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ANTIFERTILITY ACTIVITY OF MEDICINAL PLANTS ON MALE AND FEMALE REPRODUCTION

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ABSTRACT: Population control is a significant issue worldwide especially in developing countries like India. Population breakout has responsible for various deleterious effects on life surviving resources on the earth. Therefore fertility regulation is necessary for the conservation of life supporting resources as well as good reproductive life of both males and females. Various chemical methods of contraception are available today but these methods possess several side effects. Herbal medicinal plants have been used as safe alternatives of the chemical methods. Evaluation of the herbal medicinal plants has been in progress for several decades to identify effective and safe substances for fertility regulation. Several medicinal plant extract were investigated for their antifertility activity both in male and female animal models. This review covered scientific proven information on various medicinal plants used for antifertility activity in both males and females. This review provides information on botanical name along with their common name, antifertility activity, part used and their phytochemicals present in plants.

INTRODUCTION: Fertility control is a significant issue of global and national public health concern. Ever increasing human population throughout the world particularly in developing and underdeveloped countries has inevitable effects on the life supporting resources on the earth and also detrimental effects on all aspects of development especially employment, education, housing, health care, sanitation and environment ^{1, 2}. In 2005 G.C., world population is estimated to be 6.5 billion. The number is expected to increase by 2.5 billion over the next 45 year, 6.5 billion to 9 billion in 2050.

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The developing countries absorbed 95% of all population growth and only 5% contributed by the developed world ³. Population explosion is one of the serious problems in developing countries like India that would be increased about 9.2 billion by the year 2050. The census of 2005 showed that the growth rate of population in India during the previous 10 years was about 1.5%. Annually, around 18 million people are adding to our total population which leads to an extra burden on the community and finally responsible for poverty and pollution in developing countries ⁴.

Due to all these detrimental effects, fertility regulation comprising contraception and management of infertility forms an important component of reproductive health for both male and females ⁵. Several effective approaches for the induction of infertility have been investigated over a long period including hormonal, chemical and

immunological approaches ¹. The chemical approaches possess various side effects like obesity, cholethiasis, gastric trouble and carcinoma of breast and cervix, asthma and thromboembolism which decrease their popularity and utility among women. The hormonal contraceptives also have carcinogenic effects. Therefore, now a days, plant products catches the attention of many scientists as a primary source of naturally occurring fertility regulating agents because of their little and no side effects ⁴. Higher use of contraceptives methods is a direct indicator of health, population development and women empowerment ⁶. Several herbal plants possess different types of antifertility activities both in males and females ⁷.

Some herbal plants exhibiting potent antifertility activity for males:

1. Gossipium herbaceum:

Common name: Levant cotton

Family: Malvaceae

Chemical constituents: It contains Gossypol (yellow colour phenolic compound), sugar, gum, tannins and fixed oil¹.

Activities:

- i. It reduced the level of serum testosterone and luteinizing hormone in dose dependent manner ⁸.
- ii. It induced the azoospermia or oligospermia by directly acting on the testis ⁹.
- iii. It was found that it inhibits sperm motility by blocking the cAMP formation in the sperms ¹⁰.
- iv. It was acts on the pituitary gonadal axis and decreased the secretory activity of accessory sex glands ¹¹.
- v. It inhibited T-type Ca²⁺ currents in mouse spermatogenic cells ¹².

Type of extract used: Root bark tincture was showed male conceptive activity.

Animal models: Hamester ¹³ and rats ^{14, 15} were used for finding the male contraceptive potential of *Gossipium herbaceum*.

2. Carica papaya:

Common name: Papaya

Family: Caricaceaea

Chemical constituents: It contains papain, chymopapain, pectin, carposide, carpaine, pseudocarpaine, dehydrocarpines, carotenoids, cryptoglavin, cis-violaxanthin and antheraxanthin ⁷.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Activities:

- i. It suppressed cauda epididymal sperm motility and also reduced the sperm count without influencing libido of animals¹⁶.
- ii. It induced long term reversible azoospermia
- iii. It also caused sterility in rats due to total suppression of sperm motility ¹⁸.
- iv. It was also caused degeneration of germinal epithelium and germ cells, reduction in the number of leydig cells and vacuoles in the tubules ¹⁹.
- v. It altered cauda epididymal microenvironment ²⁰.
- vi. It was also reduced the contractile responses of cauda epididymal tubules and retarded the sperm transport in cauda epididymus ²¹ and also caused ultrastructural changes in testis and epididymus of rats ²².

Type of extract used: Aqueous, benzene and chloroform extracts of papaya seeds used for reversible antifertility action ⁷.

Animal models: Male rats were used for investigating the antifertility action of papaya seeds ⁷

3. Hibiscus rosasinensis:

Common name: Gudhal

Family: Malvaceae

Chemical constituents: It contains steroids, tannins, saponins and flavonoids ²³.

Activities:

i. It possessed antispermatogenic and antiandrogenic activity ^{24, 25, 26}.

Type of extract used: 50% ethanolic extract, benzene and benzene/ether extract of flowers were showed antifertilty activity ^{24, 25, 26}.

Animal models: Rats ²⁵, Nonscrotal bat ²⁴ and mice ²⁶ were used for demonstrating the above mentioned antifertility activity of *Hibiscus rosasinensis*.

4. Andrographis paniculata:

Common name: Kirayat

Family: Acanthaceae

Chemical constituents: It contains flavonoids, andrographilode, diterpenoids, phenylpropanoids, oleanolic acid and β -sitosterol ²⁷.

Activities:

- i. It caused cessation of spermatogenesis, degeneration in seminiferous tubules and regression of leydig cells. Degeneration had also been seen in accessory sex organs and its fluid content ²⁸.
- ii. It was prevented cytokinesis of the dividing spermatogenic cell lines.
- iii. It also caused decrease in sperm motility and sperm count and also provides abnormalities in sperms ²⁹.

Type of extract used: Dry leaf powder was generally used at the dose of 20 mg powder/day/rat.

Animal models: Rats were used for identifying the antifertility efficacy of *Andrographis paniculata*.

5. Tripterygium wilfordii:

Common name: Thunder god vine

Family: Celastraceae

Chemical constituents: It contains Triptolide (diterpene epoxide), Tripchlorolide and glycosides.

Activities:

It caused degenerative changes in seminiferous tubular epithelium and decrease in plasma testosterone ³⁰.

It was inhibit the Ca²⁺ channel activity in mouse spermatogenic cells ¹².

It also caused severe impairment in cauda epididymal sperm ^{31, 32}.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Type of extract: Crude extract of roots was showed antifertility action.

Animal models: Rats and mice ³³ were used.

6. Solanum surattense:

Common name: Kateli

Family: Solanaceae

Chemical constituents: It contains sterols, alkaloids (solasodine), saponins, flavonoids and glycosides ³⁴.

Activities:

- i. It caused disruptive changes in the acrosomal membrane of sperm and arrest spermicidal motility ³⁵.
- ii. It was also cause degenerative changes in seminiferous epithelium and spermatogonic elements ³⁶.

