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DEVELOPMENT AND VALIDATION OF UV SPECTROPHOTOMETRIC METHODS FOR ESTIMATION OF CELIPROLOL HYDROCHLORIDE IN TABLET FORMULATION

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Keywords:

Celiprolol Hydrochloride, UV spectrometry, zero order derivative, area under curve, validation

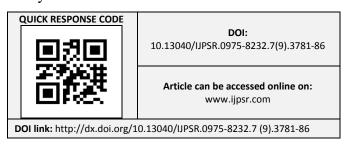
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ABSTRACT: Simple, specific, accurate and cost economic UV spectrophotometric methods were developed and validated for determination of Celiprolol Hydrochloride. Instead of using organic solvents, mixture of dilute hydrochloric acid and water was used during method development and validation. The drug was extracted from tablet formulation with a mixture of water and hydrochloric acid. Stability of drug solution and formulation was established throughout the analysis time. Zero Order Derivative and Area Under Curve methods were developed for estimation of drug and both the methods were validated as per ICH guidelines. The linearity of drug was established by performing calibration curve. The analytical range of drug was established from linearity study. Precision of the method was established with Intraday and interday analysis. Accuracy was established by conducting recovery study. Robustness of both the methods was confirmed by conducting analysis with different analysts.

INTRODUCTION: Celiprolol Hydrochloride (CPHL) is a highly cardio-selective β1 adrenergic partial β2 agonist blocker with activity (Vasodilating) ¹. CPHL is equal to other β-blockers in controlling blood pressure and heart rate increases during exercises. The drug is comparable or superior to other β-blockers and calcium channel blockers in angina ². CPHL is primarily indicated in conditions like Angina pectoris, Hypertension³, mild to moderate hypertension, and can also be given in adjunctive therapy as an alternative drug of choice in Arrhythmias and Supraventricular Arrhythmias ³⁻⁹.



It is official in Indian & British Pharmacopoeia. Chemically it is 3-[-3Acetyl-4-[(2RS)-3-[(1, 1-dimethylethyl)amino]-2-hydroxypropoxy]phenyl]-1,1-diethyl urea hydrochloride¹⁰. It is a white off, very slightly yellow, crystalline powder, freely soluble in water and in methanol, soluble in ethanol (96%) and very slightly soluble in Methylene Chloride. Analytical methods available for CPHL are Potentiometric method ¹⁰ HPLC, and Colorimeteric method ¹¹.

Literature survey revealed that no zero order and Area Under Curve UV spectrometric method was reported for estimation of drug. In the present work, the study was conducted using aqueous solvent so that waste disposal was not a problem. The present paper was aimed to provide simple, accurate, cost economic and precise method for estimation of CPHL in tablet formulation.

MATERIAL AND METHODS:

CPHL pure drug was obtained as a gift sample from Ranbaxy Goa. All other chemicals and reagents used were of analytical grade. Ethanol was purchased from Research-lab Fine Chemicals, Mumbai. Sodium Hydroxide was purchased from Research Lab, Pune. Con. Hydrochloric acid was purchased from Fine chemicals, Pune. spectrophotometer used was Jasco V-630 with UV visible detector. Other apparatus, utilized were digital electronic balance (Shimadzu BL-220H), Ultrasonicator (Equiptron), Micro Syringe (Hamilton micro lab), Glassware (Borosilicate) used were calibrated.

Selection of suitable solvent:

The solubility of drug was determined in a solvents such as Methanol, Ethanol (IP, BP) as well as in dilute HCl.

Determination of λ max of drug:

Accurately weighed quantity of 10 mg drug was dissolved in 30 ml distilled water & 2 ml conc. HCl and was transferred to 100 ml volumetric flask and volume was made with 0.01 N HCl to prepare 100 μ g/ml and kept for ultrasonication for 5 min. The resulting standard stock solution of bulk drug was further diluted with Dil. HCl to get 15 μ g/ml concentrations. The solution was scanned between 200-400 nm range against Dil. HCl as blank. After observing the UV-Spectrum of drug, 232 nm was selected as λ max for further analysis (**Fig. 1**).

