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## ISOLATION AND PARTIAL CHARACTERIZATION OF PROTEASE FROM *BACILLUS HALODURANS* (AJ302709) FROM ALKALINE LONAR LAKE

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Lonar Lake,  
Haloalkaliphilic, *Bacillus*, protease

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**ABSTRACT:** Alkaline protease alone account for 60% of the total world enzyme market with its improved properties such as higher optimum temperature and alkaline pH. Alkaline Lonar Lake, situated in the Buldhana District of the Maharashtra State, India, is a unique ecosystem and harbors various haloalkaliphilic bacterial species which produces biotechnologically important thermo-haloalkaliphilic enzymes. The present study deals with the production and partial characterizations of enzyme protease isolated from the extremophilic Lonar Lake bacterium. A total six bacterial strains were isolated on Horikoshi medium from the water, sediment and matt samples collected from Lonar crater. Out of these, one bacterium showing prominent proteolytic activity was characterized on the basis of morphological, cultural and biochemical parameter and identified as *Bacillus halodurans* (AJ302709) by 16S rRNA sequencing. The alkaline protease produced by *Bacillus halodurans* showed optimum activities at pH 8, at temperature 70°C. It proves that the enzyme alkaline protease produced by this bacillus has potential applications in food, pharmaceutical and detergent industries.

**INTRODUCTION:** Protease is an enzyme occurring everywhere in nature being inside or on the surface of living organisms such as plants, animals and microbes. These enzymes carry out proteolysis by hydrolysis of the peptide bond that exists between two amino acids of a polypeptide chain <sup>1</sup>. The proteases are available in the market are mostly derived from microbial sources. Many bacteria like *Bacillus cerus*, *Bacillus fermus*, *Bacillus pseudofirmus* and *Enterococcus caseliflavus* are well known for the production of thermostable and alkali stable protease <sup>2</sup>.

Proteases are industrially important enzymes used in the detergent, food, pharmaceutical, leather industries and also have application in silver recovery from photographic plates and in peptide synthesis which account for about 60% of total industrial enzyme sales <sup>3</sup>. The alkaline enzyme has better resistance to alkali and some other denaturing chemicals in the reaction mixture and has a higher affinity towards proteinaceous substrates. The microorganism growing in alkaline and thermostable habitats produces the alkaline-thermostable protease which may have with special characteristics <sup>4</sup>. Very less study had been done on protease from *Bacilli* of Lonar Lake, which can withstand high temperature and high pH and has wide applications in different industries <sup>5</sup>.

In present study, aim was to deal with the isolation, characterization, production and optimization of a

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protease from bacterial strain isolated from the alkaline Lonar Lake and which could be useful in the food, pharmaceutical, detergent and leather industry.

## MATERIALS AND METHODS:

### Screening of bacterial proteolytic alkaliphiles:

A total of twelve samples (sediment, matt and water) were collected from alkaline Lonar Lake was screened for proteolytic enzymes. About 1g of sediment and matt sample was transferred to 100mL sterilized distilled water in 250 mL conical flask and agitated (100 rpm) at 37<sup>0</sup>C for 15 min on rotary shaker. The sample was then heated at 80<sup>0</sup>C for 15 min to destroy all the vegetative microbial cells. The suspension was then diluted to 10<sup>7</sup> dilutions. One ml of each diluted sample was lawn into Petri plates containing Horikoshi medium (A, B, C and D) and incubated at 37<sup>0</sup>C for 72h and four time repeated sub culturing was made in the same medium. After enrichment, it was inoculated on Nutrient agar (pH 10) and incubated at 37<sup>0</sup>C for 24h for isolation pure bacterial culture.

The isolated bacterial colonies were screened for proteolytic activities on Skim milk agar medium. The inoculated plates were incubated at 37<sup>0</sup>C for 72h and observed for zones of clearance, indicating proteolytic activities<sup>4</sup>. The bacterial isolates with prominent zones of clearance on skim milk agar medium were identified based on cultural, morphological and biochemical characterization and finally by 16S rRNA sequencing at Agharkar research institute, Pune.

