



Received on 23 June, 2016; received in revised form, 26 July, 2016; accepted, 02 August, 2016; published 01 December, 2016

REVERSE PHASE-HIGH PERFORMANCE LIQUID CHROMATOGRAPHY METHOD FOR ESTIMATION OF BIOACTIVE COMPOUNDS FROM DIFFERENT FRACTIONS OF NEW POLYHERBAL FORMULATION OF *CLINACANTHUS NUTANS* AND *ELEPHANTOPUS SCABER*

Muhammad Shahzad Aslam ^{*1}, Muhammad Syarhabil Ahmad ¹, Awang Soh Mamat ¹, Muhammad Zamharir Ahmad ² and Faridah Salam ²

School of Bioprocess Engineering ¹, Universiti Malaysia Perlis, Kompleks Pusat Pengajian Jejawi 3 (KPPJ3), Kawasan Perindustrian Jejawi, 02600, Arau, Perlis, Malaysia.

Biotechnology and Nanotechnology Research Center ², Malaysian Agricultural Research and Development Institute, 43400 Serdang, Selangor, Malaysia.

Keywords:

Clinacanthus nutans,
Elephantopus scaber, Polyherbal formulation, Herb-Herb combination, Flavonoids, Apigenin, luteolin, b-sitosterol, vanillin, gallic acid, tetra methoxy flavone, quercetin, rutin, benzoic acid, gallic acid, HPLC

Correspondence to Author: Muhammad Shahzad Aslam

PhD Scholar, School of Bioprocess Engineering, Universiti Malaysia Perlis, Kompleks Pusat Pengajian Jejawi 3 (KPPJ3), Kawasan Perindustrian Jejawi, 02600, Arau, Perlis, Malaysia.


Email: aslammuhammadshahzad@gmail.com

ABSTRACT: Background: Polyherbal formulation is the trend of modern pharmacognosy and holistic approach in the treatment of incurable diseases such as cancer. Medicinal plants contain many bioactive compounds that give different pharmacological activities. Therefore, it is important to investigate the quality of new polyherbal formulation by identification of bioactive markers. **Objective:** To identify apigenin (1), luteolin (2), b-sitosterol (9), 3'-hydroxy-5, 6, 7, 4'-tetra methoxy flavone (8), quercetin (3), vanillin (7), gallic acid (5), benzoic acid (6) and rutin (4) as biomarker from different fractions of the new polyherbal formulation of *Clinacanthus nutans* and *Elephantopus scaber* by RP-HPLC gradient method. **Methods:** A Shimadzu HPLC was utilized to perform the analysis which was equipped with an autosampler, column oven, and UV/VIS detector. An HPLC column used was Merck LiChrospher Start RP 18 column (250mm, 4.6 mm i.d, 5µm pore size). The temperature was maintained at 40.0 °C throughout the study. The mobile phase isocratic method with acetonitrile 100%. The flow rate was 0.5 mL min⁻¹. Absorbance was observed at λ = 360 nm. **Results:** After identification of bioactive compounds it was established that 3'-hydroxy-5, 6, 7, 4'-tetra methoxy flavone, gallic acid, benzoic acid and vanillin was found in a polyherbal formulation that was not present before in *Clinacanthus nutans* and *Elephantopus scaber*. Other bioactive compounds such as Apigenin, Luteolin, b-sitosterol, quercetin, and rutin were present in both medicinal herbs as well as inside polyherbal fractions. Vanillic acid was the oxidative product of vanillin that was found in *Elephantopus scaber*. In the polyherbal formulation, vanillin was found that shows the transformation of vanillic acid into vanillin. **Conclusion:** Formation of new bioactive compounds from a polyherbal formulation that was not present before in both medicinal plant. Identification of biomarkers helps to increase the quality of the new product for future.

INTRODUCTION: The use of medicinal plants in human health has been documented since ancient times and they provide a useful source of new therapeutics ¹. Knowledge of Traditional medicinal plant leads to the discovery of new medicines ².

These plants can now be found in herbal products and as part of the traditional Malaysian health care system because of their therapeutic efficacy ³. Natural phenolic compounds play an important role in cancer prevention and treatment. Phenolic compounds from medicinal herbs and dietary plants include phenolic acids, flavonoids, tannins, stilbenes, curcuminoids, coumarins, lignans, quinones, and others ⁴.

Elephantopus scaber and *Clinacanthus nutans* is a well-known medicinal plant in Malaysia and enriched with numerous bioactive compounds ^{5, 6}.

QUICK RESPONSE CODE 	DOI: 10.13040/IJPSR.0975-8232.7(12).4778-86
	Article can be accessed online on: www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.7(12).4778-86	

The objective of our study is the identification of biomarkers from the polyherbal formulation to improve the quality of the new product and compare the bioactive compounds of individual herb (i.e. *Clinacanthus nutans* and *Elephantopus scaber*) from the literature to find similarities and differences from polyherbal fractions.

METHOD AND MATERIAL:

Plant material: The leaves of *Clinacanthus nutans* and *Elephantopus scaber* were collected from Institute Of Sustainable Agrotechnology, Sg. Chuchuh, Universiti Malaysia Perlis (UniMAP) and washed using clean water. After that, the leaves were dried in a dryer at the temperature of 35-40°C for two days. Once dried, the leaves were ground into a fine powder by using a mechanical grinder.

Preparation of Plant Extract: Soxhlet extraction was used in this experiment to extract the herbs. For Soxhlet extraction, a powder sample is weighted approximately. A powdered mixture containing equal proportions of two herbs (5 g each) was extracted with 100 ml of aqueous ethanol 50% for 12-hour extraction. The extract solution was then evaporated by using a rotary evaporator to remove the solvent in the extract solution and dried in an oven at 35-40 °C for 12 hours.

The extract was fractionated using different solvents viz. ethyl acetate, n-butanol, and water. The supernatant was filtered using Whatman No. 1 sheet, pooled and concentrated using vacuum rotary evaporator. The concentrated solutions were then dried in an oven at 35 °C to get the dry form of respective fractions.

Chemicals: Methanol (Fischer scientific, USA), ethanol, Benzoic acid (HmbG, Germany). Rutin, b-sitosterol, vanillin, gallic acid (Sigma-Aldrich, USA). Apigenin, luteolin, quercetin, gallic acid, Merck (Darmstadt, Germany) whereas 3'-hydroxy-5, 6, 7, 4'- tetra methoxy flavone (Indofine Chemical Company, New Jersey, USA).

