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# CYTOTOXIC ACTIVITY AND PHYTOCHEMICAL INVESTIGATION OF *PHYLLANTHUS EMBLICA L.* LEAVES

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Key words:

Phyllanthus emblica L., Extraction methods, cytotoxic effect, MTT assay flavonoids.

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**ABSTRACT:** The aim of this study is to investigate the cytotoxic effect of various extracts from Phyllanthus emblica L. According to Ayurveda this plant has potent anticancer activity and hence, this study is targeted at screening potential cytotoxic extracts using a yeast system. Yeast has been used as a model organism for many preclinical drug screens. In this study phytochemical extraction has been done by three methods (sonication, low and elevated temperature systems) using six different solvent systems (aqueous, methanol, ethyl acetate, petroleum ether, chloroform: methanol and water: methanol). The viability studies have been done using Methyl Thiazole Tetrazolium (MTT) assay. Aqueous extract at elevated temperature and water: methanol extract showed viability less than vinblastine. Further phytochemical analysis was done to identify major group of compounds present in each extract. It was concluded that flavonoids are present in extracts by doing TLC. HPTLC using standards was carried out and showed that at least two chemotherapeutic agents kaempferol and rutin were present. The data further indicates that kaempferol is major constituent in the extracts.

**INTRODUCTION:** Medicinal Plants have been known to mankind since ages, India itself has a number of indigenous systems of medicine based on plants and plant products. The usage of plants as traditional health remedies is the most popular among 80% of the world's population in Asia, Latin America and Africa and is reported to have minimal side effects.<sup>1</sup> However; modern science requires that we move from empirical uses of plants in medicine to scientifically substantiated applications.

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Further, one can assume that several different active principles (or their precursors) are present in extracts, since the history of their medicinal use demonstrates that the same plant has been used for several different medical conditions. Hence it makes good sense to separate and identify active components. This will enable us to isolate important plant compounds for specific applications in medicine.

*Phyllanthus emblica L.* belongs to the family Phyllanthaceae and preliminary research has demonstrated its *in vitro* antiviral and antimicrobial properties.<sup>2</sup> While there is evidence that its extracts induce apoptosis, <sup>3</sup> it has also been suggested that it may prove to have potential activity against some cancers.<sup>4, 5, 6</sup> Experimental evidence showing that its bark leaves and fruits have shown potential efficacy against cancer besides diabetes, and inflammation too exists.<sup>7</sup>

According to World Health Organization statistics, breast cancer is the most prevalent form of cancer disease in women. Our review of the literature indicated several plants that have been used for breast cancer cures<sup>8</sup>, but not all of them have resulted in purifying a single active principle or a mixture of known active principles. Hence, identifying the active principle involved in the plant based therapies for breast cancer from some of the other plants will enable us to ensure a wider range of medications that could be used to specify one that can help a given patient for chemotherapy, since it is known that a given chemotherapy for a specific cancer does not have identical effects on every patient. The aim of this study is to substantiate the medicinal effect of selected plants with respect to breast cancer by the use of cell culture technology.

As an experimental organism, yeast is a model of simplicity. Its 90-minute population doubling time allows for relatively short experiments allowing for examining exposure to drugs over many population doublings within minimal experimental time spans. In addition, yeast requires very simple and inexpensive media, can grow either in liquid culture or on solid media, and can be readily manipulated to express foreign genes.

Yeast was the first eukaryotic organism to have its genome fully sequenced and an extensive annotation of most yeast genes has been compiled. In addition, a vast array of tools allows the efficient and accurate manipulation of yeast genes. Genomewide functional analyses, including the analysis of transcriptional responses to a range of stimuli and comprehensive maps of protein–protein interactions, further increase the ease of studying of yeast cell and systems biology.

