



Received on 10 October, 2013; received in revised form, 29 November, 2013; accepted, 10 February, 2014; published 01 March, 2014

IMPACT OF ALCOHOL ON RAT HEART AND AORTIC TISSUE ACETYLCHOLINE CONTENT LEVELS *IN VITRO* AND *IN VIVO*

M. Yugandhar*, M. Rajeswara Rao

Department of Zoology, S.V. University, Tirupati – 517502, Andhra Pradesh, India

Keywords:

Acetylcholine, Alcohol, Aortic Tissue, Heart

Correspondence to Author:

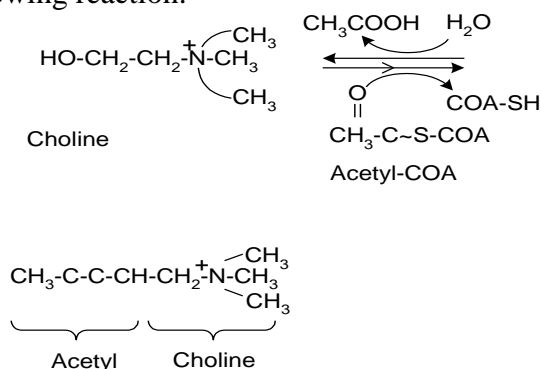
Dr. M. Yugandhar

C/o Prof. M. Rajeswara Rao,
Department of Zoology, S.V.
University, Tirupati – 517502,
Andhra Pradesh, India

E-mail: dryugandar.sv@gmail.com

ABSTRACT: Acetylcholine satisfies all criteria for a transmitter substance. Its most widely recognized action sites are the endings of vertebrate motor nerves, the endings of the automatic pre ganglionics and the parasympathetic post ganglionics. It appears also to be the transmitter of motor nerves in all advanced invertebrate animals except the arthropods where it seems to separate at the endings of sensory nerves. Acetylcholine is concerned with various visceral functions in both invertebrates and vertebrates. The impact of 20% ethanol (w/v) at selected doses of 5gm/kg wt over 5 and 10 weeks *in vivo* and 100-1000 μ l *in vitro* on the heart and aortic tissue acetylcholine content activity was reported. Alcohol is selected doses appeared to inhibit the rat and aortic tissue based Ach levels both *in vitro* and *in vivo*.

INTRODUCTION: The acetylation of the nitrogenous alcohol choline gives rise to acetylcholine, now recognized as neurotransmitter in all major groups of animals. The biosynthesis is catalyzed by choline acetylase and readily reversed by acetyl cholinesterase as indicated in the following reaction.



Claude Bernard launched the history of transmitter pharmacology in 1857, when he discovered that nerve stimulation failed to excite muscles that were poisoned with a plant alkaloid, curare, even though the muscle remained responsive to direct stimulation. Bernard Correctly concluded that this substance blocked excitatory events at the neuromuscular junction. It is now known that curare acts by competing with Ach for receptor sites on the motor end plate, with highly purified preparations of curare (d-tubocurarine) muscular excitability is gradually reduced in accordance with the dose; thus, this substance has found a valuable place in clinical medicine as well as physiology.

Effects on basal or stimulated Ach release have been described in mammalian brain^{1, 2, 3, 4, 5, 6}, *Aplysia ganglios*^{7, 8} and smooth muscle^{9, 10, 11, 12, 13}, through prejunctional or retrograde actions, other concern the physiological role of NO on the skeletal muscle of the adult neuromuscular junction^{14, 15, 16, 17, 18}, on its development¹⁹ or in pathological situations.

	QUICK RESPONSE CODE DOI: 10.13040/IJPSR.0975-8232.5(3).892-95
	Article can be accessed online on: www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.5(3).892-95	

Likewise NO and Ach are interrelated with regard to their physiological functioning^{20,21}.

MATERIAL AND METHODS: Albino rats of the weight range 125 ± 5 gm were selected for the current study. Animals were maintained at a constant temperature of $15 \pm 5^\circ\text{C}$. The rats were divided into three groups of seven each and were maintained in separate cages. They were fed *ad libitum* with commercial rat feed supplied by Kamadhenu Agencies, Bangalore, India.

Treatment of Animals: For *in vitro* studies 20% alcohol 100-1000 μl was used. For *in vivo* studies group II rats were gavaged with 5g/kg body of alcohol 20% (w/v) over 5 weeks (weekly doses). Group III rats were gavaged with 5gm/kg body weight of alcohol over 10 weeks (weekly doses) and group I animals acted as normal control ones. After treatment of animals, they were anesthetized with chloroform and were dissected. The tissues (heart and Aortic arches – cleared) were isolated sliced weighted and were transferred into clean test tubes. The tubes were kept in boiling water bath for

5 minutes to inactivate the acetyl cholinesterase (AChE) enzyme activity and to release bound Ach. The study of by Ach content was estimated by the method of Hestrin as described²².

Statistical Analysis: For each parameter, the mean of individual observation (for both control and experimental groups were taken into consideration). Statistical significance of the data was analyzed through two was ANOVA (analysis of Variance); SNK (Student – Newman - Keuls) test and regression analysis.

