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### ANTIMICROBIAL AND ANTIOXIDANT ACTIVITY OF *CHLAMYDOMONAS REINHARDTII* SP.

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#### ABSTRACT

##### Keywords:

Algae,  
*Chlamydomonas reinhardtii*,  
GC-MS,  
antimicrobial,  
antioxidant,  
phenolic content

The present investigation was undertaken to study the antimicrobial and antioxidant activity in algae (*Chlamydomonas reinhardtii* CC 124). The extractions of metabolites were done by two methods (glass bead and steel ball) using methanol: chloroform: water (5:2:2) as solvents. The presences of metabolites were identified by thin layer chromatography and structural analysis was done by Gas Chromatography and Mass Spectrometry. Antimicrobial activities of the extracts were assessed against bacterial and fungal strains (*Bacillus subtilis*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Candida albicans*, *Aspergillus niger*, *Aspergillus flavus*) by disc diffusion method and compared with standard antibiotics. The results, of both extracts showed more potent antimicrobial activity in (*B. subtilis*, *P. aeruginosa*, *K. pneumoniae*) bacterial species and (*C. albicans*) fungal strain. The antioxidant activity was tested by DPPH (2, 2-Diphenyl-1-picrylhydrazyl) free radical method and determination of H<sub>2</sub>O<sub>2</sub> radical scavenging activities were also performed. The presence of antioxidants was compared with commercial antioxidants, such as BHT (Butylated Hydroxy Toluene), BHA (Butylated Hydroxyl Anisole). The phenolic content was compared with gallic acid. From the present study, it can be concluded that the two different extracts of the algae has the antimicrobial and antioxidant activity.

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**INTRODUCTION:** *Chlamydomonas* is a genus of green alga, belonging to unicellular flagellates and has been used as a model organism in molecular biology, especially studies of flagellar motility, chloroplast dynamics, biogenesis, and genetics. *Chlamydomonas* species are known to produce high levels of galactose containing polysaccharides that can be hydrolyzed to produce monosaccharides<sup>1</sup>.

The production of extracellular antibiotic metabolites by marine algae has been well studied in recent years<sup>2-3</sup>. Various strains of cyanobacteria are known to produce intracellular and extracellular metabolites with wide biological activities such as antialgal, antibacterial, antifungal and antiviral activity<sup>4</sup>. Both cell extracts and extracts of the growth media of

various unicellular algae (e.g. *Chlorella vulgaris*, *Chlamydomonas pyrenoidosa*) has proved to have antibacterial activity *in vitro* against both Gram-positive and Gram-negative bacteria<sup>5</sup>. It has also been reported that a wide range of antifungal activities were obtained from extracts of green algae, Diatoms and Dinoflagellates<sup>6</sup>. Brown-algal polyphenols and phlorotannins worked as antioxidants, antibacterial and anti-algal compounds<sup>7</sup>. Antioxidant compounds play an important role against various diseases (e.g., chronic inflammation, atherosclerosis, cancer and cardiovascular disorders) and ageing processes, which explains their considerable commercial potential in medicine, food production and the cosmetic industry<sup>8</sup>.

In the earlier publication interest in employing antioxidants from natural sources was considerably enhanced by consumer preference for natural products and concern about potential toxic effects of synthetic antioxidants. The potential of metabolite extracts of *C. reinhardtii* CC 124 for practical applications has not been previously studied in detail, thus this study provides important information concerning the antimicrobials and antioxidants present in *C. reinhardtii* CC 124.

## MATERIALS AND METHODS:

**Algal material:** *Chlamydomonas reinhardtii* strains CC-124 were acquired from Centre for Plant Molecular Biology (CPMB), Tamilnadu Agriculture University (TNAU), and Coimbatore and maintained in Tris-acetate-phosphate (TAP) medium at 25°C under constant illumination with cool white fluorescent light and kept in an Erlenmeyer flask for 125 rpm for two weeks. The cultures ( $5 \times 10^5$  cells/ml) were inoculated in fresh medium (TAP) and maintained for 48 hours. After the incubation period the cells were centrifuged. The harvested cells were washed twice with sterile 20 mM Tris (pH 7.0), supplied with 300 mM CaCl<sub>2</sub>, 400 mM MgCl<sub>2</sub>, and 7 mM KCl, and resuspended at a starting density of  $2.5 \times 10^6$  cells/ml in 20 ml of Tris-buffered media under standard growth conditions for 24 hours<sup>9</sup>.