### Preparation and Partial characterization of crude enzyme protease:

The 100 mL Yeast extract casein medium was inoculated with isolated culture and incubated at 37<sup>0</sup>C in shaking incubator. After 72h incubation, the broth was centrifuged at 5000-8000 rpm for 15 min. The supernatant served as crude enzyme source. The standard graph with tyrosine was prepared. Estimation of proteases was carried out with casein and the absorbance was read at 650 nm<sup>2, 6</sup>. The proteolytic activity was defined as the amount of the enzyme that released 1µg of tyrosine per minute under the assay conditions. Partial characterization of protease was determined by assaying the enzyme activity at different

parameters such as pH (7.0 to 10.5), temperature (55<sup>0</sup>C to 80<sup>0</sup>C), substrate concentration (5 mg/mL to 40 mg/mL) and enzyme concentration<sup>7</sup>.

## RESULT AND DISCUSSION:

In the present study, a total of 12 different bacterial species were isolated from water, sediment and matt samples from Lonar Lake. Out of 12, six isolates were showed maximum casein hydrolysis activity on skim milk agar at pH 8. Out of them one isolate DHT19 was selected for further study since it showed prominent proteolytic zone of 25mm. These isolate was characterized by cultural, morphological and biochemically by commercially available Hi-media Rapid detection kit. The isolate DHT 19 was Gram positive, rod shape and motile. Growth was detected at different pH (7 to 12) and salt concentration of NaCl (1 to 8%).

The growth of isolate DHT 19 was found to be optimum at 40<sup>0</sup>C to 55<sup>0</sup>C temperature. Phylogenetic analysis based on 16S rRNA gene sequences indicated that strain DHT19 is *Bacillus Halodurans*. From the data it showed that, the maximum proteolytic activity was observed at pH 8 (**Fig.1**) and at 70<sup>0</sup>C (**Fig.2**) by *Bacillus halodurans*. Optimum pH for proteolytic activity of protease producing bacteria was observed between pH 8-10.5. The optimum enzyme concentration required for maximum activity of protease 1.54ug/mL (**Fig.4**) and substrate concentration was found 1.5µg/mL (**Fig.3**).

These indicate that for the growth of protease producing bacteria, alkaline environment is more suitable. This idea is also supported by Narwal *et al.*<sup>8</sup>. The phylogenetic position indicated the Lonar lake bacterial strains were related to phylum Firmicutes and belongs to genera Bacillus and are *Bacillus flexus*, *Bacillus pseudofirmus* and *Bacillus cereus*<sup>9, 10</sup>.

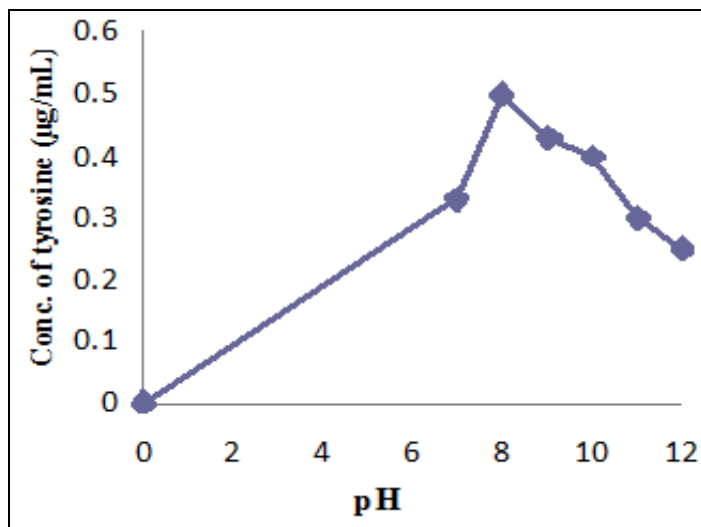
**CONCLUSION:** In the present study, different bacterial species were isolated from water, sediment and matt sample of Lonar Lake. Out of them, one bacterial strain DHT19 was found protease producer and screened for production and the partial characterizations of protease. The bacterial isolates were characterized and identified as *Bacillus halodurans*. Alkaline protease

production was maximum at pH 8 and the activity was 1.5unit/mL. The isolated *Bacillus halodurans* strain produces the protease enzymes which were

