

PHARMACEUTICAL SCIENCES



Received on 26 May, 2015; received in revised form, 10 July, 2015; accepted, 30 September, 2015; published 01 December, 2015

EFFECT OF FIVE DIFFERENT CULTURE MEDIA ON MYCELIAL GROWTH OF AGROCYBE AEGERITA

Nagalakshmi Muthu and Krishnakumari Shanmugasundaram*

Department of Biochemistry, Kongunadu arts and science college (Autonomous), Coimbatore – 641 029, Tamil Nadu, India.

Keywords:

Agrocyba aegerita, Mycelium growth, Medium, Culture

Correspondence to Author: Dr. S. Krishnakumari

Associate Professor, Department of Biochemistry, Kongunadu Arts and Science College, Coimbatore – 641029, Tamil Nadu, India

E-mail: krishnashanmugambc@gmail.com

ABSTRACT: A preliminary experiment was carried out to analyse the growth performance of *Agrocyba aegerita* mushroom cultures using chemosynthetic media. This study is mainly aimed to different media such as Yeast Malt Agar Medium (YMA), Saboraud's Dextrose Agar Medium (SDA), Glucose Peptone Agar Medium (GPA), Malt Extract Agar Medium (MEA) and Potato Dextrose Agar Medium (PDA) on the growth of mycelium of the mushroom. Studies revealed that the joint portion of cap and stripe produced vigorous mycelium growth in minimum time; the average maximum growth was obtained on Malt Extract Agar (MEA) than Potato Dextrose agar (PDA) medium.

INTRODUCTION: Mushrooms have been recorded as a source of vegetable and medicines for human beings throughout the world. They are generally divided into edible and non-edible. Mushroom fruit bodies are well known food items since ancient times and became important as nutraceutical and pharmaceutical agent due to the capability of producing many useful secondary metabolites, high protein content with essential amino acids. vitamins, minerals and exopolysaccharides Agrocyba aegerita mushroom with high economic, nutritional, and pharmaceutical value. Mushrooms have become attractive as a functional food and as source for the development of drugs and nutraceuticals ^{2, 3} namely for antioxidant ⁴⁻¹⁰ and antimicrobial compounds 11.



In nature mushrooms grow wild in every country from snowy mountains to sandy deserts on all types of soils, pastures, forests, cultivated fields or water lands. They appear in all seasons, chiefly during the rainy weather, wherever organic matter or its decomposition products are available ¹². Many fungi and their mycelium biomass are reported as good source of food, protein supplement, lipid source and many more metabolites ^{13, 14, 15}.

The media listed above, with the exception of compost agars, are all devoid of insoluble material and are rich in simple sugars, few of which are encountered by the mushroom on its 'natural' substrate. The insoluble fraction, which includes lignin's, cellulose, hemicelluloses, protein and microbial biomass, is preferentially used by the mushroom mycelium for growth ¹⁶.

Mushrooms are amongst the most popular food items accepted the world over. The increased consumer demand over the years has resulted in production of mushrooms in large proportions ¹⁷ through cultivation which is a highly efficient

method for recycling the agricultural residues so as to produce nutritious food ¹⁸.

Today China is the major producer of mushroom in the world ¹⁹. In the present study, stock culture of vigorous growing mycelia of *Agrocyba aegerita*, was cultivated on different media (i.e.) YMA, PDA, SDA, GPA and MEA and their mycelial growth rate was determined. The purpose of the present study was to determine the most suitable media, for the growth of the mushroom.

MATERIAL AND METHODS:

Collection and Maintenance of A. Aegerita Culture:

The pure culture of *A. aegerita* was obtained from the Directorate of Mushroom Research (DMR), Chambaghat, Solan, Himachal Pradesh and were used for mass culture production.

Media preparation:

Following media were used for the purpose:

PDA- Potato Dextrose Agar Medium:

Potato dextrose agar - 39g

Water -1000ml

MEA - Malt Extract Agar Medium:

Malt extract - 30 g Agar-Agar - 15 g

Water - 1000 ml

GPA - Glucose Peptone Agar Medium:

Peptone - 20 g

Dextrose - 10 g

Nacl - 5 g

Agar-Agar - 15 g

Water - 1000 ml

YMA - Yeast Malt Agar Medium:

Malt - 20 g
Yeast - 2 g
Agar-Agar - 15 g
Water - 1000 ml

SDA - Saboraud's Dextrose Agar Medium:

Dextrose - 40 g

Agar-Agar - 15 g
Peptone - 10 g
Water - 1000 ml

All five media were prepared according to above mentioned composition.