Type of extract used: 50% ethanolic extract of roots ³⁶, alcoholic extract of seeds ³⁷ were utilized.

Animal models: Male rats ³⁶ was used for checking the antifertility potential of *Solanum surattense*.

7. Embelia ribes:

Common name: Laksmana, amalaki

Family: Myrsinaceae

Chemical constituents: It contains alkaloids, quinine, proteins, saponins, triterpenes, coumarins, resins, tannins and Embelin (2, 5-dihydroxy 3-undicyl-1, 4-benzoquinone) ^{38, 39}.

Activities:

- i. It was affected the sperm motility, quantity and quality of semen and lowered the hormonal level ⁴⁰.
- ii. It produced profound morphological and histological changes in testis ^{38, 41}.

Type of extract used: Extract of berries was used as a fertility regulating agent.

Animal models: Male bonnet monkeys ⁴⁰ were utilized.

8. Stephania hernandifolia:

Common name: Aknadi Family: Menispermaceae

Chemical constituents:

i. It contains alkaloids (hernandin, hernsubanine), proteins and tannins ⁴².

Activities:

i. It showed deminution of the activities of testicular androgenic key enzymes and plasma testosterone with spermatogenesis 43

Type of extract used: Aqueous extract of leaf was used.

Animal models: Rats ⁴³ were used for demonstrating the antifertility activity of *Stephania hernandifolia*.

9. Catharanthus roseus:

Common name: Sadabahar

Family: Apocynaceae

Chemical constituents: It contains tannins, steroids, saponin glycosides, cardiac glycosides, anthraquinone glycosides and flavonoids ⁴⁴.

Activities:

- ii. It produced pathological changes in the principle and apical cells of caput and nuclear cells of cauda causing impairment of epididymal functions ⁴⁵.
- iii. It affected spermatogenic cell lines other than spermatogonia 46,47.

Type of extract used: Extract of leaf ⁴⁷ was used for carried out male contraception.

Animal models: Male rats ⁴⁷ were used for investigating the male conception activity of *Catharanthus roseus*.

10. Abrus precatorius:

Common name: Indian liquorice, chirmi

Family: Fabaceae

Chemical constituents: It contains glycosides, alkaloids, tannins and flavonoids ⁴⁸.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Activities:

- i. In Srilanka, ayurvedic physicians have been claimed that seeds of A. procatorius inhibit conception in humans when taken orally ⁴⁸.
- ii. It altered the sperm morphology, reduced sperm motility and metabolism ⁴⁹.
- iii. It reduced testicular weight and sperm count and also causes degeneration in the testis during the later stages of spermatogenesis

Type of extract used: 50% ethanolic extract of seeds was used for antifertility action in males ⁵⁰.

Animal models: Rats^{51, 52}, rabbits and presbytis monkeys⁵³ were used for finding the above mentioned activities.

11. Azadirachta indica:

Common name: Neem

Family: Meliaceae

Chemical constituents: It contains flavonoids, saponins, phenols and chymophenols ⁵⁴.

Activities:

- i. It caused disturbances in the structure and functions of testis and spermatozoa ⁵⁴.
- ii. It also produced histopathological and biochemical changes in the caput and cauda 55
- iii. It reduced the serum testosterone level ⁵⁵.
- iv. It produced mass atrophy in spermatogenic elements ⁵⁶ and arrested the spermatogenesis stage (late XII).
- v. It caused morphological changes in the head of the sperm and its acrosome due to androgen deficiency ⁵⁷.
- vi. It was also showing the spermicidal activity

Type of extract: Seed oil ⁵⁹, Neem oil ⁶⁰ and ethanolic extract of leaves ⁶¹ were established for male infertility.

Animal models: Rats ^{62, 59, 63}, mouse ⁶⁰, monkeys ⁶⁴ and humans ⁶⁴ were used for evaluating the antifertility activity in males.

12. Aegle marmelos:

Common name: Bael fruit tree

Family: Rutaceae

Chemical constituents: It contains marmin, fagarine, skimmianine, aegelin, lupeol, citral, cineol, citronella, cumin aldehyde, eugenol and marmesinine ⁶⁵. Marmin and fagarine is especially responsible for male infertility.

Activities:

- i. It was significantly reduces the reproductive organs weight and serum testosterone level
- ii. (It also reduced sperm density, motility, viability and sperm acrosomal integrity ^{67,} 68, 69
- iii. It was also responsible for exfoliation of elongated spermatids, nuclear chromatin condensation, degeneration and prominent spaces detected within the germinal epithelium which indicated testicular cytotoxicity and necrosis ⁶.

Type of extract used: Methanolic extract of aegle marmelos bark, leaves, seeds and fruits were used for male infertility ^{67, 68, 69, 6}.

Animal models: Rats ⁶ were used as preclinical model for evaluating the antifertility activity in males.

13. Apium graveolens:

Common name: Celery

Family: Apiaceae

Chemical constituents: It contains essential oils, sesquiterpenes, flavonoids, coumarins and furocoumarins ⁷⁰.

Activities: It arrested spermatogenesis as well as decrease sperm count, sperm motility, blood

testosterone concentration, weight of testes and seminal vesicles and diameter and viability of seminiferous tubules ⁷⁰.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Type of extract used: Ethanolic extract of *Apium graveolens* seeds was administered for male infertility ⁷⁰.

Animal models: Rats ⁷⁰ were used for evaluating the antifertility activity of *Apium graveolens*.

Some herbal plants exhibiting antifertility activity in females:

Several medicinal plants associated with antifertility activity in females are exists in India. These plants produce antifertility activity by acting through various mechanisms:

- (a) Estrous cycle disruptors
- (b) Anti-estrogenic agents
- (c) Anti-implantation agents
- (d) Abortifacient agents

Estrous cycle disruptors:

1. Rivea hypocrateriformis:

Common name: Night glory, vaividang

Family: Convolvulaceae

Chemical constituents: It contains alkaloids, glycosides, saponins, tannins and phenolic compounds ⁷¹.

Activities:

- i. After administration of extract, the level of cholesterol increased due to the inhibition of steroidogenesis.
- ii. The myometrium and endometrium thickness and diameter were found to be increased indicating the uterotrophic effect.
- iii. The number of graffian follicles declined and increased in number of atretic follicle indicating antiovulatory effect ⁷².

Type of extract used: Ethanolic extract of *Rivea hycrateriformis* at dose level of 200-400 mg/kg/body weight in rats ⁷² disrupts the estrous cycle.

Animal models: Rats were used to explore the antifertility potential of *Rivea hypocrateriformis* 72 .

2. Momordica charantia:

Common name: Bitter melon, karela

Family: Cucurbitaceae

Chemical constituents: The principle phytochemicals which causes antifertility are steroids, triterpinoids, reducing sugars, sugars, alkaloids, phenolic compounds, flavonoids and tannins ⁷³.

Activities:

- i. It exhibited irregular pattern of estrous cyclicity and increases the length of estrous cycle ⁷⁴.
- ii. The disruption of the estrous cycle was found to be executed by disturbance in ovarian function and estrous cyclicity through interplay of ovarian and extraovarian hormones ⁷⁴.
- iii. It may be inhibiting the estrogen production or competing for its receptor ⁷⁵.