The sample was filtered through a 0.45µm nylon membrane filter into an HPLC vial prior to HPLC analysis. Solvent mixtures were filtered through a 0.45µm nylon membrane filter and degassed before use.

Instrumentation: A Shimadzu HPLC was utilized to perform the analysis which was equipped with an autosampler, column oven, and UV/VIS detector.

An HPLC column used was Merck Licrochart Purospher Start RP 18 column (250mm, 4.6 mm i.d, 5µm pore size). The temperature was maintained at 40°C throughout the study.

Chromatography conditions: The mobile phase was an isocratic system of Acetonitrile 100% (v/v), The flow rate was 0.5 mL min⁻¹. Absorbance was monitored at λ = 360 nm.

Preparation of stock solution: A standard stock solution was prepared by dissolving 5 mg of standard in ethanol, yielding 12.25 mL of a concentration stock = 0.41 mg mL⁻¹. Dilution of 5.0 mL was prepared by aliquoting 5.0 mL of the standard stock solution and diluted with the ethanol to yield 10 mL of standard solutions containing 200µg mL⁻¹ of the standard.

RESULT AND DISCUSSION:

Method development: The main objective of the chromatographic method was to identify different bioactive compound inside a new polyherbal formulation such as Apigenin, luteolin, b-sitosterol, vanillin, gallic acid, tetra methoxy flavone, quercetin, rutin, benzoic acid, gallic acid.

This new polyherbal formulation contains an equal amount of *Clinacanthus nutans* and *Elephantopus scaber* in the treatment of the wound.

After identification of compounds from polyherbal formulation it was found that apigenin, luteolin, b-sitosterol, quercetin, rutin was found in *Elephantopus scaber*^{7, 8} whereas *Clinacanthus nutans* contains b-sitosterol⁹, apigenin¹⁰. 3'-hydroxy-5, 6, 7, 4'- tetra methoxy flavone, gallic acid, benzoic acid and vanillin was not found in both medicinal plants.