Sterilization of medium:

The flasks having media were sterilized in the autoclave at 15lb/sq. inch pressure for one hour and then poured in 90 mm Petri dishes under the laminar flow hood to avoid contamination. Media were cooled to 37°C. The joint, stalk and veil of the fresh mushrooms were inoculated on culture media.. Radial growth of mycelium of different portions was observed until the Petri dishes were filled with it. The experiment was repeated for 5 times. The plates was incubated at 37°C and observed for 15 days during which the mycelial vegetative growth and mycelial density of *Agrocyba aegerita* were recorded.

The mycelial density was rated as described ²⁰ as follows:

+ = Very Scanty mycelial density

2+ = Scanty mycelial density

3+ = Moderate mycelial density

4+ = Abundant mycelial density

5+ = Very abundant mycelial density

The growth rate is given by the formula below:

Growth rate=Colony diameter on the last day (cm) /Number of day's measurement was taken after inoculation

Daily mycelial growth was determined using a ruler across the Petri-dish horizontally.

RESULTS AND DISCUSSION:

The result in (**Fig; Table 1** and **2**) on the 15th day showed that Agrocyba aegerita had the highest mycelial colony diameter 8.82 ± 0.27 cm, density (5+) and growth rate (1.5cm/day) on MEA media followed by PDA media with 8.00 ± 0.86 cm, 5+ and 1.3 cm/day being colony diameter, mycelial density and growth rate respectively.

On YMA media colony diameter was 7.76 ± 1.46 cm, mycelial density 5+ and growth rate was

E-ISSN: 0975-8232; P-ISSN: 2320-5148

 $1.3\,$ cm/day. Furthermore, the growth on GPA media had $6.78\pm0.11\,$ cm as colony diameter, 4+ as mycelial density and 1.1cm/day as the growth rate. The least growth was recorded on SDA media

was poor with $6.20 \pm 0.38 \text{cm}$ as colony diameter, 3+ as mycelial density and 1.0 cm/day as growth rate.

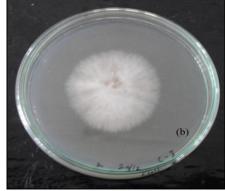
TABLE 1: MYCELIAL DENSITY AND GROWTH RATE OF AGROCYBA AEGERITA IN DIFFERENT MEDIUM

S. No	Medium	Average Mycelial Density On 15 th Day Of Incubation	Growth Rate cm/day			
1.	Saboraud's dextrose agar	3 +	1.0			
2.	Potato dextrose agar	5 +	1.3			
3.	Yeast malt agar	5 +	1.3			
4.	Glucose peptone agar	4 +	1.1			
5.	Malt extract agar	5 +	1.5			