Type of extract used: Methanolic extract of *Momordica charantia* seeds caused disturbances in the estrous cycle ⁷⁴.

Animal model: Rats ⁷⁴ were used for evaluating the female antifertility activity of *Momordica charantia*.

3. Aspilia Africana:

Common name: wild sunflower

Family: compositae

Chemical constituents: The phytochemicals are saponins, tannins, flavonoids and cardiac glycosides ⁷⁶.

Activities:

- i. It caused the alteration in estrous cycle by the prolonged proestrous and a reduced diestrous and estrous phase ⁷⁷.
- ii. It reduced the number of ova observed in oviduct ⁷⁷.
- iii. It caused the inflammation of the fallopian tube, degeneration in the ovarian cortex in the stroma cells of the ovary ⁷⁷.

iv. It also caused the disruption of the endometrium of the uterus⁷⁷.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Type of extract used: Extract of *Aspilia africana* leaves was used for antifertility action ⁷⁷.

Animal models: Rats ⁷⁷ were utilised for finding the antifertility activity of *Aspilia africana*.

4. Anethum graveolens

Common name: Dill, sowa

Family: Umbelliferae

Chemical constituents: It contains tannins, glycosides, saponins, steroids, terpenoids and reducing sugars ⁷⁸.

Activities:

i. It increased the duration of diestrous phases and total time of the estrous cycle ⁷⁹.

Type of extract used: Ethanolic extract of Anethum graveolens was administered as estrous cycle disruptor ⁷⁹.

Animal models: Rats ⁷⁹ were used for demonstrating the antifertility activity of *Anethum graveolens*.

5. Cissampelos pareira:

Common name: Abuta, harjeuri

Family: Menispermaceae

Chemical constituents: Phytochemical analysis showed the presence of terpenoids, alkaloids, tannins, amino acid proteins and carbohydrates⁸⁰.

Activities:

- ii. It altered the estrous cycle pattern and prolonged the length of estrous cycle with significant increase in the duration of diestrous stage ⁸¹.
- iii. It also altered the secretion of luteinizing hormone (LH), follicle stimulating hormone (FSH), prolactin and estradiol ⁸¹.

Type of extract used: Extract of Cissampelos pareira leaves were used for antifertility activity ⁸¹.

Animal models: Rats ⁸¹ were used for checking the antifertility activity of *Cissampelos pareira* leaves.

6. Curcuma longa:

Common name: Haldi

Family: Zingiberaceae

Chemical constituents: Phtochemical analysis revealed the presence of flavonoids, aminoacids and alkaloids 82.

Activities:

- i. It caused the suppression of ovulation by the inhibition of estrous phase ⁸³.
- ii. It showed anti-estrogenic activity which either block the estrogen receptors or diminishing the estrogen synthesis due to decrease in cholesterol metabolism ⁸³.

Type of extract used: Extract of *Curcuma longa* was used for the suppression of estrous cycle ⁸³.

Animal models: Rats ⁸³ were used for investigating the female antifertility activity of *Curcuma longa*.

7. Acacia leucophloea

Common name: Reonja

Family: Mimosaceae

Chemical constituents: It contains tannins, flavonoids, terpenes and alkaloids ⁸⁴.

Activities:

- i. It increased the proestrous phase while estrous and metaestrous phase decreases ⁸⁵.
- ii. It caused decreases in the weight of ovary 85.
- iii. It increased the cholesterol content 85.

Type of extract used: Alcoholic extract of *Acacia leucophloea* roots was used for estrous cycle disruptors ⁸⁵.

Animal models: Rats ⁸⁵ were used to evaluate the antifertility activity of *Acacia leucophloea*.

Anti-estrogenic agents:

1. Butea monosperma:

Common name: Flame of the forest, dhak

Family: Fabaceae

Chemical constituents: It contains glucose, glycine, glycoside and an aromatic hydroxyl compound ⁸⁶.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Activities:

- i. It was significantly reduces the weight of the ovaries and increases the level of cholesterol ⁸⁷.
- ii. It inhibited the activity of G-6-PDH indicating anti-steroidogenic activity ⁸⁷.

Type of extract used: Petroleum ether and chloroform extract of *Butea monosperma* roots were used for antifertility action ⁸⁷.

Animal models: Mice ⁸⁷ were utilised for finding the antifertility activity of *Butea monosperma* extract.

2. Piper bitle:

Common name: Betel leaf, paan

Family: Piperaceae

Chemical constituents: It showed the presence of carbohydrates, alkaloids, gums, oils, steroids, glycosides, tannins, phenols, vitamins, organic acids and inorganic constituents ⁸⁸.

Activities:

It showed anti-estrogenic activity by decreasing the weight of the ovary ⁸⁹.

It reduced the circulating level of estrogen, fertility and number of litters ⁸⁹.

It also caused decline in serum glucose concentration, enzyme activity of acid phosphatise, SGOT and SGPT and increment in the level of cholesterol and ascorbic acid ⁸⁹.

Type of extract used: Ethanolic extract of *Piper bitle* leaves showing anti-estrogenic activity ⁸⁹.

Animal models: Rats ⁸⁹ were used for evaluating the anti-estrogenic activity of *Piper bitle* .

3. Cassia fistula:

Common name: Golden shower, amaltas

Family: Caesalpiniaceae

Chemical constituents: It showed the presence of anthraquinone glycosides, flavonoids, phenolic compounds and carbohydrates ⁹⁰.

Activities:

- i. It showed strong anti-estrogenic activity in presence of a strong estrogen like estradiol valerate and significantly reduces the estrogen induced uterotrophic effect ⁹¹.
- ii. It was also prevents pregnancy when extract administered to the mated female rats ⁹¹.

Type of extract used: Aqueous extract of *Cassia fistula* seeds was used for anti-estrogenic activity ⁹¹

Animal models: Rats ⁹¹ were used to explore the anti-estrogenic potential of *Cassia fistula* seeds.

4. Ocimum gratissimum:

Common name: Tulsi

Family: Lamiaceae

Chemical constituents: It contains the presence of alkaloids, phenolics, glycosides, resins, steroids and tannins ⁹².

Activities:

- i. It showed slight anti-estrogenic activity when given along with strong estrogen i.e. estradiol valerate ⁹³.
- ii. It declined the fertility index, number of uterine implants and live foetuses when administered in mated female rats ⁹³.

Type of extract used: Acetone extract of *Ocimum gratissimum* stem was administered for antiestrogenic activity ⁹³.

Animal models: Rats ⁹³ are used for finding antiestrogenic activity of *Ocimum gratissium*.

Anti-implantation agents:

1. Ficus religiosa

Common name: Pipal

Family: Moraceae

Chemical constituents: Phytochemical screening revealed the presence of n-hexadecanoic acid, 9, 12-octadecadienoic acid, 9, 12, 15-octadecatrienoic acid and butyl 9, 12, 15-octadecatrienoate ⁷⁵.