**Extraction of metabolites using Glass bead and Steel ball:** 1ml of the cell suspension was mixed with 1ml of 70% ice cold methanol. The mixture was centrifuged for 10 minutes at 14,000 rpm at 4°C. The supernatant was discarded and the pellet was removed carefully and stored. The pellet was mixed with 0.5ml of solvent mixture in the ratio of 5:2:2 (methanol: chloroform: water) in ice cold condition<sup>10</sup>. According to Oliver Fiehn *et al.*, (2006) the sample was extracted with slight modifications. 100µl of the extract was mixed with single glass beads (3mm) and steel ball (5mm) and crushed thoroughly. The extracts were collected and centrifuged at 14,000 rpm for 2 minutes. The supernatant was collected and mixed with the solvent mixture and centrifuged at 14,000 rpm for 2 minutes. The upper phases of the two samples were lyophilized and stored in -80°C<sup>11</sup>.

**Thin Layer Chromatography:** The presence of metabolites was checked using TLC. To separate the compound present in the dried extract (glass bead and steel ball method) two solvent systems were used. Solvent 1 (chloroform: methanol: water (65:25:4v/v/v)) and Solvent 2 (chloroform: methanol: concentrated ammonia (65:35:5 v/ v/ v)). To confirm the presence of the compounds the TLC plates were exposed to iodine vapor for 5 minutes<sup>12</sup>.

**GC-MS analysis:** The algal samples (glass bead and steel ball method) were analyzed by GC-MS [SHIMADZU instrument, model: GC-MS QP2010]. The GC silica column dimension was 30m × 0.25mm × 0.25µm DB-5ms. The conditions for GC were an injector temperature of 200°C, column temperature of 80°C and programmed to rise up to 300°C at the rate of

10°C/min and stand for 10 minutes. The ion source temperature was 200°C and the interface temperature was 240°C. Helium gas was engaged as a carrier at the rate of 1ml/min. The spectra were obtained in EI mode with 70eV ionization energy. The compounds were identified by comparing with the standards or the mass spectra were matched with the inbuilt library (NIST/Wiley).

**Antimicrobial Assay:** Antibacterial and antifungal activities of algae were tested by well diffusion method. The antimicrobial activity of microalgae was compared with antibiotics like (Amikacin, Amoxicillin, Streptomycin, and Erythromycin). Six microbial strains were used in this study namely *Bacillus subtilis*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Candida albicans*, *Aspergillus niger* and *Aspergillus flavus*. The extracts (glass bead and steel ball method) were dissolved in 2% DMSO (Dimethyl sulfoxide) (200mg/ml) and subjected to agar plates in various concentrations (20µl, 40µl, 60µl and 80µl). The inoculated plates were incubated for 24 hours at 37 °C and zone of inhibition was measured<sup>13-14</sup>.

$$\text{Zone of inhibition (Z)}_{\text{mm}} = \frac{\text{Diameter of the zone (mm)}}{\text{Diameter of the zone (mm)}}$$

### Antioxidant Property Assay:

**DPPH (2, 2-Diphenyl 1-2 picrylhydrazyl) Assay:** DPPH (2, 2-diphenyl-1-picrylhydrazyl) radical scavenging activity were determined for the extracts (glass bead and steel ball method). 1ml of methanolic solution of DPPH radicals ( $3 \times 10^{-4}$  mol/l) were added to 2.5 ml of sample and measured immediately (without incubation) after 20 minutes of incubation period. The absorbance recorded spectrophotometrically at 517 nm. A decrease by 50% of the initial DPPH concentration was defined as the  $IC_{50}$  (Inhibition Concentration). The  $IC_{50}$  value ( $\mu\text{g/ml}$ ) was determined for all the extracts. BHT was used as the reference compound ( $IC_{50} = 0.021 \mu\text{g/ml}$ )<sup>15-16</sup>. The capacity of the scavenging free radicals were calculated as follows