Finally, the high degree of conservation with mammalian cells in terms of gene sequence and protein and pathway function makes yeast very relevant for anticancer drug discovery. <sup>9</sup> Therefore this study involves use of genetically modified yeast having its cell cycle altered used as prescreen for the extracts.<sup>10</sup>

Plant materials were collected from the plant repositories and botanical gardens at St. Xavier's College campus. The sample used by us was originally bought from government nursery, Gandhinagar and planted at green house of Xavier Research Foundation. The leaves were air dried under shade and crushed finely. This powder was then used for all extraction purposes.

# **Preparation of extracts:**

The extraction was carried out for three different experimental conditions: i) lower temperatures  $(25^{\circ}C)$  ii) elevated temperature (example-  $60^{\circ}C$  for methanol, near its boiling point) iii) sonication. The solvents selected were i) water ii) methanol iii) ethyl acetate iv) chloroform:methanol: (1:1) v) water:methanol (1:1). The entire range of solvents was used for each of the experimental conditions. The selection of solvents was based on azeotropic range from polar to non-polar. For each of the conditions 1gram of powder was weighed and suspended in 20ml of selected solvent. The cold extraction procedure involved continuous shaking and changing of solvent every 24 hours until it was rendered colourless. However for the elevated temperature and sonication based methods solvent was changed every one hour until solvents were rendered colourless. These were pooled and the solvent was allowed to evaporate. The residue was resuspended in Dimethyl Sulfoxide (DMSO) to make a stock of 10 mg/ml and stored at  $4^{\circ}$ C.

# Cell viability assay:

The cytotoxicity of extracts in yeast cells was determined using the Methyl thiazol tetrazolium (MTT) assay.<sup>11</sup> Yeast cells at an OD660 of 0.820 (approx  $1.3 \times 10^7$  cells) were transferred to 96-well microplate. Extracts at concentration of 50 µg, 100µg, 150 and 200µg/100µl were constituted form the stored stocks mentioned above using growth medium and then added to the wells. It may be noted that no precipitates were seen and it was verified. The final concentration of DMSO was below 1%. The incubation with extracts was done for 16 hours and 24 hours. Vinblastine was used as positive control at same concentration as the extracts and cells without any drug as negative control. Cells were also incubated with DMSO as vehicle control. After exposure of 24 and 16 hours, 20 µl of MTT dye (2.5mg/ml) was added to each well. The plate was then incubated in dark for four hours. In order to dissolve the formazan crystals 100  $\mu$ l of DMSO was added after four hours and the plate agitated for five minutes. The absorbance was read at 570nm using a Thermofischer Multiskan GO microplate reader. Based on the absorbance cell viability was calculated and the ones showing cytoxoicity more than the positive control were considered to be effective.

$$Cell viability = OD of sample-OD of blank$$
$$OD of yeast cells -OD of blank$$

### **Phytochemical analysis:**

In order to identify phytochemicals in extracts preparative thin layer chromatography (TLC) was done.<sup>12</sup> All the extracts were subjected to TLC in two solvent systems i) ethyl acetate:methanol:water (100:13.5:10) and ii) toluene:ethyl acetate (93:7). Solvent system i) seperates polar compounds and ii) seperates non-polar compounds. Each TLC plate was then treated with various spraying reagents for detection of phytochemicals.<sup>12</sup>

Further Thin High Performance Layer Chromatography (HPTLC) was done using flavonoid standards (Rutin, quercitin, kaempferol etc.) with solvent system ethyl acetate: formic acid: glacial acetic acid: water (100:10:10:13) specific for flavonoid separation.<sup>12</sup> HPTLC was done on Silical Gel precoated F 240 plates. Sample application was done using Linomat5 application system and detection in Camag TLC scanner. Absorbance was measured at 366 nm.