Activities:

i. It dereased the thickness of surface epithelium, diameter of uterine glands, diameter of gland cells and thickness of the layer of myometrium ⁷⁵.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Type of extract used: An Extract of *Ficus religiosa* fruit was used for anti-implantation effect 75

Animal models: Goat ⁷⁵ was utilised for finding the anti-implantation activity of *Ficus religiosa*.

2. Calotropis procera:

Common name: Sodom apple, aak

Family: Asclipiadaceae

Chemical constituents: It contains alkaloids, flavonoids, tannins, saponins and cardiac glycosides ⁹⁴.

Activities: (i) It possessed strong anti-implantation activity ⁹⁵.

Type of extract used: Ethanolic extract of *Calotropis procera* roots was used for anti-implantation effect ⁹⁵.

Animal models: Rats ⁹⁵ were used to explore the female anti-fertility potential of *Calotropis procera*.

3. Terminalia belerica:

Common name: Baheda

Family: Combretaceae

Chemical constituents: It showed the presence of phytosterols, carbohydrates, flavonoids, phenolic compounds and tannins ⁹⁶.

Activities:

ii. It caused loss of implantation due to antizygotic, blastocytotoxic or anti-implantation activity of *Terminalia belerica* ⁹⁷.

Type of extract used: Ethanolic extract of Terminalia belerica bark was used for anti-implantation activity ⁹⁸.

Animal models: Rats ⁹⁸ were used for checking the anti-implantation activity of *Terminalia belerica*.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

4. Physalis alkekengi:

Common name: Bladder cherry, kakanaj

Family: Solanaceae

Chemical constituents: It contains tannins, saponins, alkaloids, flavonoids and glycosides ⁹⁹.

Activities:

i. It was act as antagonist of female sexual hormones i.e. estrogen and progesterone which involves in the maintenance of implanted embryo ⁹⁷.

Type of extract used: Extract of *Physalis alkekengi* plant was given for inhibition of implantation ⁹⁷.

Animal models: Rats ⁹⁷ were used for investigating the anti-implantation activity of *Physalis alkekengi*.

5. Leonotis ocymifolia:

Common name: Sun-Bird flower

Family: Lamiaceae

Chemical constituents: It contains phenols, flavonoids, alkaloids, saponins, glycosides and tannins 100.

Activities:

i. It reduced the number of implants significantly ¹⁰¹.

Type of extract used: Ethanolic extract of *Leonotis ocymifolia* leaves were used ¹⁰¹.

Animal models: Rats ¹⁰¹ were used for finding anti-implantation activity of *Leonotis ocymifolia*.

6. Alianthus excelsa

Common name: Indian tree of heaven

Family: Simaroubaceae

Chemical constituents: It contains quassinoids, glaucarubinone, alianthin, β -sitisterol and malanthin 102, 103, 1-p-deoxy-13-formylalianthiol and alianthin 104, 105, excelsin, 13, 18-dehydroexcelsin, glaucarubin, glaucarbol, 13, 18-dehydro 15-isovalerate and trihydroxy triucal 7-ene 106, 107, 108, 109, 110, 111

Activities:

i. It possessed strong anti-implantation activity ¹¹².

ii. It increased uterine weight when administered with ethinyl estradiol indicating its significant anti-estrogenic activity 112.

Type of extract used: Hydroalcoholic extract of *Alianthus excelas* stem bark was given for its anti-implantation activity ¹¹².

Animal models: Rats ¹¹² were used for evaluating the anti-implantation activity of *Alianthus excelsa*.

7. Atrabotrys odoratissimus:

Common name: Nag champa

Family: Annonaceae

Chemical constituents: It contains hydroxy-9-methoxypeterocarpan, nonacosanoic acid 2', 3'-dihydroxypropyl ester, pentacosanoic acid 2', 3'-dihydroxypropyl ester and docosanoic ¹¹³.

Activities:

It possessed strong anti-implantation activity and also produced the disturbances in the duration of the various stages of estrous cycle ¹¹⁴.

It reduced the number of implantation sites¹¹⁴.

Type of extract used: Benzene, ethanol and water extract of *Atrabotrys odoratissimus* leaves were used for causing infertility in females via anti-implantation activity ¹¹⁴.

Animal models: Rats ¹¹⁴ were used for evaluating the anti-implantation activity of *Atrabotrys odoratissimus*.

Abortifacient agents:

1. Coriandrum sativum:

Common name: Dhania

Family: Apiaceae

Chemical constituents: It showed the presence of carbohydrates, proteins, phenolic compounds, tannins and flavonoids¹¹⁵.

Activities: (i) It caused significant decrease in progesterone level of serum on day-5 of progesterone indicating abortifacient activity¹¹⁶.

Type of extract used: Aqueous extract of *Coriandrum sativum* seeds was used as abortifacient agent.

Animal model: Rats ¹¹⁶ were used to evaluate the abortifacient activity of *Coriandrum sativum*.

2. Melia azedarach:

Common name: Chinaberry

Family: Meliaceae

Chemical constituents: It contains alkaloids, tannins, saponins, phenols, glycosides, steroids, terpenoids and flavonoids ¹¹⁷.

Activities:

- i. It caused loss of implantation ¹¹⁸.
- ii. It was also significantly reduces myometrial thickness, uterine gland diameter, luminal diameter of uterine glands and luminal epithelium cell height ¹¹⁸.

Type of extract used: Extract of *Melia azedarach* seeds were used as abortifacient ¹¹⁸.

Animal models: Rats ¹¹⁸ were used for finding the abortifacient potential of *Melia azedarach*.

3. Trianthema portulacastrum:

Common name: Horse purslane

Family: Aizoceae

Chemical constituents: Phytochemical analysis showed the presence of alkaloids, flavonoids, saponins, phenolic compounds and terpenoids 119, 120

Activities:

- i. It possessed significant abortifacient activity ¹²⁰.
- ii. It also produced significant increases in uterine weight, diameter of the uterus and thickness of endometrium which indicates its mild anti-estrogenic activity ¹²⁰.

Type of extract used: Extract of *Trianthema portulacastrum* was used for abortifacient activity 120

Animal models: Rats ¹²⁰ were utilised for finding the abortifacient activity of *Trianthema portulacastrum*.

4. Balantis roxburghii:

Common name: Desert date

Family: Zygophyllaceae

Chemical constituents: It contains alkaloids, saponins, tannins, flavonoids, phenolic compounds, gum and mucilage ^{121, 122}.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

Activities:

It showed significant abortifacient activity ¹²¹.

It was also significantly increases the uterine weight diameter of uterus, thickness of endometrium and height of endometrial epithelium 121

Type of extract used: Ethanolic extract of *Balanties roxburghii* fruits was given for abortion ¹²¹.

Animal models: Rats ¹²¹ were used for evaluating the abortifacient activity of *Balanties roxburghii*.

5. Cannabis sativa

Common name: Hemp, bhang

Family: Cannabinaceae

Chemical constituents: It showed the presence of flavonoids, phenolic compounds, alkaloids, steroids, saponins, terpenoids, tannins and reducing sugars^{123, 124}.