3+ = Moderate, 4+ = Abundant, 5+ = Very abundant



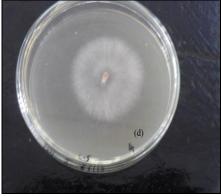
(a) Saboraud's dextrose agar



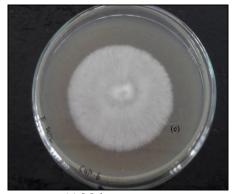
(b) Potato dextrose agar



(c) Yeast malt agar



(d) Glucose peptone agar



(e) Malt extract agar

FIG.1: MYCELIAL GROWTH IN DIFFERENT MEDIA

E-ISSN: 0975-8232; P-ISSN: 2320-5148

TABLE 2: MYCELIAL GROWTH PATTERN OF AGROCYBA AEGERITA IN DIFFERENT MEDIUM

S. No	Medium	Colony diameter in different days (in cm)									
		6 th	$7^{\rm th}$	8 th	9 th	10 th	11 th	12 th	13 th	14 th	15 th
1.	Saboraud's dextrose agar	0.58	1.22	2.06 ±	3.30 ±	3.60 ±	3.90 ±	4.40 ±	4.90 ±	5.50 ±	6.20 ±
		± 0.19	± 0.40	0.39	0.27	0.29	0.31	0.30	0.31	0.24	0.38
2.	Potato dextrose agar	$0.92 \pm$	$1.72 \pm$	$2.32 \pm$	$2.89 \pm$	$3.80 \pm$	$4.70 \pm$	$5.46 \pm$	$6.24 \pm$	$6.94 \pm$	$8.00 \pm$
۷.		0.19	0.36	0.31	0.27	0.25	0.34	0.38	0.24	0.09	0.86
3.	Vanet malt agar	$0.74 \pm$	$1.30 \pm$	$1.78 \pm$	$2.66 \pm$	$3.56 \pm$	$4.54 \pm$	$5.44 \pm$	$6.44 \pm$	$7.14 \pm$	$7.76 \pm$
3.	Yeast malt agar	0.36	0.26	0.31	0.60	0.95	1.44	1.66	1.67	1.56	1.46
4.	Glucose peptone agar	$0.80 \pm$	$1.84 \pm$	$2.36 \pm$	$3.00 \pm$	$3.54 \pm$	$4.04 \pm$	$4.66 \pm$	$5.40 \pm$	$6.16 \pm$	$6.78 \pm$
		0.23	0.21	0.21	0.15	0.21	0.21	0.32	0.37	0.29	0.11
5.	Malt extract agar	$0.90 \pm$	$1.78 \pm$	$2.98 \pm$	$4.14 \pm$	$4.84 \pm$	$5.64 \pm$	$6.50 \pm$	$7.36 \pm$	$8.34 \pm$	$8.82 \pm$
		0.16	0.19	0.46	0.46	0.27	0.34	0.34	0.31	0.38	0.27

Among the five solid growth media, malt extract agar media found to support fastidious growth of mushroom cultures with the presence of higher amount of carbohydrates particularly maltose, protein and other medium. This is followed by potato dextrose agar medium, yeast malt agar medium, saboraud's dextrose agar medium and glucose peptone agar medium. Though the mycelial diameter of the mushroom cultures in saboraud's dextrose agar medium, glucose peptone agar medium were in par with rest of the growth media, the density of the mycelia were low in these two growth media.

Hence, malt extract agar medium was found to be an ideal media for growth of these mushroom cultures. Mycological peptone present in the malt agar rapidly gives a luxuriant growth with typical morphology and pigmentation.

Mycelium is an important part for mushroom production as well as for production of several secondary metabolites used for therapeutic purpose. The mycelium growth depends on several factors such as growth media, pH, temperature, nutrient elements and some environmental factors ²¹. Growth medium is the most important factor because it supplies necessary nutrient for the growth of mushroom mycelium.

Different media such as potato dextrose agar, yeast extract agar, malt extract agar, lamberts agar and compost extract agar are mostly used for the growth of mycelium ²². Mycelium growth is the best tool to identify necessary nutrients for the production of fruiting bodies as mycelium growth requires short time in comparison with fruiting bodies development ²³.

CONCLUSION: Since mushrooms are good source of bioactive compounds of anticancer, antifungal and anti-diabetic in nature, the mycelia may be used for the large scale production of the compounds as mushrooms are seasonal. To make the bioactive production technology cost effective, present study may be useful in order to obtain more biomass ultimately to have bioactive compounds in hand. Further standardization regarding quantification of substrate as nutritional source for biomass production and its cost economics is required to reach more constructive conclusion

ACKNOWLEDGEMENTS: Our sincere gratitude to Tamil Nadu State Council for Science and Technology (TNSCST), Chennai, Tamil Nadu, India (F.No. **TNSCST** S & PROJECT/BS/RJ/2013 - 2014) and Management of Kongunadu Science College Arts and (Autonomous), Coimbatore, TamiNadu, India for the financial support offered to carry out the research work.