$$\text{DPPH} = 100 \left( 1 - \frac{A_s - A_b}{A_c} \right)$$

$A_s$  = sample absorbance of the algal extract in the DPPH solution,

$A_b$  = blank absorbance of the sample extract without DPPH solution

$A_c$  = absorbance of the control solution containing only DPPH (1 ml of DPPH radical of  $3 \times 10^{-4}$  mol/L concentration + 2.5 ml of methanol).

**$H_2O_2$  radical scavenging assay:** The  $H_2O_2$  radical scavenging assay in the *C. reinhardtii* was determined using BHA (Butylated Hydroxyl Anisole)<sup>17</sup>, as a standard. Samples at different concentration in ethanol was added to  $H_2O_2$  solution (0.6 ml, 40mM) and allowed to stand for 10 minutes. The absorbance was measured at 230 nm and a blank solution containing phosphate buffer without  $H_2O_2$  was used. The percentage of  $H_2O_2$  scavenging of both the extracts and standard compounds were calculated ( $IC_{50} = 0.011 \mu\text{g/ml}$ ). The  $H_2O_2$  radical scavenging activity was calculated as follow

$$\text{Scavenging effect} = \frac{\text{Control} - \text{Sample}}{\text{Sample}} \times 100$$

**Total Phenolic Content:** The total phenolic content in the *C. reinhardtii* extracts were determined spectrophotometrically according to the Folin-Ciocalteu<sup>18</sup>, method using gallic acid as a standard (the concentration range: 0.025 to 0.5  $\mu\text{g/ml}$ ). 200 $\mu\text{l}$  of the sample was mixed with 9 ml of distilled water, and 1 ml of Folin-Ciocalteu reagent and 10 ml of 7% sodium carbonate was added. After 90 minutes of incubation, the absorbance was determined at 765 nm. The total phenolic content was expressed as GAE (Gallic Acid Equivalent) in milligram per gram extract<sup>19</sup>. The standard absorbance at 765 nm =  $0.431 C_{\text{gallic acid}} (\mu\text{g/ml}) - 9.33 \times 10^{-3}$ ,  $R^2 = 0.9992$ .

### RESULTS AND DISCUSSION:

#### Detection of metabolite compounds by Thin Layer Chromatography:

The metabolites present in the two extracts were identified by thin layer chromatography. The results showed the presence of more compounds in steel ball method extraction than the glass bead extract. Similar work was done in TLC and the presence of compounds in *C. reinhardtii* CC 124 were reported as Gal-acyl<sub>2</sub> Gro, acyl<sub>2</sub> Gro-Me<sub>3</sub>Hse, PtdGro, PtdEtn, SQui-acyl<sub>2</sub> Gro, Gal -acyl<sub>2</sub> Gro, PtdIns<sup>20</sup>.

#### Analysis of *C. reinhardtii* CC 124 metabolites by using GC-MS analysis:

The composition of the volatile compounds of the *C. reinhardtii* glass bead and steel ball extract was determined by GC/MS. Different groups of compounds identified such as hydrocarbons, phenols, alcohols and esters are shown in **Table 1** and **2**. The glass bead extract has 10 compounds and the steel ball extract has 11 compounds **fig. 1** and **2**. The compounds which were identified through mass spectrometry were found to exhibit the biological and pharmacological activity. In recent years, many studies on volatile compounds from *C. reinhardtii* CC 124 have been published<sup>10-11</sup>.

Previous publications reported that the compounds such as 1-Nonadecene, 1-Octadecene, 1-Tetradecene, Diisooctyl phthalate, and 1-Heptacosanol were found in both algae and plants shows anticancer, antioxidant and antimicrobial activity<sup>21-22</sup>.

