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A REVIEW ON HEPATOPROTECTIVE ACTIVITY

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ABSTRACT: Liver, the largest gland functioning as an organ of storage, manufacturing and biotransformation is a vulnerable target for injury. Chronic alcohol consumption, exposure to toxic chemicals and certain drugs like paracetamol, tetracycline, antitubercular drugs, chemotherapeutic agents, NSAIDS, damage the liver cells (hepatocytes) in long run. Drug induced liver injury is a major health problem, the manifestations of which are highly variable, ranging from asymptomatic elevation of liver enzymes to fulminant liver failure. Modern medicine has provided us many drugs that alleviate liver diseases but compared to it herbal medicine is preferred because the latter is cost effective and considered to be a safe approach for treatment with minimal side effects. Through the decades many scientists, researchers have reported hepatoprotective activity of many medicinal plants mostly in the form of plant extracts. The present review is aimed at compiling data on different medicinal plants with hepatoprotective activity on various models of hepatotoxicity.

INTRODUCTION: The Liver is the largest gland in a human body, situated in the right side of upper abdominal cavity. The cells of the liver called hepatocytes plays vital functions like;

1. Synthesis of proteins, bile,
2. Stores glycogen, vitamins, iron,
3. Metabolises toxic chemicals and drugs.

Drug metabolism/biotransformation is a process of detoxification in which a substance is chemically modified into a less toxic form under the influence of enzymatic system.

The capacity of the liver to carry out the several oxidative metabolisms is associated with the high cellular content of cytochrome P450¹. As liver being the central organ of metabolism it is highly vulnerable target for injury from drugs and chemicals, the manifestations of which are highly variable, ranging from asymptomatic elevation of liver enzymes to fulminant hepatic failure².

Herbal medicine: Modern medicines have a little to offer for alleviation of hepatic diseases and it is chiefly the plant based preparations which are employed for their treatment of liver disorders³. Herbal medicines are the most lucrative form of traditional medicine on which about 80% of the population depends (WHO traditional medicine facts sheet no 134. Dec 2008). These have gained importance and popularity in recent years because they are easily available, safe, efficient and cost effective.

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In India, more than 87 plants are used in 33 patented & proprietary multi-ingredient plant formulations & about 40 polyherbal commercial formulations reputed to have hepatoprotective action are being used. It has been reported that about 160 phytoconstituents from 101 medicinal plants have hepatoprotective activity⁴.

Many herbs have been used to alleviate various liver diseases, of which the most popular ones include Silymarin from *Silybum marianum*, *andrographolide* and *neoandrographolide* from *Andrographis paniculata*, curcumin from *Curcuma longa*, picroside and kutkoside from *Picrorrhiza kurroa*, phyllanthin and hypophyllanthin from *Phyllanthus niruri*, glycyrrhizin from *Glycyrrhiza glabra*, etc.⁵.

Hepatotoxins and their mechanism of hepatotoxicity: In this review, the authors mainly concentrated on hepatotoxins like Carbon tetrachloride, Paracetamol, D-Galactosamine and Thioacetamide.

1. Carbon tetrachloride: The hepatotoxicity of CCl₄ is due to the formation of the highly reactive trichloromethyl free radical in the body which attacks the polyunsaturated fatty acids of the membrane of endoplasmic reticulum. Carbon tetrachloride poisoning leads rapidly to cessation of movement of large quantities of triglycerides from the liver to the plasma leading to fatty liver⁶.

If the damage is severe it leads to an abnormal increase in liver enzymes followed by hepatocellular necrosis. There is an influx of monocytes into the liver during acute and chronic CCl₄ induced hepatotoxicity causing an increase of Reactive Oxygen Species (ROS) production and a rise in Kupffer cell leukotriene production in the liver leading to imbalance between cytoprotective and cytotoxic prostanoids⁷.

2. Paracetamol: Paracetamol is metabolically activated by cytochrome P450 to a reactive metabolite that covalently binds to protein⁸. The reactive metabolite responsible for hepatotoxicity is N-acetyl-p-benzoquinone-imine which reacts with N-acetyl cysteine⁹.

Although considered safe at therapeutic doses, in overdose, it produces a centrilobular hepatic necrosis that can be fatal¹⁰. Various mechanisms leading to paracetamol toxicity includes

- a. Increased formation of superoxide anions which cause lipid peroxidation (oxidative stress) via hydrogen peroxide formation¹¹.
- b. Decreased glutathione concentrations in centrilobular cells¹².

3. D-galactosamine: Galactosamine administration induces an inflammatory response in liver that biochemically and histologically resembles viral hepatitis¹³. A single administration causes hepatocellular necrosis and fatty liver¹⁴.

It causes appearance of specific lesions in liver cells, characterized by inhibition of nuclear RNA and protein synthesis¹⁵.

4. Thioacetamide: Thioacetamide, originally used as a fungicide is a potent hepatotoxic and is bioactivated by CYP450 and/or flavin-containing monooxygenase (FMO) systems to sulfine (sulfoxide) and sulfene (sulfone) metabolites, which causes centrilobular necrosis^{16, 17}.

This metabolite causes liver fibrosis. Thioacetamide interferes with the movement of RNA from the nucleus to cytoplasm which may cause membrane injury¹⁸.

In the present study, the authors have reviewed the hepatoprotective activity of medicinal plants evaluated in carbon tetrachloride, paracetamol, D-galactosamine and thioacetamide induced hepatotoxicity.

TABLE 1: HEPATOPROTECTIVE ACTIVITY OF MEDICINAL PLANTS IN CARBON TETRACHLORIDE INDUCED HEPATOTOXICITY

Plant	Family	Part used	Type of extract	Test dose	Carbon tetrachloride dose
<i>Casuarina equisetifolia</i> , ¹⁹	Casuarinaceae	Leaf & bark	Methanol	i.p 250mg/kg, 500mg/kg	i.p 3ml/kg
<i>Cajanus cajan</i> , ¹⁹	Papilionaceae	Whole plant	Methanol	i.p 250mg/kg, 500mg/kg	i.p 3ml/kg
<i>Glycosmis pentaphylla</i> , ¹⁹	Rutaceae	Leaf, bark	Methanol	i.p 250mg/kg, 500mg/kg	i.p 3ml/kg
<i>Bixa orellana</i> , ¹⁹	Bixaceae	Seed	Methanol	i.p 250mg/kg, 500mg/kg	i.p 3ml/kg
<i>Physalis minima</i> , ¹⁹	Solanaceae	Whole plant	Methanol	i.p 250mg/kg, 500mg/kg	i.p 3ml/kg
<i>Argemone Mexicana</i> , ¹⁹	Papaveraceae	Leaf & flower	Methanol	i.p 250mg/kg, 500mg/kg	i.p 3ml/kg
<i>Caesalpinia bonduc</i> , ¹⁹	Caesalpiniaceae	Leaf & bark	Methanol	i.p 250mg/kg, 500mg/kg	i.p 3ml/kg
<i>Carthamus tinctorius</i> , ²⁰	Compositae	Flower	Methanol	p.o 200mg/kg	p.o 1ml/kg
<i>Ardisia solanacea</i> , ²¹	Myrsinaceae	Leaves	Alcohol	p.o 100mg/kg, 200mg/kg	s.c 2ml/kg
<i>Delonix regia</i> , ²²	Caesalpiniaceae	Aerial parts	Methanol	p.o 400mg/kg	p.o 2ml/kg
<i>Aphanamixis polystachya</i> , ²³	Meliaceae	Leaves	Ethanol	p.o 50mg/kg	i.p 2ml/kg
<i>Coriandrum sativum</i> , ²⁴	Apiaceae	Whole plant	Pulverised plant powder	p.o 5%, 10%, 15%	i.p 1ml/kg
<i>Solanum pubescens</i> , ²⁵	Solanaceae	Whole plant	Ethanol	p.o 500mg/kg	i.p 1ml/kg
<i>Plumbago zeylanica</i> , ²⁶	Plumbaginaceae	Aerial parts	Methanol	p.o 35mg/kg, 70mg/kg	i.p 0.1ml/kg
<i>Cardiospermum helicacabum</i> , ²⁷	Sapindaceae	Stem	Petroleum ether, methanol, water, chloroform, ethylacetate	p.o 200mg/kg, 400mg/kg	p.o 1.5ml/kg
<i>Luffa acutangula</i> , ²⁸	Cucurbitaceae	Leaves	Ethanol	p.o 200mg/kg, 400mg/kg, 600mg/kg	p.o 1ml/kg
<i>Epaltes divaricata</i> , ²⁹	Compositae	Whole plant	Aqueous	p.o 0.9g/kg	i.p 0.5ml/kg
<i>Tagetes erecta</i> , ³⁰	Asteraceae	Flowers	Ethanol	p.o 400mg/kg	p.o 1875g/kg
<i>Zizphus rotundifolia</i> , ³¹	Rhamnaceae	Leaves	Ethanol	p.o 650mg/kg	p.o 1.25ml/kg
<i>Millettia aboensis</i> , ³²	Fabaceae	Roots	Ethanol, aqueous	p.o 215mg/kg, 431mg/kg	i.p 0.7ml/kg
<i>Ficus carica</i> , ³³	Moraceae	Leaves	Methanol Hexane, methanol, chloroform, ethylacetate, acetone	p.o 500mg/kg	p.o 1.5ml/kg
<i>Alchornea cordifolia</i> , ³⁴	Euphorbiaceae	Leaves	Petroleum ether, chloroform, alcohol, aqueous	p.o 300mg/kg	i.p 1.25ml/kg
<i>Morus alba</i> , ³⁵	Moraceae	Leaves	Petroleum ether, chloroform, alcohol, aqueous	p.o 125mg/kg, 150mg/kg, 175mg/kg, 250mg/kg, 300mg/kg, 350mg/kg	s.c 2ml/kg
<i>Leucophyllum frutescens</i> , ³⁶	Scrophulariaceae	Aerial parts	Methanol	p.o 100mg/kg, 200mg/kg	p.o 2ml/kg
<i>Carissa carandas</i> , ³⁷	Apocynaceae	Roots	Ethanol	p.o 100mg/kg, 200mg/kg,	i.p 0.7ml/kg