RESULTS: The data in **table 1 & 2** shows the trends obtained for Ach levels of the heart and aortic tissues under selected doses of alcoholic stress. Alcohol appeared to inhibit the heart and aortic tissues Ach levels *in vitro* and *in vivo* and the changes were found to be statistically significant ($P < 0.001$) for 500 μl and 1000 μl of 20% alcohol (w/v) concentration in vitro and 5g/kg wt over 5 and 10 week periods treated rat tissues *in vivo* and the percent depletion was found to be more for the higher doses of alcohol employed in this study.

TABLE 1: IMPACT OF ALCOHOL ON RAT HEART AND ACETYLCHOLINE CONTENT LEVELS *IN VITRO*

Name of the tissue / cellular preparation	Ach levels			
	Control	Control + 100 μl of 20% alcohol	Control + 500 μl of 20% alcohol	Control + 1000 μl of 20% alcohol
Heart soluble fractions				
A V	0.1630	1.157	0.827	0.590
S D	0.81	0.280	0.075	0.082
P C		-29.010	-49.255	-63.084
t-test		*	*	*
Endothelial cellular soluble fractions				
A V	1.217	1.493	1.797	2.577
S D	0.085	0.140	0.106	0.290
P C		-22.653	-47.653	-11.737
t-test		*	*	*

(Values expressed as μmol Ach/gm wet wt of tissue). AV: average; SD: Standard deviation; PC: Percent Change over control; NS: Not Significant; * $p < 0.001$

TABLE: 2 IMPACT OF ALCOHOL ON RAT HEART AND ACETYLCHOLINE CONTENT LEVELS *IN VIVO*

Name of the tissue / cellular preparation	Ach levels		
	Control	Alcohol Treated ones	
		5 weeks	10 weeks
Heart soluble fractions			
A V	1.834	1.621	1.643
S D	0.056	0.044	0.091
P C		-11.604	-10.436
t-test		*	*
Endothelial cellular soluble fractions			
A V	1.651	1.763	2.546
S D	0.058	0.120	0.075
P C		-30.129	-35.129
t-test		*	*

(Values expressed as μ mol Ach/gm wet wt of tissue). Each value is the mean \pm SD of 7 samples. AV: Average; SD; Standard variation; PC: Percent change over the control; * P <:0.01

DISCUSSION: They² have cited that alcohol alters the Ach levels and Ach further evoke significant membrane depolarization in rat major pelvic ganglion. Ach evoking large long-term effects on NMDA receptors in rats has been reported by⁴. They³ studied the effect of ethyl alcohol on the contractile activities of canine gastric corpus and fundus circular muscles and they reported that alcohol alters the normal functioning of Ach²³, showed ethanol as to modulate the receptor c-protein function in the rat cerebral cortex. They⁷ have studied the biphasic effect of alcohol activity on membrane associated AchE activity their studies on alcohol versus *Torpeda californica* support impaired activity of the key enzyme AchE that regulates Ach levels by alcohol, Alcohol developing pathological symptoms in experimental models with reference to Ach was also worked out by²⁴. All these reports clearly depict that alcohol impairs the normal functioning of Ach in different experimental models.

In the current study, depletion in the rat heart and aortic tissues *in vitro* and *in vivo* Ach levels were observed under alcoholic stress shown in tab (1&2). The changes observed were perfectly in a dose and time – dependent manner. As the Ach binding to its Ach receptor on ET cells accelerate intracellular Ca²⁺ levels which in turn regulates NO release in SMC^{25,26}. The lowered levels of Ach in the heart and aortic tissues by alcohol in the present investigation may not be enough to stimulate its receptors and this may impart lead to impairment of NO production in the cardiovascular system (Due to higher costs of assay kits receptor binding studies were not under taken in the present study).

REFERENCES:

1. Yugandhar *et al.* Impact of alcohol on the cardiovascular nitric oxide pathway in rats.
2. Prast, H. and Philippu, A. Nitric Oxide releases acetylcholine in the basal forebrain. *Eur. J. Pharmacol.* 1992; 216: 139-140.
3. Guevara – Guzman, R., Emson, P.C. and Kendrick, K.M. Modulation of *in vivo* striatal transmitter release by nitric oxide and cyclic GMP. *J. Neurochem.* 1994; 62: 807-810.
4. Ohkuma, S., Katsura, M., Chen, D.Z., Guo, J.L. and Kuriyama, K. c. Hydroxyl radical scavengers enhance nitric oxide-evoked acetylcholine release from mouse cortical neurons. *Mol.Brain. Res.* 1995; 34: 347-350.
5. Ohkuma, S. Katsura M., Guo, J.L., Hasegawa, T. and Kuriyama, K.1995b. involvement of peroxynitrite in N-