TABLE 1: GC-MS PROFILE OF GLASS BEAD EXTRACT

Retention time	Compound name	Area (%)
9.702	1-Tetradecene	5.84
11.261	Phenol, 2, 4-di-tert-butyl-	15.18
12.233	1-Pentadecene	10.96
12.540	N-(1, 1-Dimethylpropyl)-2, 2, 3-trimethylaziridine-1-carboxamide	1.54
14.506	1-Octadecene	20.18
16.564	1-Nonadecene	14.74
18.430	9-Tricosene, (Z) -	10.64
20.158	n-Eicosanol	5.73
21.404	Diisooctyl phthalate	12.16
21.748	1-Eicosanol	3.03

TABLE 2: GC-MS PROFILE OF STEEL BALL EXTRACT

Retention time	Compound name	Area (%)
6.890	6-Methyl-1-heptanol	2.34
9.722	1-Dodecanol	9.39
11.281	2, 4-Ditert-Butylphenol	5.07
12.249	(9E)-9-Icosene	3.04
14.513	1-Nonadecene	18.88
16.564	Behenic alcohol	7.12
18.437	Lignoceric alcohol	13.18
20.157	Eicosyl trifluoroacetate	9.15
21.429	Bis (2-ethylhexyl) phthalate	2.70
21.744	1-Heptacosanol	5.77
23.221	Nonadecyl pentafluoro propionate	3.34