<i>Sesamum indicum</i> , ³⁸	Pedaliaceae	Seeds	Methanol	400mg/kg p.o 200mg/kg, 400mg/kg, 800mg/kg	s.c 2ml/kg
<i>Flacourtie indica</i> , ³⁹	Flacourtiaceae	Leaves	Aqueous	p.o 250mg/kg, 500mg/kg	p.o 1.5ml/kg
<i>Hippophae rhamnoides</i> , ⁴⁰	Elaeagnaceae	Leaves	Aqueous	p.o 100mg/kg, 400mg/kg	p.o 1ml/kg
<i>Apium graveolens</i> , ⁴¹	Apiaceae	Seeds	Acetone, methanol, petroleum ether	p.o 250mg/kg	p.o 1.5ml/kg
<i>Croton oblongifolius</i> , ⁴¹	Euphorbiaceae	Whole plant	Acetone, methanol, petroleum ether Water,	p.o 200mg/kg	p.o 1.5ml/kg
<i>Hypericum japonicum</i> , ⁴²	Hypericaceae	Whole plant	petroleum ether, chloroform	p.o 0.5, 1.5, 4.5g/kg	i.p 10ml/kg
<i>Cinnamomum zeylanicum</i> , ⁴³	Lauraceae	Bark	Ethanol	p.o 0.05g/kg, 0.005g/kg, 0.01g/kg, 0.1g/kg	i.p 0.5ml/kg
<i>Launea intybacea</i> , ⁴⁴	Asteraceae	Aerial parts	Aqueous	p.o 200mg/kg	p.o 3ml/kg
<i>Mimosa pudica</i> , ⁴⁵	Mimosaceae	Leaves	Methanol	p.o 200mg/kg	i.p 1.25ml/kg
<i>Polygala javana</i> , ⁴⁶	Polygalaceae	Whole plant	Ethanol	p.o 100mg/kg, 200mg/kg	i.p 2.5ml/kg
<i>Marsilea minuta</i> , ⁴⁷	Marsileaceae	Whole plant	Methanol, toluene, aqueous, n- butyl alcohol	p.o 50mg/kg, 100mg/kg	p.o 1ml/kg
<i>Ficus bengalensis</i> , ⁴⁸	Moraceae	Leaves	Ethanol	p.o 100mg/kg, 200mg/kg, 400mg/kg	i.p 2ml/kg
<i>Chenopodium album</i> , ⁴⁹	Chenopodiaceae	Aerial parts	Ethyl acetate, methanol	p.o 300mg/kg	i.p 1ml/kg
<i>Psidium guajava</i> , ⁵⁰	Myrtaceae	Leaves	Methanol	p.o 250mg/kg, 500mg/kg	i.p 1ml/kg
<i>Luffa acutangula</i> , ⁵¹	Cucurbitaceae	Fruits	Hydroalcohol	p.o 100mg/kg, 200mg/kg	i.p 1ml/kg
<i>Rhododendron arboreum</i> , ⁵²	Ericaceae	Leaves	Ethanol	p.o 40mg/kg, 60mg/kg, 100mg/kg	s.c 1ml/kg
<i>Diteracanthus patulus</i> , ⁵³	Acanthaceae	Leaves	Methanol	p.o 250mg/kg, 500mg/kg	i.p 1ml/kg
<i>Cuscuta reflexa</i> , ⁵⁴	Cuscutaceae	Aerial parts	Ethanol	p.o 1g/kg	i.p 0.1ml/kg
<i>Crassocephalum crepidioides</i> , ⁵⁵	Asteraceae	Whole plant	Aqueous	i.p 5ml/kg	s.c 2ml/kg
<i>Glycyrrhiza glabra</i> , ⁵⁶	Fabaceae	Root	Crude powder	p.o 1000mg/kg	s.c 3ml/kg
<i>Gundelia tourenfortii</i> , ⁵⁷	Asteraceae	Footstalks	Hydroalcohol	i.p 100mg/kg, 300mg/kg	i.p 3ml/kg
<i>Coptidis rhizome</i> , ⁵⁸	Ranunculaceae	Whole plant	Aqueous	p.o 400mg/kg, 600mg/kg, 800mg/kg	i.p 1ml/kg
<i>Carica papaya</i> , ⁵⁹	Caricaceae	Seeds	Aqueous	p.o 100mg/kg, 200mg/kg, 400mg/kg	i.p 1.5ml/kg
<i>Cichorium intybus</i> , ⁶⁰	Asteraceae	Leaves	70% ethanol	p.o 50mg/kg, 100mg/kg, 200mg/kg	i.p 3ml/kg

<i>Scoparia dulcis</i> , ⁶¹	Scrophulariaceae	Whole plant	Petroleum ether, diethylether, methanol Acetone, methanol, chloroform	p.o 50mg/kg, 200mg/kg, 800mg/kg	i.p 2ml/kg
<i>Indigofera tinctoria</i> , ⁶²	Leguminosae-papilionatae	Whole plant		p.o 500mg/kg	0.5mM perfusion
<i>Solanum trilobactum</i> , ⁶³	Solanaceae	Whole plant	Methanol	p.o 150mg/kg, 200mg/kg, 250mg/kg	i.p 1ml/kg
<i>Pterocarpus marsupium</i> , ⁶⁴	Papilionaceae	Stem bark	Aqueous, methanol	p.o 25mg/kg	i.p 0.1ml/kg
<i>Pterocarpus santalinus</i> , ⁶⁵	Fabaceae	Stem bark	70% ethanol	p.o 30mg/kg, 45mg/kg	i.p 0.1ml/kg
<i>Curculigo orchioides</i> , ⁶⁶	Amaryllidaceae	Rhizomes	Methanol	p.o 70mg/kg	s.c 1ml/kg
<i>Phoenix dactylifera</i> , ⁶⁷	Palmae	Fruit	Aqueous	Ad libitum in place of rat chow	i.p 2ml/kg
<i>Asteracantha longifolia</i> , ⁶⁸	Acanthaceae	Whole plant	Aqueous	p.o 900mg/kg	i.p 0.5ml/kg
<i>Strychnos potatorum</i> , ⁶⁹	Loganiaceae	Seeds	Aqueous, seed powder	p.o 100mg/kg, 200mg/kg	s.c 3ml/kg
<i>Vitex trifolia</i> , ⁷⁰	Verbenaceae	Leaves	Aqueous, ethanol	p.o 20mg/kg, 30mg/kg p.o 100mg/kg,	i.p 0.1ml/kg
<i>Capparis spinosa</i> , ⁷¹	Capparidaceae	Root bark	Ethanol	200mg/kg, 400mg/kg, 800mg/kg	p.o 0.2mg/kg
<i>Lawsonia alba</i> , ⁷²	Lythraceae	Bark	50% ethanol	p.o 250mg/kg, 500mg/kg	p.o 1ml/kg
<i>Carissa opaca</i> , ⁷³	Apocynaceae	Leaves	Methanol	p.o 200mg/kg	i.p 0.5ml/kg

TABLE 2: HEPATOPROTECTIVE ACTIVITY OF MEDICINAL PLANTS IN PARACETAMOL INDUCED HEPATOTOXICITY

Plant	Family	Part used	Type of extract	Test dose	Paracetamol dose
<i>Azima tetracantha</i> , ⁷⁴	Salvadoraceae	Leaves	Ethanol	p.o 250mg/kg, 500mg/kg	p.o 2g/kg
<i>Dragea volubilis</i> , ⁷⁵	Asclepiadaceae	Fruits	Petroleum ether	p.o 100mg/kg, 200mg/kg	p.o 650mg/kg
<i>Coccinia indica</i> , ⁷⁶	Cucurbitaceae	Fruits	aqueous	p.o 200mg/kg, 400mg/kg	p.o 2g/kg
<i>Sida rhombifolia</i> , ⁷⁷	Malvaceae	Whole plant	Ethanol	p.o 100mg/kg, 200mg/kg	p.o 2g/kg
<i>Psidium guajava</i> , ^{78, 50}	Myrtaceae	Leaves	Ethanol ⁶⁰ Methanol ³²	p.o (200mg/kg, 400mg/kg) ⁶⁰ , (250mg/kg, 500mg/kg) ³²	p.o 835mg/kg ⁶⁰ , 1g/kg ³²
<i>Solanum pubescens</i> , ⁷⁹ <i>Tabebuia rosea</i> , ⁷⁹	Solanaceae Bignoniaceae	Leaves	Methanol	p.o 300mg/kg	p.o 3g/kg
<i>Ichnocarpus frutescens</i> , ⁸⁰	Apocynaceae	Whole plant	Methanol, chloroform	p.o 500mg/kg	p.o 3g/kg
<i>Vanilla planifolia</i> , ⁸¹	Orchidaceae	Beans	Ethanol	p.o 250mg/kg	p.o 750mg/kg
<i>Nilgirianthus ciliatus</i> , ⁸²	Acanthaceae	Bark	Methanol	p.o 500mg/kg	p.o 500mg/kg
<i>Phyllanthus niruri</i> , ⁸³ <i>Aegle marmelos</i> , ⁸³	Phyllanthaceae Rutaceae	Leaves	Aqueous	p.o 440mg/kg	p.o 2.5g/kg
<i>Aloe vera</i> , ⁸³ <i>Eclipta alba</i> , ⁸³	Xanthorrhoeaceae Asteraceae	Leaves	Aqueous	p.o 440mg/kg	p.o 500mg/kg
<i>Solanum indicum</i> , ⁸³	Solanaceae	Leaves	Aqueous	p.o 440mg/kg	p.o 500mg/kg

