄ and hydrolysed at 80°C and aliquot was used for sialic acid estimation. To the remaining solution, 0.1 M NaOH was added. The aliquots were used for fucose, hexose, and hexosamine estimation.

Estimation of hexose: Protein-bound hexoses were estimated by the method of Dubois and Gilles¹⁷. To 0.1 ml of plasma or defatted tissue sample, 5.0 ml of 95% ethanol was added, mixed and then centrifuged. The precipitate was dissolved in 1.0 ml of 0.1 N NaOH. Subsequently, 1.0 ml of distilled water and 1.0 ml of standards (20 - 100µg) were set up along with the test. To all the tubes, 8.5 ml of orcinol-sulphuric acid reagent was added and kept in a water bath for exactly 15 min at 90°C. The tubes were cooled in tap water and the color developed was read at 540 nm against a blank.

Estimation of hexosamine: Hexosamine was estimated by the method of Wagner¹⁸. Briefly, the reaction mixture contained, 0.5 ml plasma/1.0 ml aliquot, 2.5 ml of 3 M HCl and boiled over 6 h and neutralized with 6 M NaOH. To 0.8 ml of neutralized sample added 0.6 ml of acetyl acetone reagent and boiled for 30 min. The mixture was treated with 2.0 ml of Ehrlich's reagent. The absorbance was read at 540 nm.

Estimation of sialic acid: Sialic acid was determined by the method of Warren¹⁹. In brief, 0.5 ml of aliquot/plasma was treated with 0.5 ml of water, 0.25 ml of periodic acid and incubated at 37°C for 30 min. To the reaction mixture added 0.2 ml of sodium meta arsenate and 2.0 ml of thiobarbituric acid were added and heated for 6 min and added 5.0 ml of acidified butanol. The absorbance was read at 540 nm.

Estimation of fucose: Fucose was estimated by the method of Winzler²⁰. Briefly, 1.0 ml of precipitated glycoprotein from platelet membrane and 1.0 ml of processed serum were dissolved in 1 ml of 0.1M NaOH and placed in an ice-bath and 4.5 ml of cold H₂SO₄ was added and mixed well. The tubes were heated in a boiling water bath for 3min and cooled and then 0.1 ml of 3% cysteine was added and mixed immediately. The tubes were allowed to stand at room temperature for 60-90 min. The absorbance of the solution at 396 and 430nm was measured in a spectrophotometer and the difference in the absorbance was taken for the calculation.

Statistical analysis: All data were expressed as mean \pm SD for six rats in each group. The statistical analysis was done by one-way analysis of variance (ANOVA) followed by Duncan's Multiple Range Test (DMRT) using SPSS (Statistical Program for Social Sciences, SPSS Corporation, Chicago, IL) version 12.0 for Windows, $p < 0.05$ were considered as significant and included in the study²¹.

RESULTS AND DISCUSSION:

Effect of diosmin on plasma glucose and insulin levels: Fig. 1 shows the levels of plasma glucose and plasma insulin of different experimental groups. The diabetic control rats showed a significant increase in the level of blood glucose with significant decrease in the level of plasma insulin. Oral administration of diosmin to diabetic rats significantly reversed the above biochemical changes. In our previous study, we have reported that diosmin at 100 mg/kg body

weight showed better effect than 25 and 50 mg/kg body weight, therefore the 100 mg/kg body weight was used in this study.

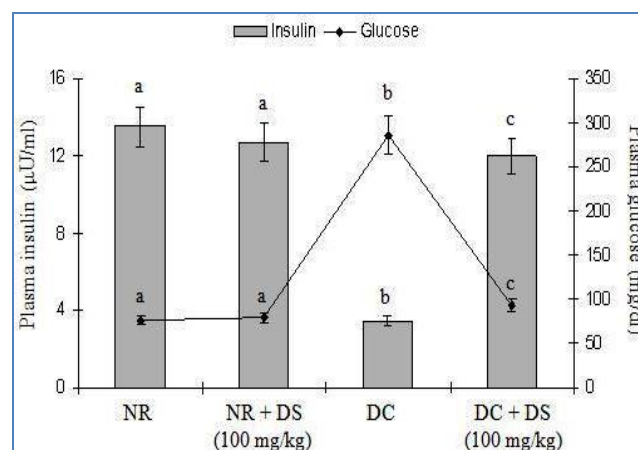


FIG. 1: EFFECT OF DIOSMIN ON PLASMA INSULIN AND GLUCOSE LEVELS IN CONTROL AND DIABETIC RATS

Changes in the levels of plasma glucose and insulin in normal control and experimental rats

Values are given as mean \pm SD from 6 rats in each group

Values not sharing a common superscript letter (a-c) differ significantly at $p < 0.05$ (DMRT)

NR- normal; DS- diosmin; DC- diabetic control

Effect of diosmin on plasma and tissue glycoproteins: The levels of plasma and tissue glycoproteins in normal and experimental rats are shown in the Table 1, 2 and 3. There was a significant increase in the level of plasma glycoproteins in diabetic rats. In liver and kidney of diabetic rats, the level of hexose, hexosamine and fucose were significantly increased whereas the level of sialic acid was significantly decreased. Oral administration of diosmin significantly reversed the changes in plasma, liver and kidney glycoproteins of diabetic rats.