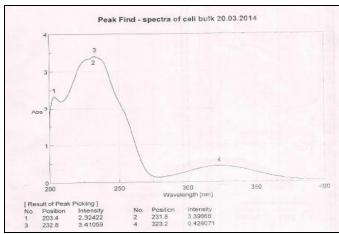


FIG. 1: λ MAX OF CPHL

Extraction of CPHL from tablet formulation: 11

Accurately weighed quantity 10mg of drug was transferred to a volumetric flask and dissolved in

the mixture of 30 ml water and 2.0 ml of conc. HCl. The sample was centrifuged, filtered and diluted with 0.01 N HCl up to the mark to prepare 100 μ g/ml solution. Similarly, tablet powder equivalent to 10 mg of drug was also processed using water and conc. HCl to prepare 100 μ g/ml solution. The absorption spectrums of series of dilution of CPHL extracted from tablet were scanned from 200 nm to 400 nm and compared with that of bulk solution (**Fig. 2 & 3**).

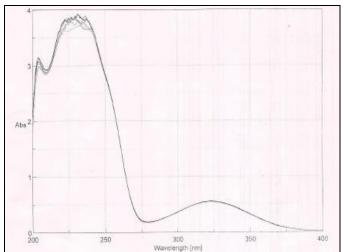


FIG. 2: OVERLAY SPECTRA OF CPHL BULK

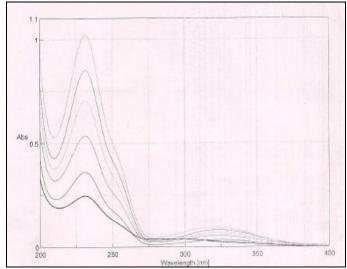


FIG. 3: OVERLAY OF CPHL TABLET

Stability of solution: 12-13

Series of dilutions of tablet stock solutions were prepared separately to get 3-18 $\mu g/ml$ of working standards. The stability of these solutions was checked by measuring the solutions at 232 nm at two different time intervals, i.e. initially and after 72 hrs. The overlain spectra of tablet solution,

obtained at two different times were compared to verify the stability of solution. (**Fig.4** and **5**).

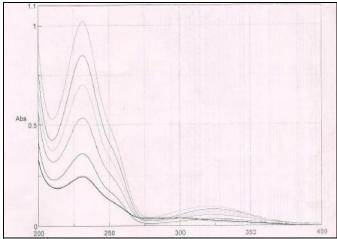


FIG. 4: SPECTRA OF TABLET FORMULATION AT (HOURS

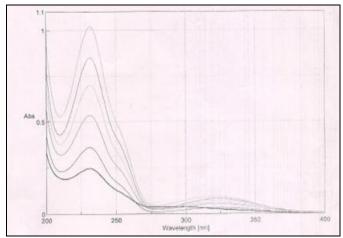


FIG. 5: SPECTRA OF TABLET FORMULATION AT 72 HOURS

Preparation of standard and test solutions of tablet formulation:

The quantity of tablet powder equivalent to 10 mg drug was dissolved drug tablet powder was dissolved in 30ml distilled water & 2 ml conc. HCl and was transferred to 100 ml volumetric flask and volume was made with 0.01 N HCl and kept for ultrasonication for 5 min. It was filtered and volume was made up to the mark to prepare

concentration of 100 μ g/ml. From the resulting standard stock solution, series of dilutions were prepared to get 9, 12, and 15 μ g/ml of laboratory samples.

Estimation of drug concentration in test solutions:

Method A- Zero order derivative spectrometry ¹⁴⁻¹⁸ In this method, calibration curve was obtained by plotting absorbance of working standards (3-18 μ g/ml) against concentrations at 232 nm to get zero order spectra of CPHL(**Fig. 6** and **Table 1**).

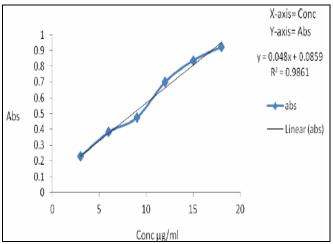


FIG. 6: CALIBRATION CURVE OF CPHL

TABLE 1: CALIBRATION CURVE OF CPHL

Concentration(µg/ml)	Absorbnace
3	0.2293
6	0.3829
9	0.4738
12	0.6989
15	0.8362
18	0.9213

The regression equation and correlation coefficient was obtained from the calibration curve. The absorbance of laboratory solutions of tablet (9, 12, and 15 μ g/ml) was measured separately against 0.01 N HCl blank. From the regression equation, the concentrations of laboratory solution of tablet formulation were estimated. (**Table 2**)