<i>Maytenus emarginata</i> , ⁸³	Celastraceae	Leaves	Aqueous	p.o 440mg/kg	p.o 500mg/kg
<i>Aerva lanata</i> , ⁸⁴	Amaranthaceae	Whole plant	Hydroalcohol	p.o 200mg/kg	p.o 3g/kg
<i>Aerva sanguinolenta</i> , ⁸⁵	Amaranthaceae	Leaves	Ethanol	p.o 200mg/kg, 400mg/kg	p.o 640mg/kg
<i>Gymnosporia emerginata</i> , ⁸⁶	Clasteraceae	Whole plant	Methanol	p.o 300mg/kg	p.o 3g/kg
<i>Marsdenia volubillis</i> , ⁸⁶	Asclepiadaceae	Whole plant	Methanol	p.o 500mg/kg p.o 100mg/kg,	p.o 3g/kg
<i>Carissa carandas</i> , ⁸⁷	Apocynaceae	Roots	Ethanol	200mg/kg, 400mg/kg	p.o 2000mg/kg
<i>Asparagus racemosus</i> , ⁸⁸	Liliaceae	Roots	Aqueous	p.o 150mg/kg, 200mg/kg	p.o 500mg/kg
<i>Kigelia Africana</i> , ⁸⁹	Bignoniaceae	Leaves	Methanol	p.o 200mg/kg	p.o 3g/kg
<i>Anogeissus acuminata</i> , ⁸⁹	Combretaceae	Leaves	Methanol	p.o 300mg/kg	p.o 3g/kg
<i>Abelmoschus moschatus</i> , ⁹⁰	Malvaceae	Seeds	Aqueous	p.o 300mg/kg	p.o 500mg/kg
<i>Cyperus articulatus</i> , ⁹¹	Cyperaceae	Rhizome	Methanol	p.o 200mg/kg, 400mg/kg	p.o 640mg/kg
<i>Thymus capitatus</i> , ⁹²	Lamiaceae	Essential oils	Steam distillation	p.o 50mg/kg	p.o 500mg/kg
<i>Salvia officinalis</i> , ⁹²	Lamiaceae	Essential oils	Steam distillation	p.o 50mg/kg	p.o 500mg/kg
<i>Solidago microglossa</i> , ⁹³	Compositae	Leaves	Ethanol	p.o 100mg/kg, 200mg/kg	p.o 250mg/kg
<i>Macrotyloma uniflorum</i> , ⁹⁴	Fabaceae	Seeds	Methanol	p.o 200mg/kg, 400mg/kg	p.o 2g/kg
<i>p.o 100mg/kg, 200mg/kg, 300mg/kg, 400mg/kg</i>					
<i>Tridax procumbens</i> , ⁹⁵	Compositae	Whole plant	Ethanol	p.o 2g/kg	
<i>Dipteracanthus patulus</i> , ⁵³	Acanthaceae	Leaves	Methanol	p.o 250mg/kg, 500mg/kg	p.o 2g/kg
<i>Orthosiphon stamineus</i> , ⁹⁶	Lamiaceae	Leaves	Methanol	p.o 100mg/kg, 200mg/kg	p.o 2g/kg
<i>Desmodium oojeinense</i> , ⁹⁷	Fabaceae	Bark	Ethanol	p.o 100mg/kg, 200mg/kg, 400mg/kg	p.o 2g/kg
<i>Trianthema portulacastrum</i> , ⁹⁸	Aizoaceae	Whole plant	Alcohol, aqueous	p.o 100mg/kg	p.o 3g/kg
<i>Acacia chatechu</i> , ⁹⁹	Mimosaceae	Heartwood	Ethanol	p.o 250mg/kg p.o 100mg/kg, 200mg/kg	p.o 250mg/kg
<i>Feronia limonia</i> , ¹⁰⁰	Rutaceae	Fruits	Ethanol	p.o 500mg/kg	
<i>Tecomella undulate</i> , ¹⁰¹	Bignoneaceae	Leaves	Methanol	p.o 100mg/kg, 200mg/kg	p.o 500mg/kg
<i>Balsiospermum montanum</i> , ¹⁰²	Euphorbiaceae	Roots	Chloroform, alcohol, aqueous	p.o 200mg/kg	p.o 2g/kg
<i>Cyathea gigantean</i> , ¹⁰³	Cyatheaceae	Leaves	Methanol	p.o 100mg/kg, 200mg/kg	p.o 1g/kg
<i>Asteracantha longifolia</i> , ⁶⁸	Acanthaceae	Whole plant	Aqueous	p.o 900mg/kg	p.o 300mg/kg
<i>Operculina turpethum</i> , ¹⁰⁴	Convolvulaceae	Roots	Ethanol	p.o 200mg/kg	p.o 3g/kg
<i>Tamarindus indica</i> , ¹⁰⁵	Caesalpiniaceae	Fruits, seeds, leaves	Aqueous	p.o 350mg/kg, 700mg/kg	p.o 1g/kg
<i>Berberis tinctoria</i> , ¹⁰⁶	Berneridaceae	Leaves	Methanol	p.o 150mg/kg, 300mg/kg	p.o 750mg/kg
<i>Azadirachta indica</i> , ¹⁰⁷	Meliaceae	Leaves	70% ethanol	p.o 500mg/kg	p.o 2g/kg
<i>Ceiba pentandra</i> , ¹⁰⁸	Bombacaceae	Stem bark	Ethylacetate	p.o 400mg/kg	p.o 3g/mg
<i>Plumbago zeylanica</i> , ¹⁰⁹	Plumbaginaceae	Roots	Petroleum ether	p.o 300mg/kg	p.o 400mg/kg
<i>Phyllanthus emblica</i> , ¹¹⁰	Euphorbiaceae	Fruits	Aqueous	p.o 100mg/kg, 200mg/kg	p.o 2g/kg

TABLE 3: HEPATOPROTECTIVE ACTIVITY OF MEDICINAL PLANTS IN D-GALACTOSAMINE INDUCED HEPATOTOXICITY

Plant	Family	Part used	Type of extract	Test dose	D-galactosamine dose
<i>Pittosporum neelgherrense</i> , ¹¹¹	Pittosporaceae	Stem bark	Methanol	p.o 100mg/kg, 200mg/kg, 300mg/kg	i.p 400mg/kg
<i>Sphaeranthus amaranthoides</i> , ¹¹²	Compositae	Whole plant	Ethanol	p.o 500mg/kg	i.p 500mg/kg
<i>Olenlandia herbacea</i> , ¹¹³	Rubiaceae	Whole plant	Methanol	p.o 100mg/kg, 200mg/kg	i.p 200mg/kg
<i>Calotropis gigantea</i> , ¹¹⁴	Asclepiadaceae	Root bark	Ethanol	p.o 200mg/kg, 400mg/kg	i.p 400mg/kg
<i>Coldenia procumbens</i> , ¹¹⁵	Boraginaceae	Whole plant	Methanol, chloroform	p.o 200mg/kg, 400mg/kg	i.p 400mg/kg
<i>Portulaca oleracea</i> , ¹¹⁶	Portulacaceae	Whole plant	Methanol, petroleum ether	p.o 200mg/kg, 400mg/kg	i.p 400mg/kg
<i>Betula utilis</i> , ¹¹⁷	Betulaceae	Bark	Ethanol, aqueous	p.o 100mg/kg, 200mg/kg	i.p 400mg/kg
<i>Pterocarpus santalinus</i> , ¹¹⁸	Fabaceae	heartwood	chloroform	p.o 200mg/kg, 400mg/kg	i.p 400mg/kg
<i>Santolina chamaecyparissus</i> , ¹¹⁹	Asteraceae	Whole plant	Ethanol	p.o 250mg/kg	i.p 400mg/kg
<i>Macrotyloma uniflorum</i> , ⁹⁴	Fabaceae	Seeds	Methanol	p.o 200mg/kg, 400mg/kg	i.p 400mg/kg
<i>Polygala arvensis</i> , ¹²⁰	Polygalaceae	Leaves	Chloroform	p.o 200mg/kg, 400mg/kg	i.p 400mg/kg
<i>Enicostemma axillare</i> , ¹²¹	Gentianaceae	Whole plant	Ethylacetate	p.o 100mg/kg, 200mg/kg	i.p 200mg/kg
<i>Solanum tuberosum</i> (purple potato), ¹²²	Solanaceae	Tubers	Formic acid, distilled water	p.o 400mg/kg	i.p 250mg/kg
<i>Crassocephalum crepidioides</i> , ⁵⁵	Asteraceae	Whole plant	Aqueous	i.p 5ml/kg	i.p 400mg/kg
<i>Garcinia kola</i> , ¹²³	Guttifera	Seeds	Methanol 50%ethanol, butanol, chloroform	p.o 100mg/kg	1.p 800mg/kg
<i>Fumaria indica</i> pugsley, ¹²⁴	Fumariaceae	Whole plant	50%ethanol, butanol, chloroform	p.o 200mg/kg, protopine 50mg/kg	i.p 400mg/kg
<i>Leucas lavandulaefolia</i> , ¹²⁵	Labiatae	Leaves	Methanol	p.o 100mg/kg	i.p 800mg/kg
<i>Indigofera tinctoria</i> , ⁶²	Leguminaceae	Whole plant	Acetone, methanol, chloroform	p.o 500mg/kg	5mM perfusion