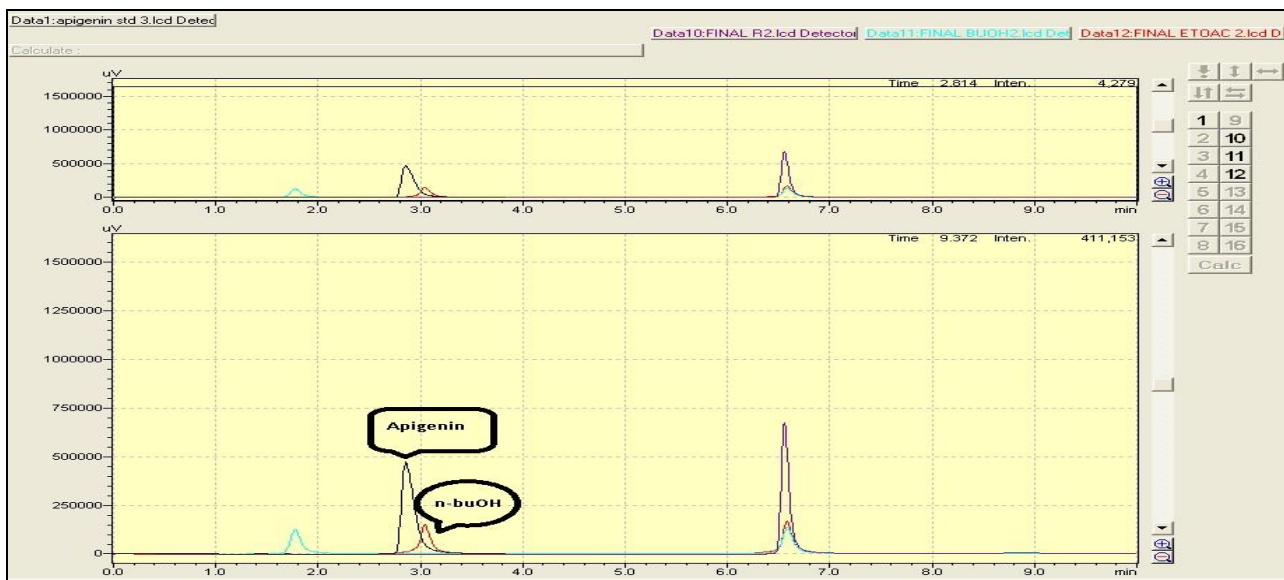


FIG. 1: CHROMATOGRAM FOR STANDARD APIGENIN VS DIFFERENT FRACTIONS

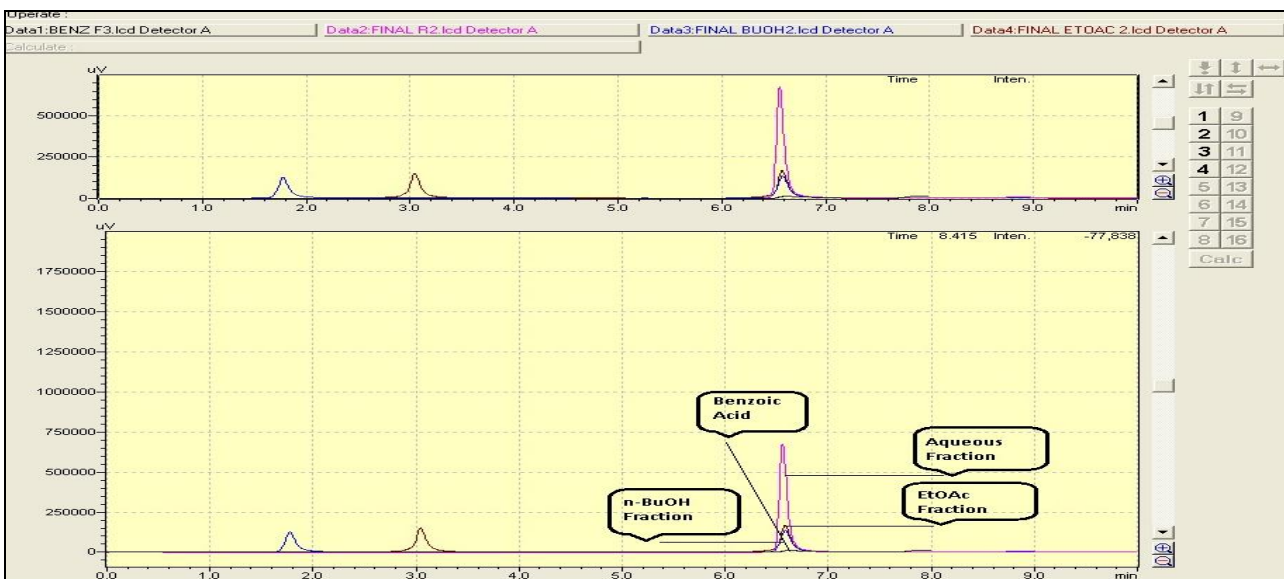


FIG. 2: CHROMATOGRAM FOR STANDARD BENZOIC ACID VS DIFFERENT FRACTIONS

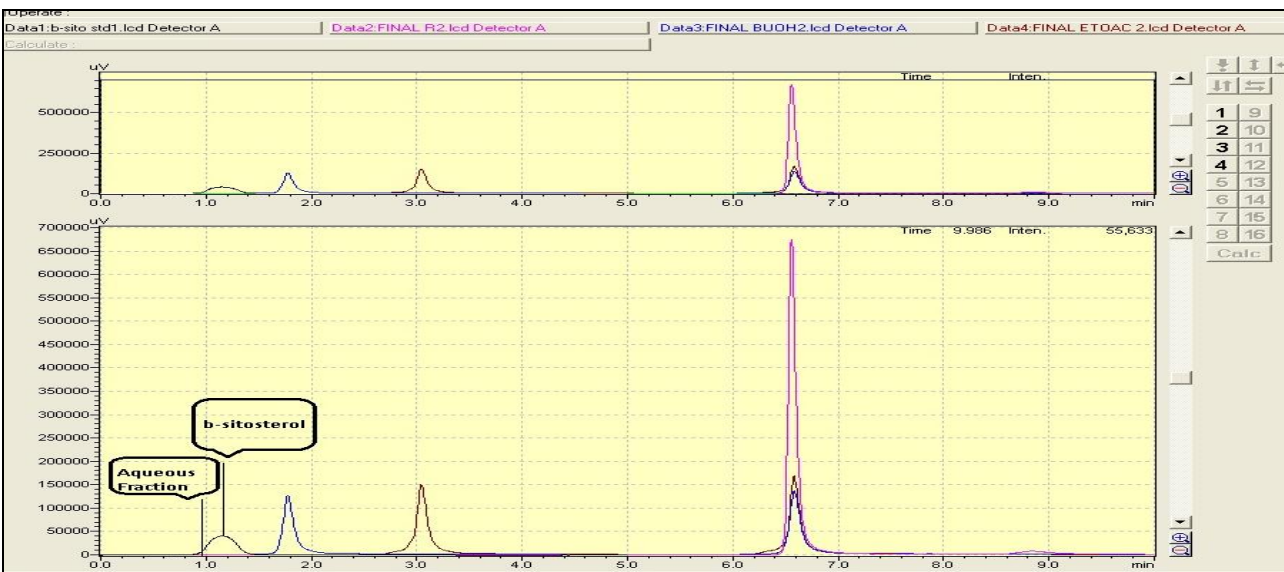


FIG. 3: CHROMATOGRAM FOR STANDARD β -SITOSTEROL VS DIFFERENT FRACTIONS

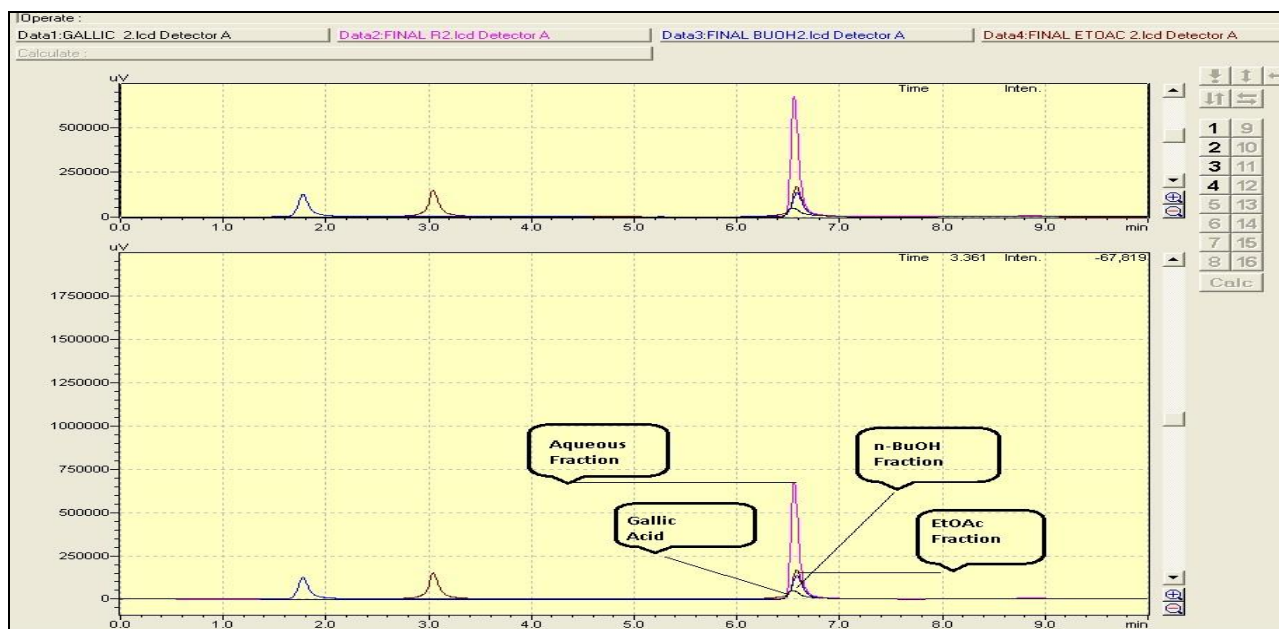


FIG. 4: CHROMATOGRAM FOR STANDARD GALLIC ACID VS DIFFERENT FRACTIONS

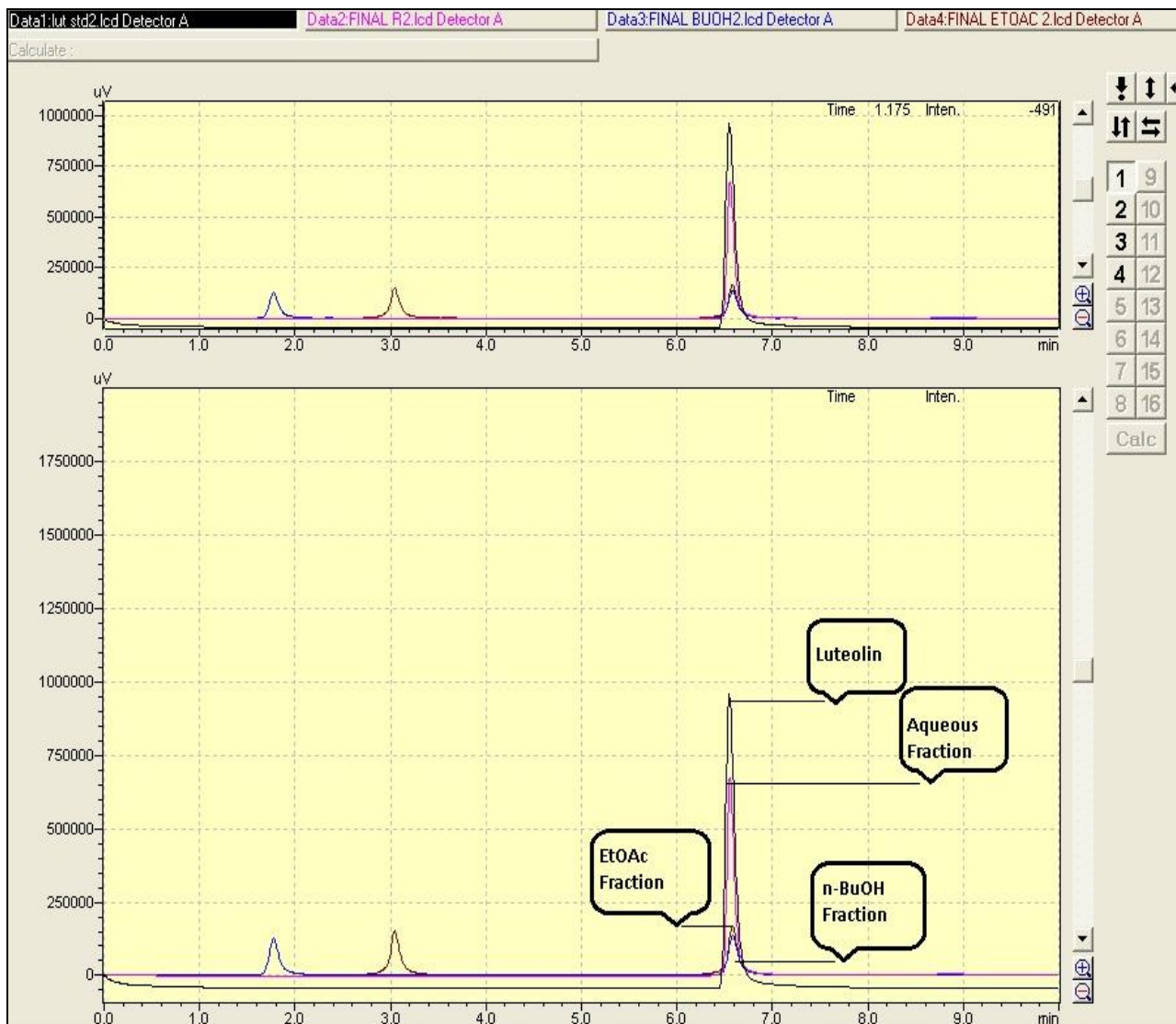


FIG.5: CHROMATOGRAM FOR STANDARD LUTEOLIN VS DIFFERENT FRACTIONS

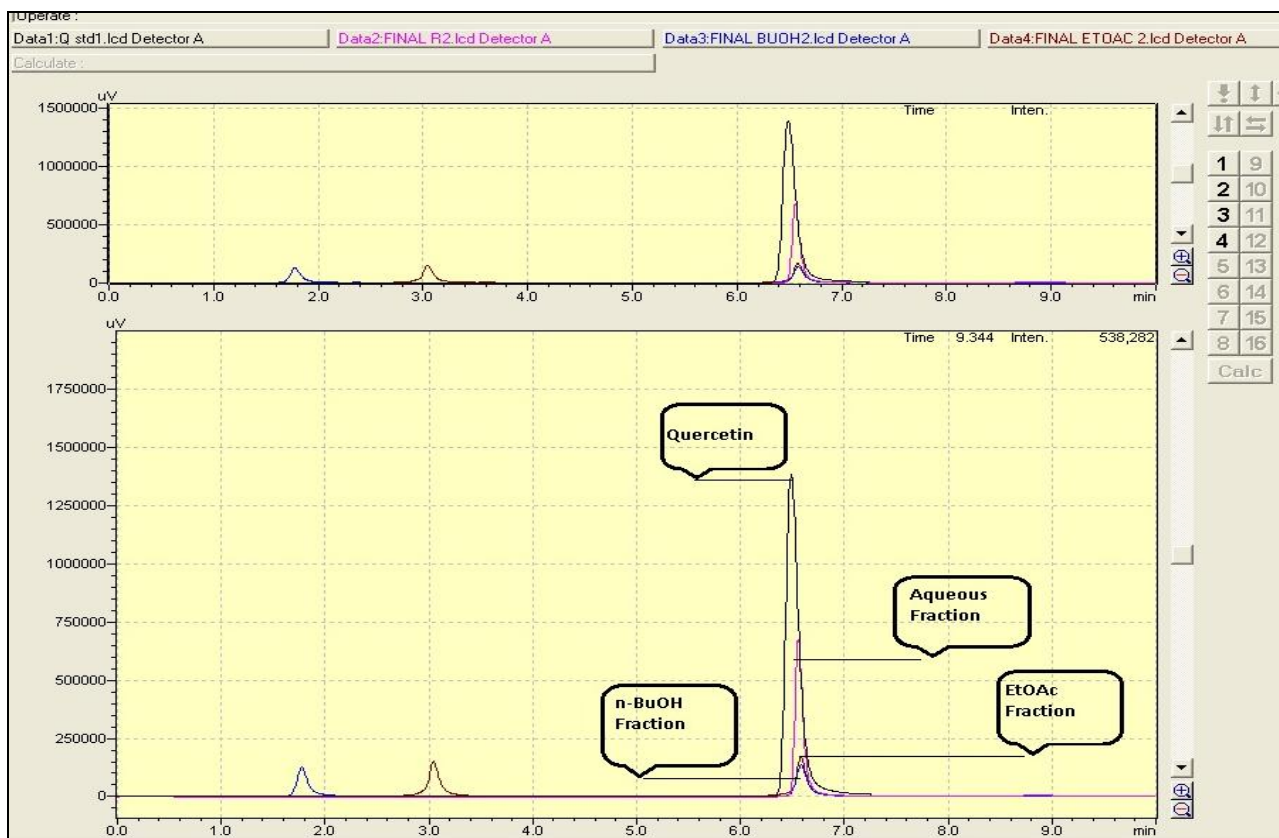