TABLE 1: EFFECT OF DIOSMIN (DS) ON PLASMA GLYCOPROTEIN LEVELS IN NORMAL AND EXPERIMENTAL RATS

GROUPS	HEXOSE	HEXOSAMINE	SIALIC ACID	FUCOSE
	(mg/dl)			
Normal	93.26 \pm 7.10 ^a	76.38 \pm 5.82 ^a	52.58 \pm 4.00 ^a	27.16 \pm 2.07 ^a
Normal + diosmin (100 mg/kg)	91.20 \pm 6.98 ^a	74.93 \pm 5.74 ^a	51.24 \pm 3.92 ^a	25.99 \pm 1.99 ^a
Diabetic control	97.50 \pm 7.46 ^b	97.50 \pm 7.46 ^b	68.52 \pm 5.25 ^b	40.58 \pm 3.11 ^b
Diabetic + diosmin (100 mg/kg)	84.91 \pm 6.50 ^c	84.91 \pm 6.50 ^c	57.59 \pm 4.41 ^c	31.25 \pm 2.39 ^c

Each value is mean \pm S.D. for 6 rats in each group. ^{a-c}. In each column, means with different superscript letter differ significantly at $p < 0.05$ (DMRT)

TABLE 2: EFFECT OF DIOSMIN (DS) ON LIVER GLYCOPROTEIN LEVELS IN NORMAL AND EXPERIMENTAL RATS

GROUPS	HEXOSE	HEXOSAMINE	SIALIC ACID	FUCOSE
	(mg/g defatted tissue)			
Normal	27.13 ± 2.07 ^a	10.59 ± 0.81 ^a	8.79 ± 0.67 ^a	16.98 ± 1.29 ^a
Normal + diosmin (100 mg/kg)	26.02 ± 1.99 ^a	9.47 ± 0.73 ^a	9.31 ± 0.71 ^a	17.59 ± 1.35 ^a
Diabetic control	47.41 ± 3.63 ^b	18.30 ± 1.40 ^b	4.01 ± 0.31 ^b	27.90 ± 2.91 ^b
Diabetic + diosmin (100 mg/kg)	33.25 ± 2.54 ^c	13.27 ± 1.02 ^c	6.29 ± 0.48 ^c	20.03 ± 1.53 ^c

Each value is mean ± S.D. for 6 rats in each group. ^{a-c}. In each column, means with different superscript letter differ significantly at p<0.05 (DMRT)

TABLE 3: EFFECT OF DIOSMIN (DS) ON KIDNEY GLYCOPROTEIN LEVELS IN NORMAL AND EXPERIMENTAL RATS

GROUPS	HEXOSE	HEXOSAMINE	SIALIC ACID	FUCOSE
	(mg/g defatted tissue)			
Normal	22.83 ± 1.74 ^a	15.57 ± 1.19 ^a	8.23 ± 0.63 ^a	12.89 ± 0.98 ^a
Normal + diosmin (100 mg/kg)	21.03 ± 1.61 ^a	14.80 ± 1.13 ^a	8.86 ± 0.68 ^b	13.54 ± 1.04 ^b
Diabetic control	41.40 ± 3.17 ^b	30.03 ± 2.30 ^b	4.01 ± 0.31 ^c	28.44 ± 2.18 ^c
Diabetic + diosmin (100 mg/kg)	27.35 ± 2.09 ^c	18.52 ± 1.66 ^c	6.52 ± 0.50 ^d	16.16 ± 1.24 ^d

Each value is mean ± S.D. for 6 rats in each group. ^{a-c}. In each column, means with different superscript letter differ significantly at p<0.05 (DMRT)

The objective of all diabetes treatment and management is to maintain an adequate blood glucose concentration. Four major classes of oral hypoglycemic agents have been used extensively²². Each class of drug works on different mechanisms of actions, including stimulation of insulin secretion, reduction of hepatic gluconeogenesis, increase in insulin receptor sensitivity and delay of digestion and absorption of carbohydrate, respectively. Unfortunately, these agents could produce severe hypoglycemia, weight gain or gastrointestinal disturbances.

Therefore, it is necessary to look for new solutions to manage this health problem. The search for newer antidiabetic agents represents a challenge to the medical profession. India is a country with a vast reserve of natural resources and a rich history of traditional medicine²³. Naturally occurring phytochemicals with antidiabetic activities are relatively nontoxic, inexpensive and available in an ingestive form. Therefore, they are commonly used to prevent morbidity and mortality from chronic diseases in countries where low or middle-income populations²⁴.