TABLE 4: HEPATOPROTECTIVE ACTIVITY OF MEDICINAL PLANTS IN THIOACETAMIDE INDUCED HEPATOTOXICITY

Plant	Family	Part used	Type of extract	Test dose	Thioacetamide dose
<i>Pisonia aculeate</i> , ¹²⁶	Nyctaginaceae	Whole plant	Methanol	p.o 250mg/kg, 500mg/kg	s.c 50mg/kg
<i>Phyllanthus niruri</i> , ¹²⁷	Phyllanthaceae	Whole plant	Ethanol	p.o 200mg/kg	i.p 200mg/kg
<i>Vitex negundo</i> , ¹²⁸	Lamiaceae	Leaves	Ethanol	p.o 100mg/kg, 300mg/kg	0.03% in drinking water
<i>Orthosiphon stamineus</i> , ¹²⁹	Lamiaceae	Leaves	Ethanol	p.o 100mg/kg, 200mg/kg	i.p 200mg/kg
<i>Momordica tuberosa</i> , ¹³⁰	Cucurbitaceae	Tubers	Ethanol	p.o 20mg/kg, 40mg/kg	s.c 100mg/kg
<i>Tinispora crispa</i> , ¹³¹	Menispermaceae	Stem	Ethanol	p.o 100mg/kg, 200mg/kg	i.p 200mg/kg
<i>Zizyphus jujube</i> , ¹³²	Rhamnaceae	Fruits	Methanol	p.o 250mg/kg,	s.c 100mg/kg

				500mg/kg, 1000mg/kg	
<i>Phoenix dactylifera</i> , ¹³³	Aracaceae	Fruits	Aqueous	p.o 4ml/kg	i.p 400mg/kg
<i>Gardenia gummifera</i> , ¹³⁴	Rubiaceae	Roots	Methanol	p.o 125mg/kg, 250mg/kg	s.c 100mg/kg
<i>Albizia lebbeck</i> , ¹³⁵	Fabaceae	Leaves	70% Ethanol	p.o 100mg/kg, 200mg/kg	s.c 100mg/kg
<i>Wedelia calendulaceaee</i> , ¹³⁶	Compositae	Leaves	Methanol	p.o 100mg/kg, 200mg/kg, 400mg/kg	s.c 100mg/kg
<i>Ceiba pentandra</i> , ¹³⁷	Bombacaceae	Root	Methanol	p.o 200mg/kg, 400mg/kg	s.c 50mg/kg
<i>Ipomoea aquatica</i> , ¹³⁸	Convolvulaceae	Leaves	Ethanol	p.o 250mg/kg, 500mg/kg	i.p 200mg/kg
<i>Boerhaavia diffusa</i> , ¹³⁹	Nyctaginaceae	Stem, leaves	Alcohol, aqueous	p.o 150mg/kg, 300mg/kg	s.c 50mg/kg
<i>Anisochilus carnosus</i> , ¹³⁹	Nyctaginaceae	Leaves	Alcohol, aqueous	p.o 200mg/kg, 400mg/kg	s.c 50mg/kg
<i>Phyllanthus acidus</i> , ¹⁴⁰	Euphorbiaceae	Leaves	Ethanol, aqueous	p.o 200mg/kg, 400mg/kg	s.c 100mg/kg
<i>Artemisia aucheri</i> , ¹⁴¹	Compositae	Flowered branches	Ethanol	i.p 100mg/kg, 200mg/kg	i.p 50mg/kg
<i>Nigella sativa</i> , ¹⁴²	Ranunculaceae	Seed oil		p.o 5ml/kg, 10ml/kg	i.p 20mg/kg
<i>Curcuma longa</i> , ¹⁴³	Zingiberaceae	Rhizome	Ethanol	p.o 250mg/kg, 500mg/kg	i.p 200mg/kg
<i>Capparis deciduas</i> , ¹⁴⁴	Capparaceae	Root bark	Ethanol	p.o 300mg/kg, 600mg/kg	s.c 400mg/kg
<i>Bosenbergia rotunda</i> , ¹⁴⁵	Zingiberaceae	Rhizomes	Ethanol	p.o 250mg/kg, 500g/kg	i.p 200mg/kg
<i>Feronia elephantum</i> , ¹⁴⁶	Rutaceae	Leaves	Aqueous	p.o 400mg/kg, 800mg/kg	i.p 300g/kg

CONCLUSION: From this study, it is clear that many medicinal plants possess significant hepatoprotective activity. Our review will help researchers to choose different herbs and blend it to a formulation which could be an effective treatment for various liver diseases.

REFERENCES:

- Coon MJ, Ding XX, Pernecke SJ, & Vaz AD: Cytochrome P450: Progress and predictions. FASEB Journal 1992; 6(2): 669-673.
- Rajesh Thatavarthi, Putta Rajesh Kumar and Sreedevi: Racemethionine Hepatoprotective Activity against Rifampicin Induced Hepatotoxicity in Albino Rats. Der Pharmacia Lettre, 2011, 3(2):396-406.
- Orhan DD, Orhan N and Ergun F.: Hepatoprotective effect of *Vitis vinifera* L. Leaves on carbon tetrachloride induced acute liver damage in rats. Journal of Ethnopharmacology 2007; 112: 145-151.
- TS Mohamed Saleem, C Madhusudhana Chetty, S Ramkanth, VST Rajan, K Mahesh Kumar, Gauthaman K: Hepatoprotective Herbs – A Review International Journal of research in Pharmaceutical Sciences 2010; 1(1): 1-5.
- Arvind S Negi, JK Kumar, Suaiib Luqman, Karuna Shanker, MM Gupta, SPS Khanuja: Recent Advances in Plant Hepatoprotectives: A Chemical and Biological Profile of Some Important Leads. Medicinal Research Reviews (2008); 28(5): 746 -772.
- Richardo et al: Carbon Tetrachloride Hepatotoxicity- Pharmacological review 1967; 19:145-208.
- Alric L, M Fort, et al.: Study of host and virus related factors associated with spontaneous hepatitis C virus clearance. Tissue antigens 2000; 56(2): 154-58.
- Mitchell JR, Jollow DJ, Potter WZ, Gillette JR and Brodie BB. : Acetaminophen-induced hepatic necrosis. IV. Protective role of glutathione. Journal of Pharmacology and Experimental Therapeutics 1973; 187: 211–217.
- Huggette Houghton PJ, Hylands PJ, Mensah AY, Hensel A, Deters AM: In vitro tests and ethnopharmacological investigations: wound healing as an example. Journal of Ethnopharmacology 2005; 100(1-2): 100-107.