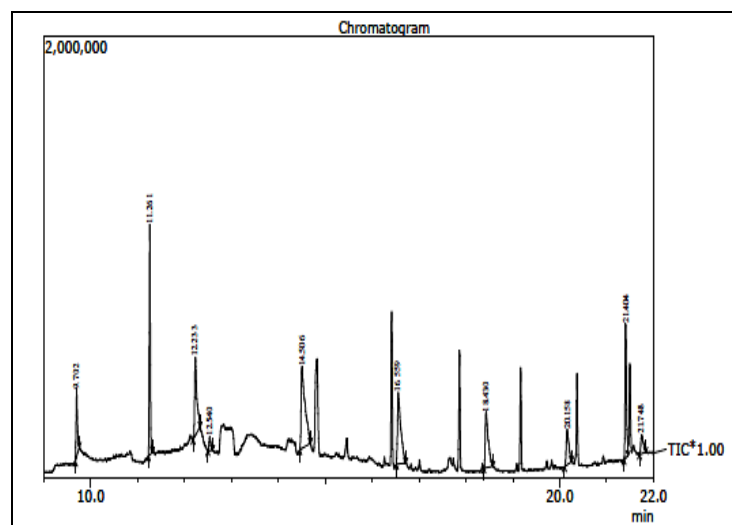


FIG 1: GC- MS PROFILE FOR GLASS BEAD EXTRACT OF C. REINHARDTII CC124

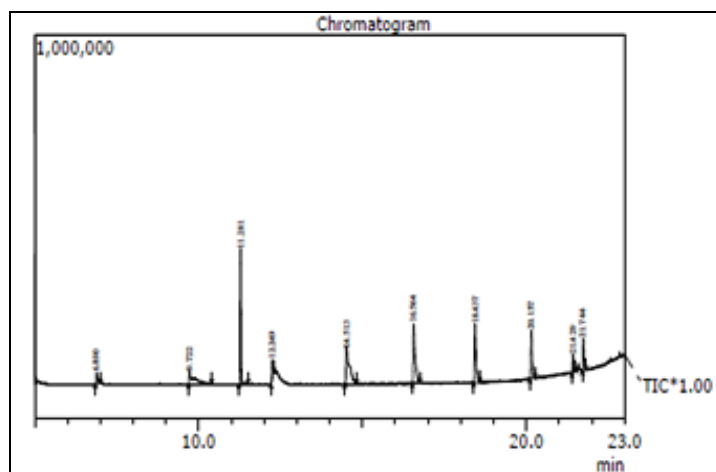


FIG 2: GC- MS PROFILE FOR STEEL BALL EXTRACT OF C. REINHARDTII CC 124

**Antimicrobial Activity of crude extract:** The antimicrobial activity of the different extracts of *C.reinhardtii* was determined by agar well diffusion method using bacterial and fungal strains and compared with antibiotics (Amikacin, Amoxicillin, Streptomycin, and Erythromycin). *C. reinhardtii* CC 124 showed the inhibition zone for microorganisms such as *B. subtilis*, *P. aeruginosa*, *K. pneumoniae*, *C. albicans* and no inhibition was observed in *A. niger* and *A. flavus*. The results of antimicrobial activity of glass bead extract and steel ball extract and zone of inhibition for antibiotics were tabulated in **Table 3 & 4**. Similar results were given, in brown algae and green algae<sup>13-14</sup>. The antimicrobial activity in previous reports confirms that *C. reinhardtii* CC 124 possess antimicrobial effects<sup>23-24</sup>.

**Antioxidant Activity of crude extract DPPH (2, 2-Diphenyl 1-2 picrylhydrazyl) Assay:** Chemicals which are able to change the color of DPPH free radical from purple to yellow can be considered as antioxidants and therefore, a radical scavenger<sup>15</sup>. The *C. reinhardtii* CC 124 extract were tested for antioxidant activity using various concentrations of the glass bead and steel ball extracts ranging from 10-50µg/ml and compared with the standard BHT. The 50 µg/ml concentration showed the IC<sub>50</sub> value of 66.13% for glass bead and 71.13% for steel ball. The result shows the presence of strong antioxidant in steel ball extract than the glass bead extract, which is equal to the Standard BHT. The absorbance of algal extracts and standard BHT, percentage inhibition was calculated and were tabulated in **Table 5**.

TABLE 4: ANTIMICROBIAL ACTIVITY OF STEEL BALL EXTRACT OF *C. REINHARDTII* CC 124 BY WELL DIFFUSION METHOD

	Volume (µl)	<i>B. subtilis</i> (mm)	<i>P. aeruginosa</i> (mm)	<i>K. pneumoniae</i> (mm)	<i>C. albicans</i> (mm)	<i>A. niger</i> (mm)	<i>A. flavus</i> (mm)
Extract of <i>C. reinhardtii</i> metabolites in various concentration	20	12	10	15	9	-	-
	40	16	15	20	13	-	-
	60	21	20	23	16	-	-
	80	26	24	30	21	-	-
Antibiotic Activity against Microorganism (10 µg)	Amikacin	-	17	26	12	-	-
	Amoxicillin	31	30	13	18	-	-
	Streptomycin	12	9	17	7	-	-
	Erythromycin	23	21	-	20	-	-

TABLE 5: ANTIOXIDANT ACTIVITY OF GLASS BEAD AND STEEL BALL EXTRACT OF *C. REINHARDTII* CC 124 BY DPPH METHOD

Concentration (µg/ml)	Glass bead extract of <i>C.reinhardtii</i> % Inhibition	Steel ball extract of <i>C.reinhardtii</i> % Inhibition	Standard BHA (Butylated Hydroxyl Anisole) % Inhibition
10 µg/ml	19.54	25.04	54.93
20 µg/ml	47.16	48.9	124.03
30 µg/ml	62.79	69.89	199.33
40 µg/ml	72	88.72	278.20
50 µg/ml	90.69	107.34	359.01

**Determination of H<sub>2</sub>O<sub>2</sub> radical scavenging activity for antioxidant assay:** The scavenging activities were determined for glass bead and steel ball extract range from 10-50µg/ml and compound with BHA standard. The concentration of 50µg/ml showed higher scavenging activity in glass bead sample 90.69% and steel ball has 97.34%. The activity was found to be approximately linear with R<sup>2</sup> (Correlation coefficient)

value for glass and steel ball respectively 0.978 and 0.995. The absorbance of algal extracts and standard BHA at 230nm were recorded, percentage inhibition were calculated and tabulated in **Table 6**. In earlier studies the algal extract of brown algae species showed good antioxidant activity. Among the previous results, Hexane extracts of *D. dichotoma var. implexa* were found to have good antioxidant activity<sup>19-18</sup>.