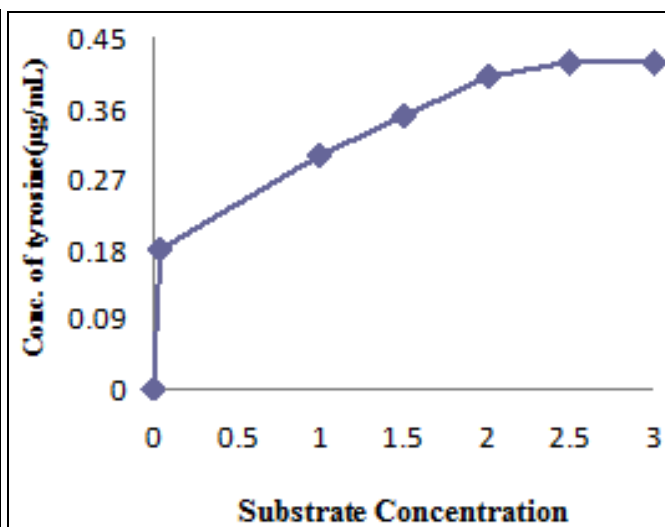
thermostable, alkaliphilic and has potential to produce good quality protease which can be used in food, pharmaceutical and the detergent industries.

**TABLE 1: CULTURAL, MORPHOLOGICAL AND BIOCHEMICAL CHARACTERISTICS OF PROTEASE PRODUCING *BACILLUS HALODURANS* (T) (AJ302709)**

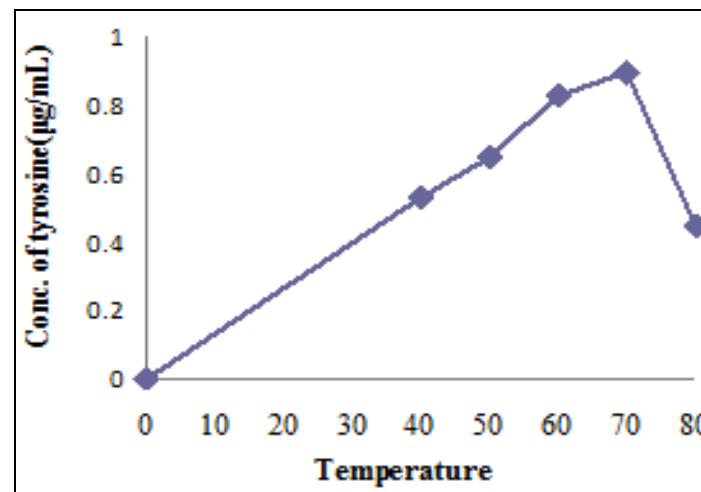
Gram character	+	Glucose	+	$\alpha$ -Methyl-D-glucoside	-
Shape of Bacteria	LR	Dextrose	+	Rhamnose	-
Arrangement of Cell	Single	Galactose	-	Cellobiose	-
Spore	+	Raffinose	-	Melezitose	-
Motility	+	Trehalose	-	$\alpha$ -Methyl-D-mannoside	-
Catalase	-	Melibiose	-	Xylitol	-
Oxidase	-	Sucrose	-	ONPG	-
Voges Proskauer	-	L- Arabinose	-	Esculin hydrolysis	-
Citrate utilization	+	Mannose	-	D-Arabinose	-
Nitrate reduction	-	Inositol	-	Malonate Utilization	+
Arginine	+	Sorbitol	-	Sorbose	-
Lactose	-	Mannitol	-	Inulin	-
Xylose	-	Adonitol	-	Sodium gluconate	-
Maltose	-	Arabitol	-	Glycerol	-
Fructose	-	Erythritol	-	Salicin	-