TABLE 2: RESULT FOR LABORATORY ANALYSIS OF TABLET FOR METHOD A & B

Co	nc C	Conc. Found			RSD			
(μg/i	ml) N	Mean ±SD		%w/v		%		
	Method A	Method B	Method A	Method B	Mtd A	Mtd B		
9	9.0919 ± 0.0271	9.0827±0.0048	100.6	100.9	0.2980	0.0531		
12	$2 11.999 \pm 0.0181$	12.081 ± 0.0128	99.99	100.5	0.1508	0.1059		
15	$5 15.0231 \pm 0.0100$	15.084 ± 0.0073	100.1	100.5	0.0665	0.0483		

Method B-Area under curve (AUC): ¹⁹⁻²⁴ Selection of analytical wavelength:

For the selections of analytical wavelength, 15 μ g/ml solution of drug was prepared and scanned in the spectra measurement mode across 200-400 nm range. From the spectra of drug, area under the curve in the range of 231-241 nm was selected for further analysis (**Fig. 1**).

Estimation of drug concentration in laboratory samples:

The calibration curve was obtained by plotting AUC of working standards in the range of 3-18 μ g/ml against concentration (**Fig. 7**). The laboratory solutions of tablet (9, 12, and 15 μ g/ml) were measured separately against 0.01 N HCl blank. From the regression equation, the concentration of drug in the laboratory solutions of tablet was estimated (**Table 2**).

Validation of method A and Method B for bulk and tablet formulation:

The analytical methods were validated as per ICH guidelines ¹² for the parameters linearity, range, accuracy, precision, limit of detection, limit of quantitation, and robustness.

Range and Linearity study:

TABLE 4: RESULT FOR ACCURACY STUDY

Conc. (µg/ml)		Total	Conc. found (µg/ml) ±SD		% Recovery		RSD %	
		conc.	(n=3)		Methods		Methods	
Гакеп	Standard	μg/ml	Method A	Method B	A	В	A	В
	added							
8	6.4	14.4	14.17±0.02	14.12 ± 0.01	98.42	98.12	0.14	0.070
8	8	16	16.06±0.01	16.07 ± 0.01	100.3	100.4	0.062	0.070
8	9.6	17.6	17.05 ± 0.01	17.54 ± 0.01	100.9	99.67	0.057	0.057
		Faken Standard added 8 6.4 8 8	Γaken Standard added μg/ml 8 6.4 14.4 8 8 16	Conc. (n= Gaken Standard added μg/ml Method A 8 6.4 14.4 14.17±0.02 8 8 16 16.06±0.01	Gaken Standard added μg/ml Method A Method B 8 6.4 14.4 14.17±0.02 14.12±0.01 8 8 16 16.06±0.01 16.07±0.01	$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$		Conc. μg/ml added conc. μg/ml added (n=3) Methods Method Method B Method A Method B A B A 8 6.4 14.4 14.17±0.02 14.12±0.01 98.42 98.12 0.14 8 8 16 16.06±0.01 16.07±0.01 100.3 100.4 0.062

Precision:

Precision of the method was studied as intra-day and inter-day variations. Intra-day precision was established by analyzing three concentrations (8, 10, and 18 μ g/ml) of tablet formulation in triplicate, three times a day by the proposed methods, to estimate the concentration of drug in solutions (**Table 5**).

TABLE 5: RESULT FOR PRECISION STUDY

Precision	Conc.	Conc. Found		Content % w/v		RSD %		
	(µg/ml)	Mean \pm SD (n=3)		Met	Methods		Methods	
		Method A	Method B	A	В	A	В	
Interday	8	8.087 ± 0.009	7.99 ± 0.008	101	99.96	0.1112	0.1001	
	10	10.08±0.003	10.07 ± 0.004	100.8	100.7	0.0382	0.0397	
	18	18.09 ± 0.006	18.08 ± 0.004	100.5	100.5	0.0331	0.0221	
Intraday	8	7.978 ± 0.010	7.99 ± 0.012	99.72	99.93	0.1253	0.1501	
	10	10.07 ± 0.012	10.04 ± 0.025	100.6	100.4	0.1121	0.2490	
	18	18.08 ± 0.089	18.06 ± 0.008	100.3	100.3	0.4922	0.0442	