TABLE 6: DETERMINATION OF H<sub>2</sub>O<sub>2</sub> RADICAL SCAVENGING ACTIVITY (ANTIOXIDANT ASSAY) OF GLASS BEAD AND STEEL BALL EXTRACT OF *C. REINHARDTII* CC 124

Concentration (µg/ml)	Glass bead extract of <i>C.reinhardtii</i> % Inhibition	Steel ball extract of <i>C.reinhardtii</i> % Inhibition	Standard BHT (Butylated Hydroxy Toluene) % Inhibition
10 µg/ml	4.27	6.47	74.67
20 µg/ml	19.73	21.33	77.33
30 µg/ml	36.81	45.27	80.00
40 µg/ml	52.67	61.33	85.67
50 µg/ml	66.13	71.13	93.20

**Estimation of Total Phenolic Content:** The extract which has high antioxidant activity shows the presence of high phenolic content. The extracts were determined for the presence of total phenolic content by Gallic acid standard. The results for both the extracts of *C. reinhardtii* CC 124 are valued as 42.2 mg/ml and 51mg/ml **Table 7**. From the phenolic content it was noted that *C. reinhardtii* has high

antioxidant activity. The previous result in plant extracts with a high phenolic content also shows high flavonoid content<sup>25</sup>.

TABLE 7: TOTAL PHENOLIC CONTENT OF *C. REINHARDTII* CC 124

Extract	Total phenolic content, GAE (mg/ml)
Glass bead	42.2
Steel ball	51

**CONCLUSION:** Marine organisms have several active chemicals such as antioxidant and antimicrobial compounds. The present study proves that the steel ball extracts of *C. reinhardtii* CC 124 showed profound antimicrobial, antioxidant effect and have potential uses in medicine. Metabolites identified during this study represent different structural classes of natural products. This indicates that the marine derived algae have a wide range of biosynthetic capabilities for the production of novel drugs. These compounds might have still undiscovered biological activities.

The work can be further modified by complete purification of the metabolites and can be used in pharmacological studies for the potential discovery of new natural bioactive compounds. Much work remains to explore the metabolic characteristics of *C. reinhardtii* CC 124 in relation to defense as well as symbiosis between marine algae and their associated organisms. Finally, to our knowledge, this is the first time that antimicrobial, and antioxidant property, have been described for *C. reinhardtii* CC 124.