FIG.6: CHROMATOGRAM FOR STANDARD QUERCETIN VS DIFFERENT FRACTIONS

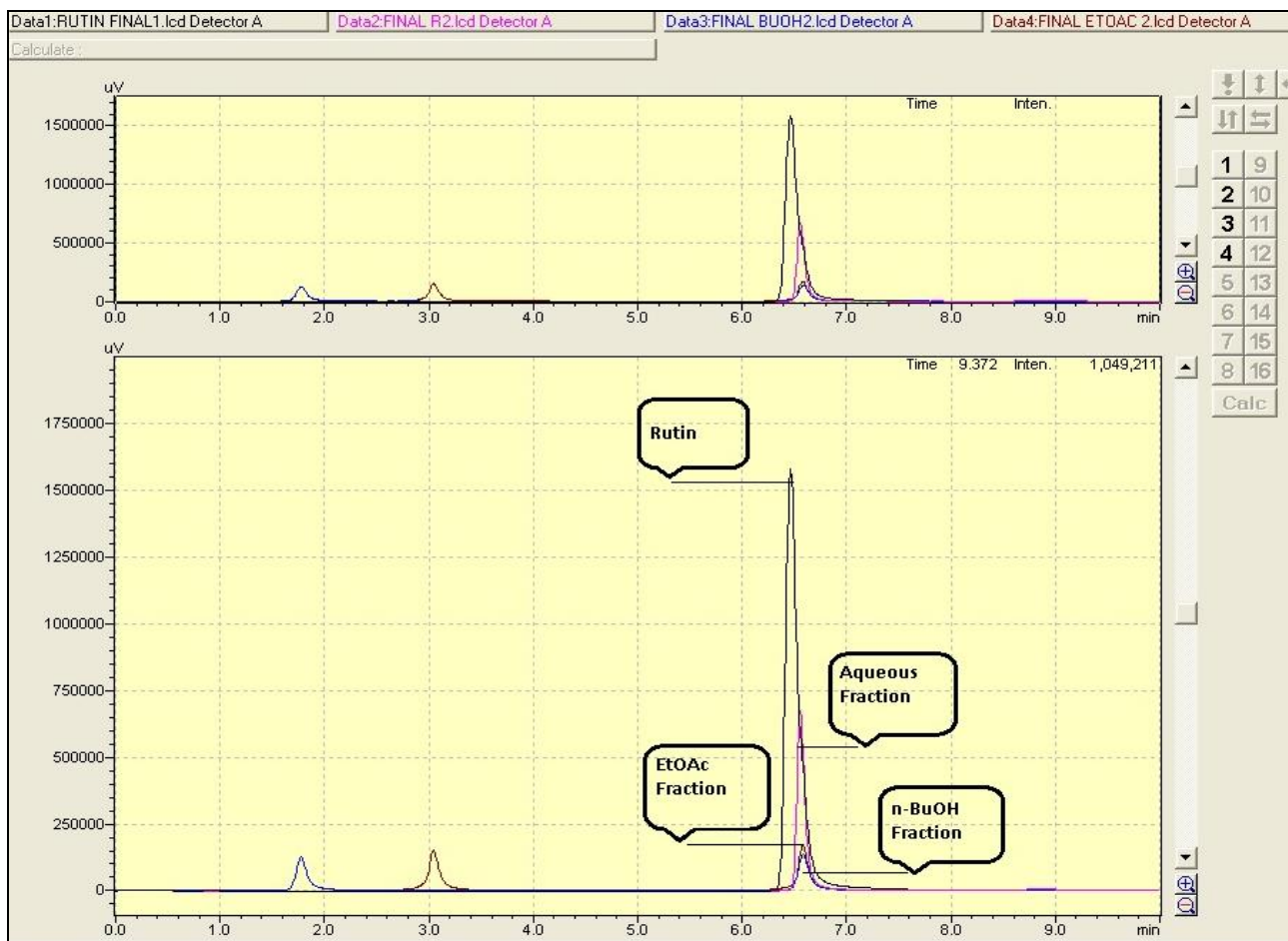


FIG. 7: CHROMATOGRAM FOR STANDARD RUTIN VS DIFFERENT FRACTIONS

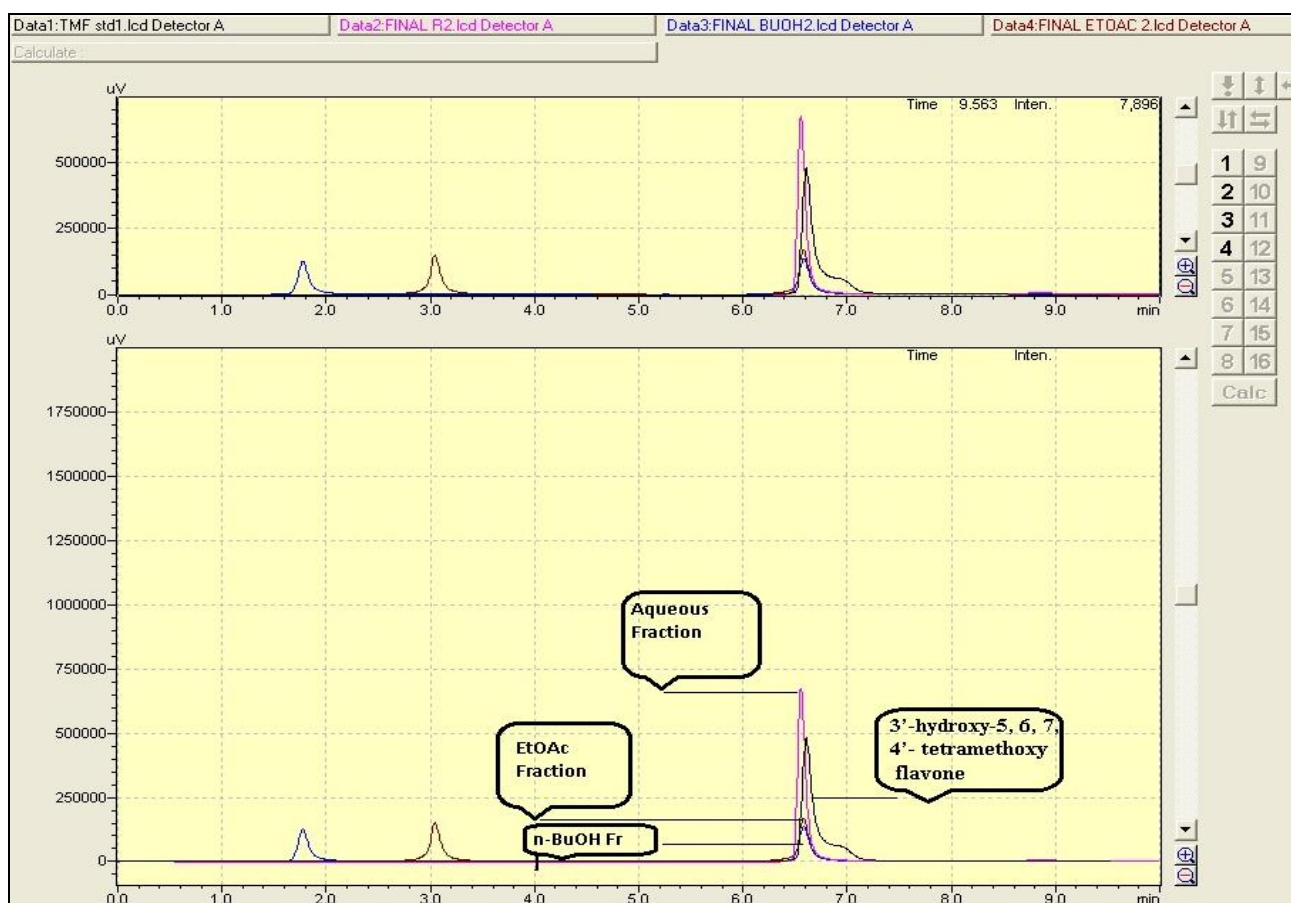


FIG. 8: CHROMATOGRAM FOR STANDARD 3'-HYDROXY-5, 6, 7, 4'- TETRAMETHOXY FLAVONE VS DIFFERENT FRACTIONS

TABLE 1: RETENTION TIME FOR STANDARDS

Bioactive compounds	Retention tim (Rt)	Area	Height	%Area	% Height
Apigenin	2.854	4361364	471520	100	100
Luteolin	6.545	7717110	1008205	99.959	99.976
b-sitosterol	1.147	724840	40330	96.567	97.375
Quercetin	6.484	12649427	1383209	99.987	99.991
Rutin	6.460	14405905	1578157	99.931	99.980