Activities:

- i. It was possessed strong abortifacient activity. It produced significant decrease in ovarian and uterine weight whereas non-significant increase in body weight ¹²³.
- ii. It also caused slight increment in serum progesterone level and decrement in serum estrogen level ¹²³.
- iii. The level of LH and FSH was found to be significantly reduced after administration of extract ¹²³.

Type of extract used: Alcoholic extract of *Cannabis sativa* leaves was used for abortifacient effect ¹²³.

Animal models: Rats ¹²² were used for finding abortifacient potential of *Cannabis sativa*.

CONCLUSION: This review summarized scientific proven literature about phytochemical constituents, antifertility activities and type of extract used of various herbal medicinal plants for

both males and females which being traditionally used. The present review also covered animal models used to explore the antifertility activity of the above mentioned plants. These herbal medicinal plants act as antifertility agents via various mechanisms in both males and females. Future research is also required to make preparation of these herbal plants in scientific manner to make them safe and effective.

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REFERENCES:

- Gupta RS and Sharma R: A review on medicinal plants exhibiting antifertility activity in males. Natural product radiance 2006; 5:389-410.
- Mamatha A, Elumalai A, Eswaraiah MC, Mathangi N: An updated review on antifertility plants-2012. International Journal of Pharmacotherapy 2012; 2:4-6.
- Thakur DS, Kumar P, Kujur A, Kumar P and Kumar R: Contribution of Male Contraception in World Population. Journal of Pharmaceutical Sciences and Research 2010; 1:384-393.
- Bhowmik D, Umadevi M, and Kumar PKS and Duraivel S: Medicinal Plants with Potential Antifertility Activity. Journal of Medicinal Plants Studies 2013; 1:26-33.
- Allag IS and Rangari K: Extragenomic action of steroids on spermatozoa: Prospects for regulation of fertility. Health Population 2002; 25:38-44.
- Agrawal SS, Kumar A, Gullaiya S, Dubey V, Nagar A, Tiwari P, Dhar P and Singh V: Antifertility activity of methanolic bark extracts of Aegle marmelos (l.) in male wistar rats. DARU Journal of Pharmaceutical Sciences 2012, 20:94.
- Saravanan K, Priya G and Renuka C: Medicinal plants with potential antifertility activity- A review of sixteen years of herbal medicine research (1994-2010). International Journal of PharmTech Research 2012; 4:481-494.
- 8. Hadley MA, Lin YC and Dym M: Effect of gossypol on reproductive system of male rats. Journal of Andrology 1981; 2:190-199.
- Taitzoglou IA, Tsantarliotou M, Koretas D and Kokolis NA: Gossypol induced inhibition of plasminogen activator activity in humans and ovine acrosomal extract. Andrologia 1999; 31: 355-359.
- Zavos PM and Zavos ZPN: The inhibitory effect of gossypol on human sperm motility characteristics: Possisible modes of reversibility of those effects. Tohoku Journal of Experimental Medicine 1996; 179:167-175.
- 11. Nair IN and Bhigwade DA: Effect of gossypol on reproductive axis: Ultrastructural and biochemical studies. Indian Journal of Experimental Biology 1990; 28:724-732.
- 12. Bai J and Shi Y: Inibition of t-type Ca²⁺ in mouse spermatogenic cells by gossypol, an antifertility compound. European Journal of Pharmacology 2002; 440:1-6.
- 13. Yuan YY and Shi QX: Inhibition of hamster sperm acrosomal enzymes by gossypol is closely associated with

the decrease in fertilization capacity. Contraception 2000; 62:203-209.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

- 14. Chaddha S, Sanyal SN and Kanwar U: Reversibility of the effects of gossypol acid, an antispermatogenic/antifertility agent on the intestinal structure and functions of male albino rats. Research in Experimental Medicine 1989; 189:205-219.
- 15. Ye WS, Dan L, Guo Y, Qian XJ, Ying J and Xue SP: The antifertility effect of gossypol plus testosterone and estrogen. Yao Xue Xue Bao 1996; 31:313-315.
- Lohiya NK and Goyal RB: Antifertility investigations on the crude chloroform extract of *Carica papaya* Linn in male albino rats. Indian Journal of Experimental Biology, 1992; 30: 1051-1055.
- 17. Lohiya NK, Manivannan B, Mishra PK, Pathak N, Sriram S, Bhande SS and Panneerdoss S: Chloroform extract of *Carica papaya* seeds induces long term reversible azoospermia in langur monkey. Asian Journal of Andrology 2002; 4:17-26.
- 18. Pathak N, Miashra PK, Manivannan B and Lohiya NK: Sterlity due to inhibition of sperm motility by oral administration of benzene chromatographic fraction of the chloroform extract of the seeds of *Carica papaya* in rats. Phytomedicine 2000; 7:325-333.
- 19. Udoh P and Kehinde A: Studies on antifertility effect of pawpaw seeds (*Carica papaya*) on the gonads of male albino rats. Phytotherapeutic Research 1999; 13:226-228.
- 20. Verma RJ and Chinoy NJ: Effect of papaya seed extract on the microenvironment of cauda epididymis. Asian Journal of Andrology 2001; 3:143-146.
- 21. Verma RJ and Chinoy NJ: Effect of papaya seeds extract on the contractile response of cauda epididymal tubules. Asian Journal of Andrology 2002; 4:77-78.
- 22. Manivannan B, Mishra PK, Pathak N, Sriram S, Bhande SS, Panneerdoss S and Lohiya NK: Ultrastructural changes in the testis and epididymis of rats following treatment with the benzene chromatographic extract of the seeds *Carica papaya*. Phtyotherapy Research 2004; 18:285-289.
- Soni D, Gupta A, Solanki R and Jana GK: Antiimplantation and pregnancy interruption activity of japakusuma (*Hibiscus rosa-sinensis*) in albino rats. International Journal Research in Ayurveda and Pharmacy 2013; 4: 493-514.
- 24. Gupta I, Tank R and Dixit VP: Fertility regulation in males: Effect of *Hibiscus rosa-sinensis* and Malvaviscous flower extract on male albino rats. Proceedings of the National Academy of Sciences of India 1985; 55: 262-267.
- 25. Singhvi MM and Lall SB: Effect of *Hibiscus rosa-sinensis* on testicular dehydrogenase of *Rhino Poma Kinneari* Wroughton. Current Science 1981; 50:360-362.
- 26. Reddy CM, Murthy DRK and Patil SB: Antispermatogenic and androgenic activities of various extract of *Hibiscus rosa-sinensis* in albino mice. Indian Journal of Experimental Biology 1997; 35:1170-1174.
- Xue C and Wang ZT. Chemical constituents from roots of Andrographis paniculata. Yao Xue Xue Bao 2011; 46: 317-321.
- Akbarsha MA, Manivannan B, Hamid KS and Vijayan B: Antifertility effects of *Andrographis paniculata* (Nees) in male albino rats. Indian Journal of Experimental Biology 1990; 28:421-426.
- Akbarsha MA and Murugaian P: Aspects of male reproductive toxicity/male antifertility property of andrographilode in albino rats: Effects on the testis and cauda epididymidal spermatozoa. Phytotherapy Research 2000; 14:432-435.