#### Cell viability:

The water, methanolic and water:methanol extracts showed cell viability below the positive control (vinblastine). The cell viability for vinblastine was 77%. i.e. 77% cells of population were viable after exposure to the extract. Cell viability can be correlated to cytotoxic effect of the compound. If the compound is cytotoxic it will decrease the cell viability compared to control. Hence after 24 hours exposure of extracts to cell a decrease in viability below 77% was considered to be cytotoxic. Moreover a dose dependent increase in cytotoxicity is observed. (**Fig. 1**)







FIG. 1: CELL VIABILITY OF CRUDE EXTRACTS AFTER 24 HOURS EXPOSURE: i) EXTRACTS PREPARED BY SONICATION ii) EXTRACTS PREPARED AT  $25^{\circ}$ C iii) EXTRACTS PREPARED AT ELEVATED TEMPERATURE. (a= non significant, b= p<0.05, c=p<0.01, d=p<0.001)

Further, the cells were exposed for different time durations to the extracts. It was observed that with increased exposure time the cytotoxic effect increases. (**Fig. 2**) This was uniform for all extracts as there was a considerable decrease in cell viability from 16 hours exposure to 24 hours exposure.



FIG 2: CELL VIABILITY AT DOSE OF 100μg FOR 16 AND 24 HOURS OF EXPOSURE TIME: i) EXTRACTS PREPARED BY SONICATION ii) EXTRACTS PREPARED AT 25<sup>0</sup>C iii) EXTRACTS PREPARED AT ELEVATED TEMPERATURE

#### **Phytochemical analysis:**

The preliminary analysis using TLC indicated the presence of flavonoids in all extracts. The water extract at elevated temperature showed separation of more than one compounds. Further HPTLC was performed using flavonoid standards. The Rf values indicated that all extracts had either rutin or kaempferol present. The water:methanol extract showed only one Rf indicating presence of kaempferol whereas the ethyl acetate extract showed presence of rutin only. The water, methanol and chloroform:methanol extracts had rutin and kaempferol both present. (**Fig.3**)



FIG.3: ABSORBANCE PEAKS OF VARIOUS STANDARDS AND EXTRACTS AT 366 nm

TABLE 1: RF VALUES OF FLAVONOID STANDARDS ANDEXTRACTS USING SOLVENT SYSTEM ETHYL ACETATE:FORMIC ACID: GLACIAL ACETIC ACID: WATER(100:10:10:13)

Substance	Rf	Absorbance (AU)	% Area of the
			peak
Quercetin	0.72	368.7	69.83
Rutin	0.85	222.5	54.27
Kaempferol	0.68	236.4	100
Water extract	0.68	217.4	35.02
	0.82	218.2	35.06
Methanol extract	0.71	57.5	48.58
	0.83	60.9	51.42
Ethyl acetate extract	0.83	216.1	40.29
Choloroform:methanol	0.72	184.5	32.89
extract	0.82	205.7	36.66
Water:methanol extract	0.72	13	100

Plants have been utilized as medicines for thousands of years.<sup>13</sup> Isolation and characterization of pharmacologically active compounds from medicinal plants continue today. More recently, drug discovery techniques have been applied to the standardization of herbal medicines. Phyllanthus emblica L. is a promising anticancer herb form which effective compounds need to be identified. Drug discovery from medicinal plants has played an important role in the treatment of cancer and, indeed, most new clinical applications of plant secondary metabolites and their derivatives over the last half century have been applied towards combating cancer <sup>14, 15</sup>. Hence we have tried to identify active compounds form plants by using yeast as a model system. In this study we demonstrated that flavonoids from the plant are probably responsible for the cytotoxic activity. *Phyllanthus emblica L.* is known to have flavonoids present and cytotoxic effect has been demonstrated.<sup>16</sup>

**CONCLUSION:** The results indicate that water, methanol and water:methanol extract have cytotoxic effect on yeast cells. Moreover the use of different methods of extraction yields similar compounds whereas different solvents are able to extract various compounds. The study concludes that *Phyllanthus emblica L*. has various flavonoids present and possibly responsible for cytotoxic effect.