Beta-cell defect and insulin resistance are essential features of non-insulin-dependent diabetes mellitus, and both features are the focus of intensive investigation. In this context, new oral antidiabetic drug diosmin present(s) interesting therapeutic properties. In the present study, we estimated the effect of diosmin on glycoprotein metabolism in STZ-NA-induced diabetic rats. Glycation is a nonenzymatic reaction of glucose and other saccharide derivatives with proteins, nucleotides and lipids²⁵. Non-enzymatic glycation (Maillard reaction) is a complex series of reactions between reducing sugars and amino groups of proteins, which leads to browning, fluorescence and cross-linking of the proteins.

Several workers have suggested that elevated levels of plasma glycoproteins in diabetic rats could be a consequence of abnormal carbohydrate metabolism. Insulin deficiency and high levels of plasma glucose in diabetic condition may result in an increased synthesis of glycoproteins. The requirement of insulin for the biosynthesis of the carbohydrate moiety of mucoproteins from glucose is thus evident. It has been reported that

hyperglycemia leads to increased synthesis of glycoproteins and glycosylated proteins due to the fact that excess glucose is redirected to insulin dependent pathway such as synthesis of glucosamine from glucose²⁶. Hexosamines are amino sugars created by adding an amine group to a hexose. The level of hexosamine, increased significantly in the plasma, liver and kidney of diabetic rats, which may be due to insulin deficiency. In diabetic rats treated with diosmin significantly lowered hexosamine, which might be due to increased secretion of insulin.

Fucose is a member of a group of eight essential sugars that the body requires for the optimal functioning of cell-to-cell communication and its metabolism appear to be altered in various diseases such as diabetes mellitus²⁷. A rise in fucose levels could be due to increased glycosylation in the diabetic state. Treatment with diosmin had restored fucose level to near normal, which could be due to improved glycaemic status.

Sialic acid is a terminal component of the non-reducing end of the carbohydrate chains of glycoproteins and glycolipids, which are essential constituents of many hormones and enzymes present in serum and tissues. Plasma sialic acid is almost completely bound to glycoproteins and lipids. Total sialic acid in the plasma has received considerable attention as a possible marker for cardiovascular disease and mortality²⁸. In diabetes mellitus, the plasma concentration of plasma sialic acid was found to increase significantly³⁰. The decrease in the content of sialic acid in tissues may be due to the utilization for the synthesis of fibronectin, which contains sialic acid residues in the core structure. The synthesis of fibronectin was also reported to increase significantly in various tissues of diabetic patients and rats. In our studies, a significant increase in total sialic acid levels in the plasma was observed when compared with the control group. Various factors might cause an elevation in the concentration of plasma sialic acid. Among these

factors, the first is an increase in the synthesis of sialic acid in insulin-independent tissues, such as the liver and the brain, and the second is an increase in the activity of sialyltransferase, which transfers the sialic acid residues to the glycolipids and glycoproteins. In our study, administration of diosmin decreased the content of sialic acid in the plasma and increased the content of sialic acid in the tissues of STZ-NA-diabetic rats.

CONCLUSION: This study showed that diosmin reversed the abnormalities in the levels of glycoprotein components. Diosmin may have beneficial effects in diabetes mellitus, by the enhancement of insulin action, as evident by the increased level of insulin in diabetic rats treated with diosmin, which may be responsible for the reversal of glycoprotein changes. Our results are also in line with the previous report.

REFERENCES:

1. Kardeşler L, Buduneli N, Bıyıkoglu B, Çetinkalp Ş and Kutukçuler N: Gingival crevicular fluid PGE₂, IL-1 β , t-PA, PAI-2 levels in type 2 diabetes and relationship with periodontal disease. *Clinical Biochemistry* 2008; 41(10-11): 863-868.
2. Wild S, Roglic G, Green A, Sicree R and King H: Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes Care* 2004; 27(5): 1047-1053.
3. Kumar G, Banu S and Murugesan AG: Influence of Helicteres isora administration for diabetes mellitus: Its effect on erythrocyte membrane and antioxidant status. *Food and Chemical Toxicology* 2009; 47:1803-9.
4. Schuster DP and Duvuuri V: Diabetes mellitus. *Clinics in Podiatric Medicine And Surgery* 2002; 19:79-107.
5. Prakasam A, Sethupathy S, Pugalendi KV: Influence of Casearia esculenta root extract on glycoprotein components in streptozotocin diabetic rats. *Pharmazie* 2005; 60: 229-232.
6. Pari L and Murugan P: Changes in Glycoprotein Components in Streptozotocin - Nicotinamide Induced Type 2 Diabetes: Influence of Tetrahydrocurcumin from Curcuma longa. *Plant Foods for Human Nutrition* 2007; 62:25-29.
7. Szkudelski T: The mechanism of alloxan and streptozotocin action in β cells of rat pancreas. *Physiological Research* 2001; 50:536-546.
8. Ohkuwa T, Sato Y and Naoi M: Hydroxyl radical formation in diabetic rat induced by streptozotocin. *Life Sci* 1995; 56; 1789-1798.
9. Senthil kumar GP and Subramanian SP: Biochemical studies on the effect of Terminalia chebula on the levels of glycoproteins in streptozotocin-induced experimental