10. Prescott LF : Hepatotoxicity of mild analgesics British Journal of Clinical Pharmacology 1980; 10(Suppl 2): 373S-379S.
11. Coles B, Wilson I, Wardman P, Hinson JA, Nelson SD, and Ketterer B: The spontaneous and enzymatic reaction of N-acetyl-p-benzoquinonimine with glutathione: a stopped-flow kinetic study. Archives Biochemistry and Biophysics 1988; 264: 253-260.
12. Nakamura W, Hosoda S, and Hayachi K : Purification and properties of rat liver glutathione peroxidase. Biochim Biophys Acta 19974; 358: 251-261.
13. Wojcicki J, L Samochowlec & A Hinek: The effect of cernitinTM on galactosamine-induced hepatic injury in rat. Graminex: "Your Botanical source" 1984; 2: 1-5.
14. Koff RS, Gordon G, and Sabesin SM: D-Galactosamine hepatitis: 1. Hepatocellular injury and fatty liver following a single dose. Proceedings of the society of experimental biology and medicine 1971; 137:696.
15. Pickering RW, GWL James & FL Parker; An investigation of some parameters that affect the galactosamine model of hepatitis in the rat. Arzneimittelforsch. 1975; 23: 1591-1592.
16. Hunter AL, Holsher MA, Neal RA: Thioacetamide-induced hepatic necrosis. I. Involvement of the mixed-function oxidase-enzyme system. Journal of Pharmacology and Therapeutics 1977; 200:439-448.
17. Porter WR, Gudzinowcz MJ, Neal RA: Thioacetamide induced hepatic necrosis. II. Pharmacokinetics of thioacetamide and thioacetamide S-oxide in the rat. Journal of Pharmacology and Experimental Therapeutics 1979; 208:386-391.
18. Feroz Ahmad, Nahida Tabassum: Experimental models used for the study of antihepatotoxic agents. Journal of Acute Disease 2012; 85-89.
19. Rajib A, Monirul Islam KM, Musaddik A and Haque E: Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride induced hepatotoxicity in albino rats. Global Journal of Pharmacology 2009; 3(3): 116-122.
20. Yar HS, Ismail DK and Alhamed MN: Hepatoprotective effect of *Carthamus tinctorius* L. against carbon tetrachloride induced hepatotoxicity in rats. Pharmacie Globale International Journal of Comprehensive pharmacy 2012; 9(2): 1-5.
21. Pradeep KS: Hepatoprotective activity of *Ardisia solanacea* in carbon tetrachloride induced hepatotoxic albino rats. Asian Journal of Research in Pharmaceutical Sciences 2013; 3(2): 79-82.
22. Jameel A, Sunil N, Vipul D, Anuja P, Sagar K, Subodh Pal, Subhash M, and Shashikant P: Hepatoprotective activity of methanol extract of aerial parts of *Delonix regia*. Phytopharmacology 2011; 1(5): 118-122.
23. Mukul KG and Dasgupta S: Role of plant metabolites in toxic liver injury. Asian Pacific Journal of Clinical Nutrition 2002; 11(1): 48-50.
24. Mohammed HH: Protective effect of *Coriandrum sativum* plant of hepatotoxicity and nephrotoxicity induced by carbon tetrachloride in male albino rats. The 6th Arab and 3rd International Annual Scientific Conference on: Development of Higher Specific Education Programs in Egypt and the Arab World in the Light of Knowledge Era Requirements; 2011: April 13-14.
25. Pushpalatha M and Ananthi T: protective effect of *Solanum pubescens* linn on CCl₄ induced hepatotoxicity in albino rats. Mintage journal of pharmaceutical & medical sciences 2012; 1(1): 11-13.
26. Rajesh K, Sushil K, Arjun P and Jayalakshmi S: Hepatoprotective activity of aerial parts of *Plumbago zeylanica* linn against carbon tetrachloride-induced hepatotoxicity in rats. International Journal of Pharmacy and Pharmaceutical Sciences 2009; 1(1): 171-175.
27. Arjumand A, Srinivas Reddy K and Reddy CS: Hepatoprotective activity of *Cardiospermum helicacabum* stem extracts against carbon tetrachloride-induced hepatotoxicity in wistar rats. International journal of pharmaceutical sciences and nanotechnology 2009; 2(1): 488-492.
28. Ulaganathan I, Divya D, Radha K, Vijayakumar TM and Dhanaraju MP: Protective effect of *Luffa acutangula* (Var) amara against carbon tetrachloride-induced hepatotoxicity in experimental rats. Research Journal of Biological Sciences 2010; 5(9): 615-624.
29. Hewawasam RP, Jayatilaka KAPW, Pathirana C and Mudduwa LKB: Hepatoprotective effect of *Epaltes divaricata* extract on carbon tetrachloride induced hepatotoxicity in mice. Indian Journal of Medical Research, 2004: 30-34.
30. Ranjan KG, Anindya bose and mishra SK: Hepatoprotective activity of *Tagets erecta* against carbon tetrachloride-induced hepatic damage in rats. Acta poloniae pharmaceutica-Drug Research 2011; 68(6): 999-1003.
31. Parameshwar P, Reddy YN, and Aruna Devi M: Hepatoprotective and antioxidant activities of *Ziziphus rotundifolia* (Linn.) against carbon tetrachloride-induced hepatic damage in rats. International journal of pharmaceutical sciences and nanotechnology 2012; 5(3): 1775-1779.
32. Ugwueze ME, Adonu CC and Attama AA: Evaluation of the hepatoprotective activity of root extracts of *Millettia aboensis* on carbon tetrachloride [CCl₄] induced hepatotoxicity in rats. International Journal of Advanced Research 2013; 1(5): 65-70.
33. Krishna Mohan G, Pallavi E, Ravi Kumar B, Ramesh M and Venkatesh S: Hepatoprotective activity of *Ficus carica* Linn. leaf extract against carbon tetrachloride-induced hepatotoxicity in rats. DARU 2007; 15(3): 162-166.
34. Patience OO, Festus BC, Philip FU, Nneka RN, Ijeoma EA and Nkemakonam CO: Phytochemical analysis, hepatoprotective and antioxidant activity of *Alchornea cordifolia* methanol leaf extract on carbon tetrachloride-induced hepatic damage in rats. Asian Pacific Journal of Tropical Medicine 2012: 289-293.
35. Hogade MG, Patil KS, Wadkar GH, Mathapati SS and Dhumal PB: Hepatoprotective activity of *Morus alba* (Linn.) leaves extract against carbon tetrachloride induced hepatotoxicity in rats. African Journal of Pharmacy and Pharmacology 2010; 4(10): 731-734.
36. Isaias BR, Maria del Rayo CC, Pilar CR, Hector GLG, Dalila CN, Francisco JAM and Elsa MTC: Hepatoprotective effect of *Leucophyllum frutescens* on wistar albino rats intoxicated with carbontetrachloride. Annals of Hepatology 2007; 6(4): 251-254.
37. Karunakar HE and Arun BJ: Hepatoprotective effect of *Carissa carandas* Linn root extract against carbon tetrachloride and paracetamol induced hepatic oxidative stress. Indian Journal of Experimental Biology, 2009; 47(8): 660-667.
38. Nwachukwu DC, Okwuosa CN, Chukwu PU, Nkiru AN and Udeani T: Hepatoprotective Activity of Methanol Extract of the Seeds of *Sesamum Indicum* in Carbon Tetrachloride Induced Hepatotoxicity in Rats. Indian Journal of Novel Drug delivery 2011; 3(1): 36-42.
39. Gnanaprakash K, Madhusudhana Chetty C, Ramkanth S, Alagusundaram M, Tiruvengadarajan VS, Angala