3'-hydroxy-5, 6, 7, 4'- tetramethoxy flavone	6.602	4887896	483674	100	100
Vanillin	6.581	1349524	109808	99.30	99.321
Benzoic acid	6.613	220638	11270	100	100
Gallic acid	6.540	577490	49780	100	100

TABLE 2: RETENTION TIME FOR ETHYL ACETATE FRACTION OF POLYHERBAL FORMULATION

Sr. No	Retention time (Rt)	Area	Height	%Area	% Height	Compound identified
1	3.040	1504358	151270	47.39	46.879	Apigenin
2	6.577	1635270	168071	51.51	51.086	Rutin, Quercetin, Gallic acid, Luteolin, Vanillin, 3'-hydroxy-5, 6, 7, 4'- tetra methoxy flavone, Benzoic acid (Rt=6.460-6.613)

TABLE 3: RETENTION TIME FOR N-BUTANOL FRACTION OF POLYHERBAL FORMULATION

Sr. No	Retention time (Rt)	Area	Height	%Area	% Height	Compound identified
1	2.822	11455	755	0.458	0.282	Apigenin
2	6.580	1311170	136695	52.439	51.062	Rutin, Quercetin, Gallic acid, Luteolin, Vanillin, 3'-hydroxy-5, 6, 7, 4'- tetra methoxy flavone, Benzoic acid (Rt=6.460-6.613)

TABLE 4: RETENTION TIME FOR AQUEOUS FRACTION OF POLYHERBAL FORMULATION

Sr. No	Retention time (Rt)	Area	Height	%Area	% Height	Compound identified
1	0.952	2655	195	0.074	0.034	b-sitosterol
2	6.561	3414202	558580	94.746	98.472	Rutin, Quercetin, Gallic acid, Luteolin, Vanillin, 3'-hydroxy-5, 6, 7, 4'- tetra methoxy flavone, Benzoic acid (Rt=6.460-6.613)