## REFERENCES:

- Behrens J, von Kries JP., Kuhl, ML. Bruhn,Wedlich.D: Functional interaction of  $\beta$ - Catenin with the transcription factor LEF-1. *Nature*1996; 382: 3638-3642.
- Hornsey and Hide: The production of antimicrobial compounds by British marine algae, Antibiotic production marine algae. *Br. phycol. J.*1974; 9: 353-361.
- Henriquez P, Candia A, Norambeuena R, Silva M, and Zemelman R: Antimicrobial Properties of marine algae II Screening of the Chilean coast for antimicrobial activity. *Botanica*1979; 22: 451-453.
- Noaman NH, Fattah A, Khaleafa M, Zaky SH: Factors affecting antimicrobial activity of *Synechococcaleopoliansi*. *Microbiological Research* 2004; 159: 395-402.
- Pratt, Daniels, Eiler, Gunnison and Kumler: Chlorellin an antibacterial substance from Chlorella. *Science*1944; 99: 351-352.
- Cannell RJP, Kellam SJ, Owsianka AM, and Walker JM,Results of a large scale screening programme to detect antibacterial activity from freshwater algae, *Br.phycol. J.*1988; 23:147- 153.
- Kuda T, Kunii T, Goto H, Suzuki T, Yano T, Changes of radical-scavenging capacity and ferrous reducing power in chub mackerel *Scomber japonicus* and Pacific saury *Cololabis saira* during 4°C storage and retorting,*Food Chem.* 2007;103:900-905.
- Shibata T, Hama Y, Miyasaki T, Ito M, Nakamura T, J, Extracellular secretion of phenolic substances from living brown algae,*Appl. Phycol* 2006;18:787-794.
- Masamichi Akimoto, Hidenao Yamada, Kazuhisa Ohtaguchi, and Kozo Koide, Photoautotrophic cultivation of the green algae *Chlaydomomonas reinhardtii* as a method for carbon dioxide fixation and a Linolenic acid production. *JAOCS*1996; 74: 181.
- Do Yup Lee and Oliver Fiehn, High quality metabolomic data for *Chlaydomomonas reinhardtii*. *Plant Methods* 2008; 4:1-13.
- Christian Bölling and Oliver Fiehn, Metabolic profiling of *Chlaydomomonas reinhardtii* under nutrient deprivation, *Plant Physiol* 2006; 139:1995-2005.
- Vogel G, and Eichenberger W, Betaine lipids in lower plants .*Biosynthesis of DGT in lipid metabolism.Plant Cell Physiol* 1992; 33: 427-436.
- Chiheb Ibtissam, Riadi Hassane, Martinez-Lopez José, Dominguez Seglar Josè Francisco, Gomez Vidal Josè Antonio, Bouziane Hassan and Kadiri Mohamed, Screening of antibacterial activity in marine green and brown macroalgae from the coast of Morocco. *African Journal of Biotechnology.* 2009; 8: 1258-1262.
- Rajasulochana P, Dhamotharan R, Krishnamoorthy P, Murugesan S, Anti bacterial activity of the extract of marine red and brown algae.*Journal of American Science* 2009; 5: 20- 25.
- Aquino R, Morelli S, Tomaino A, Pellegrino M,Saija A, Grumetto L, Puglia C, Ventura D, Bonina F, Grumetto L, Antioxidant and photo protective activity of a crude extract of *Culcitium reflexum* leaves and their major flavonoids. *J. Ethnopharmacol* 2002; 79:183-191.
- Sanchez-Moreno C, Methods used to evaluate the free radical scavenging activity in foods and biological systems. *Food Sci. Technol. Int.* 2002; 8:121-137.
- Ruch R J, Cheng S J, and Klaunig J E,Prevention of cytotoxicity and inhibition of intercellular communication by antioxidant catechins isolated from Chinese green tea. *Carcinogenesis*1989; 10: 1003-1008.
- Waterman P G, and Mole S, Analysis of Phenolic plant Metabolites, Blackwell Scientific Publication, Oxford Blackwell Scientific Publication 1994.
- Singleton V L, Orthofer R, Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Meth.Enzymol* 1999; 299: 152-178.
- Norihirosato, Mikiotsuzuki, Yoshihiro matsuda, Tomoko ehara, Tetsuakio safune and Akihiko kawaguchi, Isolation and Characterization of Mutants Affected in Lipid Metabolism of *Chlamydomonas Reinhardtii*.*Biochem*1995; 230: 987-993.
- Yong Soo Lee, Min Hee Kang, So Yean Cho, and Choon Sik Jeong. Effects of constituents of *amomum xanthioides* on gastritis in rats and on growth of gastric cancer cells. *Arch Pharm Res*2007; 30: 436-443.
- Pravat Manjari Mishra, and Sree A, Antibacterial Activity and GCMS Analysis of the Extract of Leaves of *Finlaysonia obovata* (A Mangrove Plant). *Asian Journal of Plant Sciences*2007; 6:168-172.
- Moreau J, Pesando D, Caram B, Antifungal and antibacterial screening of Dictyotales from the French Mediterranean coast. *Hydrobiologia*1984; 116/117:521-524.
- Inci Tuney, Bilge Hilalca dirci, Dilekunal, Atakansukatar, Antimicrobial activities of the extracts of marine algae from the coast of Urla.*Turk J Biol*2006; 30: 171-175.
- Ljiljana Stanojević, Mihajlo Stanković, Vesna Nikolić, Ljubiša Nikolić, Dušica Ristić, Jasna Čanadanovic-Brunet and Vesna Tumbas, Antioxidant Activity and Total Phenolic and Flavonoid Contents of *Hieracium pilosella* L. Extracts. *Sensors* 2009; 9: 5702-5714.