- Parameswari S and Mohamed Saleem TS: Aqueous Extract of *Flacourtia indica* Prevents Carbon Tetrachloride Induced Hepatotoxicity in Rat. International Journal of Biological and Life Sciences 2010; 6(1): 51-55.
40. Gayathiri S, Vetriselvan S, Shankar Jothi, Ishwin S, Hemah Devi, Shereenjeet Kaur, and Yaashini A: Hepatoprotective activity of aqueous extract of *Hippophae rhamnoides L.* in carbon tetrachloride induced hepatotoxicity in albino wistar rats. International Journal of Biological & Pharmaceutical Research 2012; 3(4): 531-537.
41. Bahar A, Tanveer A, Manoj V and Shah AK: Hepatoprotective activity of two plants belonging to the Apiaceae and the Euphorbiaceae family. Journal of Ethnopharmacology 2002; 79: 313-316.
42. Ning W, Peibo Li, Yonggang W, Wei P, Zhong W, Suiyi T, Shaoling L, Xiao S and WeiWei Su: Hepatoprotective effect of *Hypericum japonicum* extract and its fractions. Journal of Ethnopharmacology 2008; 116(1): 1-6.
43. Akram E, Pejman M, Maryam B, Jalal Z: Hepatoprotective activity of Cinnamon ethanolic extract against CCl_4 -induced liver injury in rats. EXCLI Journal 2012; 11: 495-507.
44. Takate SB, Pokharkar RD, Chopade VV and Gite VN: Hepato-Protective activity of the aqueous extract of *launaea intybacea (Jacq) Beauv* against carbon tetrachloride-induced hepatic injury in Albino Rats. Journal of Pharmaceutical Science and Technology 2010; 2 (7): 247-251.
45. Rekha R, Hemalatha S, Akasakalai K, MadhuKrishna CH, Bavan S, Vittal and Meenakshi Sundaram R: Hepatoprotective activity of *Mimosa pudica* leaves against Carbon tetrachloride induced toxicity. Journal of Natural Products 2009; 2: 116-122.
46. Sakthidevi G, Alagammal M and Mohan VR: Evaluation of Hepatoprotective and Antioxidant Activity of *Polygala javana* DC Whole Plant - CCl_4 Induced Hepatotoxicity in Rats. International Journal of Pharmaceutical and Chemical Sciences 2013; 2(2): 764-770.
47. Divya B, Praneetha P, Swaroopa Rani V and Ravi Kumar B: Hepatoprotective Effect of Whole Plant Extract Fractions of *Marsilea minuta* Linn. Asian Journal of Pharmaceutical and Clinical Research 2013; 6(3): 100-107.
48. Manisha S, Shete RV, Kore KJ and Attal AR: Hepatoprotective activity of *Ficus bengalensis* Linn leaves. Current Pharma Research 2012; 2(2): 503-507.
49. Durga PN, Dinda SC, Swain PK, Kar B and Patro VJ: Hepatoprotective activity against CCL_4 -induced hepatotoxicity in rats of *Chenopodium album* aerial parts. Journal of Phytotherapy and Pharmacology 2012; 1(2): 33-41.
50. Chanchal KR and Amit Kumar Das: Effect of *Psidium guajava* Linn. Methanolic leaf extract on hepatoprotection. Journal of Pharmaceutical and Biomedical Sciences 2010; 1(03).
51. Vishal BJ, Vishnu NT, Anupama AS, Avinash DD and Suresh RN: Hepatoprotective activity of *Luffa acutangula* against CCl_4 and rifampicin induced liver toxicity in rats: A Biochemical and histopathological evaluation. Indian Journal of Experimental Biology 2010; 48(8): 822-829.
52. Prakash T, Snehal DF, Uday Raj S, Surendra V, Divakar Goli, Perfect S and Kotresha D: Hepatoprotective activity of leaves of *Rhododendron arboreum* in CCl_4 induced hepatotoxicity in rats. Journal of Medicinal Plants Research 2008; 2(11): 315-320.
53. Shrinivas B and Suresh RN: Hepatoprotective Activity of Methanolic Extract of *Dipteracanthus patulus* (JACQ) NEES: possible involvement of antioxidant and membrane stabilization property. International Journal of Pharmacy and Pharmaceutical Sciences 2012; 4(2): 685-690.
54. Amrita R, Ajay KS, Neelima S and Sanjeev KS: Hepatoprotective and antioxidants activity of ethanolic extract of *Cuscutta reflexa* roxb. IOSR Journal of Pharmacy 2012; 2(2): 142-147.
55. Yoko A, Tomoyuki K, Chika M, Manami M, Chiho I, Shizuka K and Toshio I: Free radical scavenging and hepatoprotective actions of the medicinal herb, *Crassocephalum crepidioides* from the Okinawa Islands. Biological and Pharma Bulletin 2005; 28(1): 19-23.
56. Rajesh MG and Latha MS: Protective activity of *Glycyrrhiza glabra* Linn. on carbon tetrachloride-induced peroxidative damage. Indian Journal of Pharmacology 2004; 36: 284-7.
57. Akram J, Fatema F, Zohreh S and Hossein N: Hepatoprotective activity of *Gundelia tournefortii*. Journal of Ethnopharmacology 2005; 101(1-3): 233-237.
58. Xingshen Ye, Xibin F, Yao T, Kwan-Ming NG, Yanbo Z, Jun T, Jiangang S and Seiichi K: Hepatoprotective effects of *Coptidis rhizome* aqueous extract on carbon tetrachloride-induced hepatotoxicity in rats. Journal of Ethnopharmacology 2009; 124(1): 130-136.
59. Adeneye AA, Olagunju JA, Banju AAF, Abdul SF, Sanusi OA, Sanni OO, Osarodion BA and Shoniki OE: The aqueous seed extract of *Carica papaya* Linn. Prevents carbon tetrachloride induced hepatotoxicity in rats. International Journal of Applied Research in Natural Products 2009; 2(2): 19-32.
60. Akram J, Mohammad JK, Zahra D and Hossein N: Hepatoprotective activity of *Cichorium intybus* L. leaves extract against carbon tetrachloride induced toxicity. Iranian Journal of Pharmaceutical Research 2006; 1: 41-46.
61. Praveen TK, Dharmaraj S, Jitendra B, Dhanabal SP, Manimaran S, Nanjan MJ and Rema R: hepatoprotective activity of petroleum ether, diethyether, and methanol extract of *Scoparia dulcis* L. against carbon tetrachloride-induced acute liver injury in mice. Indian Journal of Pharmacology 2009; 41(3): 110-114.
62. Meenakshisundaram S, Devaki T and Mohamad Rayeem: Protective effects of *Indigofera tinctorial* L. against D-galactosamine and carbon tetrachloride challenge on in situ perfused rat liver. Indian Journal of Physiology and Pharmacology 2001; 45(4): 428-434.
63. Shahjahan M, Sabitha KE, Mallika J and Shyamala Devi CS: Effect of *Solanum trilobatum* against carbon tetrachloride induced hepatic damage in albino rats. Indian Journal of Medical Research 2004; 120: 194-198.
64. Mankani KL, Krishna V, Manjunatha BK, Vidya SM, Jagadeesh singh SD, Manohara YN, Anees-UR Raheman and Avinash KR: Evaluation of hepatoprotective activity of stem bark of *Pterocarpus marsupium* Roxb. Indian Journal of Pharmacology 2005; 37: 165-8.
65. Manjunatha BK: Hepatoprotective activity of *Pterocarpus santalinus* L.F, endangered medicinal plant. Indian Journal of Pharmacology 2006; 38(1): 25-28.
66. Venukumar MR and Latha MS: *Curculigo orchioides* in carbon tetrachloride induced hepatotoxicity in rats. Indian Journal of clinical Biochemistry 2002; 17(2): 80-87.
67. AL-Qarashi AA, Hamed Ali HMM and Abdel-rahman Samy H: Protective effects of extracts from Dates (*Phoenix dactylifera L.*) on carbon tetrachloride-induced hepatotoxicity in rats. International Journal of Applied Research in veterinary Medicine 2004; 2(3): 176-180.
68. Hewawasam RP, Jayatilaka KAPW, Parthiranathan C and Mudduwa LKB: Protective effect of *Asteracantha longifolia* extract in mouse liver injury induced by carbon

- tetrachloride and paracetamol. Journal of Pharmacy and Pharmacology 2003; 55(10): 1413-1418.
69. Sanmugapriya E and Venkataraman S: Studies on hepatoprotective and antioxidant actions of *Strychnos potatorum Linn.* Seeds on carbon tetrachloride-induced acute hepatic injury in experimental rats. Journal of Ethnopharmacology 2006; 105(1-2): 154-160.
 70. Manjunatha BK and Vidya SM: Hepatoprotective activity of *Vitex trifolia* against carbon tetrachloride-induced hepatic damage. Indian Journal of Pharmaceutical Sciences 2008; 70(2): 241-245.
 71. Aghel N, Rashidi I and Mombeini A: Hepatoprotective activity of *Capparis spinosa* Root bark against carbon tetrachloride induced hepatic damage in mice. Iranian Journal of pharmaceutical research 2007; 6(4): 285-290.
 72. Ahmed S, Rahman A, Alam A, Saleem M, Athar M and Sultana S: Evaluation of the efficacy of *Lawsonia alba* in the alleviation of carbon tetrachloride induced oxidative stress. Journal of Ethnopharmacology 2000; 69(2): 157-164.
 73. Sumaira S, Muhammad RK and Rahmat AK: Hepatoprotective effects of methanol extract of *Carissa opaca* leaves on carbon tetrachloride- induced damage in rats. BMC Complementary and alternative Medicine 2011; 11: 48.
 74. Arthika S, Shanthammal Y, Sheryl Igali N, Elankini P, Ganesan R, Gaidhani SN and Pramod Reddy G: Hepatoprotective Activity of the Ethanoilc Extract of *Azima tetracantha* against Paracetamol-Induced Hepatotoxicity in Wistar Albino Rats. Journal of Advances in Pharmacy and Healthcare Research 2011; 1(2): 14-21.
 75. Pallab KH, Moulisha B, Sanjib B, Tarun KK and Ashoke KG: Hepatoprotective Activity of *Dregea volubilis* Fruit against Paracetamol-Induced Liver Damage in Rats. Indian Journal of Pharmaceutical Education and Research 2012; 46(1): 17-22.
 76. Arun KS and Eswar KK: Hepatoprotective Activity of Aqueous Fruit Extract of *Coccinia indica* against Paracetamol Induced Hepatotoxicity in rats. International Journal of Research in Pharmaceutical and Biomedical Sciences 2013; 4 (1): 179-182.
 77. Ramadoss S, Kannan K, Balamurugan K, Jeganathan NS and Manavalan R: Evaluation of Hepato-Protective Activity in the Ethanolic Extract of *Sida rhombifolia* Linn. against Paracetamol - Induced Hepatic Injury in Albino Rats. Research Journal of Pharmaceutical, Biological and Chemical Sciences 2012; 3(1): 497-502.
 78. Priscilla D Mello and Milan R: Hepatoprotective activity of *Psidium guajava* extract and its phospholipid complex in paracetamol induced hepatic damage in rats. International Journal of Phytomedicine 2010; 2: 85-93.
 79. Hemamalini K, ramya Krishna V, Anurag Bhargav DR and Uma Vasireddy DR: Hepatoprotective activity of *Tabebuia rosea* and *Solanum pubescens* against paracetamol induced hepatotoxicity in rats. Asian Journal of Pharmaceutical and Clinical Research 2012; 5(4): 153-156.
 80. Deepak KD, Veerendra CY , Siva S, Tirtha G, Rajalingam D, Pinaki S , Bhim CM and Tapan KM: Evaluation of hepatoprotective and antioxidant activity of *Ichnocarpus frutescens* (Linn.) R.Br. on paracetamol-induced hepatotoxicity in rats. Tropical Journal of Pharmaceutical Research, September 2007; 6 (3): 755-765.
 81. Geogi PG, Anitha P, Anthoni Samy A and Kanimozhi R: Hepatoprotective activity of *Vanilla planifolia* against paracetamol induced hepatotoxicity in albino rats. International Journal of Institutional Pharmacy and Life Sciences 2011; 1(3): 70-74.
 82. Usha Rani K, Amirtham D and Nataraja TS: Hepatoprotective activity of *Nilgirianthus ciliatus* (Nees) bremek in paracetamol induced toxicity in Wistar albino rats. African Journal of Internal Medicine 2013; 1(4): 026-030.
 83. Simon RP, Patel Hitesh V and Kiran K: Hepatoprotective activity of some plants extract against paracetamol induced hepatotoxicity in rats. Journal of Herbal Medicine and Toxicology 2010; 4 (2): 101-106.
 84. Manokaran S, Jaswanth A, Sengottuvelu S, Nandhakumar J, Duraisamy R, Karthikeyan D and Mallegaswari R: Hepatoprotective Activity of *Aerva lanata* Linn. Against Paracetamol Induced Hepatotoxicity in Rats. Research Journal of Pharmacy and Technology 2008; 1(4): 398-400.
 85. Asif L, Bolay B, Mousumi Das, Debmalya M, Sudipta K, Samit B and Amalesh Samanta: Hepatoprotective activity of ethanolic extract of *Aerva sanguinolenta* (Amaranthaceae) against paracetamol induced liver toxicity on Wistar Rats. NSHM Journal of Pharmacy and Healthcare Management 2012; 03: 57-65.
 86. Asma Rubab, Hemamalini K, Shashi Priya G and Uma V: Hepatoprotective Activity of *Gymnosporia emerginata* (Willd) and *Marsdenia volubillis* (Linn.F) Staph Against Paracetamol Induced Hepatotoxicity in Rats. International Journal of Current Pharmaceutical Review and Research 2013; 4(2): 36-41.
 87. Karunakar HE and Arun BJ: Hepatoprotective effect of *Carissa carandas* Linn root extract against carbon tetrachloride and paracetamol induced hepatic oxidative stress. Indian Journal of Experimental Biology, 2009; 47(8): 660-667.
 88. Fasalu Rahiman OM, Rupesh Kumar M, Tamizh Mani T, Mohamed Niyas K, Satya Kumar B, Phaneendra P and Surendra B: Hepatoprotective Activity of "Asparagus Racemosus Root" On Liver Damage Caused By Paracetamol in Rats. Indian Journal of Novel Drug delivery 2011; 3(2): 112-117.
 89. Hemamalini K, Preethi B, Anurag Bhargav and Uma V: Hepatoprotective activity of *Kigelia africana* and *Anogeissus accuminata* against paracetamol induced hepatotoxicity in rats. International Journal of Pharmaceutical and Biomedical Research 2012; 3(3):152-156.
 90. Abhishek KS, Sanjiv S and Chandel HS: Evaluation of hepatoprotective activity of *abelmoschus moschatus* seed in paracetamol induced hepatotoxicity on rat. IOSR Journal of Pharmacy 2012; 2(5): 43-50.
 91. Samaresh D, Susmita D, Nayak SS and Subas CD: Hepatoprotective activity of *Cyperus articulatus* Linn.against paracetamol induced hepatotoxicity in rats. Journal of Chemical and Pharmaceutical Research 2013; 5(1): 314-319.
 92. El-Banna H, Soliman M and Al-wabel N: Hepatoprotective Effects of Thymus and *Salvia* Essential oils on Paracetamol-Induced Toxicity in Rats. Journal of Physiology and Pharmacology Advances 2013; 3(2): 41-47.
 93. Sabir SM, Ahmad SD, Hamid A, Khan MQ, Athayde ML, Santos DB, Boligon AA and Rocha JBT: Antioxidant and Hepatoprotective activity of ethanolic extract of leaves of *Solidago microglossa* containing polyphenolic compounds. Food Chemistry 2012; 131(3): 741-747.
 94. Parmar HB, Das SK and Gohil KJ: Hepatoprotective activity of *Macrotyloma uniflorum*. Seed extract on paracetamol and d-galactosamine induced liver toxicity in albino rats. International Journal of Pharmacological Research 2012; 2(2): 86-91.