- 30. Lu QX, Shen XM, Chen K, Chen XM and Xue SP: Effects of glycosides of *Trypterigium wilfordii* Hook on the the reproductive system and major organs of male rats. Acta Academiae Medicinae Sinicae 1990; 12:203-207.
- Lue Y, Sinha HAP, Wang C, Leung A, Baravarian S, Sangsawan R, Chaichana S and Swerdloff RS: Triptolide: A potential male contraceptive. Journal of Andrology 1998; 19: 479-486.
- Hikkim AP, Lue YH, Wang C, Reutrakul V, Sangsawan R and Swerdloff RS: Post testicular antifertility action of triptolide in the male rat: Evidence for severe impairment of cauda epididymal sperm ultra structure. Journal of Andrology 2000; 21:431-437.
- 33. Zheng ZR, Fang JL and Gao HZ: Effects of total glycosides of *tripterygium wilfordii* on reproductive organs of experimental animals III dynamic observation on reproductive organs and fertility in mice. Acta Academiae Medicinae Sinicae 1986; 8:19-23.
- Reddy NM and Reddy R: Solanum Xanthocarpum chemical constituents and medicinal properties: A review. Scholar Academic Journal of Pharmacy 2014: 3:146-149.
- 35. Kanwar U, Batia A, Ranga A and Sangal SN: Effect of Solasodine on morphology, motility and glycolytic enzymes of buffalo bull spermatozoa. Indian Journal of Experimental Biology 1988; 26:941-944.
- 36. Mali PC, Chaturvedi M and Dixit VP: Antispermatogenic activity of *Solanum xanthocarpus* schrad and wandl. Root (50% Et-OH extract) in rats. Journal of Phytological Research 1996; 9:13-17.
- 37. Rao MV: Effects of Alcoholic extracts of *Solanum xanthocarpum* in adult male rats. Indian Journal of Experimental Biology 1988; 26:95-98.
- 38. Asadulla S, Ramandang, Rajesekharan: Pharmacognosy of Embelia ribes burm F. International Journal of Research in Pharmacy and chemistry 2011; 1236-1251.
- 39. Agarwal SS, Chauhan S and Mathur R: Antifertility effects of embelin in male rats. Andrologia 1986; 2:125-131.
- 40. Pundare TV, Kholkute SD, Gurjar A, Joshi UM, Dittatreya MB, Shith AR, Swamy XR, Jayaraman S and Munshi R: Semen analysis and hormonal levels in bonnet macaques administered *Embelia ribes* berries, an indigenous plant having contraceptive activity. Indian Journal of Experimental Biology 1979; 17:935-936.
- 41. Gupta S, Sanyal SN and Kanwar U: Antispermatogenic effect of embellin, a potent benzoquinone on male albino rats *in vivo* and *in vitro*. Contraception 1989; 39:307-320.
- 42. Deng L, Tang L, Guan H, He L, Zhang J, Liu J, Hao X, Zhang Y. A new hasubanan alkaloid from stephania hernandifolia. Chinese Herbal Medicines 2014; 6: 70-72.
- 43. Ghosh D, Jona D and Debnath JM: Effects of leaf extract of *Stephonia hernandifolia* on testicular gametogenesis and endogenesis in albino rats: A dose dependent response study. Contraception 2002; 65: 379-384.
- 44. Yadav PD, Bharadwaj NSP, Yedukondalu M, Methushala CH and Ravi Kumar A. Phytochemical evaluation of Nyctanthes arbortristis, Nerium oleander and Cantharanthus roseus. Indian Journal of Research in Pharmacy and Biotechnology 2013; 1:333-338.
- 45. Averal HI, Stanley A, Murugaian P, Polanisamy A and Akborsha MA: Specific effect of vincristine on epididymis. Indian Journal of Experimental Biology 1996; 34:53-56.
- Murugavel T, Ruknudin A, Thangauelu S and Akbarsha MA: Antifertility effect of Vinca rosea (linn.) leaf extract on male albino mice. A sperm parametric study. Current Science 1989; 58:1102-1103.

- Murugavel T and Akbarsha MA. Antispermatogenic effect of *Vinca rosea* linn. Indian Journal of Experimental Biology 1991; 29:810-812.
- 48. Jayaweera DMA: Leguminosea, In Medicinal plants (indigenous and exotic) used in Ceylon part 3, the national science council of Srilanka, Colombo, 1981; 135-260.
- Raji U and Bolarinwa AF: Antifertility activity of *Quassia amosa* in male rats in vivo study. Life Science 1997; 61:1067-1074.
- 50. Kulshrestha SS and Mathur RS: Effect of steroidal fraction of seeds of *Abrus precatorius* linn on rat testis. Indian Journal of Experimental Biology 1990; 28:752-756.
- 51. Bajaj A, Mathur RS, Wadhwa M and Bahel S: Effect of steroidal fraction of Abrus precatorius on testis of albino rats. Geobios 1981; 8:29-31.
- 52. Dixit VP, Sinha R and Gupta I: Inhibition of sperm production and sperm dynamics in *Abrus precatorius* treated males. The Indian Zoologist 1987; 11:115-118.
- 53. Sinha R, Gupta I, Tark R and Dixit VP: Antispermatogenic activity of *Abrus precatorius* linn. seed extract in Langur monkey (Presbytis entellus entellus). In National symposium on the use of primates in biochemical research Jaipur, India, 1986; 53.
- 54. Shaikh PD, Manivannan B, Pathar KM, Kasturi M and Ahmed RN: Antispermatic activity of *Azadirachta indica* leaves in albino rats. Current Science 1993; 64: 688-689.
- 55. Kasturi M, Manivannan B, Ahmed RN, Shaikh PD and Pathar KM: Changes in epididymal structure and function of albino rat treated with *Azadirachta indica* leaves. Indian Journal of Experimental Biology 1995; 33:725-729.
- 56. Joshi AR, Ahmed RN, Pathar KM and Manivannan B: Effect of Azadirachta indica leaves on testis and its recovery in albino rats. Indian Journal of Experimental Biology 1996; 34:1091-1094.
- 57. Aladakatti RH and Ahmed RN: Effect of *Azadirachta indica* leaves on rat spermatozoa. Indian Journal of Experimental Biology 1999; 37:1251-1254.
- 58. Khillare B and Shrivastava TG: Spermicidal activity of *Azadirachta indica* (Neem) leaf extract. Contraception 2003; 68:225-229.
- 59. Sharma JD, Jha RK, Gupta I and Jain P: Anti-androgenic properties of neem seed oil *Azadirachta indica* in rat and rabbit. Ancient Science of Life 1987; 1:30-38.
- 60. Deshpandey VP, Mendulkar KN and Sadre NL: Antifertility activity of *Azadirachta indica* in mice. Journal of Postgraduate of Medicine of Bombays 1980; 26:167-170.
- 61. Choudhary CN, Singh JN, Verma SK and Singh BP: Antifertility effect of leaf extract of some plants in male rats. Indian Journal of Experimental Biology 1991; 28:714-716.
- 62. Khare AK, Sharma MK and Tiwari JP: Antifertility activity of Neem oil in rabbits and rats. Probe 1984; 23:90-94.
- 63. Riar SS, Bardhan J, Thomas P, Kain AK and Parshad R.:Mechanism of antifertility action of neem oil. Indian Journal of Medical Research 1988; 88:339-342.
- Sinha KC, Riar SS, Bardhan J, Thomas P, Jain AK and Jain RK: Antiimplantation effect of neem oil. Indian Journal of Medical Research 1984; 80:708-710.
- 65. Maity P, Hansda D, Bandyopadhyay U and Mishra DK: Biological activities of crude extracts and chemical constituents of Bael, *Aegle marmelos* (L.) Corr. Indian Journal of Experimental Biology 2009; 47:849-861.
- 66. Uttam KD, Rajkumar M, Debasis J and Debidas G: Effect of aqueous extract of leaf of *Aegle marmelos* on testicular activities in rats. Iranian Journal of Pharmacology and Therapeutics 2006; 5:21-25.