- methyl-D-aspartate and sodium nitroprusside-induced release of acetylcholine from mouse cerebral cortical neurons. *Mol. Brain. Res.*, 31: 185-193.
6. Ohkuma, S., Katsura, M., Chen, D.Z., Guo, J.L. and Kuriyama, K. 1995c. Hydroxyl radical scavengers enhance nitric oxide-evoked acetylcholine release from mouse cortical neurons. *Mol.Brain. Res.* 34: 347-350.
7. Meulemans, A., Mothet, J.P., Schirar, A., Fossier, P., Tauc, L. and Baux, G.. A nitric oxide synthase activity is involved in the modulation of acetylcholine release in *Aplysia* ganglion neurons. A histological voltammetric and electro-physiological study. *Neuroscience.* 1995; 69: 985-995.
8. Mothet, J.P., Fossier, P., Tauc, L. and Baux, G. Opposite actions of nitric oxide on cholinergic synapses: which pathways? *Proc. Natl. Acad. Sci., USA.* 1996; 93: 8721-8726.
9. Belvisi, M.G., Stretton, D. and Barnes, P.J. Nitric Oxide as an endogenous modulator of cholinergic neurotransmission in guineapig airways. *Eur.J.Pharmacol.* 1991; 198: 219-221.
10. Sekizawa, K., Fukushima, T., Ikarashi, Y., Maruyama Y. and Sasaki, H. The role of nitric oxide in cholinergic neurotransmission in rat trachea. *Br.J. Pharmacol.* 1993; 110: 816-820.
11. Wiklund, C.U., Olgard, C., Wiklund, P. and Gustafsson, L.E. Modulation of cholinergic and substance P-like neurotransmission by nitric oxide in the guinea-pig ileum. *Br.J.pharmacol.* 1993; 110: 833-839.
12. Baccari, M.A., Bertini, M. and Calamai, F. Effects of L-Ng-nitroarginine on cholinergic transmission in the gastric muscle of the rabbit. *NeuroReport.* 1993; 4: 1102-1104.
13. Kilbinger, H. and Wolf, D. Increase by NO synthase inhibitors of acetylcholine release from guinea-pig myenteric plexus. *Naunyn-Schmideberg's Arch. Pharmacol.* 1994; 349: 543-545.
14. Nakana, M., Schmidt, H.H.H.W., Pollock, J.S., Forstermann, U. and Murad, F. Cloned human brain nitric oxide synthase is highly expressed in skeletal muscle. *FEBS. Lett.*1993; 316: 175-180.
15. Kobzik, L., Reid, M.B., Bredt, D.S. and Stamler, J.S. Nitric Oxide in skeletal muscle. *Nature.* 1994; 372: 546-548.
16. Lindgren, C.A. and Laird, M.V. Nitroprusside inhibits neurotransmitter release at the frog neuromuscular junction. *NeuroReport.* 1994; 5: 2205-2208.
17. Kusner, L.L. and Kamiski, H.J. Nitric Oxide synthase is concentrated at the skeletal muscle endplate. *Brain Res.*, 1996; 730: 238-242.
18. Oliver, L., Goureau, O., Courtois, Y, and Vigny, M.. Accumulation of NO synthase (type-1) at the neuromuscular junctions in adult mice. *NeuroReport.* 1996; 7: 924-926.
19. Wang, T., Xie, Z. and LUB. Nitric oxide mediates activity dependent synaptic suppression at developing neuromuscular synapses. *Nature.* 1995; 374: 262-266.
20. Brenman, J.E., Chao, D.S., Xia, H., Aldape, K. and Bredt, D.S. 1995. Nitric oxide synthase complexed with dystrophin and absent from skeletal muscle Sarcolemma in Duchenne muscular dystrophy. *Cell.* 82: 743-752.
21. Brenman, J.E., Chao, D.S., Gee, S.H., Mc Gee, A.W., Gravan, S.E., Santillano, D.R., Wu, Z., Huang, F., Xia, H., Peters, M.F., Frochner, S.C. and Bredt, D.S. 1996. Interaction of nitric oxide synthase with the postsynaptic

- density protein PSD-95 and alpha-syntrophin mediated by PDZ domains. Cell. 84:757-767.
22. Augustinsson, K.B. In: Methods of biochemical analysis, Vol.5 (ed. Glick, D). Inter Science Publishers Inc. 1957 New York., USA. p.1.
 23. Singh S.P., Handa R.K., Depala V, Gaoy, MCILLORY, P.J. Ravindra R. The effect of ethanol on muscarinic receptor – Gprotein Coupling in the rat cortex. 1997; Vol 81: 294 – 299.
 24. Erikson C.J, The role of acetaldehyde in the actions of alcohol, 2000; vol.25, 153-325.
 25. Loweinstein CJ, Glatt CS, Bredt DS and Synder. Cloned and express macrophase nitric oxide synthase contrasts with the brain enzyme. Proceedings of the National Academy of Science of the USA; 1994; 6711-6715.
 26. Shinde, U.A., Mehta, A.A and Goyal, R.K. Nitric Oxide: a molecule of millennium. Ind. J. Ext. Biology., 2000; 38; 201-210.

How to cite this article:

Yugandhar M and Rao MR: Impact of alcohol on rat heart and aortic tissue acetylcholine content levels *in vitro* and *in vivo*. *Int J Pharm Sci Res* 2014; 5(3): 892-95.doi: 10.13040/IJPSR.0975-8232.5(3).892-95

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)