REFERENCES:

- Adebayo-Tayo BC, Jonathan SG, Popoola OO & Egbomuche RC: Optimization of growth conditions for myelial yield and exopolysaccharide production by Pleurotus ostreatus cultivated in Nigera. African Journal of Microbiology Research 5: 2011; 2130–2138
- Chang R. Functional properties of edible mushrooms. Nutr Rev1996; 54:S91-S93.
- Chang ST. Global impact of edible and medicinal mushrooms of human welfare in the 21stcentury: nongreen revolution. Int J. Med. Mushrooms 1999; 1:1-7.
- 4. Yen G-C, Hung C-Y. Effects of alkaline and heat treatment on antioxidative activity and total phenolics of extracts from Hsiantsao MesonaprocumbensHemsl.). Food Res Int 2000; 33:487-92.
- Mau JL, Lin HC, Song SF. Antioxidant properties of several specialitymushrooms. Food Res Int 2002; 35:519-26.

E-ISSN: 0975-8232; P-ISSN: 2320-5148

- Yang J-H, Lin H-C, Mau J-L. Antioxidant properties of several commercial mushrooms. Food Chem 2002; 77:229-35.
- Cheung LM, Cheung PCK, Ooi VEC. Antioxidant activity and total phenolics of edible mushroom extracts. Food Chem 2003; 81:249-55.
- 8. Cheung LM, Cheung PCK. Mushroom extracts with antioxidant activity against lipid peroxidation. Food Chem 2005; 89:403-9.
- Lo KM, Cheung PCK. Antioxidant activity of extracts from the fruiting bodies of Agrocybeaegeritavar. alba. Food Chem2005; 89:533-9.
- 10. Ferreira ICFR, Baptista P, Vilas-Boas M, Barros L. Freeradical scavenging capacity and reducing power of wild edible mushrooms from northeast Portugal: individual cap and stipe activity. Food Chem 2007; 100:1511-6.
- Barros L, Calhelha RC, Vaz JA, Ferreira ICFR, Baptista P, EstevinhoLM. Antimicrobial activity and bioactive compounds of Portuguese wild edible mushrooms methanolic extracts. EurFood Res Technol 2006. In press.
- Kapoor JN: Introduction. Mushroom cultivation. Published by Indian Council of Agricultural Research, New Delhi. 1989; 1: 1-6.
- Jong SC and Birgmingham JM: Mushroom as a source of natural flavor and aroma compounds. In: Chang ST, Buswell JA & Chiu W (eds) Mushroom Biology & Mushroom products. The Chinese University press, Hong Kong, 1993; pp. 345–366.
- Maziero R, Adami A, Cavazzoni V & Bononi VL: Exopolysaccharide and biomass production in submerged

- culture by edible mushrooms. Mushroom Science 1995;14: 887–892.
- Moore D & Chiu SW: Fungal products as food. In: Pointing SB & Hyde KD (eds) Bioexploitation of filamentous fungi. Fungal Diversity press, Hong Kong, 2001; pp. 223–251.
- Fermor TR & Wood DA: The microbiology and enzymology of wheat strawmushroom compost production. In Straw Decay and its Effect on Disposal and Utilization (ed. E. Grossbard), 1979; pp. 105-112. Wiley, Chichester, U.K.
- 17. Thakur K; Improvement of edible mushrooms. In: Advances in Horticulture. New Delhi, India.1995
- Chang ST, Lau OW, Cho KY. 1998 The cultivation and nutritive value of Pleurotus sajor-caju. European Journal Applied Microbiology and Biotechnology 1981(12), 58–62
- 19. Qei P: Manual on Mushroom Cultivation Tool Foundation Amsterdam, 1992; pp: 42-50.21.
- Kadiri M: Spawn and fruit body production of Pleurotus sajor-caju in Abeokuta , Nigeria, Niger. J. Bot 1998; 11: 125-131
- 21. Calam CT: The evaluation of mycelial growth. In: Norris J.R. and Ribbons D.W.(eds). Methods in Microbiology. Academic press, New York Vol. 1, 1971; pp. 567-591.
- 22. Pathak VN, Yadav N, Gour M. Mushroom production and processing technology. Agrobotanica. 28. 1998.
- Kalmis E and Kalyoncu. Variations in the Isolate obtained from Basidiospores of Commercial Mushroom Lentinusedodes (Shiitake). International J. Sci. & Tech. 2006; 1:99-103.

How to cite this article:

Muthu N and Shanmugasundaram K: Effect of Five Different Culture Media on Mycelial Growth of *Agrocybe Aegerita*. Int J Pharm Sci Res 2015; 6(12): 5193-97.doi: 10.13040/IJPSR.0975-8232.6(12).5193-97.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)