95. Shardul SW and Gangadhar BS: Antioxidant and hepatoprotective activity of *Tridax procumbens* Linn, against paracetamol induced hepatotoxicity in male albino rats. Advanced Studies in Biology 2010; 2(3): 105-112.
96. Maheswari C, Maryammal R and Venkatanarayanan R: Hepatoprotective Activity of "Orthosiphon stamineus" on Liver Damage Caused by Paracetamol in Rats. Jordan Journal of Biological Sciences 2008; 1(3): 105 -108.
97. Jayadevaiah KV, Ishwar Bhat K, Joshi AB, Vijayakumar MMJ and Pinkey R: Hepatoprotective activity of *Desmodium oojeinense* (ROXB) H. Ohashi against paracetamol induced hepatotoxicity. Asian Journal of Pharmaceutical and Health Sciences 2012; 2(2): 312-315.
98. Mehta RS, Shankar MB, Geetha M, Saluja AK: Preliminary evaluation of hepatoprotective activity of *Trianthemum portulacastrum* Linn. Journal of Natural Remedies 2003; 3(2): 180 – 184.
99. Sheshidhar GB, Yasmeen AM, Arati C, Sangappa VK, Pundarikaksha HP, Vijay Domble and Manjula R: Evaluation of hepatoprotective activity of ethanolic extract of *Acacia catechu* wild in paracetemol induced hepatotoxicity in albino rats. International Journal of Pharmaceutical and Biological Sciences 2013; 3(2): 264-270.
100. Arshed ID, Saxena RC and Bansal SK: Assessment of hepatoprotective activity of fruit pulp of *Feronia limonia* (Linn.) against paracetamol induced hepatotoxicity in albino rats. Journal of Natural Products Plant Resources 2012; 2 (2): 226-233.
101. Singh D and Gupta RS: Hepatoprotective Activity of Methanol Extract of *Tecomella undulate* against Alcohol and Paracetamol Induced Hepatotoxicity in Rats. Life Sciences and Medicine Research 2011; LSMR-26.
102. Raju RW, Radhika SS, Kunal MT, Kalpana SP and Sunil SJ: Screening of roots of *Baliospermum montanum* for hepatoprotective activity against paracetamol-induced liver damage in albino rats. International Journal of Green Pharmacy 2008; 2: 220-3.
103. Madhu Kiran P, Vijaya Raju A, Ganga Rao B: Investigation of hepatoprotective activity of *Cyathea gigantea* (Wal and. Ex. Hook) leaves against paracetamol-induced hepatotoxicity in rats. Asian Pacific Journal of Tropical Biomedicine 2012; 2(5): 352-356.
104. Suresh Kumar SV, Sujatha C, Syamala J, Nagasudha B and Mishra SH: Hepatoprotective effect of root extract of *Operculina turpethum* Linn. Against paracetamol-induced hepatotoxicity in rats. Indian Journal of Pharmaceutical Sciences 2006; 68: 32-5.
105. Pimple BP, Kadam PV, Badgujar NS, Bafna AR and Patil MJ: Protective effect of *Tamarindus indica* Linn. against paracetamol-induced hepatotoxicity in rats. Indian Journal of Pharmaceutical Sciences 2007; 69: 827-831.
106. Murugesh, Kanda Samy Y, Veerendra CM, Charan Bhim M and Tapan Kumar: Hepatoprotective and antioxidant role of *Berberis tinctorial* Lesch leaves on paracetamol-induced hepatic damage in rats. Iranian Journal of Pharmacology and Therapeutics 2006; 4(1).
107. Chattopadadhayay RR and Bandyopadhyay M: Possible mechanism of hepatoprotective activity of *Azadirachta indica* leaf extract against paracetamol-induced hepatic damage in rats: part III. Indian Journal of Pharmacology 2005; 37: 184-5.
108. Nirmal KB, Neeraj K and Mishra SH: Protective effect of stem bark of *Ceiba pentandra* Linn. Against paracetamol-induced hepatotoxicity in rats. Pharmacognosy Research. 2010; 2(1): 28-30.
109. Kanchana N and Mohamed Sadiq A: Hepatoprotective effect of *Plumbago zeylanica* on paracetamol- induced liver toxicity in rats. International Journal of Pharmacy and Pharmaceutical Sciences 2011; 3(1): 151-154.
110. Vidhya Malar HL and Mary Mettilda Bai S: Hepatoprotective activity of *Phyllanthus emblica* against paracetamol-induced hepatic damage in wistar albino rats. African Journal of Basic and Applied Sciences 2009; 1(1-2): 21-25.
111. Shyamal S, Latha PG, Shine VJ and Ganga Devi T: Hepatoprotective effects of *Pittosporum neelgherrense* on D - galactosamine (d-galn) induced liver injury in rats: 04-24
112. Swarnalatha L and Neelakanta Reddy P: Hepatoprotective activity of *Sphaeranthus amaranthoides* on D-galactosamine induced hepatitis in albino rats. Asian Pacific Journal of Tropical Biomedicine 2012: S1900-S1905.
113. Pandian S, Badami S and Shankar M: Hepatoprotective Activity of Methanolic Extract of *Oldenlandia herbacea* Against D – Galactosamine Induced Rats. International Journal of Applied Research in Natural Products 2008; 6 (1): 16-19.
114. Pradeep D, Tanaji N, Mahendra Singh R, Anil M and Nithin J: Hepatoprotective activity of *Calotropis gigantea* Root bark experimental liver damage induced by D-galactosamine in rats. International Journal of Pharmaceutical Sciences and Nanotechnology 2008; 1(3): 281-286.
115. Ganesan R, Venkatanarasimhan M, Sharad P, Pramod reddy G, Anandan T and Masilamani G: Hepatoprotective effect of *Coldenia procumbens* linn against D-galactosamine induced acute liver damage in rats. International Journal of Integrative sciences, Innovation and Technology 2013; 2(2): 9-11.
116. Prabhakaran V, Srinivas Ashok Kumar B, Sheshadri Shekar D, Nandeesh R, Subramanyam P and Ranganayakulu D: Evaluation of the hepatoprotective activity of *Portulaca oleracea* L. on D-galactosmaine-induced hepatic injury in rats. Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas 2010; 9(3): 199-205.
117. Duraiswamy B, Satish kumar MN, Gupta S, Rawat M, Om Porwal and Murugan R: Hepatoprotective activity of *Betula utilis* bark on D-galactosamine induced hepatic insult. World Journal of Pharmacy and Pharmaceutical Sciences 2012; 1(1): 456-471.
118. Dhanabal P, Syamala, Elango K and Suresh B: Protective and Therapeutic Effects of the Indian Medicinal Plant *Pterocarpus santalinus* on D-Galactosamine-induced Liver Damage. Asian Journal of Traditional Medicines 2007; 2(2): 51-57.
119. Dhanabal SP, Rahul Jain, Priyanka DL, Muruganantham N and Raghu PS: Hepatoprotective activity of *Santolina chamaecyparissus* Linn against D-Galactosamine Induced Hepatotoxicity in Rats. Pharmacognosy Communications 2012; 2(2): 67-70.
120. Dhanabal SP, Syamala G, Satish Kumar MN and Suresh B: Hepatoprotective activity of the Indian medicinal plant *Polygala arvensis* on D-galactosamine-induced hepatic injury in rats. Fitoterapia 2006; 77(6): 472-474.
121. Jaishree V and Shrishailappa B: Antioxidant and hepatoprotective effect of Swertiajaponin from *Enicostemma axillare* against D-galactosamine induced acute liver damage in rats. Journal of Ethnopharmacology 2010; 130(1): 103-106.
122. Kyu-Ho H, Naoto H, Ken-ichiro S, MitsuO S, Takahiro N, Hiroaki Y, Makoto H, Hideyuki C, David LT and Michihiro F: Hepatoprotective effect of purple potato extract against D-galactosamine-induced liver injury in