- 67. Chauhan A, Agarwal M, Kushwaha S and Mutreja A: Suppression of fertility in male albino rats following the administration of 50% ethanolic extract of *Aegle marmelos*. Contraception 2007; 6:474-481.
- 68. Chauhan A and Agarwal M: Reversible changes in the antifertility induced by *Aegle marmelos* in male albino rats. System Biology in Reproductive Medicine 2008, 6:240-246.
- 69. Chauhan A and Agarwal M: Assessment of the contraceptive efficacy of the aqueous extract of *Aegle marmelos* Corr. leaves in male albino rats. Human Fertility (Cambridge, England) 2009; 2:107-118.
- Al-Sanabra OMA, Qunaibi EA, Aburjai TA, Al-Qaadan FA, Shomaf MF and Disi AM: Antifertility Activity of Ethanolic Seed Extract of Celery (*Apium graveolensL.*) in Male Albino Rats. Jordan Journal of Pharmaceutical Sciences 2013; 6:30-38.
- 71. Shivalingappa H, Satyanarayan ND and Purohit MG: Antiimplantation and pregnancy interruption efficacy of *Rivea hypocrateriformis* in the rat. Journal of Ethnopharmacology 2001; 74: 245-249.
- 72. Shivalingappa H, Satyanarayan ND, Purohit MG, Sharanabasappa A and Patil SB: Effect of ethanol extract of *Rivea hypocrateriformis* on the estrous cycle of the rat. Journal of Ethnopharmacology 2002; 82:11-17.
- Britto AJ and Gracelin HS: Phytochemical analysis and antibacterial activity of *Momordica charantia* descourt, a known medicinal plant. Journal of Basic and Applied Biology 2011; 5:307-311.
- 74. Ifeanyi AC, Eboetse YO, Ikechukwu DF, Adewale OA, Carmel NC and Olugbenga OA: Effect of *Momordica charantia* on estrous cycle of sprague-dawley rats. Pacific Journal of Medical Sciences 2011; 8: 6.
- 75. Sharma RK, Goyal AK and Bhat RA: Antifertility activity of plant extracts on female reproduction: A review. Biological Sciences 2013; 3:493-514.
- 76. Essiett U and Unung I: Comparative Phytochemical and Physico-chemical Properties of Aspilia africana (Pers) C. D. Adams and Tithoniadiversifolia (Hemsl) A. gray petals as a scientific backing to their tradomedicinal potentials. International Journal of Modern Biology and Medicine 2013; 3:88-100.
- 77. Oyesola TO, Oyesola OA and Okoye CS: Effects of aqueous extract of *Aspilia africana* on reproductive functions of female Wistar rats. Pakistan Journal of Biological Sciences 2010; 13:126-131.
- 78. Dahiya P and Purkayastha S: Phytochemical analysis and antibacterial efficacy of dill seed oil against multi-drug resistant clinical isolates. Asian Journal of Pharmaceutical and Clinical Research 2012; 5:62-64.
- Monsefi M, Ghasemi M and Bahaoddini A: The effects of *Anethum graveolens* on female reproductive system of rats. Phytotherapy Research 2006; 20:865-868.
- Samanta J, Bhattacharya S and Rayat R: Phytochemical investigation and pharmacognostic standardization of *Cissampelos pareira* root. Ancient Science of Life 2012; 31:181-184.
- 81. Ganguly MK, Borthakur M, Devi N and Mahanta R: Antifertility activity of the methanolic leaf extract of *Cissampelos pareira* in female albino mice. Journal of Ethnopharmacology 2007; 111:688-691.
- 82. Deb N, Majumdar P and Ghosh AK: Pharmacognostic and Phytochemical Evaluation of the Rhizomes of *Curcuma longa* Linn, Journal of PharmaSciTech 2013; 2:81-86.
- Ghosh AK, Das AK and Patra KK: Studies on antifertility effect of rhizome of *Curcuma longa* Linn. Asian Journal of Pharmacy and Life Science 2011; 1:349-353.

- 84. Jhade D, Jain S, Jain A and Sharma P: Pharmacognostic Screening, Phytochemical Evaluation and In- Vitro free radical Scavenging Activity of Acacia leucophloea Root. Asian Pacific Journal of Tropical Biomedicine 2012; 1 S501-S505.
- 85. Ahirwar D: Antifertility activity of *Acacia leucophloea*. Der Pharmacia Lettre 2011; 3: 411-413.
- Sindhia VR and Bairwa R: Plant review: Butea monosperma. International Journal of Pharmaceutical and Clinical Research 2010; 2:90-94.
- 87. Sharm SK, Rai G and Vasudeva N: Anti-fertility Investigation of *Butea monosperma* (Lam.) Kuntze Root in Female Albino Mice. Research Journal of Medicinal Plant 2012: 6: 260-266.
- 88. Chaurasia S, Kulkarni GT and Shetty LN: Phytochemical Studies and in vitro Cytotoxicity Screening of *Piper betle* Leaf (PBL) Extract. Middle-East Journal of Scientific Research 2010; 6: 532-536.
- 89. Sharma JD, Sharma L and Yadav P: Antifertility Efficacy of *Piper betle* Linn. (Petiole) on Female Albino Rats. Asian Journal of Experimental Sciences 2007; 21:145-150.
- Pandya DJ, Patel VL, Desai TR, Lunagariya RR, Gajera SD, Mehta AJ: Pharmacognostic and phytochemical evaluation of leaves of *Cassia fistula*. International Journal of Pharmacy & Life Sciences. 2012; 3:14-24.
- 91. Yadav R and Jain GC: Antifertility effect of aqueous extract of seeds of *Cassia fistula* in female rats. Advance in Contraception 1999; 15:293-301.
- Koche DK, Kokate PS, Suradkar SS and Bhadange DG: Preliminary Phytochemitry and Antibacterial Activity of Ethanolic Extract of *Ocimum gratissimum* L. Bioscience Discovery 2012; 3:20-24.
- Sripriya S, Yuvaraj G, Nema RK, Kumar VM and Deccaraman M: Evaluation of Antifertility activity from Stem Part of *Ocimum gratissimum* in Acetone extracts. International Journal of Pharmaceutical and Clinical Research 2011; 3:41-44.
- 94. Mainasara MM, Aliero BL, Aliero AA and Dahiru SS: Phytochemical and Antibacterial Properties of *Calotropis Procera* (Ait) R. Br. (Sodom Apple) Fruit and Bark Extracts. International Journal of Modern Botany 2011; 1:8-11
- 95. Ranab AC, Kamatha and Jagadish V: Preliminary study on antifertility activity of *Calotropis procera* roots in female rats. Fitoterapia. 2002; 73:111-115.
- Manohar VR, Chandrashekar R and Rao SN: Phytochemical analysis of *Terminalia belerica* fruit pulp extracts. World Journal of Pharmacy and Pharmaceutical Sciences 2012; 1:1376-1383.
- 97. Montaserti A, Pourheydar M, Khazaei M and Ghorbani R: Anti-fertility effects of *Physalis alkekengi* alcoholic extract in female rat. Iranian Journal of Reproductive Medicine 2007; 5:13-16.
- Vishwanatha T, Satishagouda S, Patil JS and Patil BS: Antiimplantation activity of *Terminalia bellirica* bark extracts in female albino rats. Biotechnology An Indian Journal 2009; 3:66.
- Estakhr J and Javdan N: Preliminary study of phytochemical screening and antibacterial activity of Physalis alkekengi against Staphylococcus aureus. Pharmacology online 2011; 3: 97-103.
- 100. Oyedemi SO, Yakubu MT and Afolayan AJ: Antidiabetic activities of aqueous leaves extract of *Leonotis leonurus* in streptozotocin induced diabetic rats. Journal of Medicinal Plants Research 2011; 5:119-125.
- 101. Tafesse G, Mekonnen Y, Makonnen E: *In vivo* and *in vitro* anti-fertility and anti-implantation properties of *Leonotis*