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#### **REFERENCES:**

- S. Chanda, J. Parekh, Y. Vaghasiya, R. Dave, Y. Baravalia and R. Nair: Medicinal plants-from traditional use to toxicity assessment: A Review. International Journal of Pharmaceutical Sciences and Research 2015; 6(7): 2652-2670.
- 2. P.A. Dua and S.D. Patankar: Antibacterial evolution of some common medicinal plants used in Asava and Arishta preperation. International Journal of Pharmaceutical Sciences and Research 2013; 4(8): 3250-3255.
- 3. Tiejun Zhao, Qiang Sun, Maud Marques and Michael Witcher: Anticancer Properties of *Phyllanthus emblica* (Indian Gooseberry). Oxidative Medicine and Cellular Longevity 2015.
- 4. D. Sumalatha: Antioxidant and antitumor activity of *Phyllanthus emblica* in colon cancer cell lines. International Journal of Current Microbiological Application Sciences 2013; 2(5): 189-195.
- Penolazzi, L, Lampronti I, Borgatti M, Khan M, Zennaro M, Piva R and Gambari R: Induction of apoptosis of human primary osteoclasts treated with extracts from the medicinal plant *Emblica officinalis*. BMC Complementary and Alternative Medicine 2008; 8: 59.

- Ngamkitidechakul C, Jaijoy K, Hansakul P, Soonthornchareonnon N and Sireeratawong S: Antitumor effects of *Phyllantus emblica*: Induction of cancer cell apoptosis and inhibition of *in vivo* tumor promotion and *in vitro* invasion of human cancer cells. Phytotherapy Research 2010; 24(9):1405-1413.
- P. Usharani, P.V. Kishan, Nishat Fatima and C. Uday Kumar: A Comparative Study To Evaluate The effect of highly standardized aqueous extracts of *Phyllanthus Emblica*, *Withania Somnifera* and their combination on endothelial dysfunction and biomarkers in patients with Type II Diabetes Mellitus, International Journal of Pharmaceutical Sciences and Research 2014; 5(7):2687-2697.
- 8. Praful Talaviya: Anticancer Approach with Natural Product. International Journal of Pharmaceutical Sciences and Research 2011;(10): 2514-2520.
- Sanchez-Pico Angeles, J. Antonio, Gonzalez-Leon, Martin-Cordero Carmen, and Daga Rafael: Screening for natural anticancer agents using a fission yeast bioassay. Phytochemistry Letters 2014; 8: 184-189.
- Ju-Hee Lee, Ji-Hyun Yeon, Hanna Kim, Whijae Roh, Jeiwook Chan, Han-Oh Park, Dong-Myung Kim: The natural anticancer agent plumbagin induces potent cytotoxicity in MCF-7 Human breast cancer cells by inhibiting a PI-5 kinase for ROS generation. PLOS One, 2012; 7(9):1-10.
- 11. Gavamukulya Y., Abou-Elella F., Wamunyokoli F., and AEI-SHemy H: Phytochemical screening, anti-oxidant activity and *in vitro* anticancer potential of ethanolic and water leaves extracts of Annona muricata. *Asian Specific Journal of Tropical Medicine*, 2014; 7:S355-S363.
- H. Wagner and S. Bladt. Plant Drug Analysis: A Thin Layer Chromatography Atlas. Second edition. New York, Springer Publications, 2001:349-354.
- Samuelsson, G. Drugs of Natural Origin: A Textbook of Pharmacognosy. Stockholm, 5th Swedish Pharmaceutical Press, 2004.
- 14. Umashanker M. and Shruti S: Traditional Indian herbal medicine used as antipyretic, antiulcer, anti-diabetic and anticancer: A review. International Journal of Research in Pharmacy and Chemistry, 2011; 1 (4):1152-1159.
- 15. Newman, D.J., Cragg, G.M: Natural products as sources of new drugs over the 30 years from 1981 to 2010. Journal of Natural Products 2012; 75 (3): 311-335.
- 16. Ekta Singh, Sheel Sharma, Ashtosh Pareek, Jaya Dwivedi, Sachdev Yadav and Swapnil Sharma: Phytochemistry, traditional uses and cancer chemopreventive activity of amla (*Phyllanthus emblica*): The Sustainer. Journal of Applied Pharmaceutical Sciences 2011; 02 (01): 176-183.

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