Drug solutions of concentration from 3-18 μ g/ml were measured at 232 nm for reading absorbance. The calibration curve was prepared. The correlation coefficient and regression equation was obtained. The range was established from linearity study. (**Table 3, Fig. 6**)

TABLE 3: RESULT FOR VALIDATION PARAMETERS FOR BOTH METHODS

Paran	neters	Method A	Method B
Analytical V	Vavelength	232	231-241
(nr	n)		
Beer's law li	mit (µg/ml)	3-18	3-18
Regression	equation	Y=0.048X +	Y=0.1007X +
		0.0859	0.203
		$R^2 = 0.9861$	$R^2 = 0.997$
Robustness	Mean	9.974 ± 0.0228	10.061 ± 0.0092
(n=3)	conc.±SD		
	%RSD	0.2273	0.0914

Accuracy:

Accuracy of tablet solution was determined by performing recovery studies by spiking pure drug in pre-analyzed sample solution at different level i.e. 80%, 100% and 120%. The resulting solutions were analyzed by proposed methods to estimate the concentrations. The percent recovery, SD and %RSD were calculated to establish the accuracy of the method. (**Table 4**)

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Inter-day precision was determined by analyzing three concentrations 8, 10, and 18 μ g/ml of tablet formulation, in triplicate and on three consecutive days by the proposed methods to estimate the concentration of drug in solutions. For both inter and intraday precision, SD and % RSD were calculated, to ascertain the precision (**Table 5**)

Robustness:

For ascertaining robustness three different analysts analyzed the tablet solutions for estimation of drug concentration by proposed methods. Mean, SD, %RSD were calculated and reported to signify the robustness of the method (**Table 6**).

TABLE 6: RESULT FOR ROBUSTNESS

Analyst	Concentration (µg/ml)	Mean (μg/ml) ±SD %			ntent w/w	Rs 9	SD %
		A	В	A	В	A	В
1	10	10.045 ± 0.007	10.073 ± 0.008	100.4	100.7	0.0696	0.0754
2	10	9.974 ± 0.022	10.06 ± 0.009	99.74	100.6	0.2205	0.0894
3	10	10.04 ± 0.011	10.06 ± 0.012	100.4	100.6	0.1184	0.1192

RESULT AND DISSCUSSION:

The present work was aimed to provide simple, accurate, cost economic and precise analytical methods for estimation of CPHL in tablet formulation. Solvent used for analysis of tablet was mixture of hydrochloric acid and water. The analytical wavelength for method A determined and was observed 232 nm (Fig. 1), and for that of method B was 231-241 nm. The drug was extracted from tablet formulation by using the mixture of water and dilute hydrochloric acid. Scans of working standard of tablet solution was compared with that of bulk solution of drug to verify that drug has properly extracted from tablet formulation (Fig. 2 and 3). The patterns of overlain spectra of tablet solutions obtained at two different time intervals, i.e. initially and after 72 hrs were found identical which confirmed that the analytical solutions were stable throughout of analysis time (Fig. 4 & 5).

The estimation of drug concentration in laboratory samples of tablet formulation by Method A (Zero order derivative spectrometry) at 232 nm was performed with three concentrations (**Table 2**). The results of analysis were found with standard deviation from 0.0100-0.0271 and the % content was found in the range of 99.99-100.6% w/w. For method B (AUC method), the results of analysis were found with standard deviation from 0.0128-0.0073. and the % content was found in the range of 100.5-100.9% w/w.

Study of validation parameters revealed that Beer's law limit of the drug was $3\text{-}18~\mu\text{g/ml}$. The

regression equation for method A was Y=0.048X+0.0859 with $R^2=0.9861$ and for that of method B was Y=0.1007X+0.203 with $R^2=0.997$ (**Table 3**). Accuracy study (Table 4) revealed % Recovery for both the methods in the range of 98.12-100.9. The Precision (**Table 5**) was established with inter-day and intraday precision study. Both the methods were found robust (Table 6) with the standared deviation of 0.0228 and 0.0092 respectively.

CONCLUSION: The study of validation parameters revealed that both the methods were simple, accurate, precise, robost, and can be applied for routine analysis of tablet fromulation. Study conducted using aqueous solvent which made anlaytical methods cost-effective and ecofriendly.

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CONFLICT OF INTEREST: No conflict of interest

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