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SYNERGISM BETWEEN CASSIA FISTULA AND FLUCONAZOLE; "A NEW APPROACH AGAINST CANDIDIASIS"

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ABSTRACT: Natural products have great potential against microbes and can be used to overcome the problem of drug resistant pathogens. They are helpful in reducing the toxicity of existing antifungal compounds. This present in vitro study demonstrated the synergistic effectiveness of Cassia fistula (CF) leaves extracts and fluconazole (Flu) against isolated Flu resistant strains (FRSs) of Candida albicans (CA) as the growth of FRS was found to be sensitive to extracts of CF leaves. In the present study CF leaves' extracts were prepared in different solvents and their IC₅₀ values were estimated by micro dilution method. It was found a statistical significant decrease in concentration of cellular lipids, phospholipid of FRS when treated against the CF leaves or combination of CF leaves and Flu. Further, H^3 uptake analysis revealed that both methods affected the DNA synthesis as well as decrease in DNA contents of FRS. Hence, the results of this study would be significant for the development of new anti candidal drugs from CF leaves' extracts in combination with known synthetic drugs like Flu to control drug resistant CA.

INTRODUCTION: The pace of drug discovery has increased with the introduction of new targets and new inhibitors of these targets being reported each month ¹. Several authors have several studies on anti-microbial properties of natural extracts and natural products ^{2, 3, 4}. Due to the difficulty in identifying drug targets unique to fungi that are not shared with the human host, only restricted number of antifungal agents has been widely used for treating candidial systemic infections ⁵.



Fluconazole (Flu) is the most common azole drug that target lanosterol 14- α –demethylase (Erg 11) in the ergosterol biosynthesis pathway and used as antifungal agent for prevention and treatment of candidiasis ⁶. Prolonged use of above drug in recent years has contributed to the development of Flu resistance in candida species ⁷.

For this reason it has now become very important to reduce the Flu concentration by different combination methods. On this study, we assess the synergism between Flu with CF (*Cassia fistula*). CF is called as 'An Indian Laburnum', also known as the golden shower tree and is a flowering plant belongs to the family Fabaceae⁸. The species is native to the Indian Subcontinent and adjacent regions of Southeast Asia and has antifungal

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properties ⁹, hepato protective activity ¹⁰, hypo cholesteroleamic activity ¹¹, hypo glucamic activity ¹², anti plasmodial activity ¹³ and anti-tumour activity ¹⁴. Therefore, in the present study, we proposed to study the antifungal effect of CF leaves extracts in different solvents and Flu against isolated fluconazole resistant strains (FRS) of *Candida albicans* (CA). We also evaluated the intracellular changes made by CF leaves extract on isolated resistant strains CA.

MATERIALS AND METHODS: The present study has been designed to evaluate the antifungal action of *Cassia fistula* (CF) leaves extracts and commercially available drug fluconazole (Flu) on clinical isolates of fluconazole resistant strains (FRS) of *Candida albicans* (CA). The study protocol was approved by local ethical committee. The written informed consent was obtained from patients before their enrolment in the study.

Collection of plant material: CF leaves were collected from HNB Garhwal University, Srinagar (Garhwal), Uttarakhand, INDIA. This plant was identified and confirmed by Botanical Survey of India, Dehradun (BSD 112753). The leaves of the same were obtained and stored under refrigerated conditions till use.

Preparation of leaf extract: The extract was prepared by grinding leaves in solvents like Methanol, Ethanol, Dimethyl sulphoxide and Petroleum ether (wt/vol). This mixture was than centrifuged at 10000 rpm at 4°C for 10 min. The supernatant, thus obtained was used within four hours for estimation of anti-fungal activity.

Patient selection: For the isolation of flu resistant candidal species eighty samples were collected from eighty oral candidiasis patients during Aug 2012 to April 2013. Selected patients had not received any antibiotic or antifungal therapy (15 days prior to sampling) and any surgery, chemotherapy, radiotherapy or immune suppressive therapy (one year prior to sampling). Patients wearing intra oral prosthesis were also excluded from the research.

Sample collection: Required samples were obtained from the patients through oral rinse method ¹⁵. Patients were asked to rinse the mouth with 10ml of normal saline for 60 sec. Then, they

were asked to spit the oral rinse in a sterile container. The samples which could not be sent immediately to laboratory were refrigerated. $25 \ \mu$ l of oral rinse was inoculated on Sabaroud's agar plate (SDA) with in 8 hrs of collection and incubated at 37°C for 48-72 hrs.