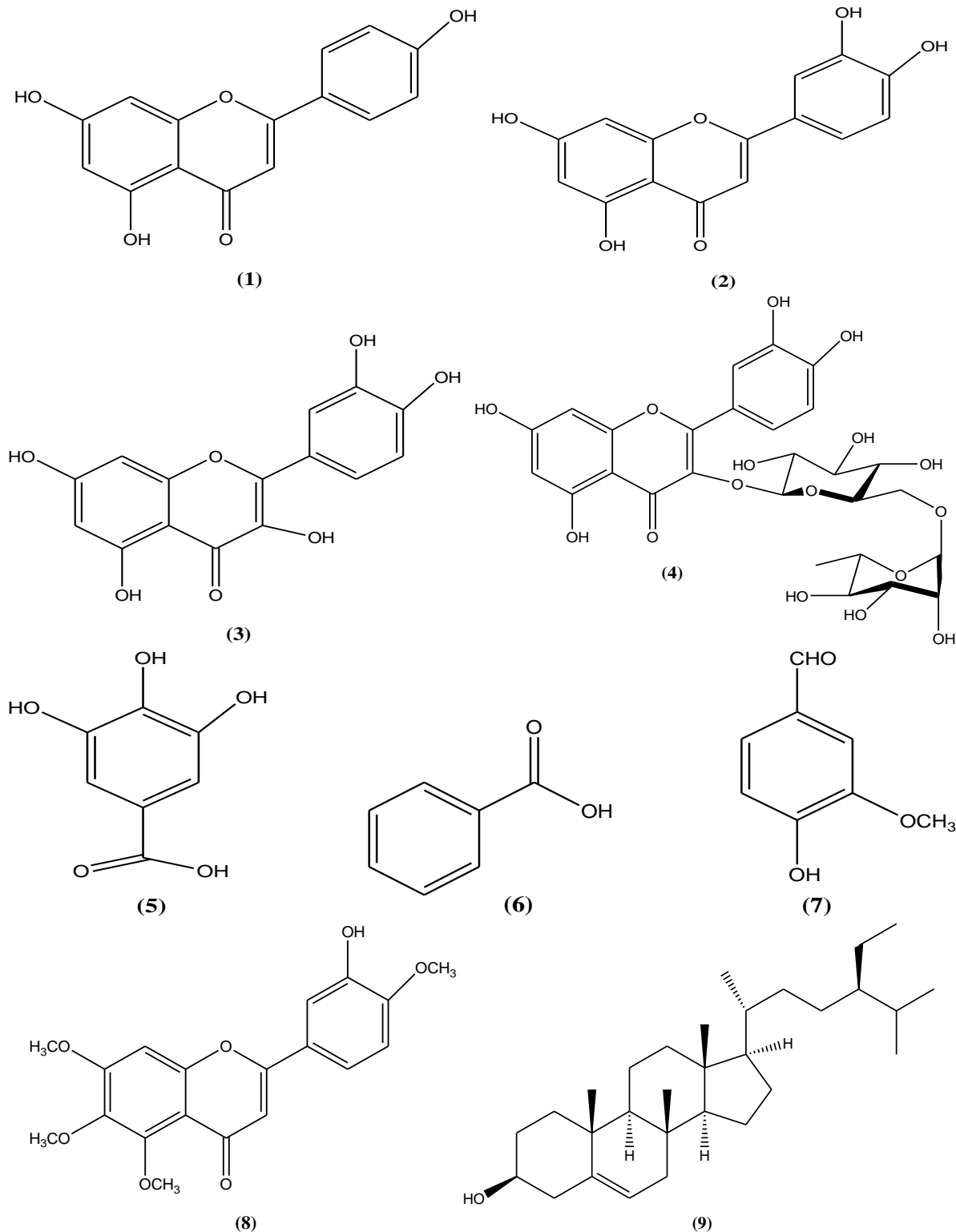
**FIG. 9: LIST OF BIOMARKERS IDENTIFIED BY HIGH PERFORMANCE LIQUID CHROMATOGRAPHY**

TABLE 5: LIST OF PHARMACOLOGICAL ACTIVITIES REPORTED IN IDENTIFIED COMPOUND

Bioactive compounds	<i>In-vitro</i> studies/ <i>In-vivo</i> studies	References
Apigenin	Anti-inflammatory, Anti-cancer (human prostate cancer, breast cancer), anti-leukemic activity, Wound healing activities, Alzheimer's Disease	11, 12, 13, 14, 15
Luteolin	Anti-tumor activity (breast cancer), Improves Cardiac Function, Oxidative stress suppression, Wound healing activity, mast cell inhibition	16, 17, 18, 19, 20
b-sitosterol	Anti-inflammatory, heart disease, hypercholesterolemia, modulating the immune system, prevention of cancer, as well as for rheumatoid arthritis, tuberculosis, cervical cancer, hair loss and benign prostatic hyperplasia.	21, 22
Quercetin	Anti-proliferation, apoptosis, Antiangiogenic activity, Wound healing activity	23, 24
Rutin	Wound healing activity, Anti-oxidant activity, apoptosis	25, 26, 27
3'-hydroxy-5, 6, 7, 4'-tetramethoxy flavone	Anti-oxidant, Anti-cancer (colon cancer, breast cancer)	27, 28, 29
Benzoic acid	Antibacterial, antifungal, anti-oxidant properties	30, 31
Gallic acid	Anti-inflammatory, anti-oxidant, Anti-metastasis effects, wound healing activity	32, 33, 34, 35

CONCLUSION: Unavailability of pharmacopoeial standard for herbal based medicinal product is a foremost problem. High-Performance liquid chromatography (HPLC) is one of the major chromatographic technique for identification of biomarker. Using HPLC helps to improve the quality of product and helps to sustain the shelf life of the product for a longer period.

ACKNOWLEDGEMENTS: Special thanks to the Institute Of Sustainable Agrotechnology, Sg. Chuchuh, Universiti Malaysia Perlis (UniMAP) for providing the samples. This research was supported by the Prototype Research Grant Scheme (PRGS; Reference no : PRGS/9013-00016) awarded by the Ministry of Higher Education (MOHE), Malaysia, Universiti Malaysia Perlis (UniMAP), Ministry of Higher Education Malaysia [Malaysian International Scholarship; Reference no. KPT.B.600-18/3 JLD 6 (31)] and Research Collaborative Effort from Malaysian Agricultural Research and Development Institute, Serdang, Malaysia.

REFERENCES:

- Siew, Y. Y. et al. Ethnobotanical survey of usage of fresh medicinal plants in Singapore. *J. Ethnopharmacol.* 2014; 155, 1450–1466.
- Inta, A., Trisonthi, P. & Trisonthi, C. Analysis of traditional knowledge in medicinal plants used by Yuan in Thailand. *J. Ethnopharmacol.* 2013; 149, 344–351.
- Alsarhan, A., Sultana, N., Al-Khatib, A., Rafiq, M. & Kadir, A. Review on Some Malaysian Traditional Medicinal Plants with Therapeutic Properties. *J. Basic Appl. Sci.* 2014; 10, 149–159.
- Huang, W.-Y., Cai, Y.-Z. & Zhang, Y. Natural phenolic compounds from medicinal herbs and dietary plants: potential use for cancer prevention. *Nutr. Cancer* 2010; 62, 1–20.
- Ho, W. Y., Ky, H., Yeap, S. K., Rahim, R. A. & Omar, A.