- rats. Bioscience. Biotechnology and Biochemistry 2006; 70(6): 1432-1437.
123. Oluwatosin AA and Edward OA: Hepatoprotection on D-galactosamine-induced toxicity in mice by purified fractions from *Garcinia kola* seeds. Basic and Clinical Pharmacology and toxicology 2006; 98(2): 135-141.
 124. Anshu R, Arvind Kumar S, Annie S and Ajay Kumar Singh R: Hepatoprotective potential of *Fumaria indica* pugsley whole plant extracts, fractions and an isolated alkaloid protopine. Phytomedicine 2008; 15(6-7): 470-477.
 125. Kotoky J, Dasgupta B and Sarma GK: Protective properties of *Leucas lavendulaefolia* extracts against D-galactosamine induced hepatotoxicity in rats. Fitoterapia 2008; 79(4): 290-292.
 126. Ambarasu C, Rajkumar B, Bhat KS, John G, Arul Amuthan A and Satish K: Protective effect of *Pisonia aculeata* on thioacetamide induced hepatotoxicity in rats. Asian Pacific Journal of Tropical Biomedicine 2012; 2(7): 511-515.
 127. Zahra AA, Mohammed AA, Mustafa K, Hapipah M Ali and Mahmood A: Gene expression profiling reveals underlying molecular mechanism of hepatoprotective effect of *Phyllanthus niruri* on thioacetamide-induced hepatotoxicity in Sprague Dawley rats. BMC Complementary and Alternative Medicine 2013; 13:160 doi: 10.1186/1472-6882-13-160.
 128. Farkaad AK, Normadiah MK, Mahmood A and Wageeh AY: Hepatoprotective role of ethanolic extract of *Vitex negundo* in thioacetamide-induced liver fibrosis in male rats. Evidence based complementary and alternative medicine 2013 doi: 10.1155/2013/739850.
 129. Mohammed AA, Mahmood AA, Salmah Ismail and Zahra A: Hepatoprotective Effects of Orthosiphon stamineus Extract on Thioacetamide-Induced Liver Cirrhosis in Rats. Evidence-Based Complementary and Alternative Medicine 2011; Article ID 103039, 6 pages.
 130. Pramod Kumar, Devala rao G, lakshmayya and Ramachandra settee S: Hepatoprotective effect of ethanol extract of tubers of *Momordica tuberosa* cogn. In thioacetamide induced hepatic damage. Pharmacologyonline 2008; 3: 181-189.
 131. Farkaad AK, Faizah O, Mahmood AA, Farida H and Pouya H: Effect of *Tinospora crispa* on Thioacetamide-induced liver cirrhosis in rats. Indian Journal of Pharmacology 2011; 43(1): 64-68.
 132. Prasanna Kumar S, Basheeruddin Asdaq SM, Prem Kumar N, Asad M and Khajuria DK: Protective Effect Of *Zizyphus Jujuba* Fruit Extract Against Paracetamol And Thioacetamide Induced Hepatic Damage In Rats. The Internet Journal of Pharmacology 2009; 7(1): DOI: 10.5580/2991.
 133. Mohamed BA, Nabil Abdel-Salam Hasona and Hanan Abdel-Hamid Selemani: Protective Effects of Extract from Dates (*Phoenix Dactylifera L.*) and ascorbic acid on thioacetamide-induced hepatotoxicity in rats. Iranian Journal of Pharmaceutical research 2008; 7(3): 193-201.
 134. Prabha SP, Ansil PN, Nitha A, Wills PJ, Latha MS: Preventive and curative effect of methanolic extract of *Gardenia gummifera* Linn. on thioacetamide induced oxidative stress in rats. Asian Pacific Journal of Tropical Disease 2012; 90-98.
 135. Shirode DS, Jain BB, Mahendra Kumar CB and Setty SR: Hepatoprotective and antioxidant effects of *Albizia lebbeck* against thioacetamide induced hepatotoxicity in rats. Journal of Chemical and Pharmaceutical sciences 2012; 5: 199-204.
 136. Pallab KH, Malaya Gupta, Upal KM, Chandi CK and Laxmanan M: Hepatoprotective effect of *Wedelia calendulacea* against thioacetamide induced liver damage in rats. Pharmacologyonline 2007; 3: 414-421.
 137. Bhushan G, Kavimani S and Rajkumar B: Protective effect of *C. pentandra* on thioacetamide-induced hepatotoxicity in rats. International Journal of Biological & Pharmaceutical Research 2012; 3(1): 23-29.
 138. Salim Said A, Mahmood A, Ahmed Salim A, Suzy MS, Siddig IA and Hamid AH: *Ipomoea aquatica* Extract Shows Protective Action Against Thioacetamide-Induced Hepatotoxicity. Molecules 2012; 17: 6146-6155. doi: 10.3390/molecules17056146.
 139. Venkatesh P, Dinakar A and Senthilkumar N: Screening of hepatoprotective and antioxidant activity of alcoholic and aqueous extracts of *Boerhaavia diffusa* and *anisochilus carnosus*. International Journal of Pharmacy and Pharmaceutical Sciences 2013; 5(2): 208-211.
 140. Nilesh KJ and Abhay KS: Protective effects of *Phyllanthus acidus* (L.) Skeels leaf extracts on acetaminophen and thioacetamide induced hepatic injuries in Wistar rats. Asian Pacific Journal of Tropical Medicine 2011: 470-474.
 141. Azam R, Shahnaz SF, Saeed CA, Hydar A, Ali Z, Maryam A and Hasan Y: The effects of *Artemisia aucheri* extract on hepatotoxicity induced by thioacetamide in male rats. Avicenna Journal of Phytomedicine 2013; 3(4): 293-301.
 142. Shamshur N and Mamta Kumari: Ameliorating effect of *Nigella sativa* oil in thioacetamide-induced liver cirrhosis in albino rats. Indian journal of Pharmaceutical education and Research 2013; 47(2): 135-139.
 143. Suzy MS, Mahmood AA, Ahmed SA, Ismail S, Salim SA and Shahram G: Hepatoprotective effect of ethanolic extract of *Curcuma longa* on thioacetamide induced liver cirrhosis in rats. BMC Complementary and Alternative Medicine 2013; 13:56.
 144. Mahesh KJ, Sunil K and Srivastava B: Hepatoprotective activity of *Capparis decidua* on liver damage caused by thioacetamide in Wistar male Rats. International Journal of Toxicological and Pharmacological Research 2010; 2(3): 92-94.
 145. Suzy M, Mehmet, Ahmed SAL and Mahmood A: Efficacy of *Bosenbergia rotunda* treatment against thioacetamide-induced liver cirrhosis in a rat model. Evidence based complementary and alternative medicine. Volume 2012, article ID: 137083.
 146. Sharma P, Bodhankar SL and Thakurdesai PA: Protective effect of *Feronia elephantum correa* leaves on thioacetamide induced liver necrosis in diabetic rats. Asian Pacific Journal of Tropical Biomedicine 2012; 2(9): 691-695.

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