Isolation of identifications of fungal species: Preliminary identification was done by Gram staining ¹⁵ and Gram positive fungal colonies were sub cultured on SDA plate. The colonies were further subjected to various confirmatory tests as per standard procedure like Lacto phenol cotton blue preparation, Germ tube test, Corn meal Tween 80, Agar culture test, carbohydrate fermentation test and slide culture ^{16, 17, 18, 19}.

Growth of isolated Candida species was maintained on Sabaroud's agar slant (Glucose 40 g/L, Peptone 10g/L, Agar – agar 20 g/L) and SDA agar slat was prepared by standardize procedure ¹⁹.

Assessment of the MIC and cellular changes: The minimum inhibitory concentration and 50% inhibitory concentration (IC₅₀) values of the fungal strains under study were determined by the micro dilution methods in accordance with the CLSI M7 – A2 protocol ²⁰. MIC was defined as the lowest concentration that did not result in any visible growth of the yeast compared with the growth in the control well. Further, intracellular changes in selected fluconazole resistant strain were studied via, total lipid ²¹, phospholipids, ergosterol, H³ uptake studies ²².

Statistics analysis: All the data provided in the study represents means \pm standard error of the mean (S.E.M.). For all other analyses ANOVA was used (p<0.05).

RESULTS AND DISCUSSION:

Isolation of Candida species: The study samples (n=80) were collected from 80 subjects with a mean age of 45.3 ± 8.01 . In our study, Candidial species were preliminary identified by Gram staining and then confirmed by Germ tube, Carbohydrate fermentation and colony formation. Our study observed that 77.77% of samples from patients (suffering from Candidiasis, Xerostomia and Mouth ulcer) showed prominent growth of CA (**Table 1**).

TABLE 1: DISTRIBUTION OF VARIOUS FUNGAL SPECIES IN ISOLATED SAMPLES:-

S. no.	Name of fungal species	No. of colonies	%age	Confirmatory test
1	*C. albicans	35	77.77	Germ tube, **CF, CM
2	C. glabrata	3	5.88	CF and ¶CM
3	C. krusei	2	3.92	CF and CM
4	C. pseudotropicalis	2	3.92	CF and CM
5	C. tropicalis	1	1.96	CF and CM
6	Penicillium marneffei	1	1.96	¥Cm and Pigment production
7	Chaetomium	1	1.96	Cm & LCB preparation

*C- Candida, **CF- Carbohydrate fermentation, ¶CM-Corn Meal and ¥Cm- Colony morphology

Effect of CF leaves extract on isolated strains: Antifungal activity was checked by micro dilution method using isolated strains of *C. albicans*, *C. glabrata*, *C. krusei*, *C. pseudotropicalis* and *C. tropicalis*. For antifungal activity, fresh leaves of CF were grind with DMSO, pet ether, methanol and ethanol in concentration dependent manner (wt/vol). It was observed that DMSO and pet ether leaves extract has very potent anti-fungal action against isolates of *C.albicans* and *C.tropicalis* with IC₅₀ of 0.8 mg/ml and 0.6 mg/ml respectively (**Table 2**).

Synergistic effect of CF leaves extract and fluconazole on Flu resistant strains of CA: A phenomenon referred to in this context as drug tolerance was observed with six isolated strains showed resistance against Flu and named as fluconazole resistance strains (FRS). The IC⁵⁰ value of these six FRS was found to be in range form 35- 58 μ g/ml (Flu). To further define whether Flu and CF leaves extract work synergistically, six FRS were examined under different combinations of Flu plus CF leaves extracts (at their IC₅₀) as depicted in **table 3**. It was found that cells of FRS that were exposed to Flu (52 μ g/ml) were not killed

during the overnight incubation (16hrs). On further, cells of FRS expose to combination treatment (Flu plus CF leaves), they did not grow up to the same density as that of control. Results of table 3 reveal that at least 75-85% growth inhibition of CA was observed even at 1/4 IC₅₀ of both drugs used in combination. It infers that both drugs are acting synergistically. Actively growing CA cells were killed rapidly and no viable cells among 5×10^5 cells were detected after 2 h. of expose to drugs in combination. Similarly, K. Amber et al 2010 showed that essential oil of Ocimum sanctum is an effective antifungal agent that inhibits Flu sensitive candida species (C.albicans, C. tropicalis, C. parapsilosis)but also Flu -resistant C. albicans, C.tropicalis isolates and C. krusei and C. glabrata which are intrinsically resistant to Flu or whose resistance is easily inducible and combination Flu/ketoconazole- Ocimum sanctum essential oil administered against the fungal strains under consideration is likely to augment the efficiency of these two azoles ²³. It is always true that basis of drug interactions are potentially multifactorial and complex.