- diabetes in rats. *Journal of Applied Biomedicine* 2008; 6: 105-115.
10. Campanero MA, Escolar M, Perez, Garcia-Quetglas E, Sadaba B and Azanza JR: Simultaneous determination of diosmin and diosmetin in human plasma by ion trap liquid chromatography-atmospheric pressure chemical ionization tandem mass spectrometry: Application to a clinical pharmacokinetic study 2010; 51:875-881.
 11. Bouskela E, Cyrino FZ and Lerond L: Effects of oral administration of different doses of purified micronized flavonoid fraction on microvascular reactivity after ischemia/reperfusion in the hamster cheek pouch. *British Journal of Pharmacology* 1997; 122: 1611.
 12. Borradaile NM, Carroll KK and Kurowska EM: Regulation of HepG2 cell apolipoprotein B metabolism by the citrus flavanones hesperidin and naringenin. *Lipids* 1999; 34: 591.
 13. Galley P and Thiollet M: A double-blind, placebo-controlled trial of a new veno-active flavonoid fraction (S 5682) in the treatment of symptomatic capillary fragility. *International Angiology* 1993; 12: 69.
 14. Unlu A, Sucu N, Tamer L, Coskun B, Yucebilgic G, Ercan B, Gul A, Dikmengil M and Atik U: Effects of Daflon on oxidative stress induced by hindlimb ischemia/reperfusion. *Pharmacological Research* 2003; 48: 11.
 15. Pari L and Srinivasan S: Antihyperglycemic effect of diosmin on hepatic key enzymes of carbohydrate metabolism in streptozotocin-nicotinamide-induced diabetic rats. *Biomedicine & Pharmacotherapy* 2010 doi:10.1016/j.biopha.2010.02.001
 16. Folch J, Lees M and Solane SGH: A simple method for isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry* 1957; 26:497-509.
 17. Dubois M and Gilles KA: *Methods in Enzymology*. Academic Press, New York, 1956; 83.
 18. Wagner WD: A more sensitive assay discriminating galactosamine and glucosamine in mixture. *Analytical Biochemistry* 1979; 94: 394-396.
 19. Warren L: The thiobarbituric acid assay of sialic acids. *J Biol Chem* 1959; 234: 1971-1975.
 20. Winzler RJ: Determination of serum glycoproteins. *Methods of Biochemical Analysis* 1955; 2: 279-311.
 21. Duncan BD: Multiple range tests for correlated and heteroscedastic means. *Biometrics* 1957; 13: 359-364.
 22. Charpentier G: Oral combination therapy for type 2 diabetes. *Diabetes/Metabolism Research and Reviews* 2002;18: 70-76.
 23. Grover JK, Vats V, Rathi SS and Dawar R: Traditional Indian anti-diabetic plants attenuate renal hypertrophy, urine volume and albuminuria in streptozotocin induced diabetic mice. *Journal of Ethnopharmacology* 2001; 76: 233-238.
 24. Gaziano TA, Galea G, and Reddy KS: Scaling up interventions for chronic disease prevention: the evidence. *The Lancet* 2007; 370 (9603): 1939-1946.
 25. Brownlee M: Biochemistry and molecular cell biology of diabetic complications. *Nature* 2001; 414: 813-820.
 26. Pari L and Ashokkumar N: Glycoprotein changes in non-insulin dependent diabetic rats: Effect of N-benzoyl-D-phenylalanine and metformin. *Therapie* 2006; 61: 125-131.
 27. Mondoia EI, Kitei M: The new healing science of glyconutrients. In: *Sugars that Heal*. Ballantine Publishing New York 2001.
 28. Moussa MA, Alsaied M, Refai TM, Abdella N, Al-Sheikh N and Gomez JE: Association of serum sialic acid with cardiovascular metabolic risk factors in Kuwaiti children and adolescents with type 1 diabetes. *Metabolism* 2004; 53:638-643.
 29. Gavella M, Lipovac V, Car A, Vucic M, Sokolic L and Rakos R: Serum sialic acid in subjects with impaired glucose tolerance and in newly diagnosed type 2 diabetic patients. *Acta Diabetol.* 2003; 40:95-100.