- R. Traditional practice, bioactivities and commercialization potential of *Elephantopus scaber* Linn. *J. Med. Plants Res.* 2009; 3, 1212–1221.
- Alam, A. et al. Clinacanthus nutans: A review of the medicinal uses, pharmacology and phytochemistry. 2016; 9, 402–409.
- Zuo, A.-X., Wan, C.-P., Zheng, X. & Rao, G.-X. Chemical Constituents of *Elephantopus scaber*. *Chem. Nat. Compd.* 2016; 52, 484–486.
- Mendhulkar, V. D. & Kharat, S. N. Hptlc assay for quercetin and rutin flavonoids in *Elephantopus Scaber* [Linn.] grown under induced heat stress condition. *Int. J. Pharma Bio Sci.* 2015; 6, B36–B52.
- Aslam, M. S., Ahmad, M. S. & Mamat, A. S. O. H. A review on phytochemical constituents and pharmacological activities of clinacanthus nutans. 2015; 7, 2–5.
- Huang, D., Guo, W., Gao, J., Chen, J. & Olatunji, J. O. *Clinacanthus nutans* (Burm. f.) Lindau Ethanol Extract Inhibits Hepatoma in Mice through Upregulation of the Immune Response. *Molecules* 2015; 20, 17405–28.
- Choi, J. S. et al. Effects of C-glycosylation on anti-diabetic, anti-Alzheimer's disease and anti-inflammatory potential of apigenin. *Food Chem. Toxicol.* 2014; 64, 27–33.
- Manuscript, A. & Proximity, I. Plant flavone apigenin inhibits HDAC and remodels chromatin to induce growth arrest and apoptosis in human prostate cancer cells: *In vitro* and *in vivo* study. *NIH Public Access* 2012; 51, 952–962.
- Budhraj, A. et al. Apigenin induces apoptosis in human leukemia cells and exhibits anti-leukemic activity in vivo. *Mol. Cancer Ther.* 2012; 11, 132–42.
- Seo, H.-S. et al. Apigenin induces apoptosis via extrinsic pathway, inducing p53 and inhibiting STAT3 and NFκB signaling in HER2-overexpressing breast cancer cells. *Mol. Cell. Biochem.* 2012; 366, 319–334.
- Zhao, L. et al. Neuroprotective, anti-amyloidogenic and neurotrophic effects of apigenin in an Alzheimer's disease mouse model. *Molecules* 2013; 18, 9949–9965.
- Lee, E.-J., Oh, S.-Y. & Sung, M.-K. Luteolin exerts anti-tumor activity through the suppression of epidermal growth factor receptor-mediated pathway in MDA-MB-231 ER-negative breast cancer cells. *Food Chem. Toxicol.* 2012; 50, 4136–43.
- Sun, D. et al. Luteolin limits infarct size and improves cardiac function after myocardium ischemia/reperfusion injury in diabetic rats. *PLoS One* 2012; 7, e33491.

18. Sun, G. et al. Oxidative stress suppression by luteolin-induced heme oxygenase-1 expression. *Toxicol. Appl. Pharmacol.* 2012; 265, 229–40.
19. Lodhi, S. & Singhai, A. K. Wound healing effect of flavonoid rich fraction and luteolin isolated from *Martynia annua* Linn. on streptozotocin induced diabetic rats. *Asian Pac. J. Trop. Med.* 2013; 6, 253–9.
20. Kritas, S. K. et al. Luteolin inhibits mast cell-mediated allergic inflammation. *J. Biol. Regul. Homeost. Agents* 27, 955–9.
21. Loizou, S., Lekakis, I., Chrousos, G. P. & Moutsatsou, P. β -Sitosterol exhibits anti-inflammatory activity in human aortic endothelial cells. *Mol. Nutr. Food Res.* 2010; 54, 551–558.
22. Soodabeh Saeidnia, Azadeh Manayi, A. R. G. Y. M. A. The Story of Beta-sitosterol- A Review. *European J. Med. Plants* 2014; 4, 590–609.
23. Gomathi, K., Gopinath, D., Ahmed, M. R. & Jayakumar, R. Quercetin incorporated collagen matrices for dermal wound healing processes in rat. *Biomaterials* 2003; 24, 2767–2772.
24. Russo, M., Spagnuolo, C., Tedesco, I., Bilotto, S. & Russo, G. L. The flavonoid quercetin in disease prevention and therapy: Facts and fancies. *Biochem. Pharmacol.* 2012; 83, 6–15.
25. Tran, N. Q., Joung, Y. K., Lih, E. & Park, K. D. In situ forming and rutin-releasing chitosan hydrogels as injectable dressings for dermal wound healing. *Biomacromolecules* 2011; 12, 2872–80.
26. Lee, K. A. et al. Antioxidant activities of onion (*Allium cepa* L.) peel extracts produced by ethanol, hot water, and subcritical water extraction. *Food Sci. Biotechnol.* 2014; 23, 615–621.
27. Chen, W. et al. Anti-tumor effect of α -pinene on human hepatoma cell lines through inducing G2 / M cell cycle arrest. *J. Pharmacol. Sci.* 2015; 127, 332–338.
28. Qiu, P. et al. Inhibitory effects of 5-hydroxy polymethoxyflavones on colon cancer cells. *Mol. Nutr. Food Res.* 2010; 54 Suppl 2, S244–52.
29. Sergeev, I. N., Li, S., Colby, J., Ho, C.-T. & Dushenkov, S. Polymethoxylated flavones induce Ca(2+)-mediated apoptosis in breast cancer cells. *Life Sci.* 2006; 80, 245–53.
30. Natella, F., Nardini, M., Di Felice, M. & Scaccini, C. Benzoic and Cinnamic Acid Derivatives as Antioxidants: Structure–Activity Relation. *J. Agric. Food Chem.* 1999; 47, 1453–1459.
31. Park, E.-S. et al. Antimicrobial activity of phenol and benzoic acid derivatives. *Int. Biodeterior. Biodegradation* 2001; 47, 209–214.
32. Kroes, B. H., van den Berg, A. J., Quarles van Ufford, H. C., van Dijk, H. & Labadie, R. P. Anti-inflammatory activity of gallic acid. *Planta Med.* 1992; 58, 499–504.
33. Aruoma, O. I., Murcia, A., Butler, J. & Halliwell, B. Evaluation of the antioxidant and prooxidant actions of gallic acid and its derivatives. *J. Agric. Food Chem.* 1993; 41, 1880–1885.
34. Ho, H.-H. et al. Anti-metastasis effects of gallic acid on gastric cancer cells involves inhibition of NF-kappaB activity and downregulation of PI3K/AKT/small GTPase signals. *Food Chem. Toxicol.* 2010; 48, 2508–16.
35. Wary, R. et al. Chitosan gallic acid microsphere incorporated collagen matrix for chronic wounds: Biophysical and biochemical characterization. *Int. J. Pharm. Pharm. Sci.* 2014; 6, 94–100.

How to cite this article:

Aslam MS, Ahmad MS, Mamat AS, Ahmad MZ and Salam F: Reverse phase-high performance liquid chromatography method for estimation of bioactive compounds from different fractions of new polyherbal formulation of *Clinacanthus nutans* and *Elephantopus scaber*. *Int J Pharm Sci Res* 2016; 7(12): 4778-86. doi: 10.13040/IJPSR.0975-8232.7(12).4778-86.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)