 TABLE 2: COMPARISON OF INHIBITION ACTIVITY OF CF LEAVES EXTRACT (IN DIFFERENT SOLVENTS)

 AGAINST ISOLATED CANDIDA SPECIES WITH THEIR IC₅₀ VALVES:

Ι	Leaves Extract	C. albicans (29*)	C. glabrata (3*)	C. kruseii (2*)	C. pseudotropicalis	C. tropicalis
1.	DMSO (mg/ml)	0.8 ± 0.06	2.8±0.65	4±0.23	1.6 ± 0.034	1.1±0.024
2.	Pet. Ether (mg/ml)	1±0.021	3.1±0.39	4.8 ± 0.14	0.8 ± 0.039	0.6 ± 0.017
3.	Methanol (mg/ml)	1.8 ± 0.083	3.8±0.58	5 ± 0.48	1.7±0.19	2.6±0.049
4.	Ethanol (mg/ml)	2±0.15	3.9±0.46	5±0.17	1.9 ± 0.28	2.8 ± 0.068
5.	Flu (µg/ml)	0.1 ± 0.024	1.8±0.26	14±0.34	0.4 ± 0.054	0.4 ± 0.15

Note -Values are mean of 8 replicates (IC₅₀ \pm SD valves), * = the number of isolates used for IC₅₀ calculation, IC₅₀- 50% inhibitory concentration of leaves extract & Flu.

Synergistic effect of CF leaves extract and fluconazole on intracellular lipids of FRS B: Further, intracellular changes were studied by estimating the concentrations of ergosterol, phospholipid and total lipid of FRS B cells (shown in **table 4**) because this strain has highest value of $IC_{50}i.e$ 52µg/ml (Table 2). These cells were grown in presence of IC_{50} of Flu, CF^{DMSO} and $CF^{Pet ether}$ extract and showed significant reduction in concentrations of total lipid, ergosterol and phopholipids. But the phospholipid and ergosterol contents were found to be statistically decreased with FRS-B were grown in combination (1/4 IC_{50} of Flu plus CF^{DMSO} and ¹/₄ IC_{50} of Flu plus CF^{PMSO}

ether). It was also observed that phospholipid and ergosterol content was 1.33 and 1.51 times decreased in FRS grown in the presence of CF^{DMSO} , Flu + CF^{DMSO} . But this effect was more pronounced (1.58 and 1.75 times) when FRSs were grown in combination of Flu + CF Pet. ether. The decreased content of lipid was because of reduced rate of synthesis of these lipids. The correlation between loss of membrane permeability and leakage of intracellular DNA studies indicate that CF kills CA by damaging the cytoplasmic membrane, leading to an impairment of all cellular functions²⁴. Similar observation by Irshad et al 2011 showed that C. fistula fruit pulp and seed extract possessed anticandidal activity. Therefore, the result of our study was significantly correlated between the MICs and ergosterol inhibition. It was concluded that the leaves extract is a promising source for anticandidal compounds. Sometimes compounds present in plant extract are responsible for their partition into the lipid bilaver of the cell

membrane, leading to an alteration of permeability and a consequent leakage of cell contents ^{3, 4, 8, 25}.

Synergistic effect of CF leaves extract and fluconazole on DNA synthesis of FRS B: To further elucidate the interaction of leaves extracts and to find out synergistic behaviour of leaves extract with the nuclear events, DNA content was measured in the isolated nuclei. Nuclear fraction was found to be significantly enriched with DNA indicative of its purity. DNA content also decreased markedly in cells grown in the presence of CF^{DMSO}. Flu + CF^{Pet. ether}, CF^{Pet ether} and Flu + CF^{Pet. ether} (shown in fig. 1). These finding were further substantiated by H^3 thymidine uptake studies, H^3 thymidine was decreased during all time period. Moreover, at 30 min significantly decline in the DNA synthesis was observed with cells grown in combination of Flu plus CF^{DMSO} (as shown on fig. 1).