- E-ISSN: 0975-8232; P-ISSN: 2320-5148
- ocymifolia in rats. African Journal of Traditional, Complementary and Alternative Medicines 2005; 2:103-112
- 102. Kapoor SK, Ahmed PL, and Zaman A: Chemical constituents of *Ailanthus excelsa*. Phytochemistry 1971; 10: 3333-3335.
- 103. Bhatia N, Sahai M and Khose RL: A chemical study on Ailanthus excels Roxb. Journal of Indian Chemical Society 1985; 62: 75-76.
- 104. Cordell GA, Ogura M and Fransworth NR: Alkaloid constituents of *Ailanthus excelsa*. Lloydia 1978; 41: 166-168.
- 105. Bhatia N, MohanY and Khose RL: Chemical studies on *Ailanthus excel* Roxb. Indian Drugs 1983; 20: 240-242.
- 106. Jain MK: Chemical examination of *Ailanthus excelsa*. Indian Journal of Chemistry 1964; 2B:40-41.
- 107. Khan SA and Shamsuddin KM: Quassinoid from *Ailanthus excelsa*. Indian Journal of Chemistry 1978; B16:1045-1046.
- 108. Ogura M, Cordell GA, Kinghorn AD, Farnsworth NR: Potential anticancer agents. VI. Constituents of Ailanthus excelsa. Lloydia 1978; 40:579-581.
- 109. Khan SA, Zuberi SS and Shamsuddin KM. Isolation and structure of excelsin, new quassionoid from *Ailanthus excelsa*. Indian Journal of Chemistry 1980; 19B:183-184.
- 110. Sherman MM, Borris RP, Ogura M, Cordell A and Farnsworth NR: Trihydroxy Tirucal 7-ene from *Ailanthus excelsa*. Phytochemistry 1980; 19:1499-1501.
- 111. Suroor AK, Sharique SZ and Shamsuddin KM. Isolation and structure of excelsin a new quassinoid from *Ailanthus excelsa*. Indian Journal of Chemistry 1980; B19:183-184.
- 112. Ravichandran V, Suresh B, Sathishkumar MN, Elango K and Srinivasan R. Antifertility activity of hydroalcoholic extract of *Ailanthus excelsa* (Roxb): An ethnomedicines used by tribals of Nilgiris region in Tamilnadu. Journal of Ethnopharmacology 2007; 112: 189-191.
- 113. Singh JP, Singh AK, Singh A and Ranjan R: Chemical constituents of *Atrabotyrys Odoratissimus*. Rasayan Journal of Chemistry 2009; 2:156-158.
- 114. Geetha M, Shankar MB, Mehta RS and Saluja AK. Antifertility activity of Artabotrys odoratissimus Roxb. and

- Couroupita guianensis Aubl. Journal of Natural Remedies 2005; 5: 121-125.
- 115. Shalini P and Smitapadma M: Screening of Antioxidant Activity of *Coriandrum sativum*. Paripex. Indian Journal of Research 2013; 3:284-285.
- 116. Al-Said MS, Al-Khamis KI, Islam MW, Parmar NS, Tariq M and Ageel AM: Post-coital antifertility activity of the seeds of *Coriandrum sativum* in rats. Journal of Ethnopharmacology 1987; 21:165-173.
- 117. Ahmed MF, Rao AS, Ahemad SR and Ibrahim M: Phytochemical studies and antioxidant activity of *Melia azedarach* Linn leaves by DPPH scavenging assay. International Journal of Pharmaceutical Applications 2012; 3:271-276.
- 118. Mandal R and Dhaliwal PK: Antifertility effect of *Melia azedarach* Linn. (dharek) seed extract in female albino rats. Indian Journal of Experimental Biology 2007; 45:853-860.
- 119. Sunder AS, Rajyalakshmi G, Bharath A and Rajeshwar Y: Antihyperglycemic activity of *Trianthema portulacastrum* plant in streptozotocin induced diabetic rats. Pharmacology online 2009; 1:1006-1011.
- 120. Pare S, Zade V and Dabhadkar D: Evaluation of potential antifertility activity of plant *Trianthema portulacastrum* in female albino rat. Int.J.A.PS.BMS. 2013; 2:007-011.
- 121. Padmashali B, Vaidya VP, Vagdevi HM and Satyanarayana ND: Antifertility efficacy of the plant *Balanites roxburghii* (balanitaceae) in female rats. Indian Journal of Pharmaceutical Sciences 2006; 68:347-351.
- 122. Singh V, Patel JR, Gaur K, Tyagi LK and Kori ML: *In vitro* antioxidant activity and phytochemical analysis of stem bark of *Balanites roxburghii* planch. Advances in Biological Research 2009; 3: 242-246.
- 123. Zade V, Wikhe M, Dabhadkar D, Dawada S and Patil U: Antifertility efficacy of *Cannabis sativa* leaves on female albino rats. International Journal of Science Inventions Today 2013; 2:107-117.
- 124. Kumar V, Tripathi MK and Kohli S: Antibacterial activity of *Cannabis sativa* against some pathogens isolated from burns of patient. International Journal of Phytomedicines and Related Industries 2011; 3:243-247.

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