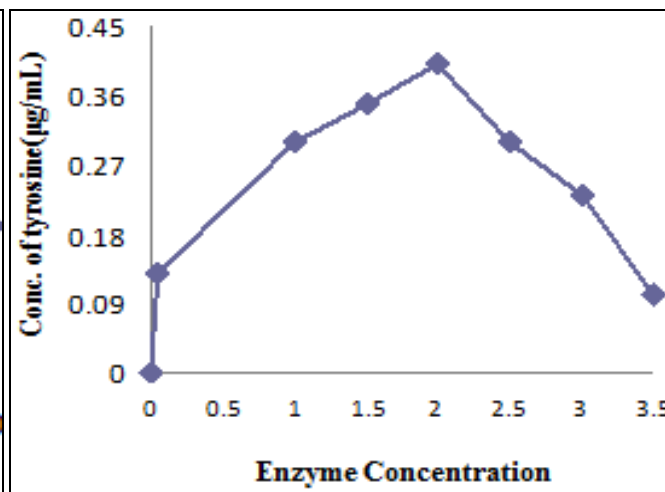
**FIG.1: EFFECT OF pH ON ACTIVITY OF PROTEASE ENZYME**



**FIG.2: EFFECT OF TEMPERATURE ON THE ACTIVITY OF PROTEASE ENZYME**



**FIG. 3: EFFECT OF SUBSTRATE CONC. ON THE ACTIVITY OF PROTEASE ENZYME**



**FIG. 4: EFFECT OF ENZYME CONC. ON THE ACTIVITY OF OF PROTEASE ENZYME**

**TABLE 2: THE 16 S rRNA GENE SEQUENCING CLOSEST PHYLOGENETIC AFFILIATION, PAIR SIMILARITY AND RIBOSOMAL DATABASE PROJECT REPORT OF ISOLATED PROTEASE PRODUCING ORGANISM DHT 19 FROM LONAR LAKE**

Strain Designation	Closest Phylogenetic affiliation	Max ident
DHT 19	<i>Bacillus halodurans</i> (T) 16S rRNA gene partial sequence (AJ302709)	99.0%

Quick Bioinformatic Phylogeny of Prokaryotes and Seewiew  
-See Legend-

0.2

Virgibacillus halodentificans T AB681753  
Virgibacillus halodentificans T AB021186  
Virgibacillus halodentificans T AY543169  
Bacillus taenensis T AY603978  
"Bacillus casamancensis" t AF519462  
Bacillus algicola T AY128462  
Bacillus algicola T FR775437  
Bacillus pseudofirmus T X76439  
Bacillus pseudofirmus T CP001878  
Bacillus pseudofirmus T JQ337958  
Bacillus marmarensis T EU621902  
Bacillus nanhausedimimis T GQ292773  
Bacillus halodurans T AJ302709  
Bacillus okuhidensis T AB047684  
Bacillus halodurans T AB021187  
Bacillus halodurans T BA000004  
ORY DHT 19  
Bacillus hemichilosilyticus T AB043846  
Bacillus ochensis T DQ226060  
Bacillus akribai T AB043858  
Bacillus krulvichiae T AB681754  
Bacillus krulvichiae T AB086897  
Bacillus wakoensis T AB043851  
"Anaerobacillus macysae" T AY032601  
Anaerobacillus arseniciselenatis T A3965469  
Anaerobacillus alkalidiazotrophicus T EU143680  
Anaerobacillus alkalicastris T DQ675454  
Bacillus alkalitriticus T EF422411  
Bacillus lignimiphilus T JQ44788  
Bacillus neizhouensis T EU925618  
Bacillus mammilyticus T AB043864

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