	FRS 1	FRS 2	FRS 3	FRS 4	FRS 5	FRS 6	
Patient (Age/Sex/Area)	32/F/ § SU	44/M/SU	49/M/SU	37/F/SU	34/M/SU	52/M/SU	
Oral Manifestation	*C	С	**U, C	U,C	U,C, $\pm X$	U,C, ±X	
CFU/mL	1674 ± 218	5920 ± 580	5388 ± 690	5448 ± 430	Confluent	Confluent	
Type of colonies	One	Two	One	One	Three	Two	
Gram staining	Р	P for both	Р	Р	N for one	P for both	
Grem tube	Р	P for one	Р	Р	P for one	P for one	
Clinically isolated	Clinically isolated fungal strains were confirmed by Carbohydrate fermentation test and Corn meal test						
Organism	¶CA	CA and † CG	CA	CA	CA	CA and †† CP	
Flu IC ₅₀ (μg/mL)	52 ± 2.8	35±1.2	40±1.5	48 ± 5.0	58±2.7	40±1.5	
CF^{DMSO} IC ₅₀ (mg/mL)	$8.0{\pm}0.6$	8.6±0.9	$10.0{\pm}1.1$	$8.0{\pm}1.2$	8.0 ± 0.8	12±1.2	
$CF^{Pet ether} IC_{50} (mg/mL)$	$9.0{\pm}0.9$	11.0 ± 0.7	13.0±0.9	9.0±1.1	10.0 ± 0.9	10.0±0.9	
Percentage growth inhibition of isolated fungal strains grown under combination of drugs and CF leaves extract in different solvents							
Flu $IC_{50} + CF^{DMSO} IC_{50}(X)$	98	95	98	95	99	98	
¹⁄₂(X)	90	92	92	90	92	95	
1⁄4(X)	70	80	78	80	79	77	
Flu $IC_{50} + CF^{Pet ether} IC_{50}(Y)$	92	98	98	90	99	90	
½ (Y)	82	85	85	85	87	85	
¹ ⁄4(Y)	74	80	75	78	75	77	

TABLE 3: SELECTED PATIENTS WITH ISOLATED FLU RESISTANT STRAIN (FRS)

*- Candidiasis, **U- Ulcer, N-Negative, P-Positive, ±X- Xerostomia, ¶ CA-*Candida albicans*, †-CG *Candida glabrata*, ††CP-*Candida pseudotropicalis*, § SU- Sub Urban

TABLE 4: TOTAL LIPID, PHOSPHOLIPID AND ERGOSTEROL CONTENT OF ISOLATED STRAIN GR	OWN IN
THE PRESENCE OF IC ⁵⁰ OF CF EXTRACT, FLU AND THEIR COMBINATIONS	

FRS 2 Cells	Total lipid	Phospholipid	Percentage of Phospholipid	Ergosterol	Percentage of Ergosterol
Control	43.6 ± 1.8	15.8±1.2	36.2	20.8±3.2	47.7
Flu IC ₅₀	28.1±1.9	9.7±0.6	34.5	8.6 ± 1.2	30.5
CF DMSO IC ₅₀	21.1±1.8	7.1 ± 0.8	33.6	7.1±0.5	21.1
CF DMSO IC ₅₀	23.2±0.7	5.9 ± 0.7	25.4	11.2±0.8	48.2
1⁄4 (X)	15.1±0.8	3.8±0.5	25.1	2.1±0.5	13.9
¼ (Y)	13.1±1.9	2.1±0.4	16.0	3.6±0.8	27.4

* Valves are expressed in mg/g of dry wt. \pm S.D, Values are mean of 8 replicates.



FIG. 1: ³H THYMIDINE UPTAKE IN CELLS INCUBATED WITH 20µm Flu, IC₅₀ of CF^{DMSO}, 1/4 (X) AND IC₅₀ of CF^{Pet Ether}, 1/4 (Y)

CONCLUSION: This research work has shown that the DMSO leaves extract of CF has potent antifungal action and this action was much more pronounced with fluconazole. Combination of both drugs induced some intracellular changes that may be responsible for reduced growth of fluconazole resistant CA.

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REFERENCES:

- 1. Akins RA. An update on antifungal targets and mechanisms of resistance in *Candida albicans*. Med. Mycol. 2005; 43: 285-318.
- Avato P, Bucci R, Tava A, Vitali C, Rosato A, Baily Z, Jurzysta M. Anti-microbial activity of saponins from Medicago spp. Structure activity relationship. Phytother. Res. 2006; 20(6): 454-57.
- 3. Burt S. Esssential oils: their antibacterial properties and potential applications in foods a review. Int. J. Food Microbiol. 2004; 94: 223-53.
- Cristani M, D'Arrigo M, Mandalari G, Castelli F, Sarpietro MG. Interaction of four monoterpenes contained in essential oils with model membranes 'Implications for their antibacterial activity. J. Agric Food Chem. 2007; 55: 6300-08.

- Sanglard D, Odds FC. Resistance to Candida species to antifungal agents molecular mechanisms and clinical consequences. Lancet Infect. Dis. 2002; 2: 73-85.
- 6. White TC, Bowden RA, Marr KA. Clinical, cellular, and molecular factors that contribute to antifungal drug resistance. Clin. Microbiol. Rev. 1998; 11: 382–402.
- 7. Sanglard D, Ischer F, Parkinson T, Falconer D, Bille J. *Candida albicans* mutations in the ergosterol biosynthetic pathway and resistance to several antifungal agents. Antimicrob. Agents chemother. 2003; 47: 2404-12.
- Mishra MP, Padhy RN. In Vitro antibacterial efficacy of 21 Indian timber-yielding plants against multidrugresistant bacteria causing urinary tract infection. Osong. Public Health Res. Perspect. 2013; 4(6):347-57.
- Irshad M, Ahmad A, Zafaryab M, Ahmad F, Manzoor N, Singh M, Rizvi MM. Composition of *Cassia fistula* oil and its antifungal activity by disrupting ergosterol biosynthesis. Nat. Prod. Commun. 2013; 8(2): 261-64.
- 10. Bhakta T, Banerje S,Subhash C. Hepatoprotective activity of *Cassia fistula* leaf extract. Phytomedicine 2001; 8(3): 220-24.
- 11. El-Saadany SS, el-Massry RA, Labib SM, Sitohy MZ. The biochemical role and hypocholesteroleamic potential of legume *Casia fistula* in hyperchlesterolaemic rats. Nahrung. 1991; 35: 807-15.
- 12. Esposito Avella M, Dias A, deGracia I, de Tello R, Gupta MP. Evaluation of tradition medicine: effects of *Cajanus cajan* L and *Cassia fistula* L on carbohydrate metabolism in mice. Rev Med Panama. 1991; 16: 39-45.
- Grace MH, Lategan C, Graziose R, Smith PJ, Raskin I, Lila MA. Antiplasmodial activity of the ethnobotanical plant *Cassia fistula*. Nat. Prod. Commun. 2012; 7(10): 1263-66.
- Gupta M, Mazumder UK, Rath N, Mukhopadhyay DK. Antitumour activity of methanolic extract of *Cassia fistula L* seeds against Ehrlich ascites carcinoma. J. ethnopharmacol 2000; 72: 151-56.

- 15. Samaranayake LP, MacFarlane TW, Lamey P-J, and Ferguson MM. A comparison of oral rinse and imprint sampling techniques for the detection of yeast, coliform and taphylococcus aureus carriage in the oral cavity. Journal of Oral Pathology 1986; 15: 7, 386–388.
- 16. Sonal S, Ankur B and Falguni D. Identification of *Candida albicans* by using different culture medias and its association in potentially malignant and malignant lesions. Contemporary Clinical Dentistry 2011; 2(3):188-193.
- Joshi KR, Solanki A, Prakash P. Morphological identification of Candida species on glucose agar, rice extract agar and corn meal agar with and without Tween-80. Indian J Pathol Microbiol. 1993; 36(1):48-52.
- Koehler AP , Chu K-P, Cheng AFB, Simple, Reliable, and Cost-Effective Yeast Identification Scheme for the Clinical Laboratory J Clinical Micro 1999;37(2):422-426
- Marsh PD and Martin M, "Oral fungal infections," in *Oral Microbiology*, Churchill Livingstone, Edinburgh, UK, 2009;166-179.
- 20. National Committee for Clinical laboratory standards. Reference method for broth dilution antifungal

susceptibility testing of yeasts. Proposed guideline M27-P.National Commite for Clinical Laboratory Standards. 1992 Villanova, Pa.

- 21. Folch J, Lees M, Sloane Stanley GH. A simple method for the isolation and purification of total lipids from animal tissues. J. Biol. Chem. 1957; 226: 497-509.
- Sadhana S, Hardeep K, Khuller G K. Cell cycle effects od the phenothiszines : trifluoperazine and chloropromazine in *Candida albicans*. FEMS Microbiology Letters 2001; 199: 185-90.
- Amber K, Aifaz A, Immaculata X, Luqman KA, Nikhat M. Anticandidal effect of *Ocimum sanctum* essential oil and its synergy with fluconazole and ketoconazole. Phytomedicine 2010; 17: 921-25.
- Irshad, Shreaz S, Manzoor N, Khan LA, Rizvi MM. Anticandidal activity of *Cassia fistula* and its effect on ergosterol biosynthesis. Pharm Biol. 2011 Jul; 49(7):727-33.
- 25. Phillipson JD. New drugs from plants It could be Yew. Phytotherapy Res. 1999; 13: 1–7.

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